



Temporal Plots of 2005-2007 Bromine Data from the Upstream San Joaquin River

Chelsea Spier¹
Remie Burks¹
Sharon Borglin¹
Randy Dahlgren²
Jeremy Hanlon¹
Justin Graham¹
William Stringfellow¹

February 2008

¹Environmental Engineering Research Program
School of Engineering & Computer Sciences
University of the Pacific
3601 Pacific Avenue, Sears Hall
Stockton, CA 95211

²UC Davis, Davis, CA

Introduction

The San Joaquin River (SJR) supports one of the most productive agricultural regions in the world and its productivity is heavily dependant on irrigated agriculture. A consequence of irrigated agriculture is the production of return flows conveyed down gradient drains that eventually discharge to surface waters. Agricultural drainage may have significant nutrient load and can impact algae growth and general water quality in the SJR. Individual farmers and agricultural organizations, such as drainage authorities, are in need of tools to manage the environmental impacts of agricultural activities (Stringfellow, 2008).

For the years 2005 through 2007, sites throughout the San Joaquin Valley watershed were sampled to assess the overall water quality in the region. One thousand nine hundred and ninety-six (1996) individual surface water samples were collected and analyzed and WQ was assessed at 113 locations in the SJR basin (Borglin et al., 2008). Samples were processed and analyzed by the Environmental Engineering Research Program (EERP) laboratory at the University of the Pacific as well as at the University of California, Davis, Dahlgren Lab. This report includes temporal plots of bromine (Br) data analyzed by the UCD Dahlgren laboratory between 2005 and 2007.

Methods

Depth integrated field samples were collected during 2005-2007 in the upper San Joaquin River in accordance with EERP Field Standard Operating Procedures Protocol Book (Graham 2008). Water samples were collected in glass 1000 mL bottles (Wheaton Science Products, Millville, NJ), 1000 mL HDPE Trace-Clean narrow mouth plastic bottles (VWR International), 250 mL HDPE Trace-Clean wide mouth plastic bottles (VWR International) as well as 40 mL trace clean vials with PTFE septa (ICHEM, Rockwood, TN). Bottles were labeled with the appropriate sample number, site name and sampling date. All bottles were rinsed with sample water prior to sample collection. Some sites required a bucket to collect sample water because of accessibility from a high bridge or platform. For these sites, the bucket was pre-rinsed with sample water and sample bottles were filled using a rinsed funnel. Care was taken to distribute water simultaneously to all sample bottles (rather than sequentially). Samples were immediately stored at 4°C after sampling (cooler temperature was recorded in the lab upon delivery) and transported to the EERP lab on the day of sampling.

Within 24 hours of collection samples were transported from the EERP laboratory to the UCD laboratory for analysis, during this time period samples were stored in coolers at 4°C.

Ion chromatography was utilized for measuring bromine (Br⁻) using a Dionex ICS-2000 Ion Chromatograph (Dionex Corporation, Sunnyvale, CA) (SM-4110 B).

Results/Discussion

Samples were measured ranging from 0.0-14.69 mg/L Br. The average concentration of Br in samples collected was 1.44 mg/L Br. These temporal plots (Figures 3-104) created an easy visual way to find outliers and double check data entry for possible mistakes.

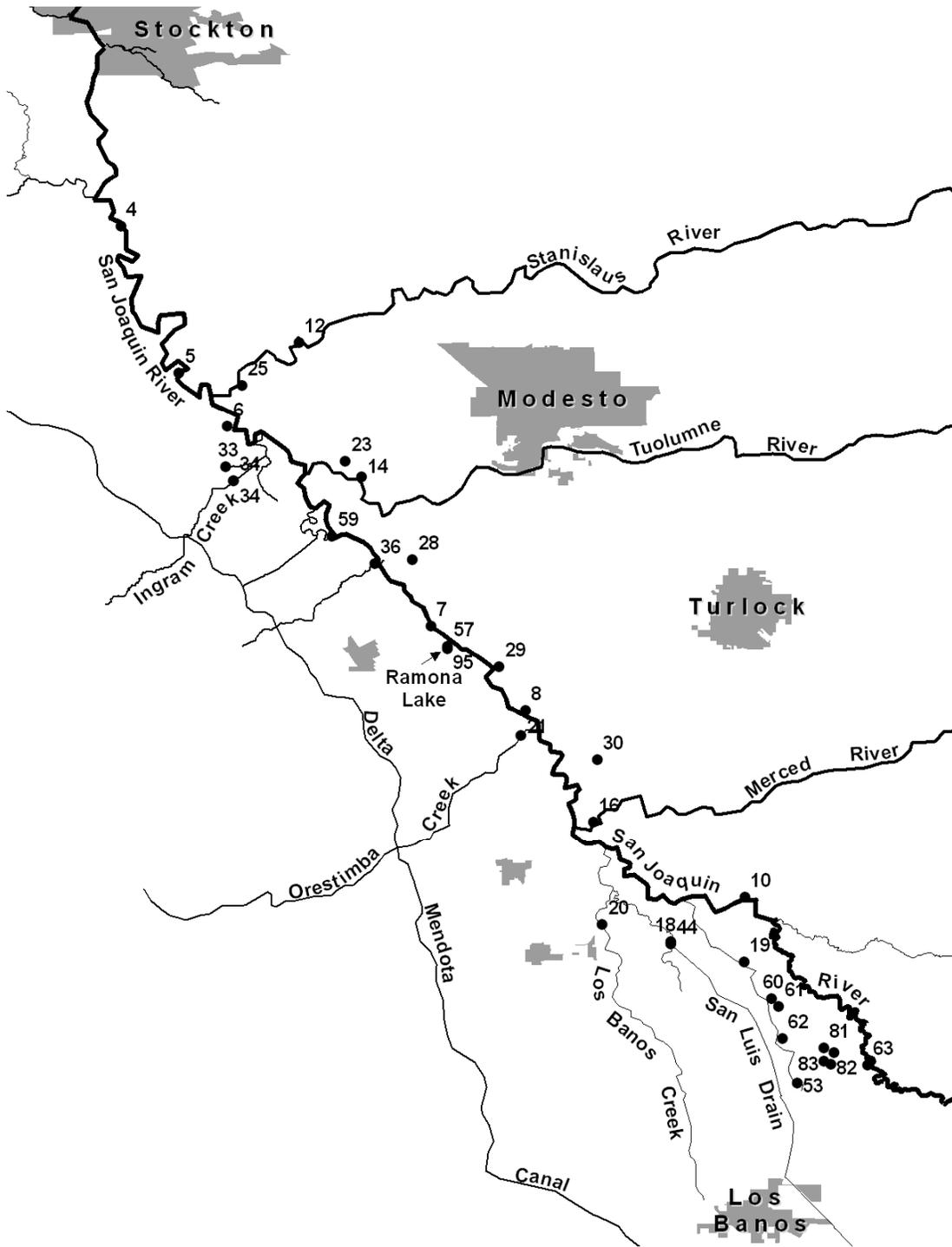
References

- American Public Health Association (APHA). 2005. Standard Methods for the Examination of Water and Wastewater, 21st Edition. American Public Health Association, Washington, DC.
- Borglin, S., W. Stringfellow, J. Hanlon. 2005. Standard Operating Procedures for the Up-Stream Dissolved Oxygen TMDL Project. LBNL/Pub-937.
- Borglin, S., Burks, R., Hanlon, J., Graham, J., Spier, C., Stringfellow, W., and Dahlgren, R., (2008) Methods overview, quality assurance, and quality control, University of the Pacific, Stockton, CA
- Borglin, S.E., Burks, R.D., Hanlon, J.S., Stringfellow, W.T. (2008) EERP Lab Protocol Book, University of the Pacific, Stockton, CA.
- Graham, J., Hanlon, J.S., Stringfellow, W.T., (2008) EERP Field Protocol Book, University of the Pacific, Stockton, CA.
- Stringfellow, W.T., et al., (2008) Evaluation of Vegetated Ditches, Ponds, and Wetlands as BMPs for Mitigating the Water Quality Impact of Irrigated Agriculture in the San Joaquin Valley, University of the Pacific, Stockton, CA
- YSI Environmental Operations Manual (2005) 6-Series Environmental Monitoring Systems, Yellow Springs, OH.

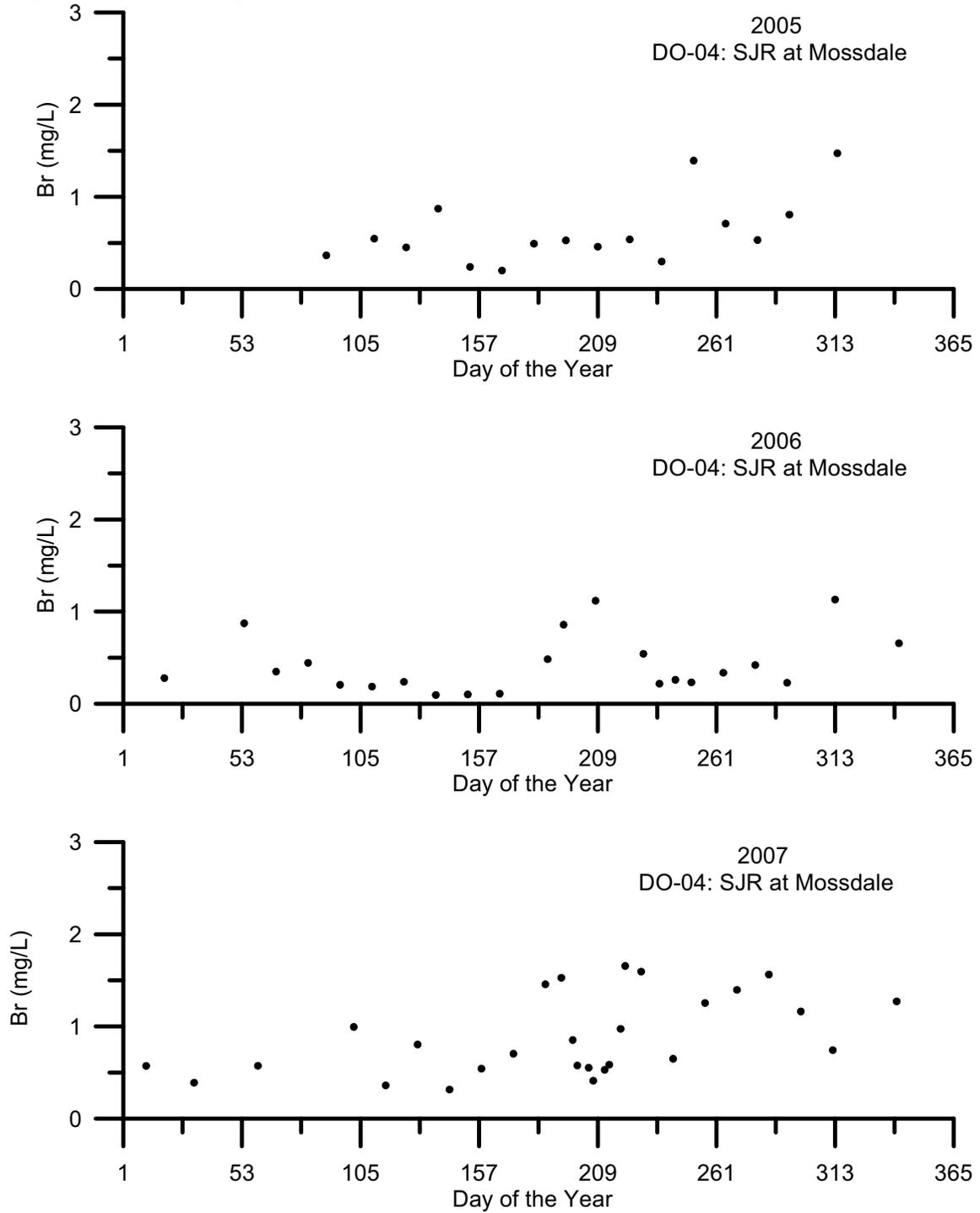
Table 1: EERP Sampling Site List

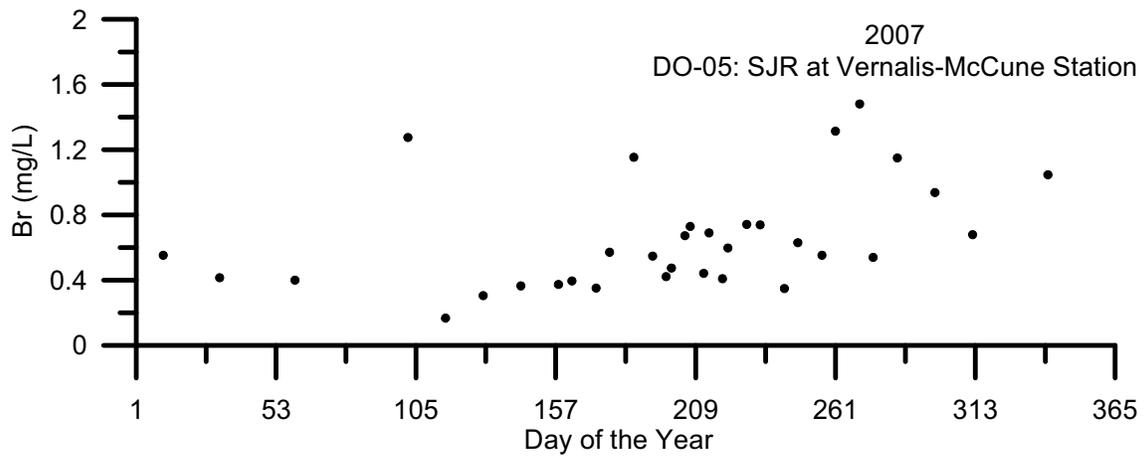
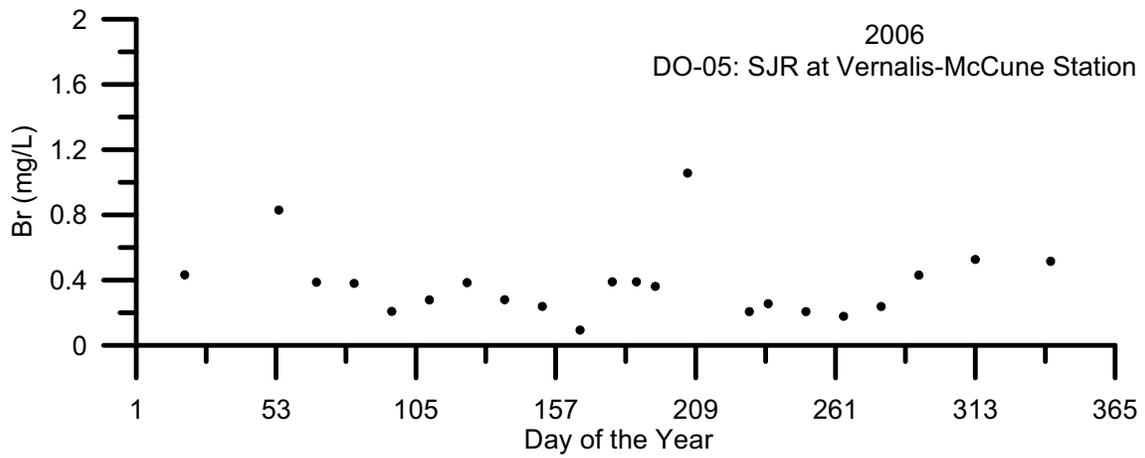
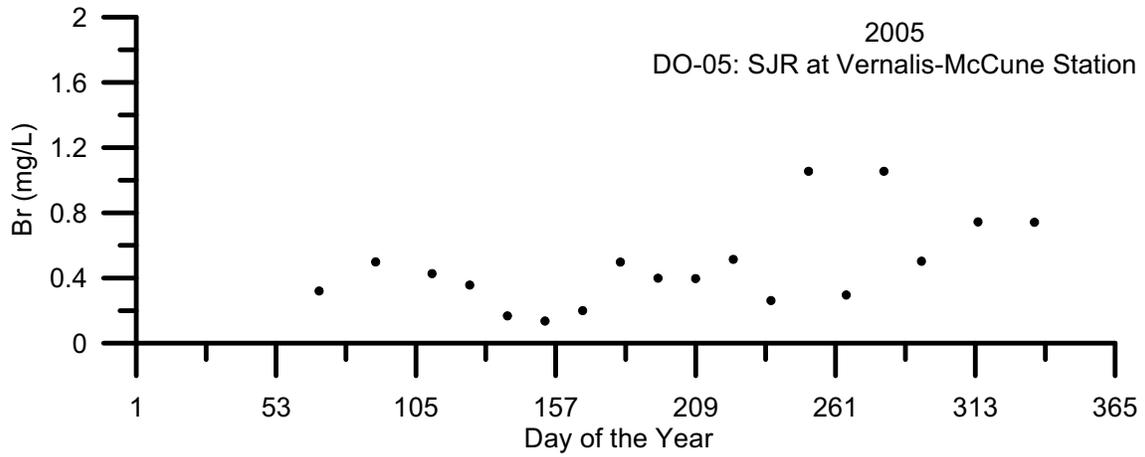
DO Number	Site Name	Type
4	SJR at Mossdale	Core sites
5	SJR at Vernalis-McCune Station (River Club)	Core sites, BMP
6	SJR at Maze	Core sites, BMP
7	SJR at Patterson	Core sites, BMP
8	SJR at Crows Landing	Core sites, BMP
10	SJR at Lander Avenue	Core sites
12	Stanislaus River at Caswell Park	Core sites
14	Tuolumne River at Shiloh Bridge	Core sites
16	Merced River at River Road	Core sites
18	Mud Slough near Gustine	Core sites, Wetland
19	Salt Slough at Lander Avenue	Core sites, Wetland
20	Los Banos Creek Flow Station	Core sites, Wetland
21	Orestimba Creek at River Road	Core sites, BMP
23	Modesto ID Lateral 5 to Tuolumne	Core sites
25	Modesto ID Main Drain to Stan. R. via Miller Lake	Core sites
28	Turlock ID Westport Drain Flow station	Core sites
29	Turlock ID Harding Drain	Core sites
30	Turlock ID Lateral 6 & 7 at Levee	Core sites
33	Hospital Creek	Intermittent, BMP
34	Ingram Creek	Core sites, BMP
36	Del Puerto Creek Flow Station	Core sites, BMP
44	San Luis Drain End	Core sites
53	Salt Slough at Wolfsen Road	Wetland
57	Ramona Lake Drain	Core sites, BMP
59	SJR Laird Park	Core sites
60	Moffit 1 South	Wetland
61	Deadman's Slough	Wetland
62	Mallard Slough	Wetland
63	Inlet C Canal	Wetland
80	South Marsh-1-Inlet	Wetland
81	South Marsh-1-Outlet	Wetland
82	South Marsh-3-Inlet	Wetland
83	South Marsh-3-Outlet	Wetland
95	Ramona drain at Ramona Lake	BMP, Intermittent

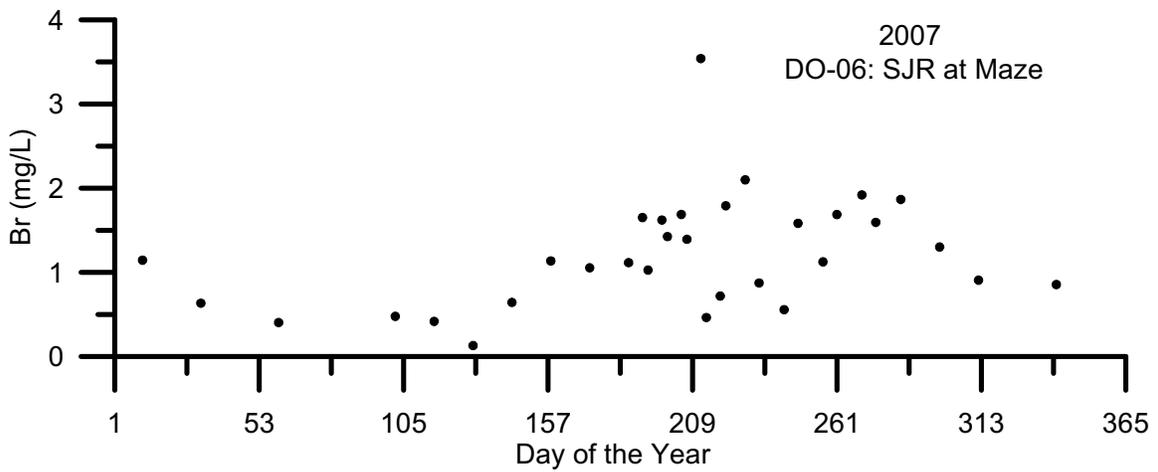
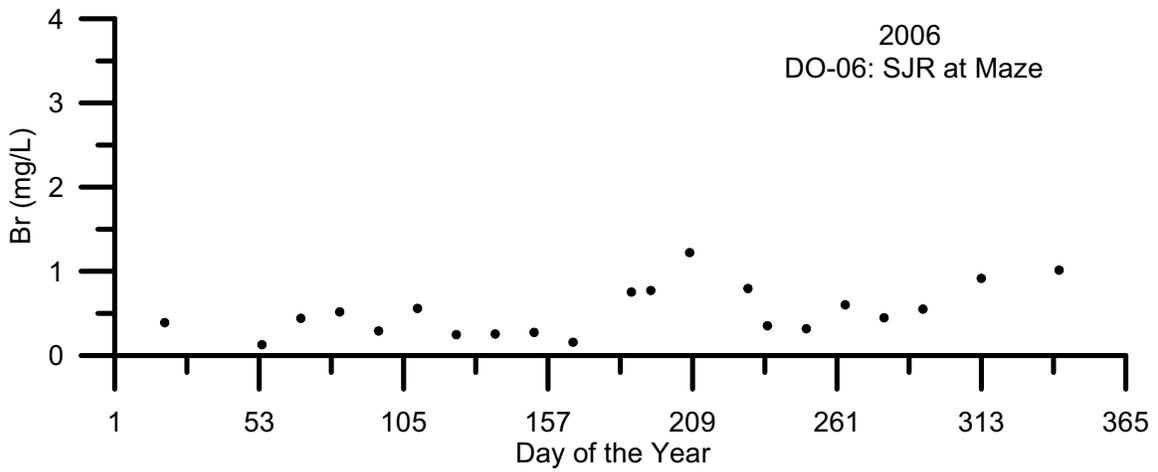
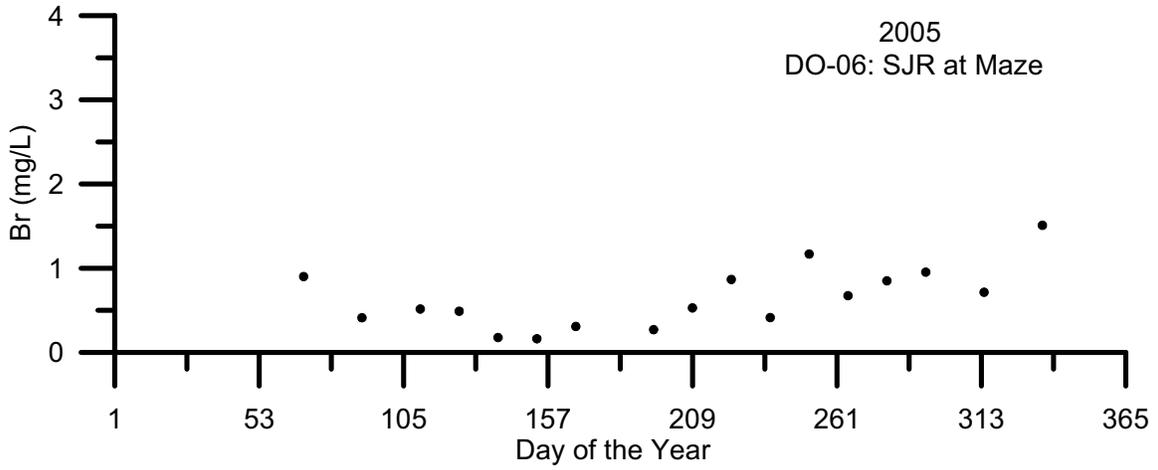
Figure 1: EERP Sampling Site Map of SJR Watershed and Tributaries

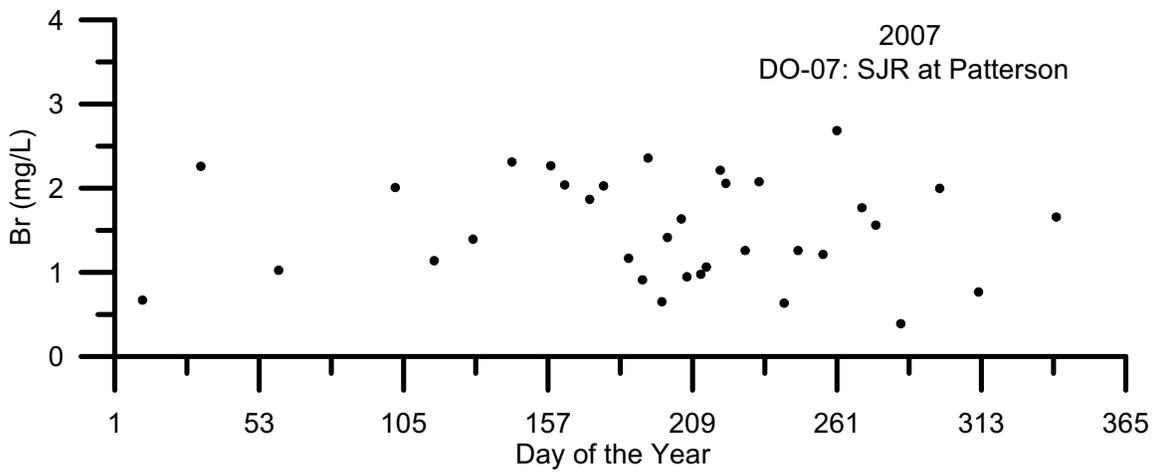
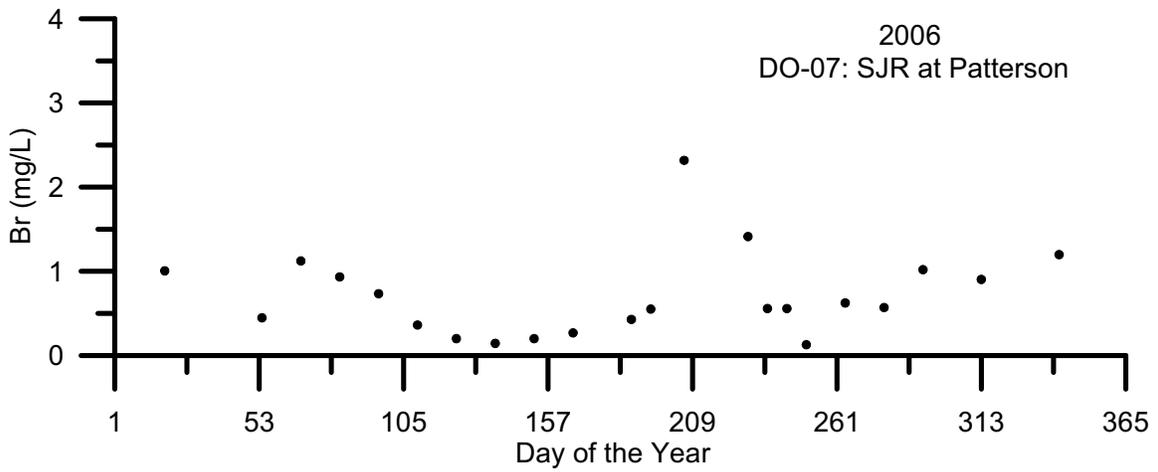
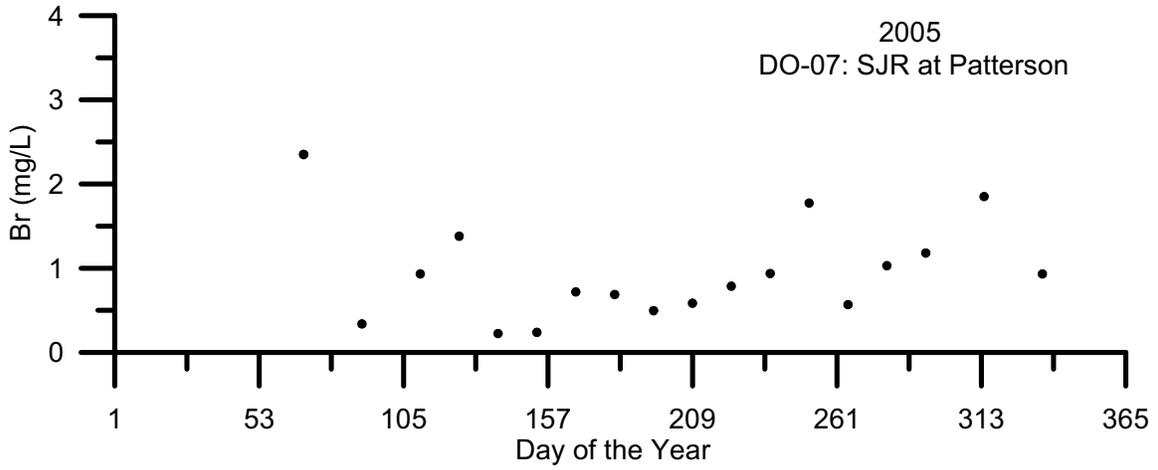


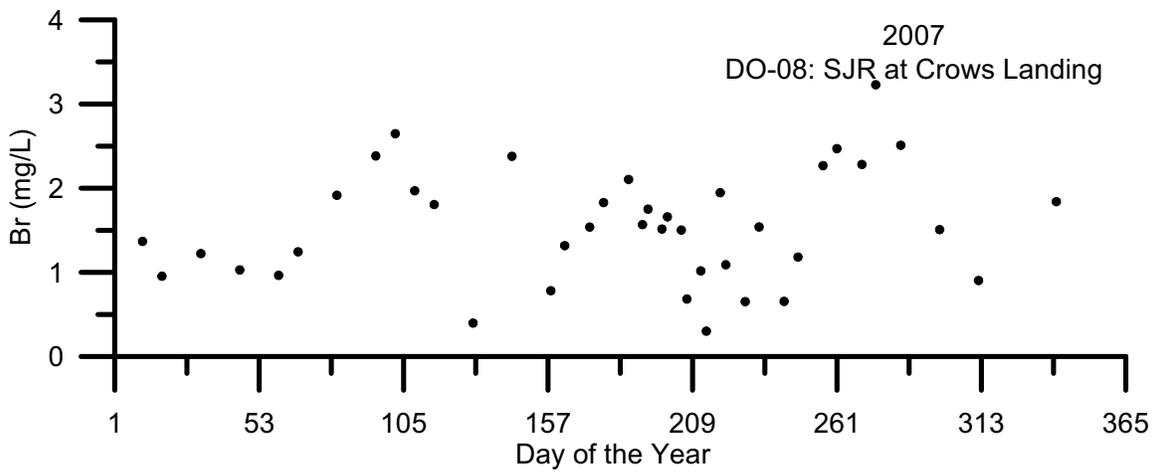
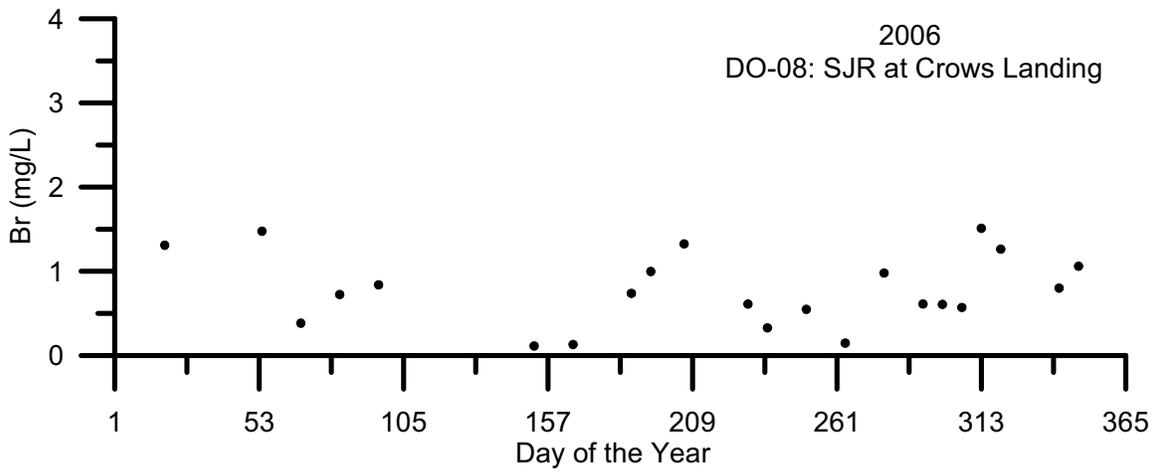
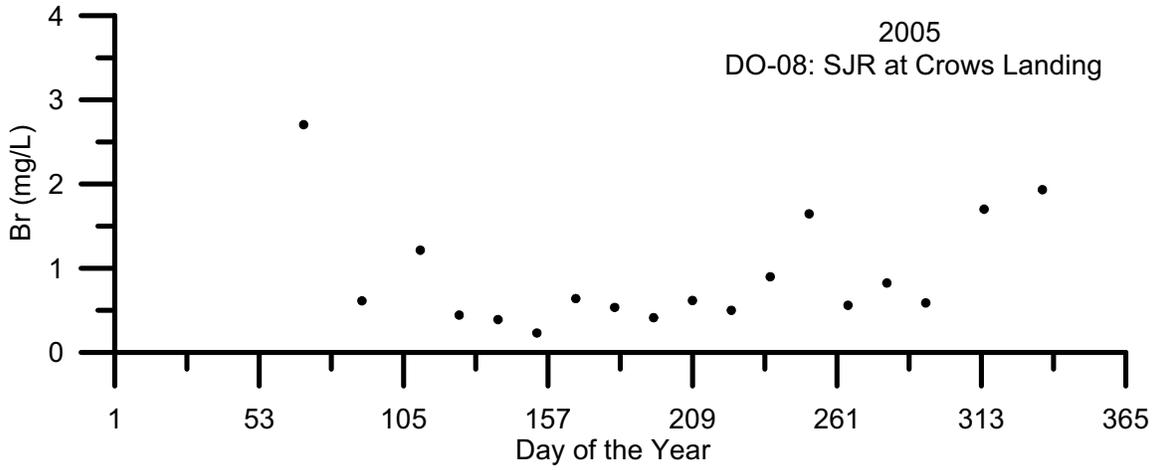
Figures 2 -103: Temporal Plots of Bromine By Site ID

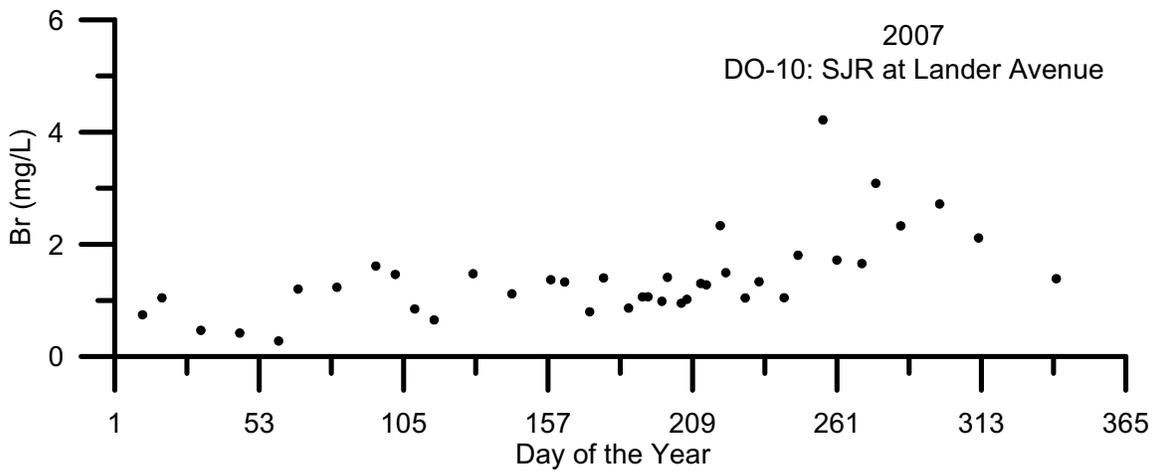
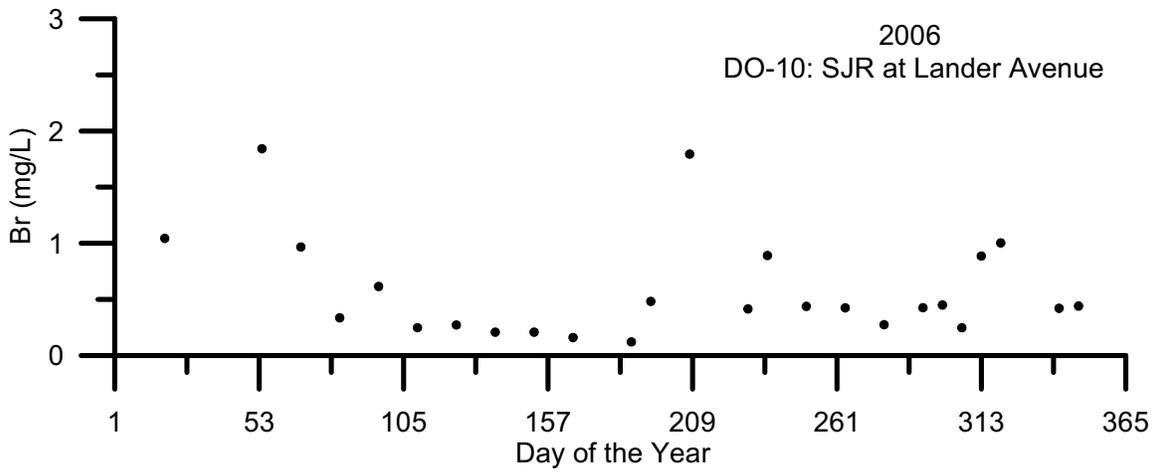
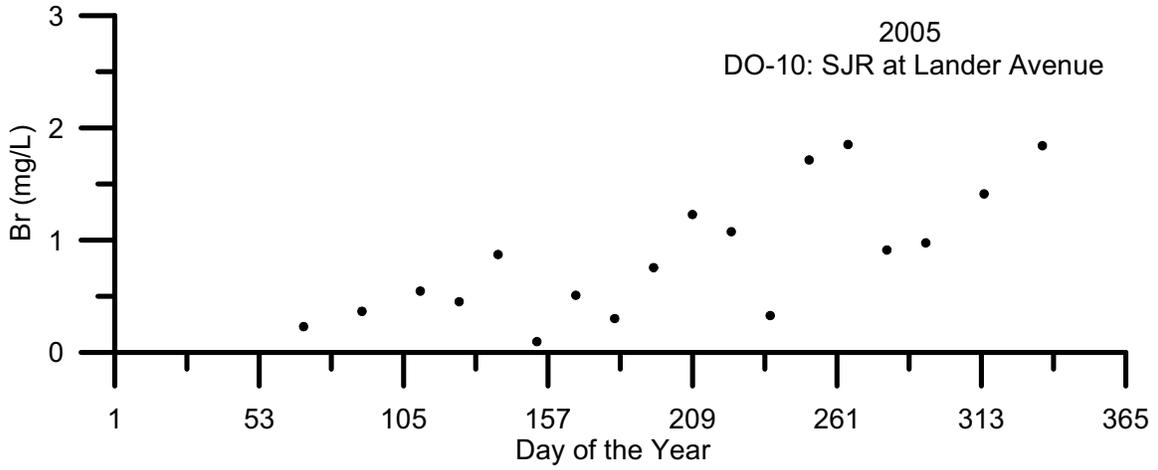


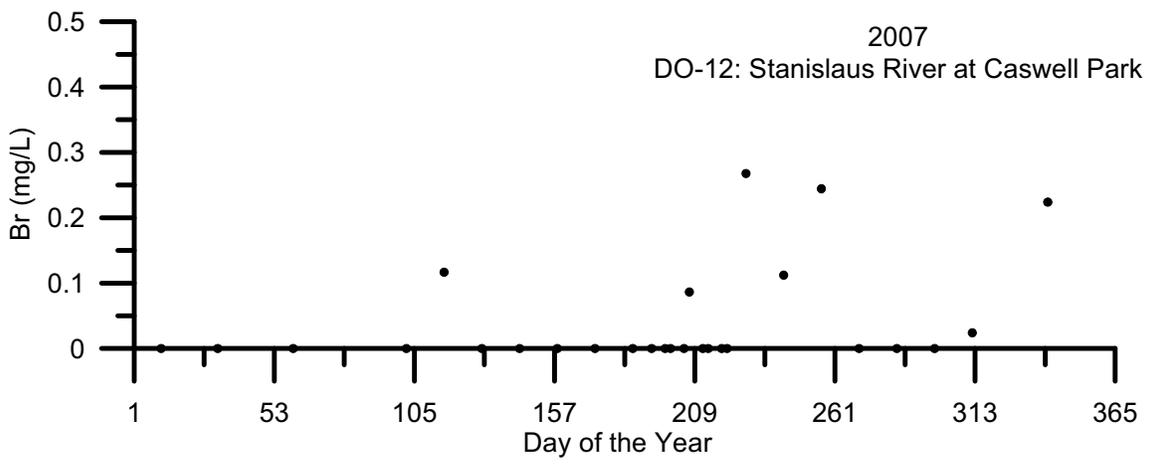
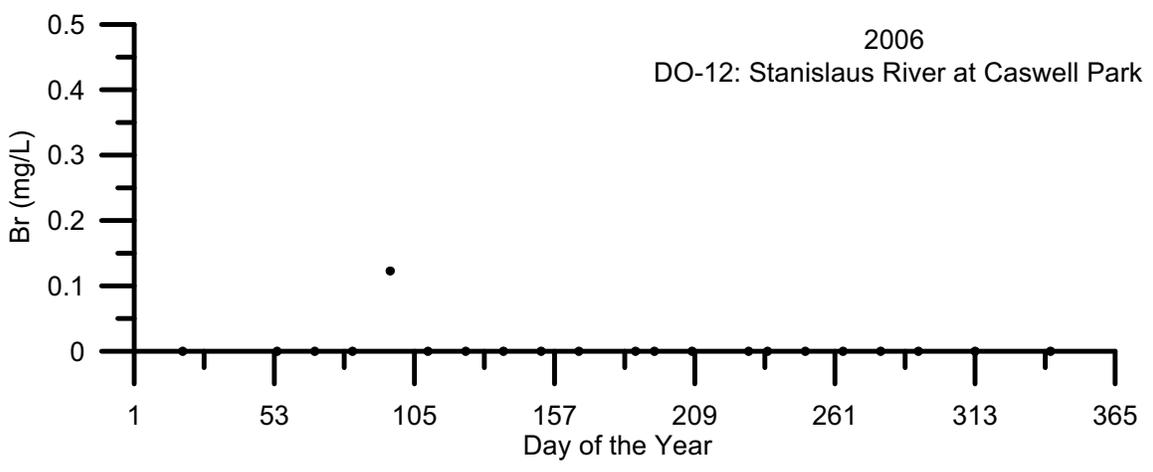
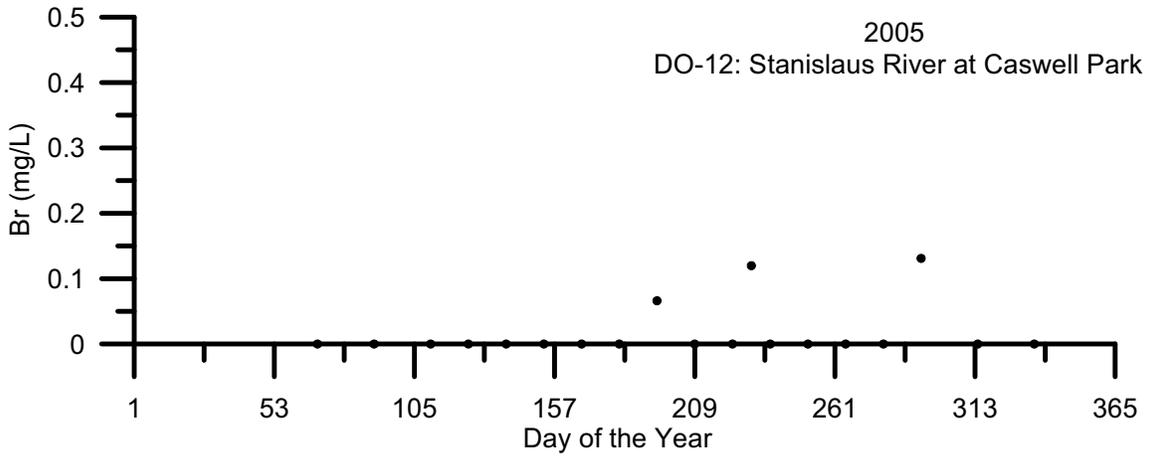


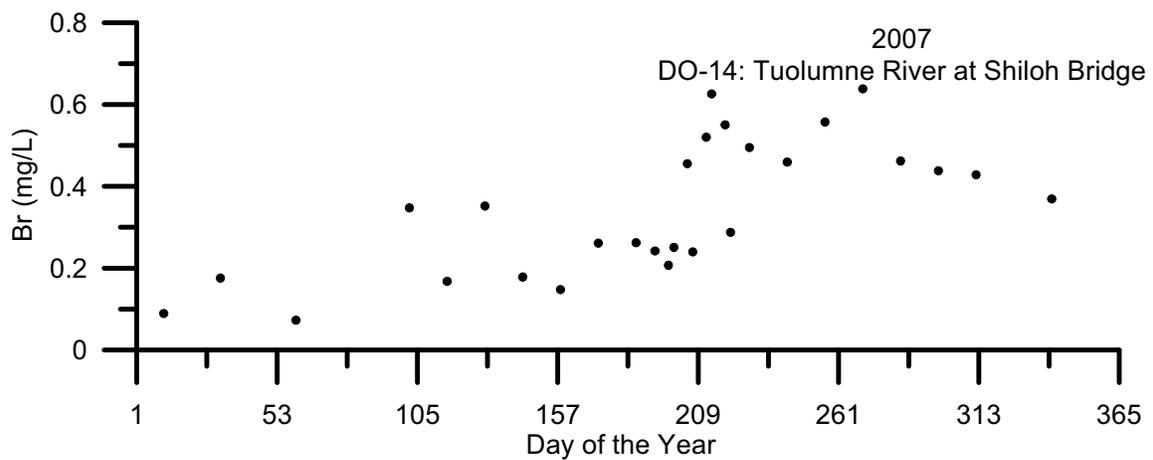
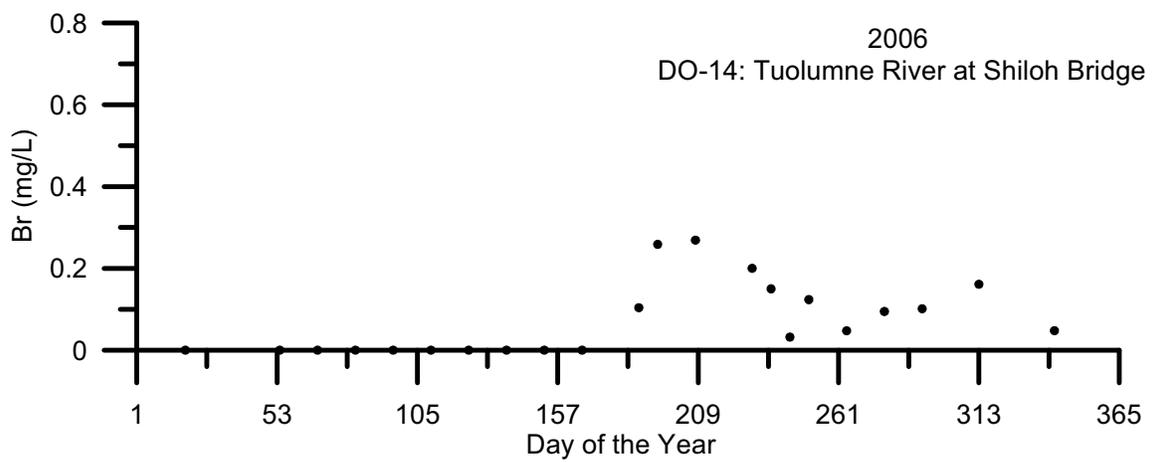
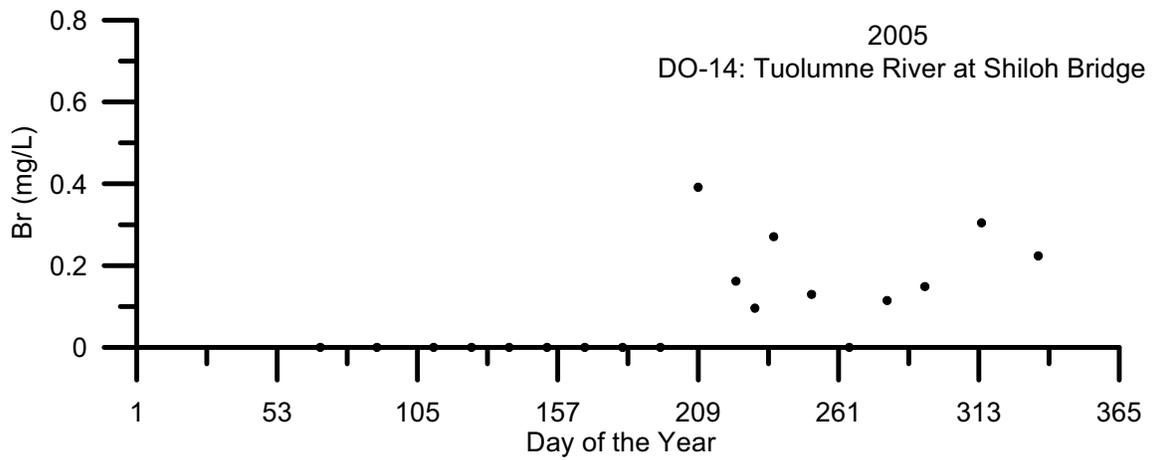


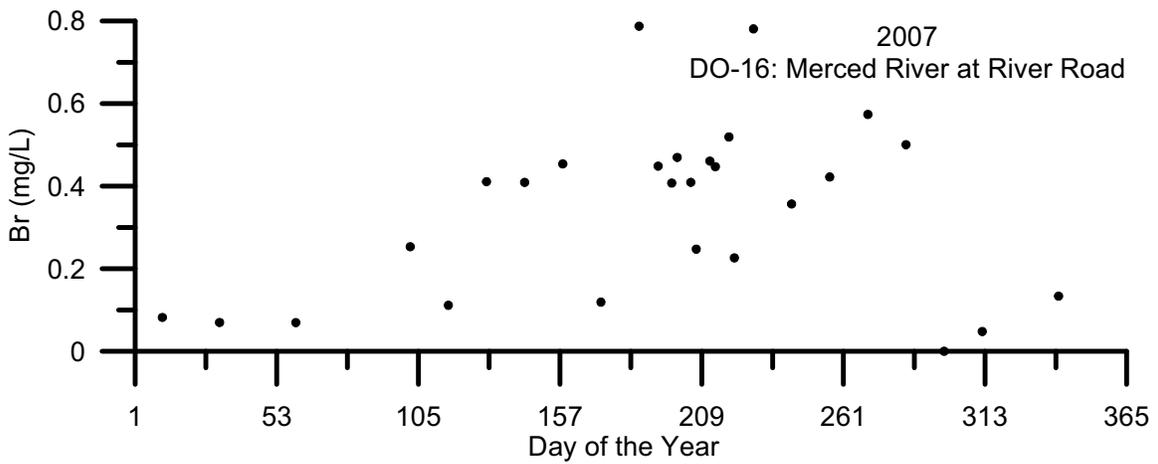
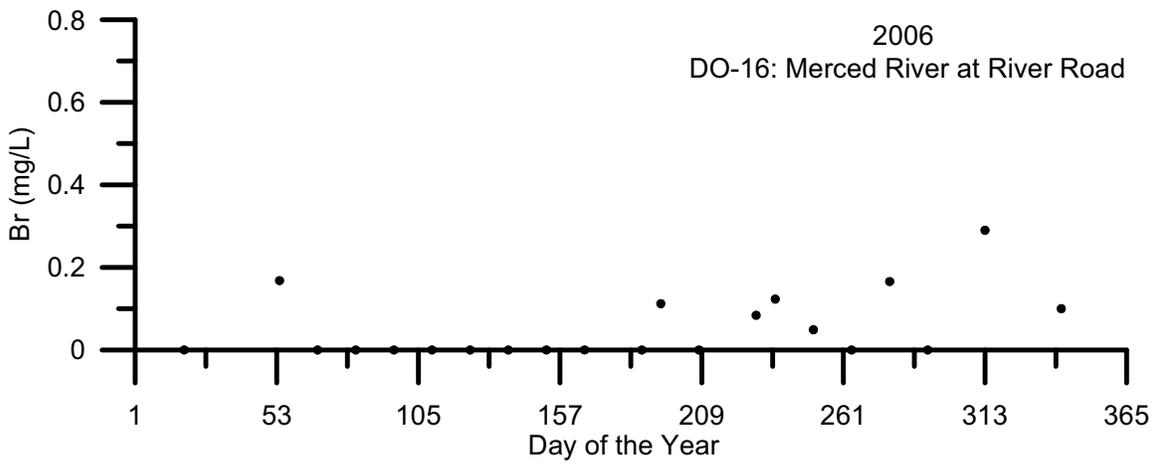
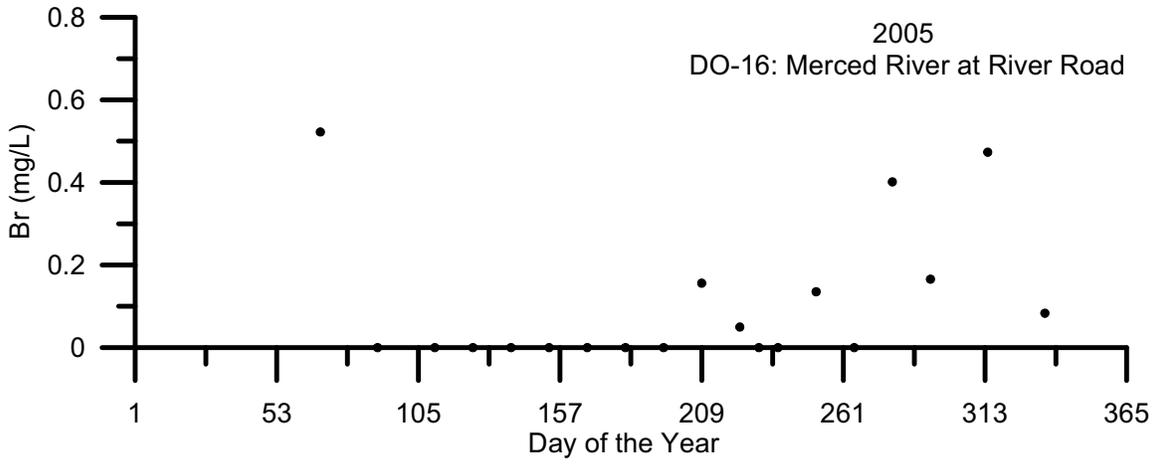


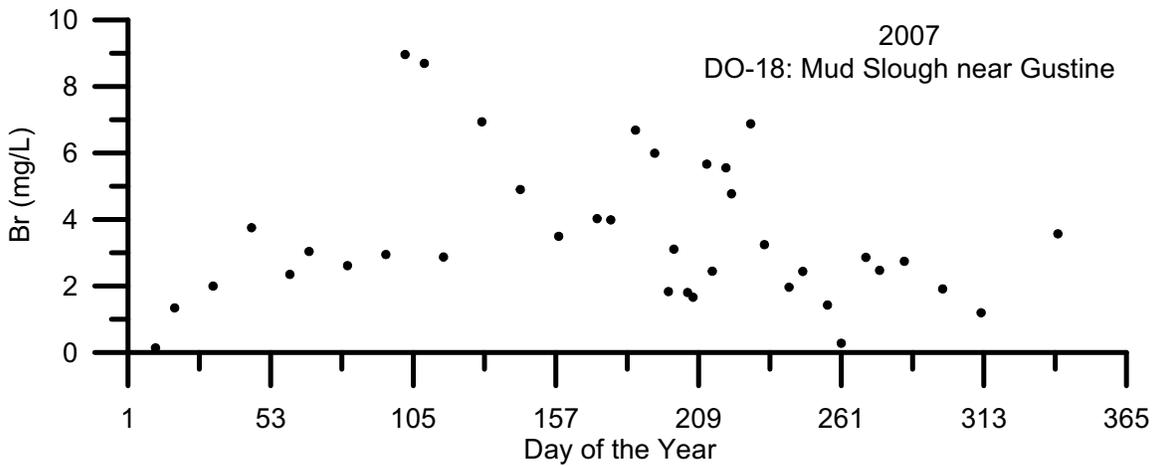
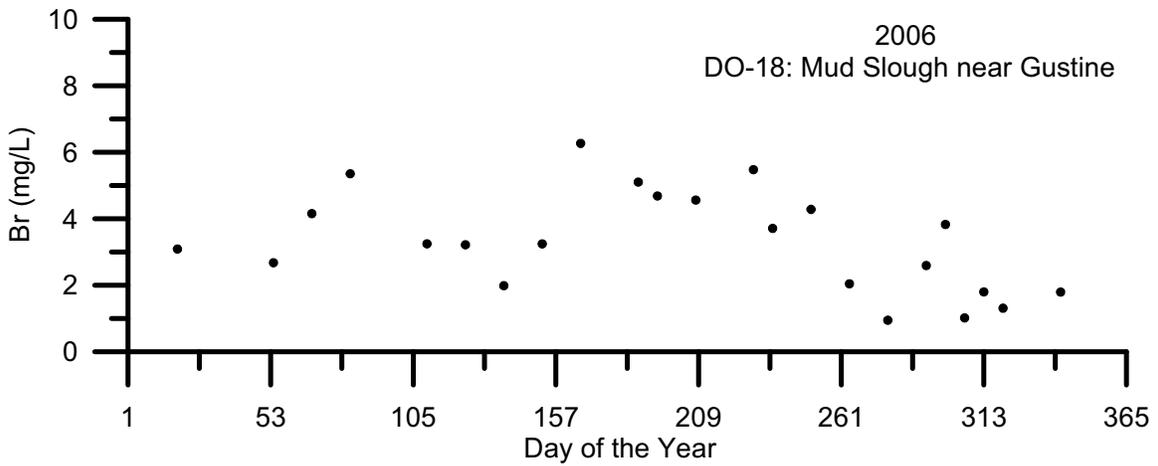
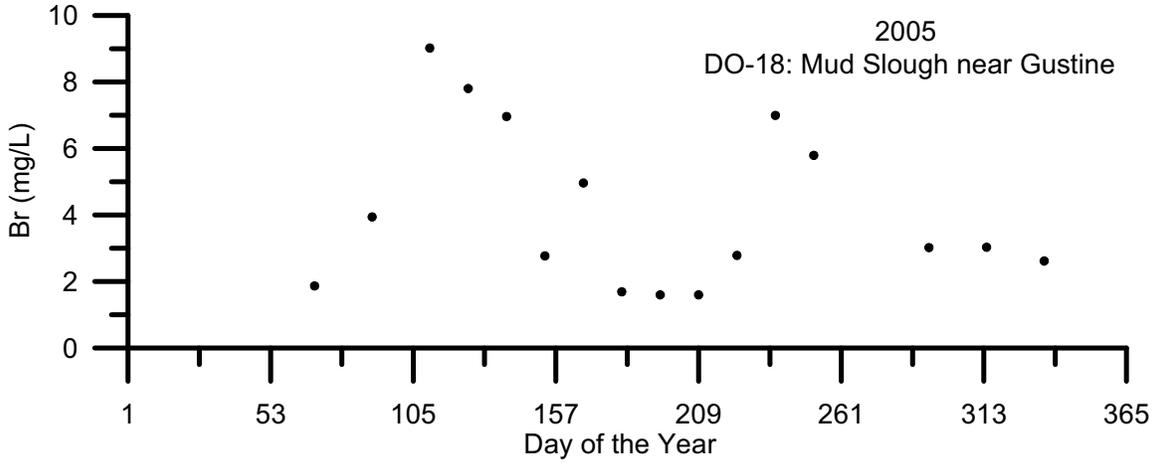


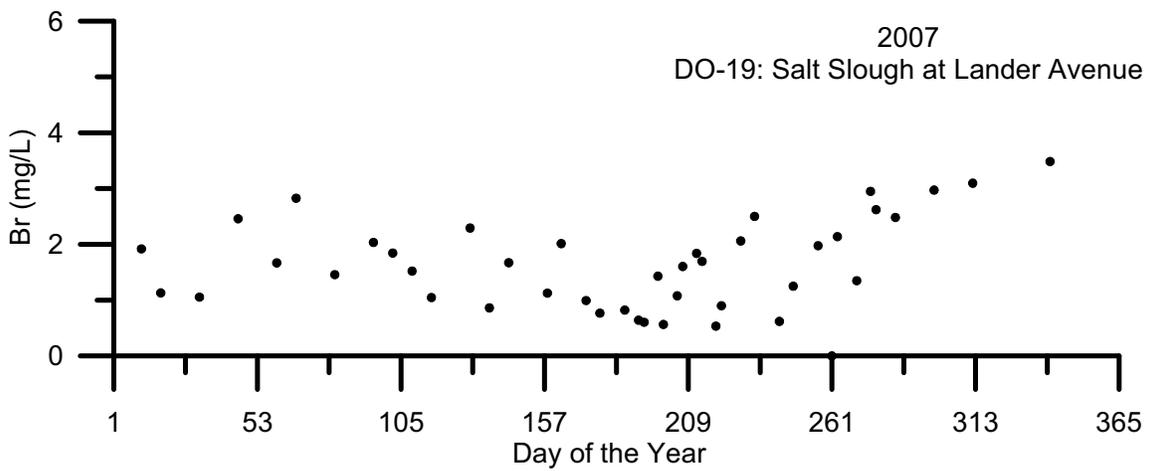
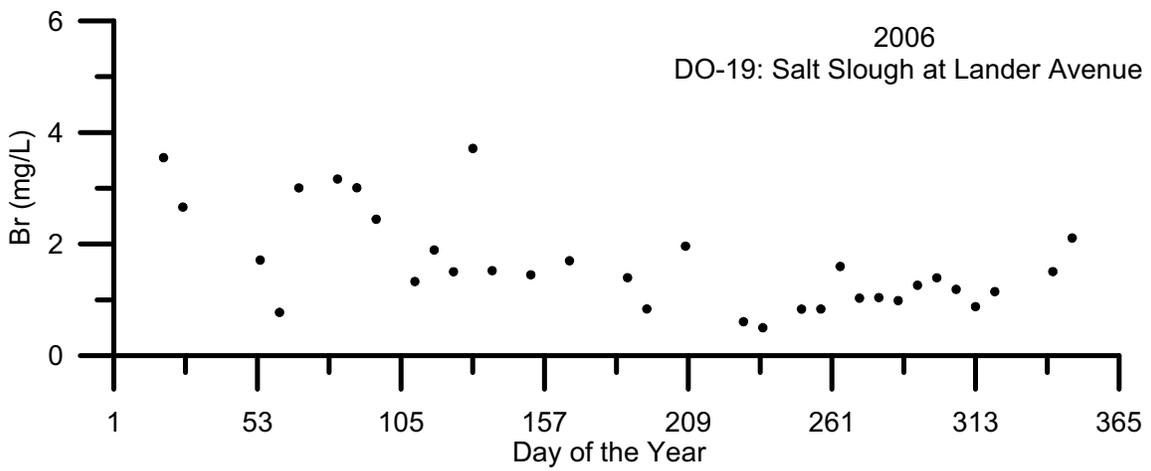
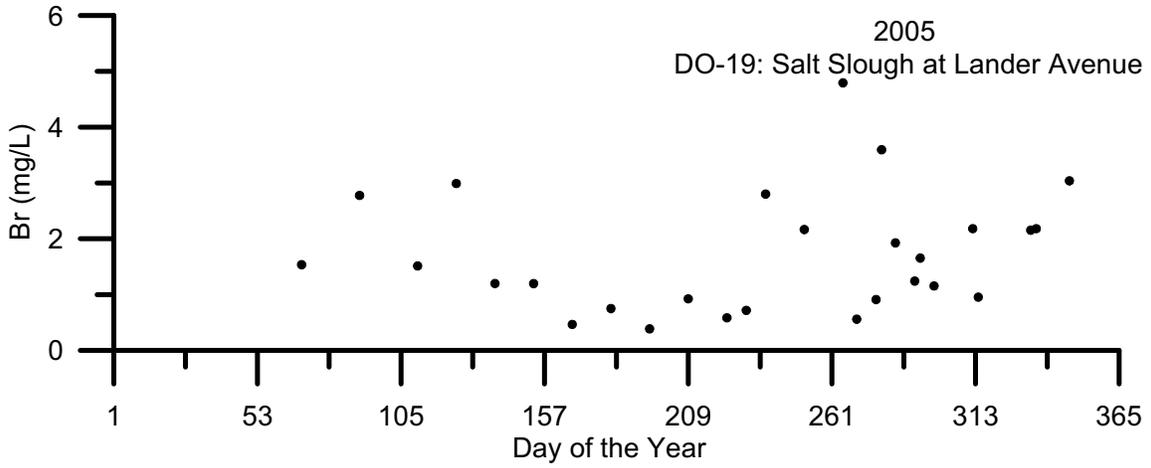


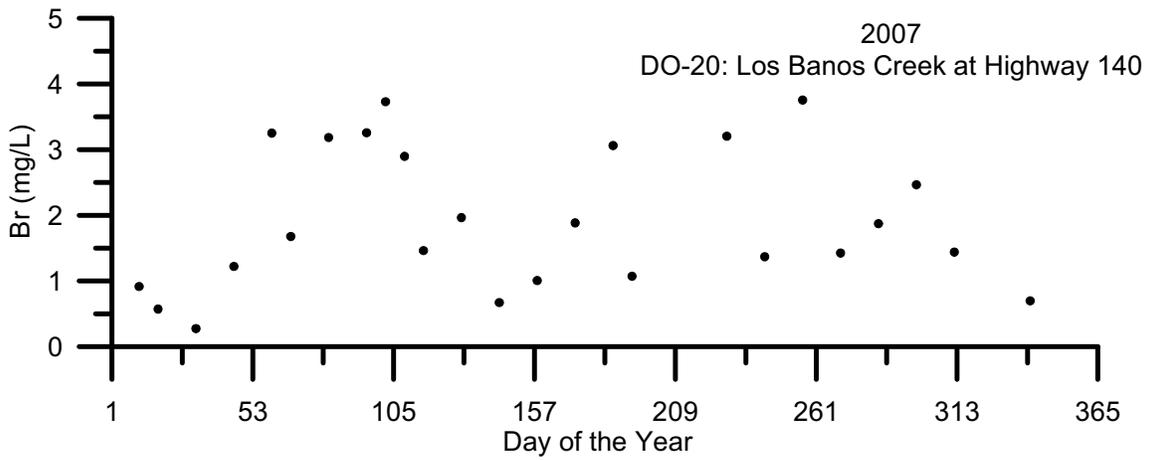
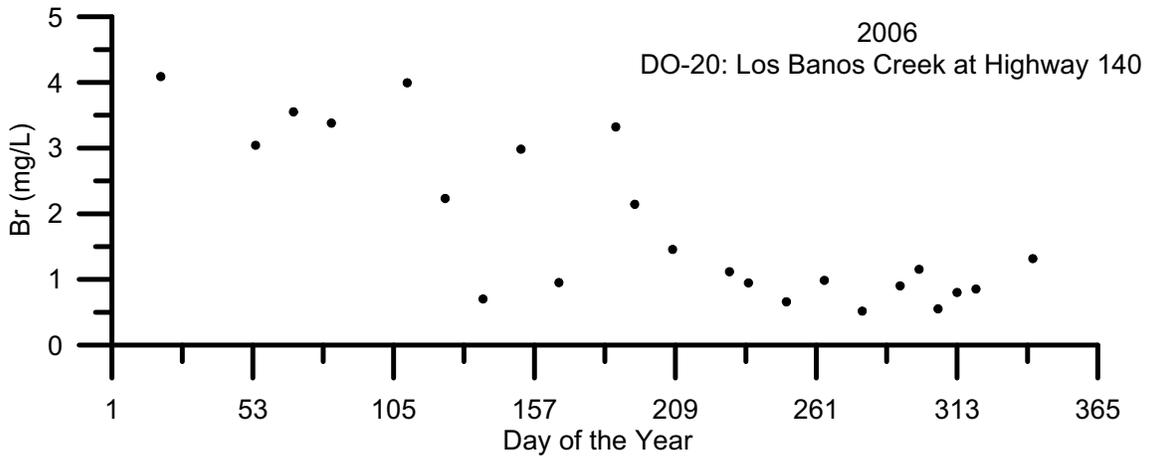
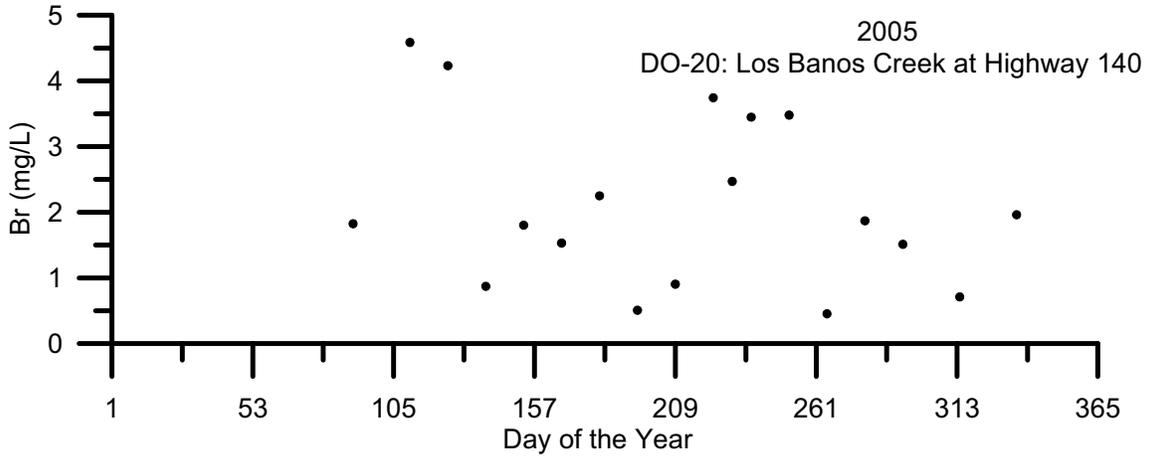


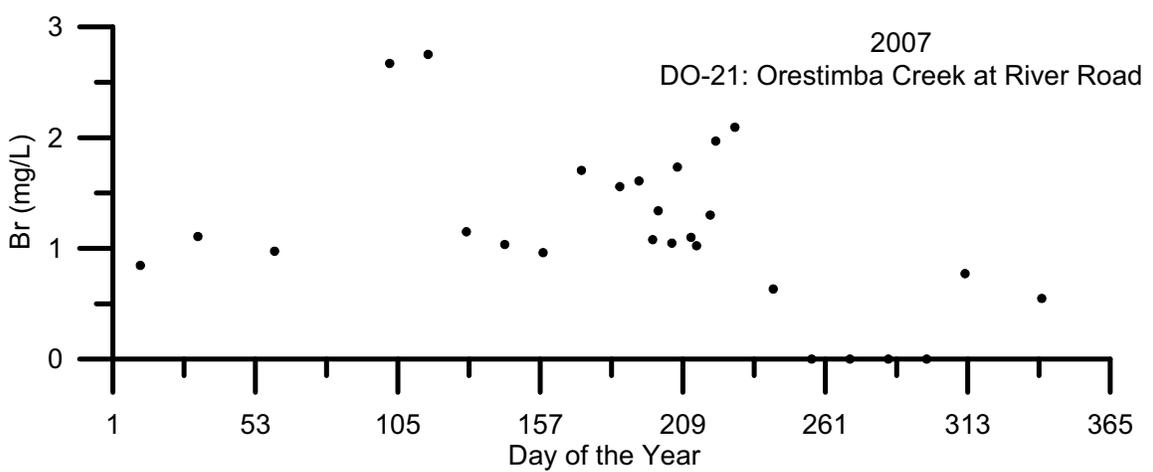
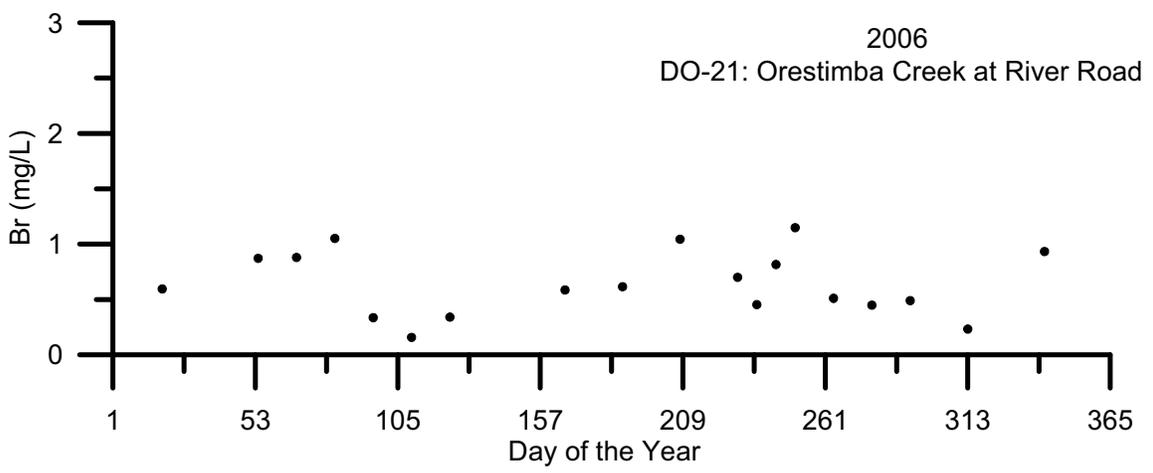
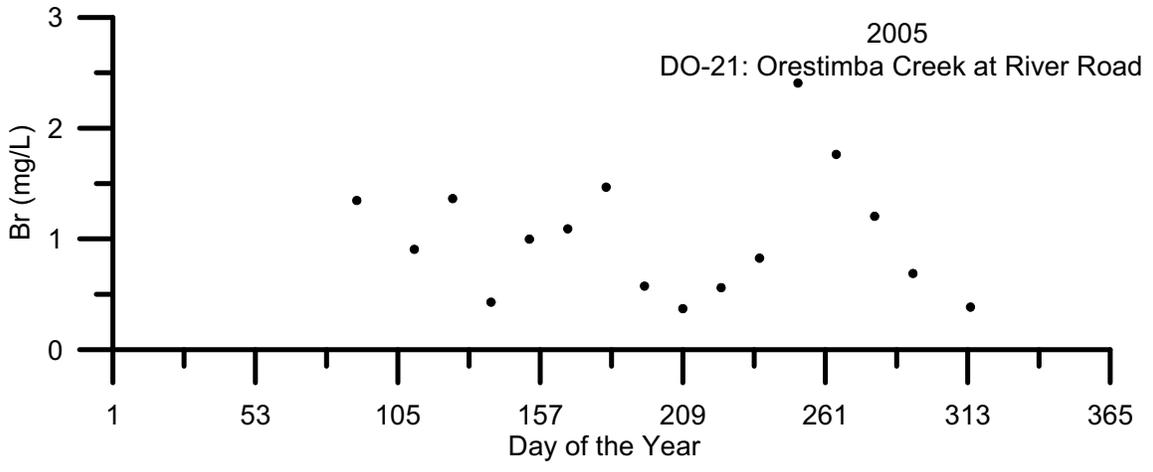


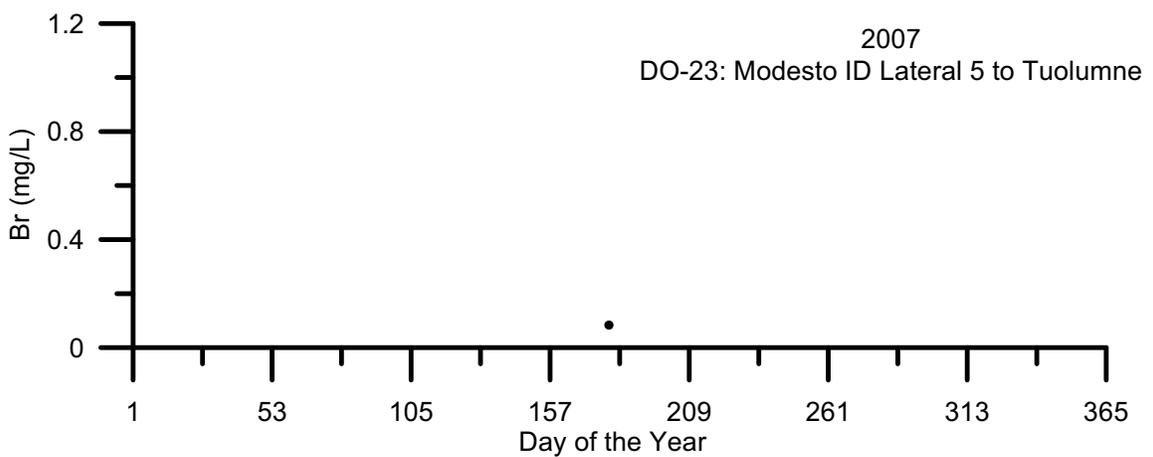
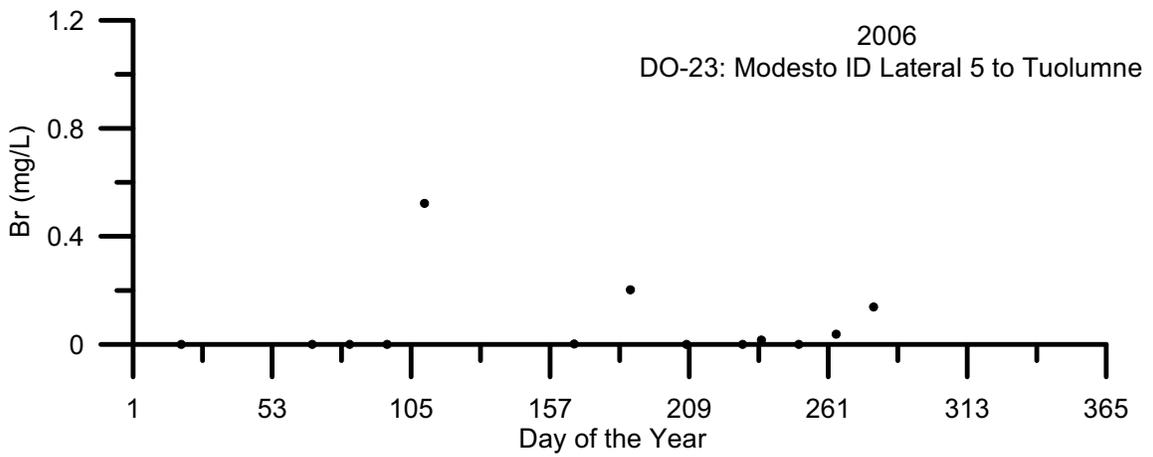
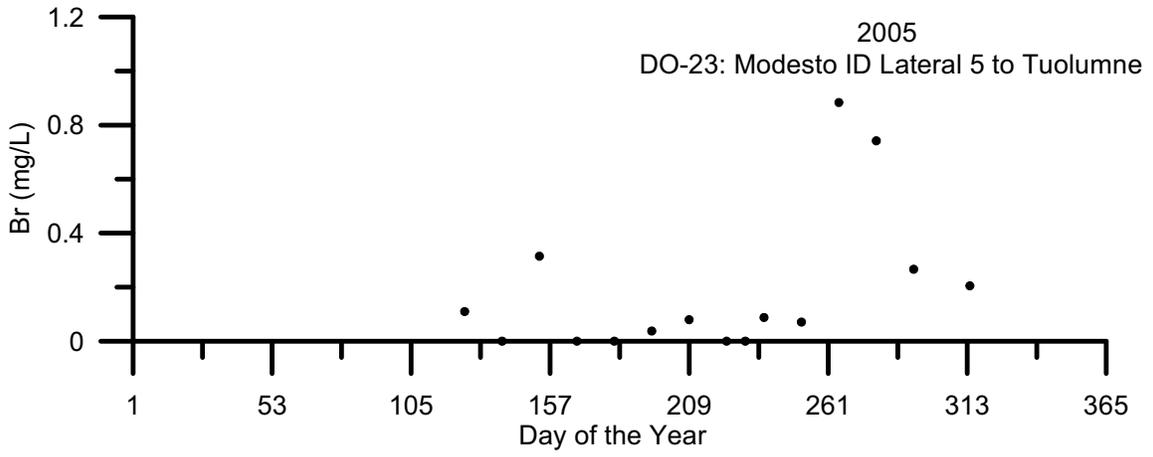


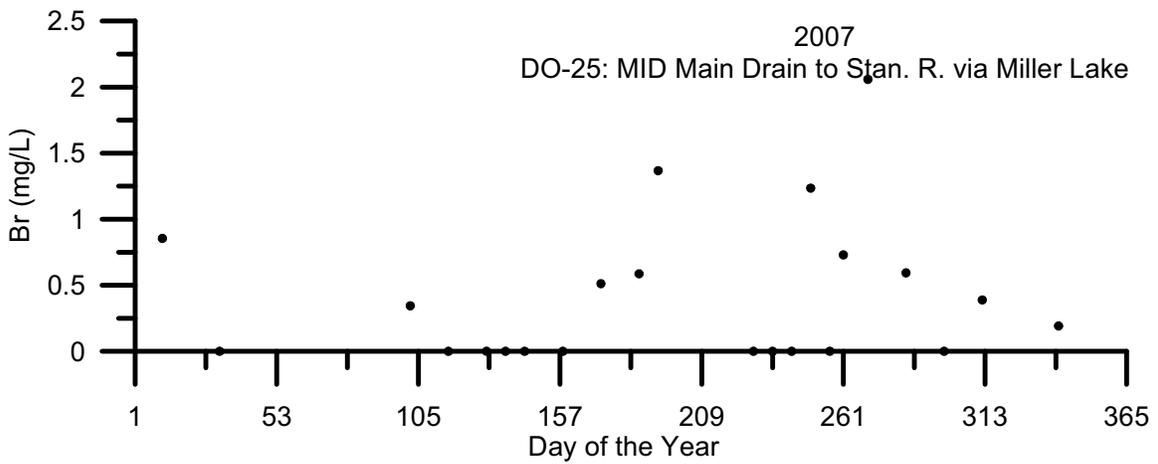
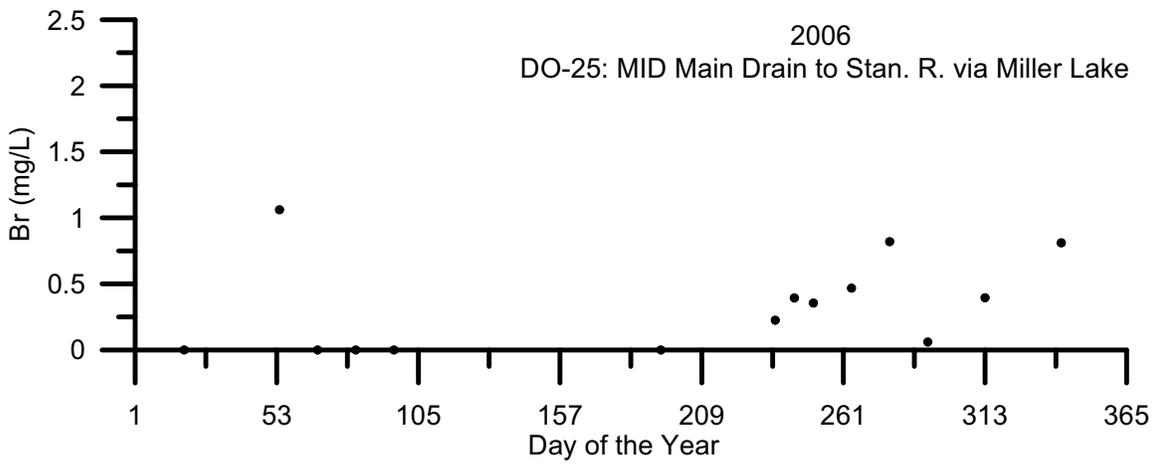
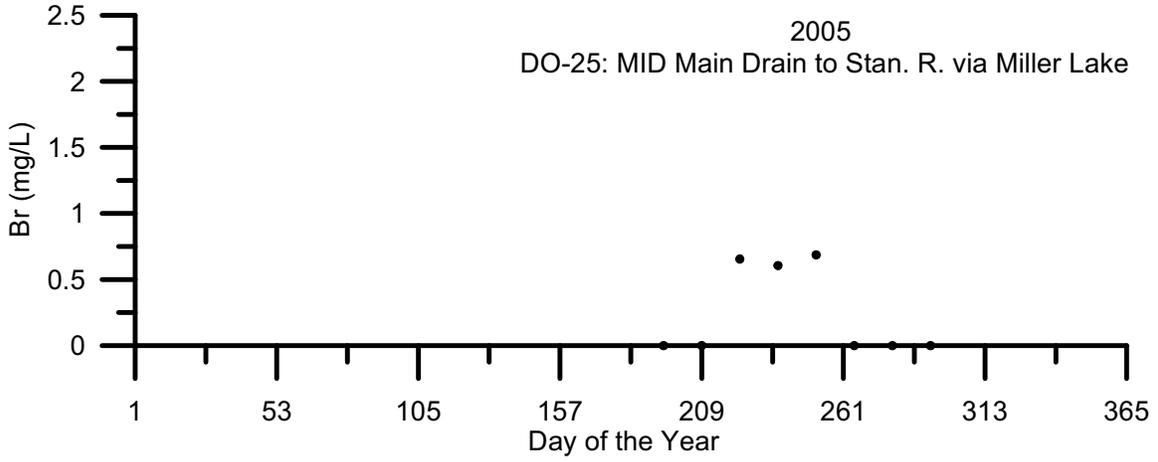


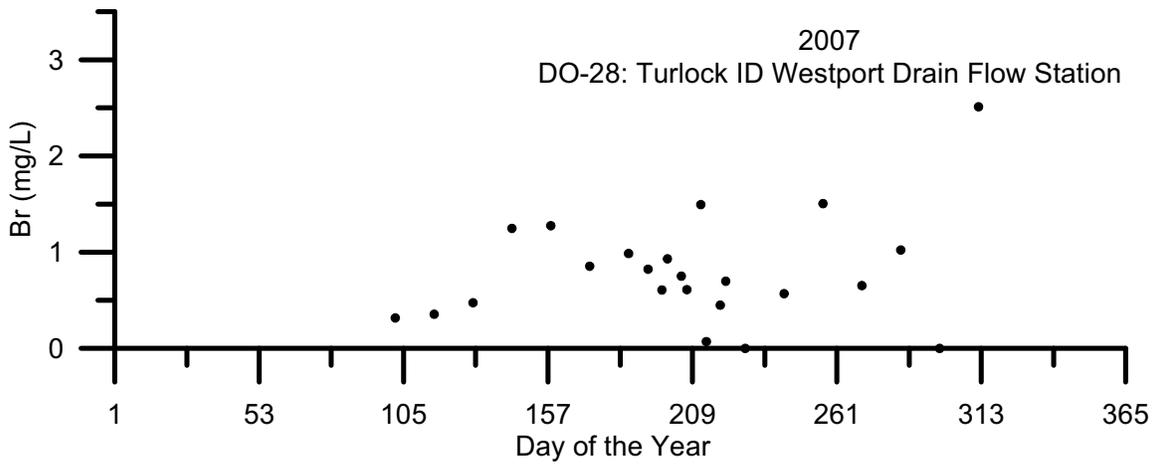
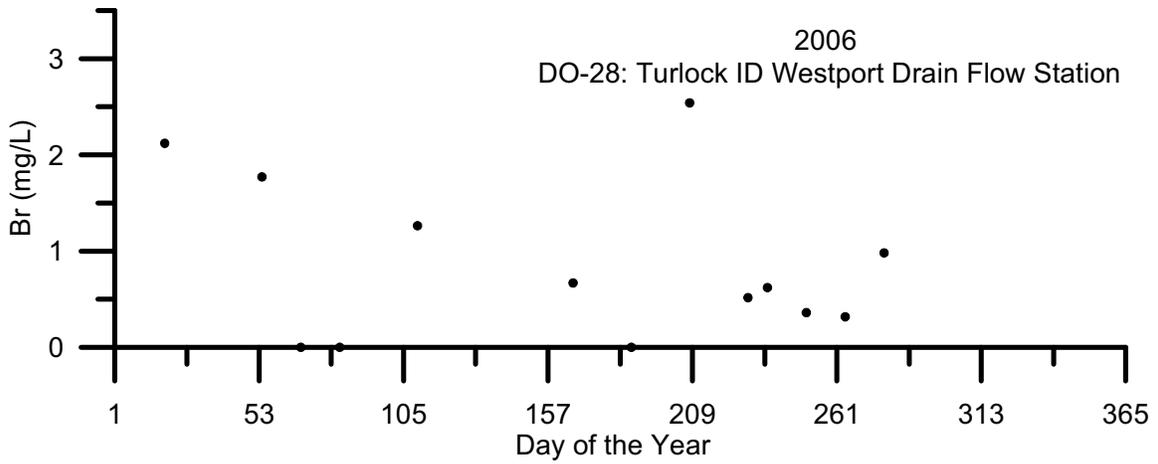
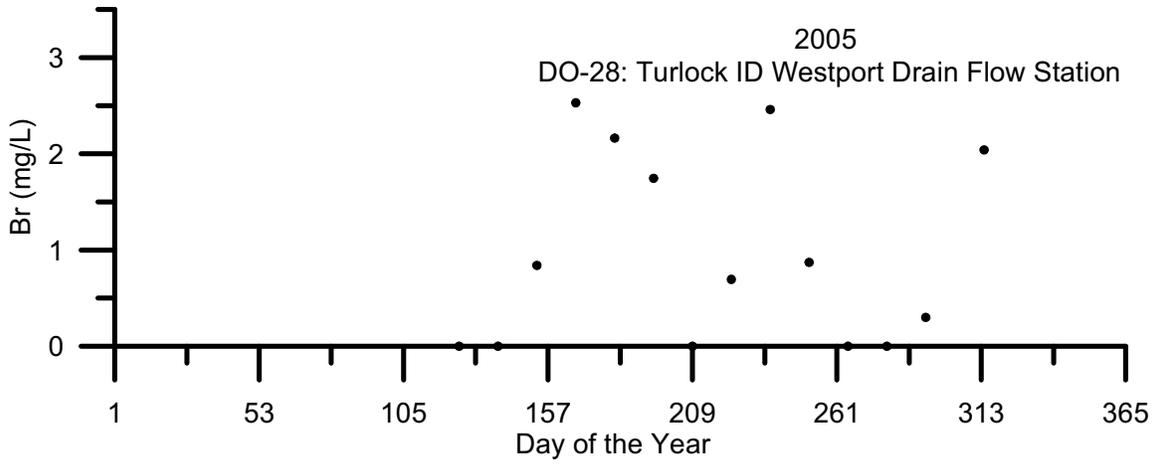


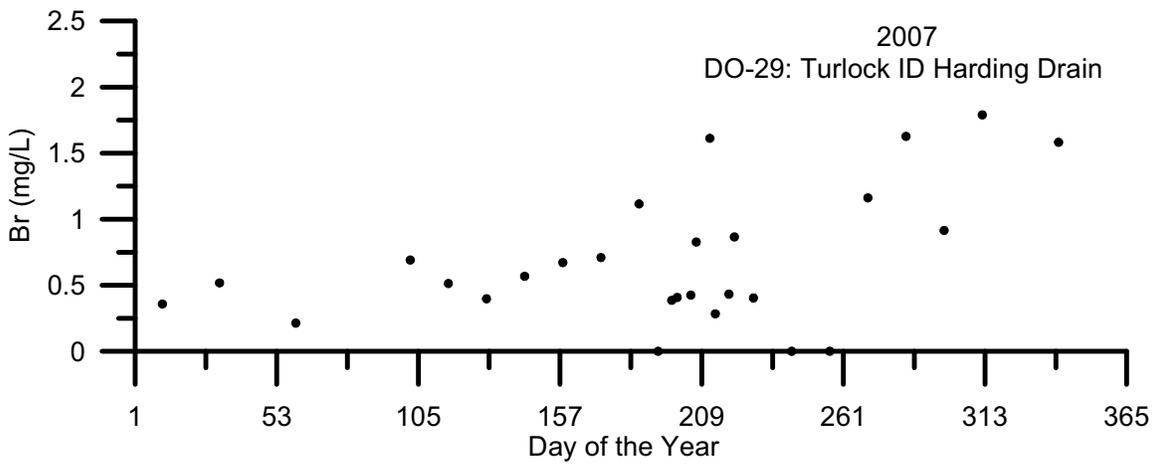
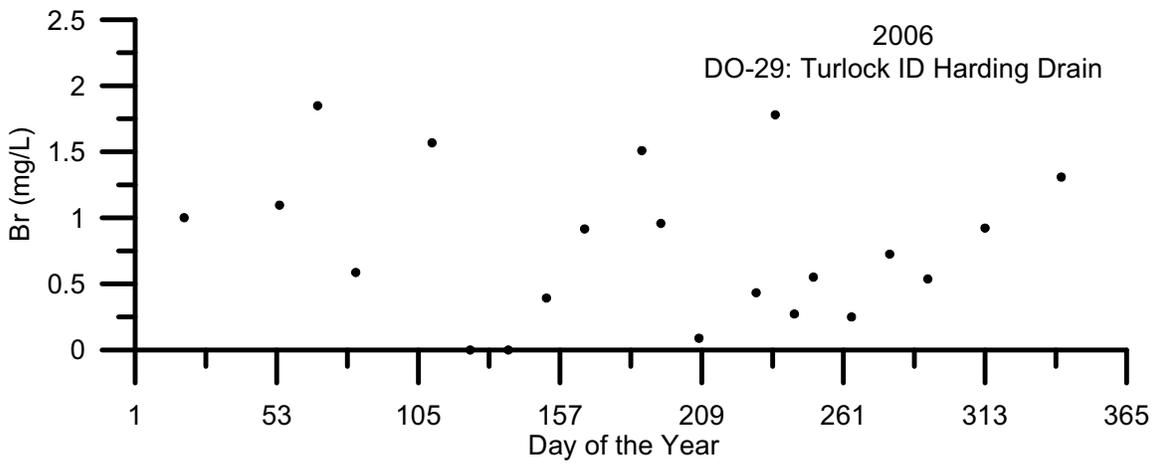
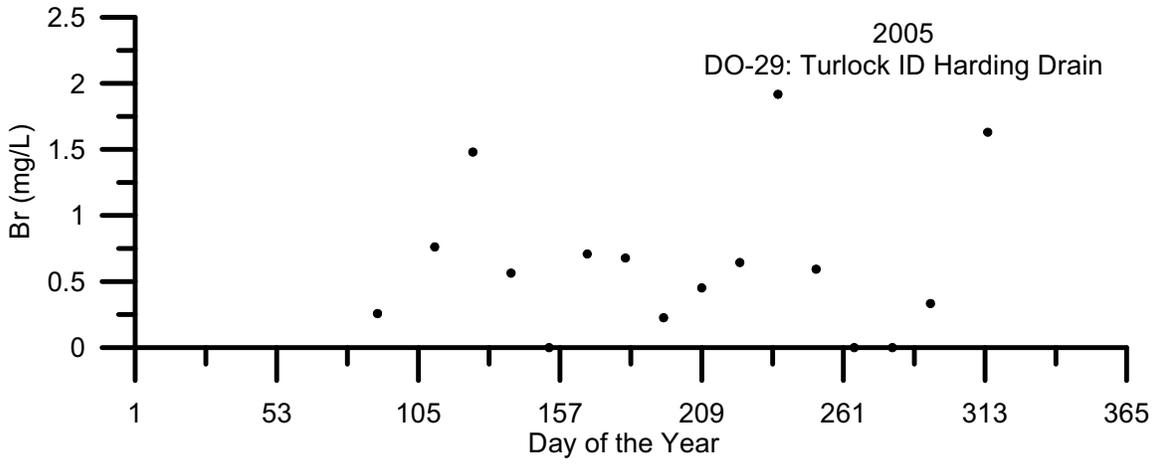


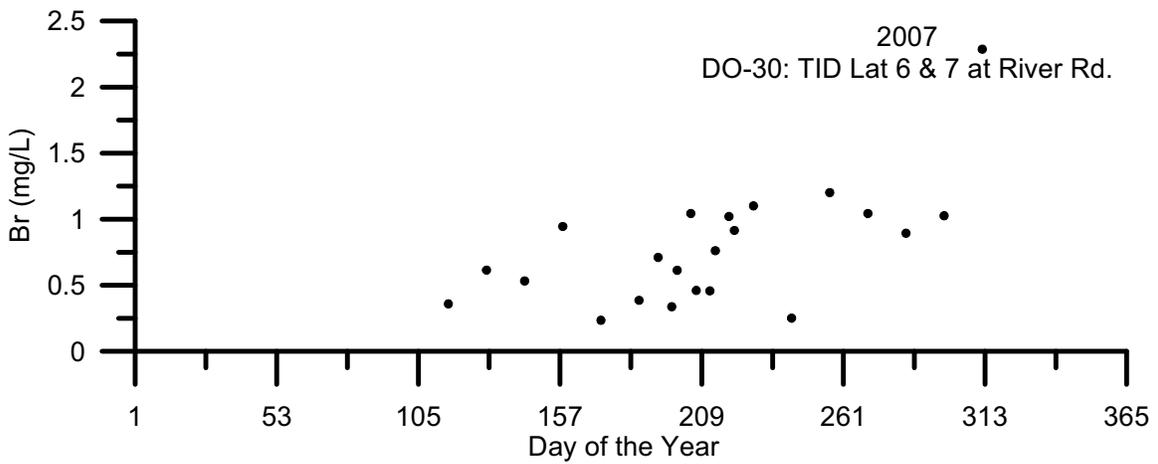
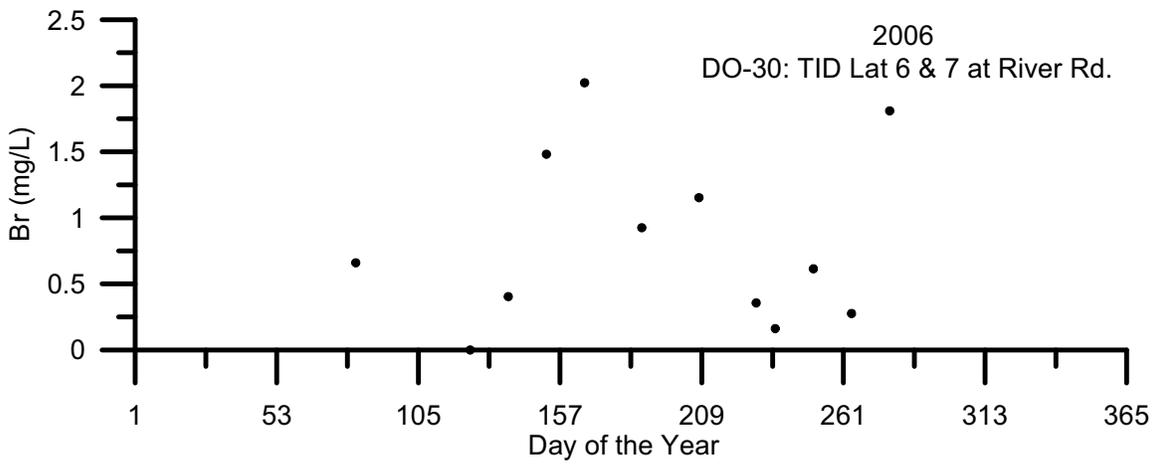
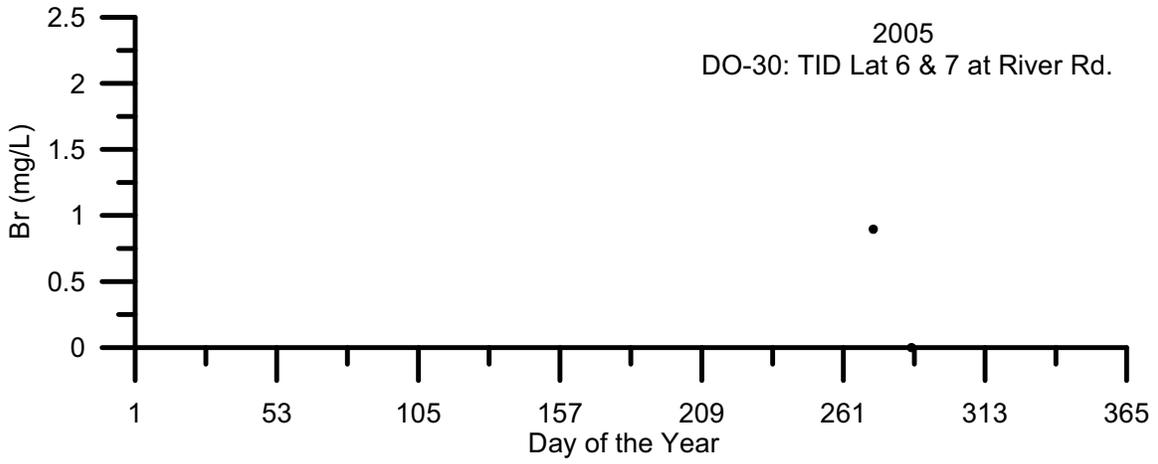


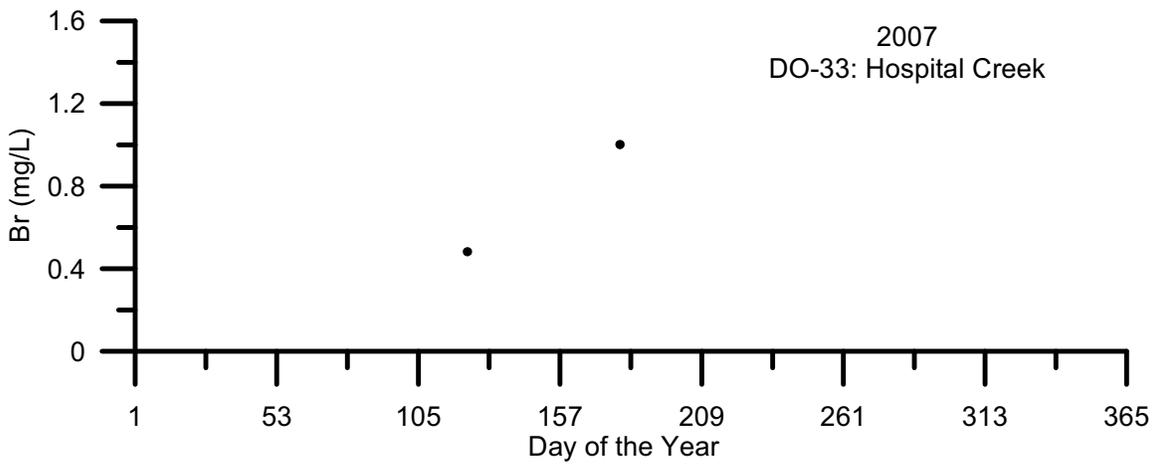
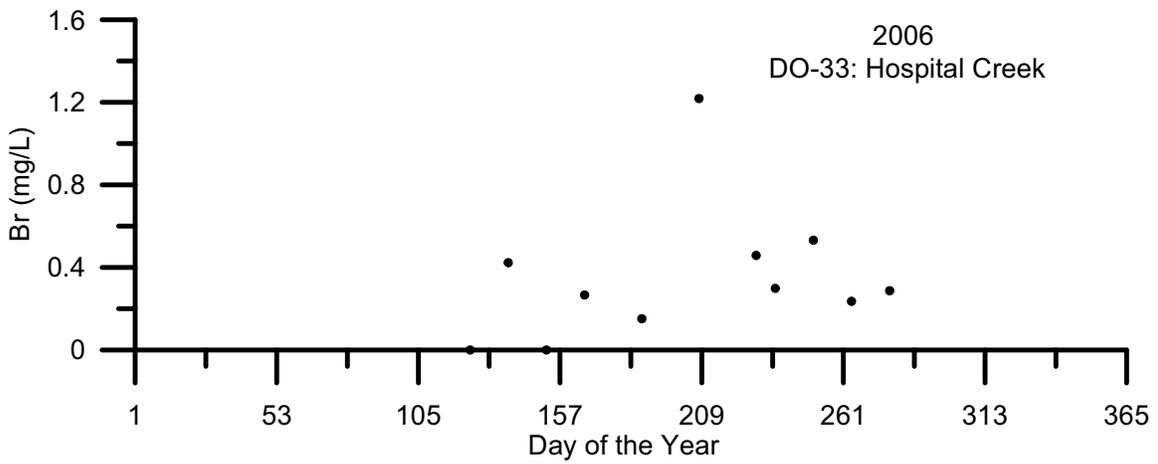
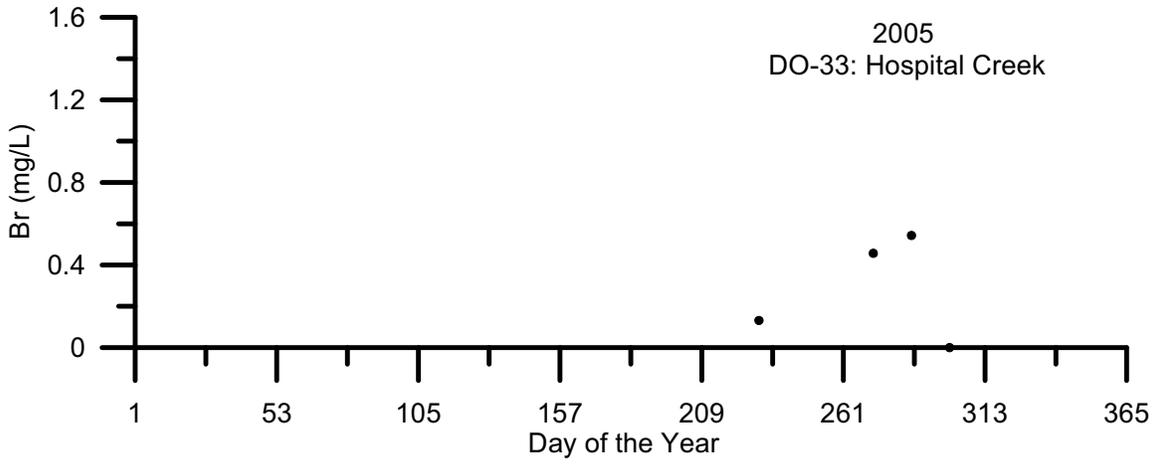


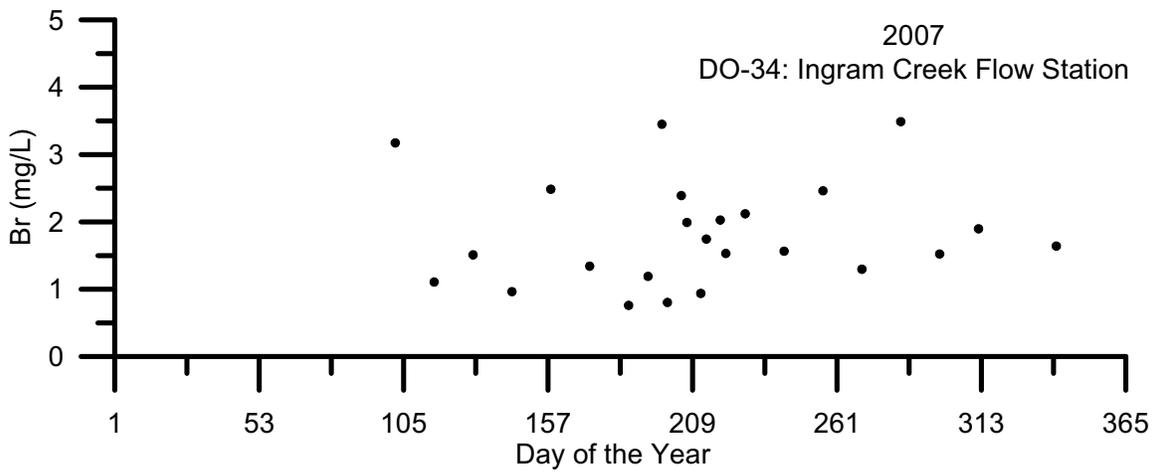
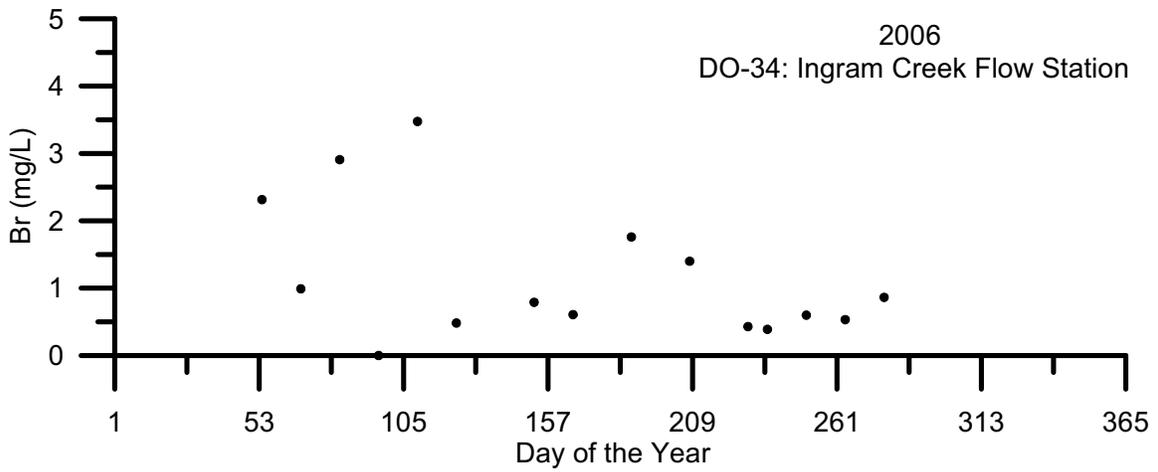
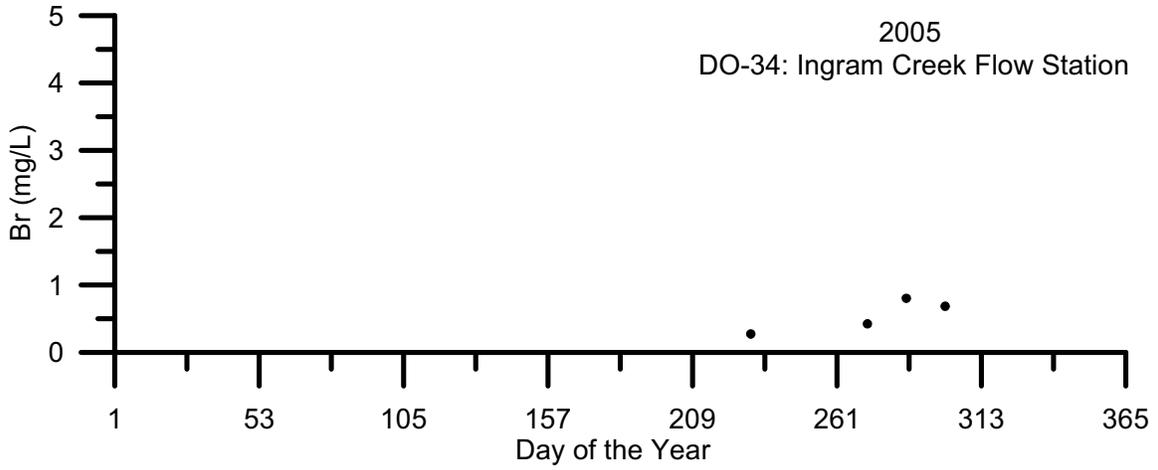


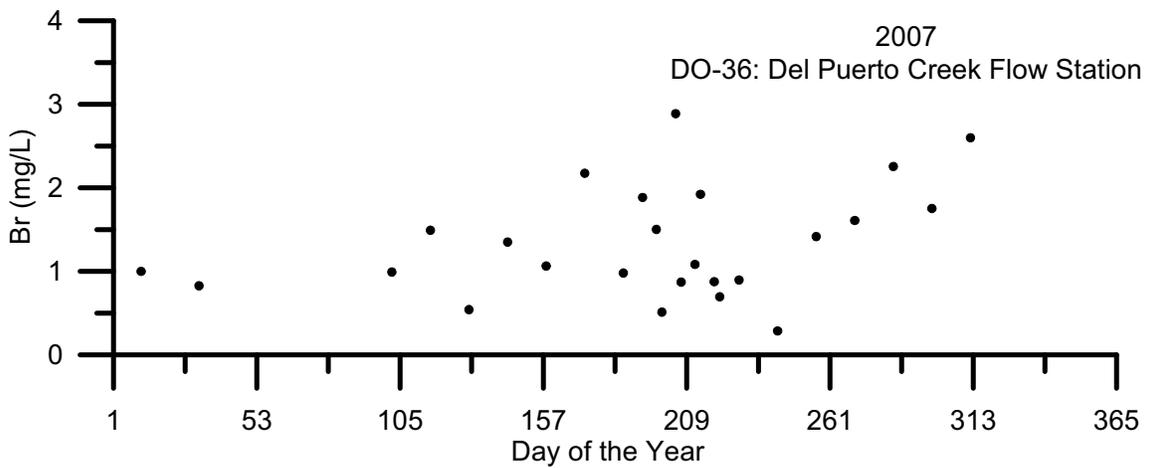
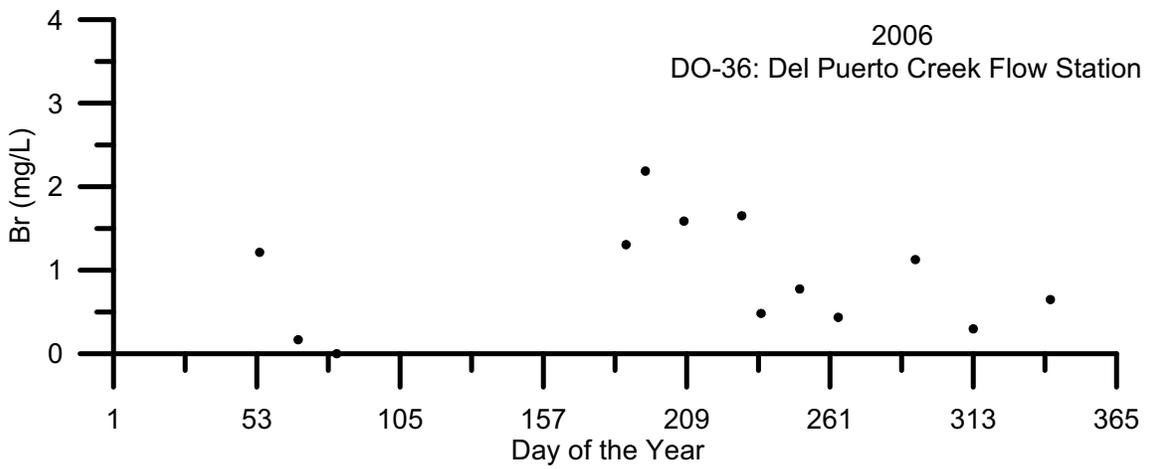
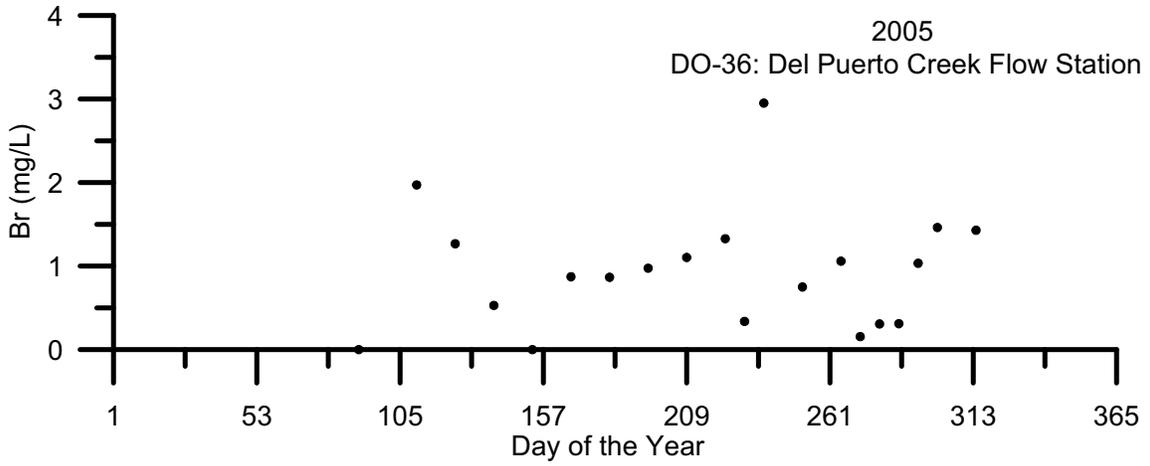


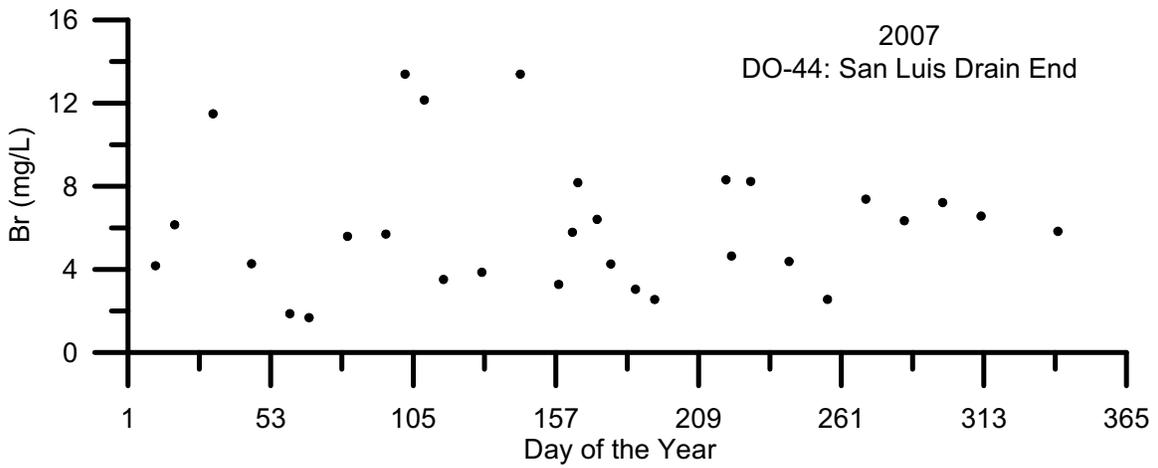
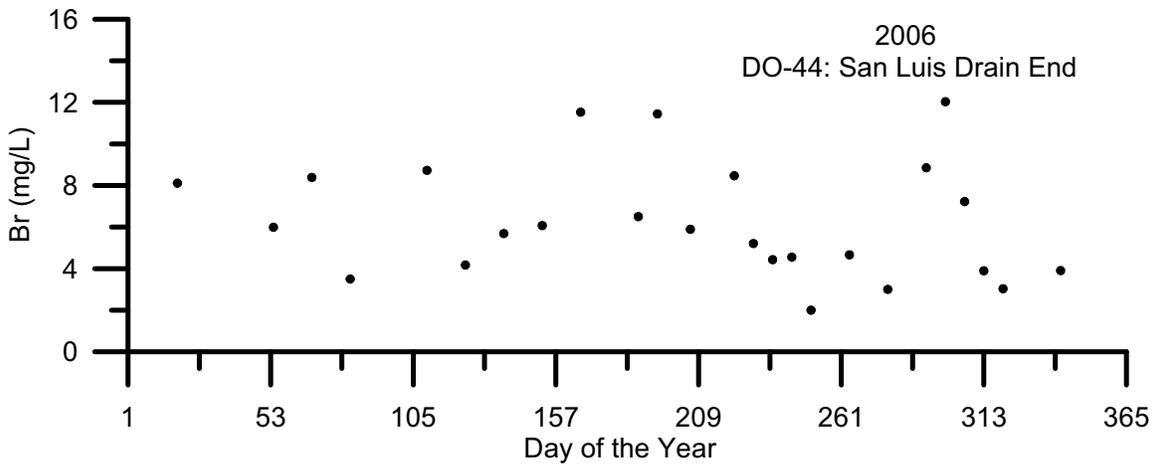
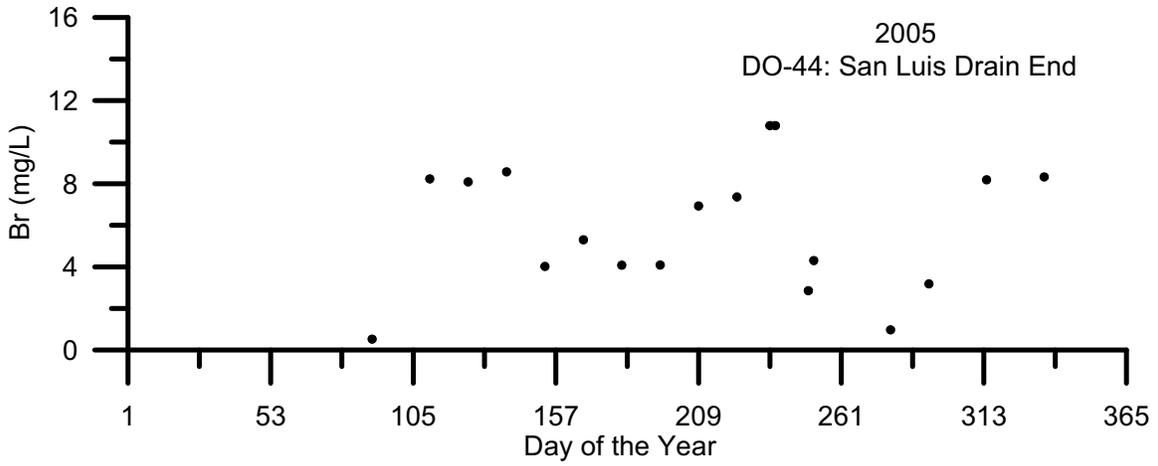


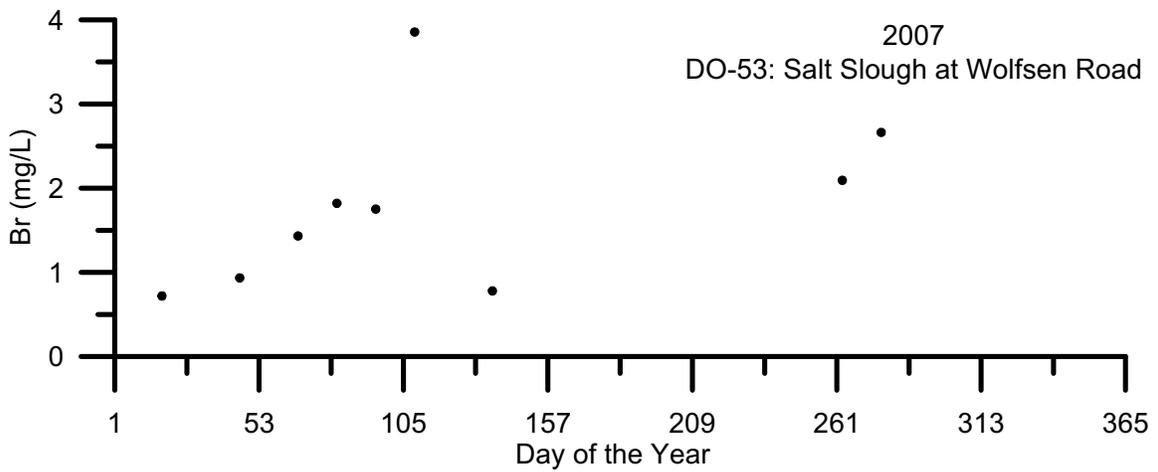
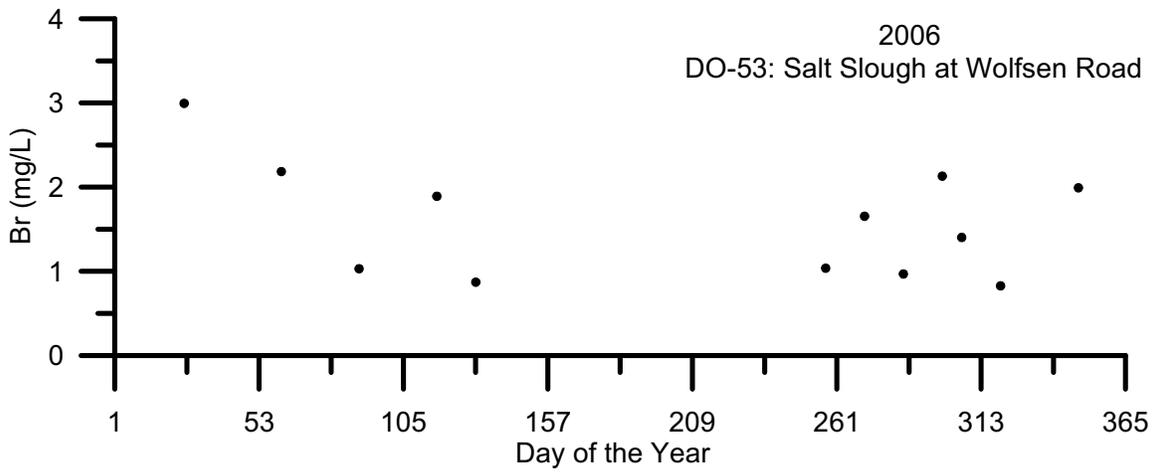
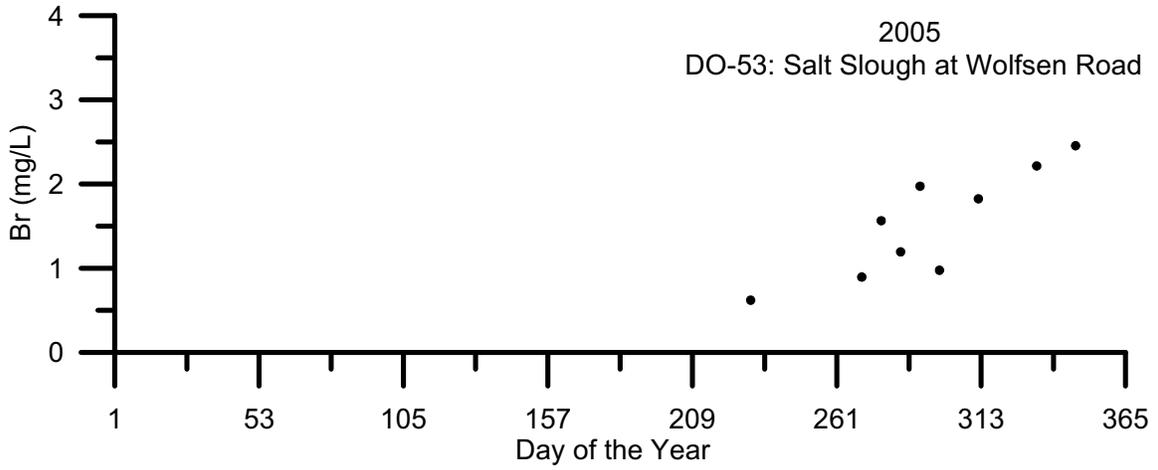


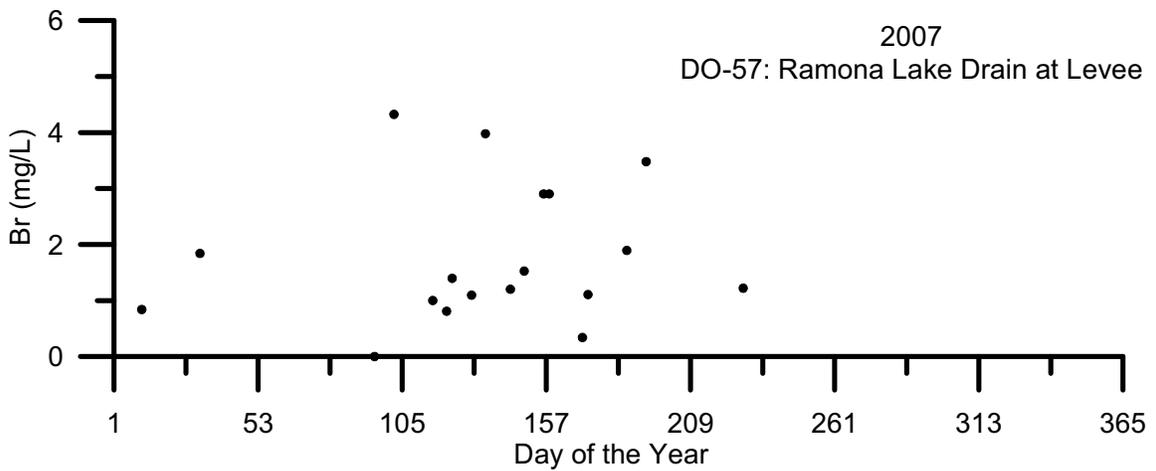
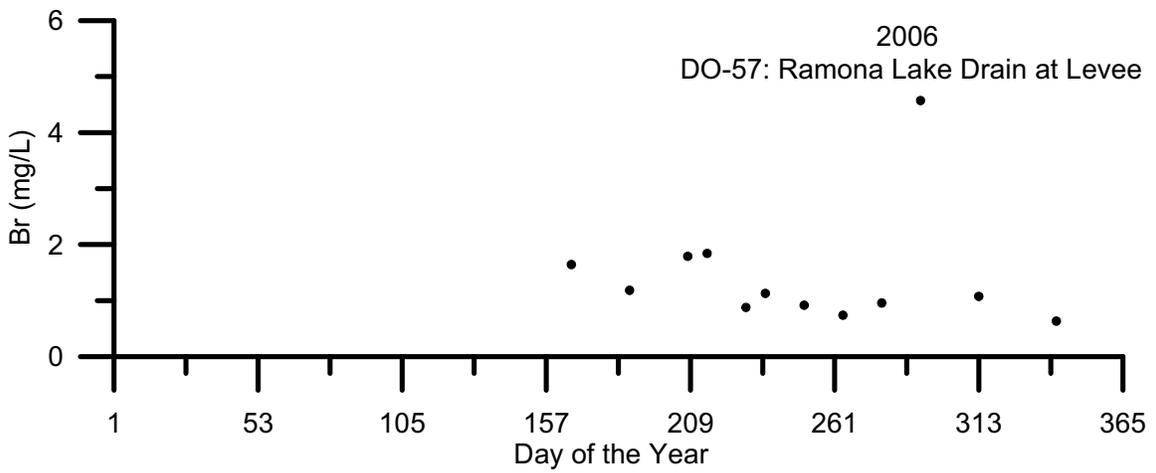
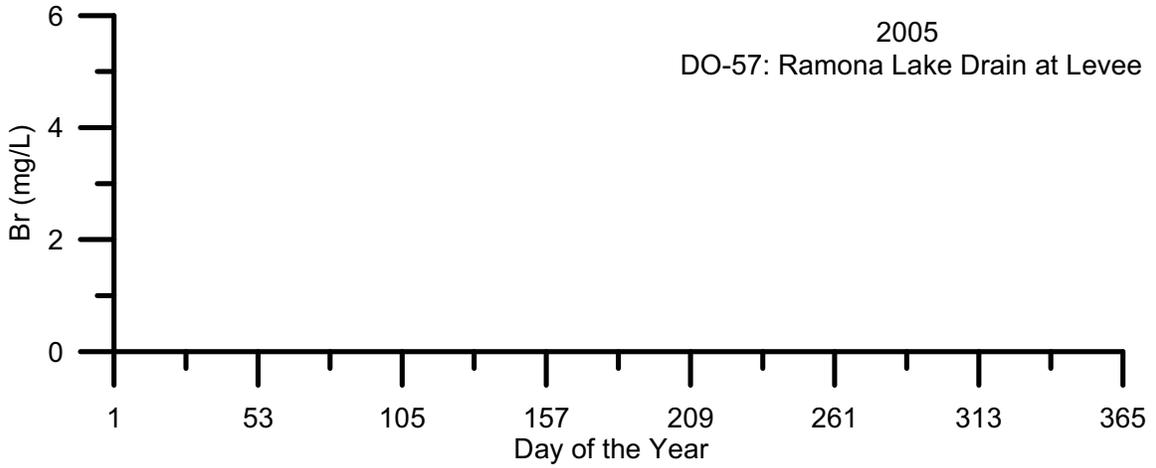


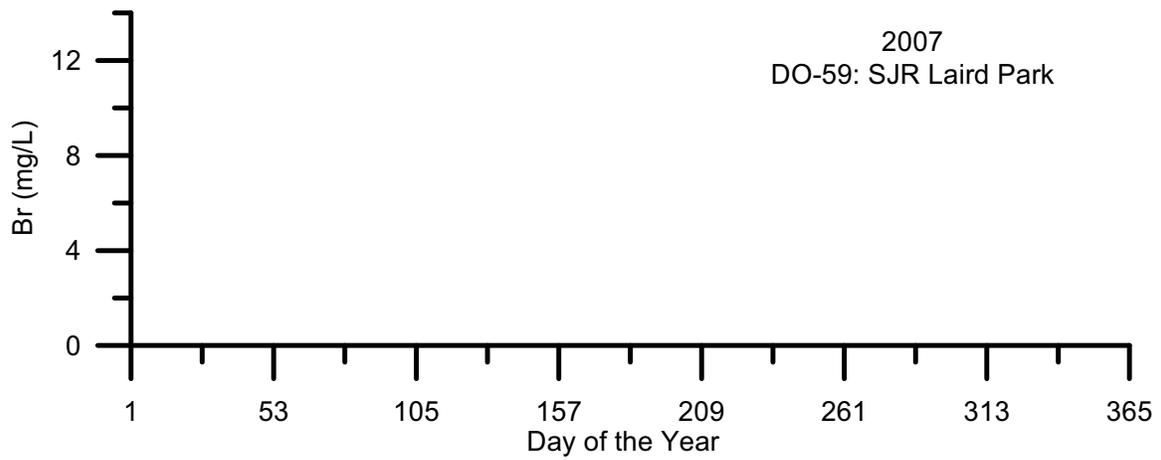
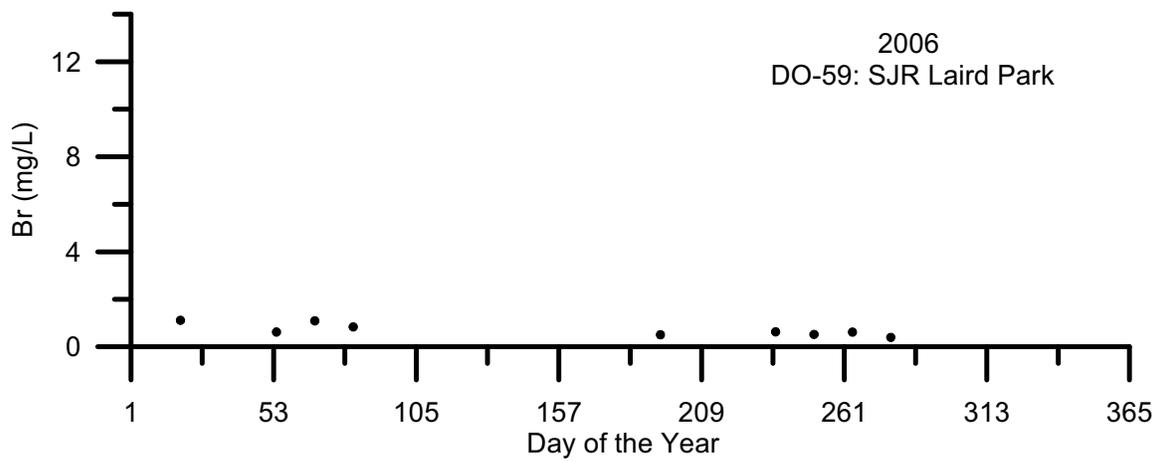
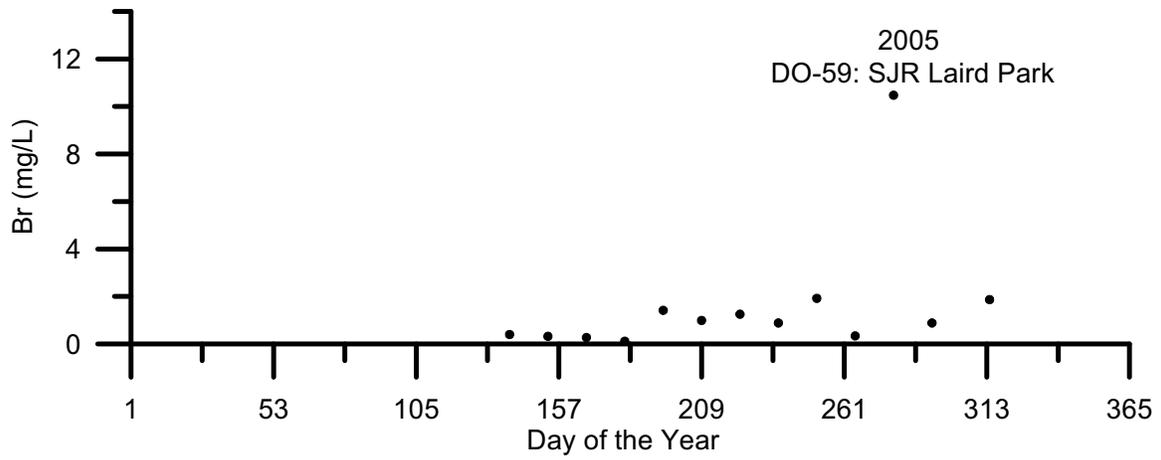


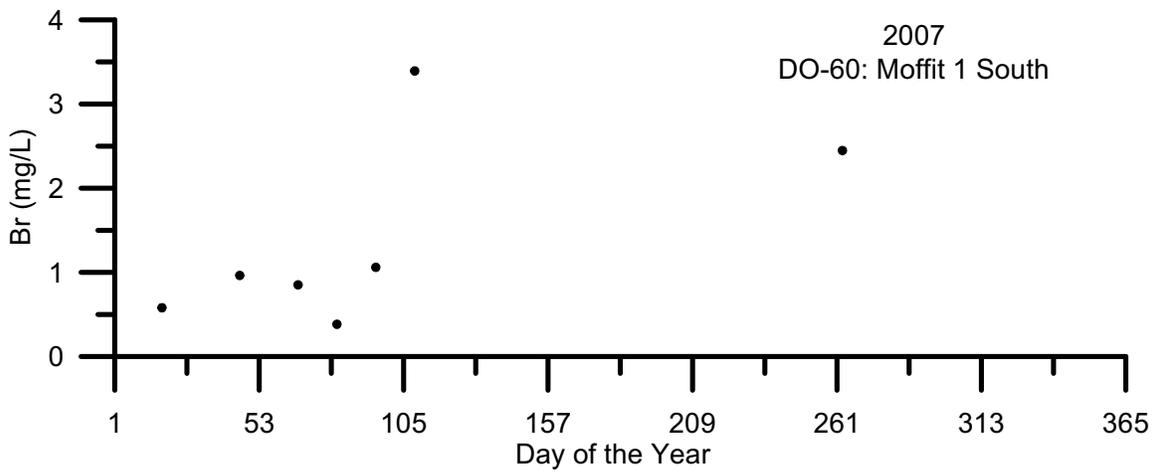
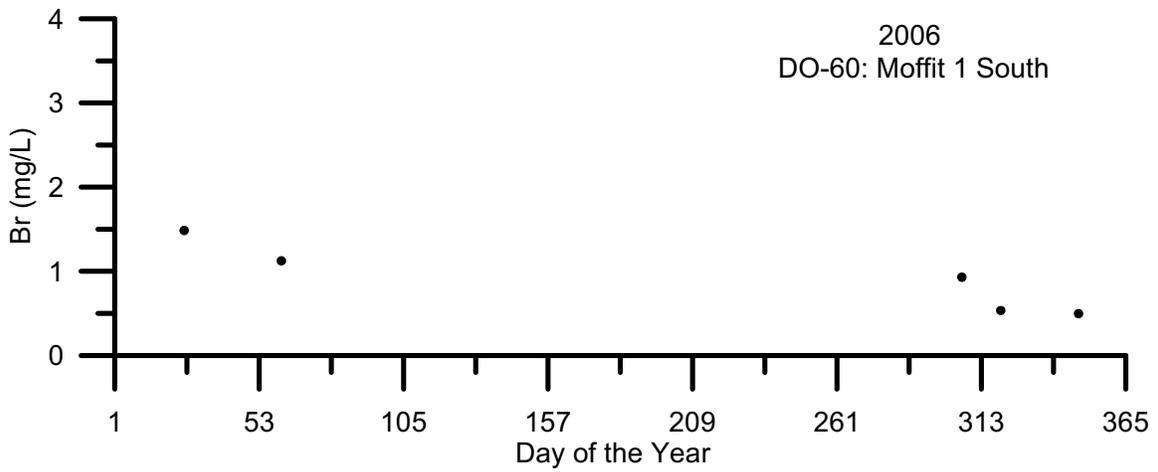
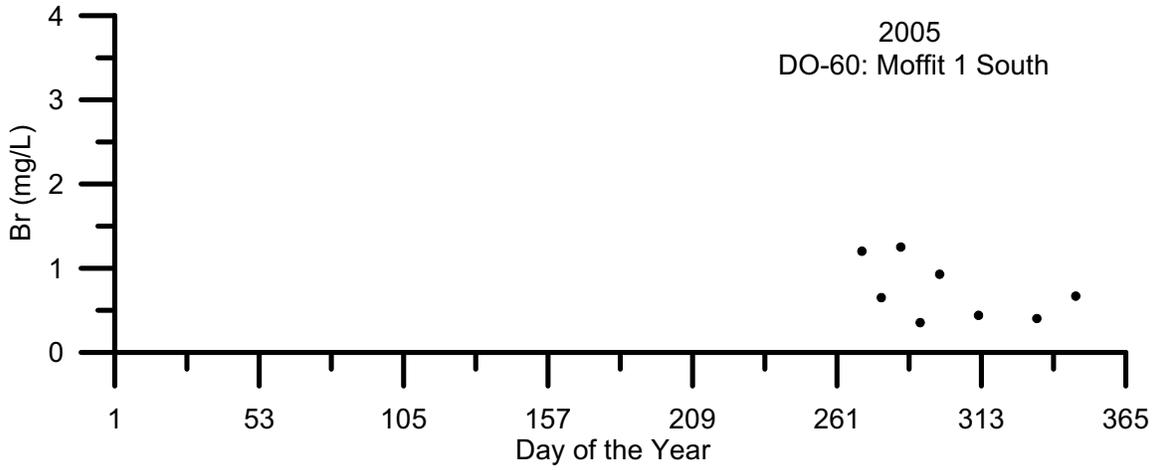


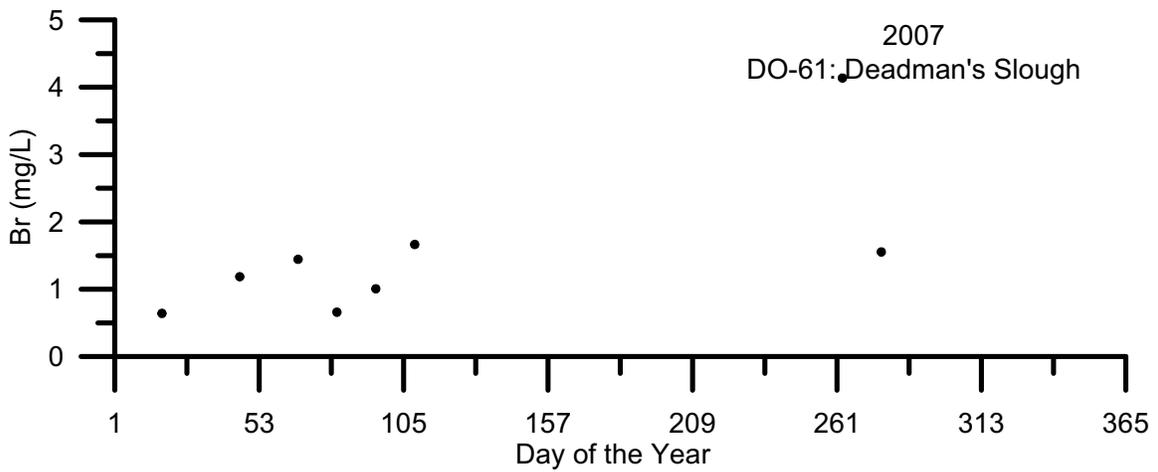
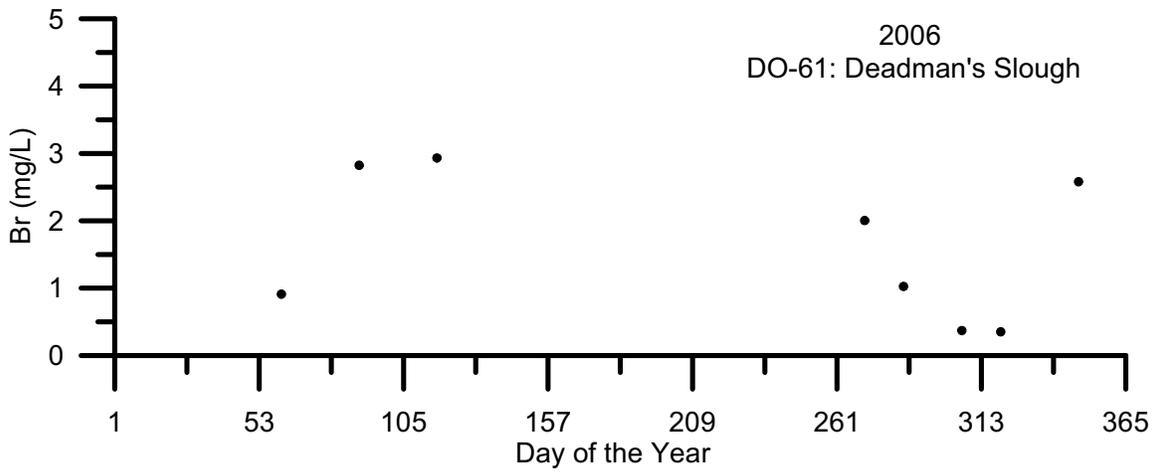
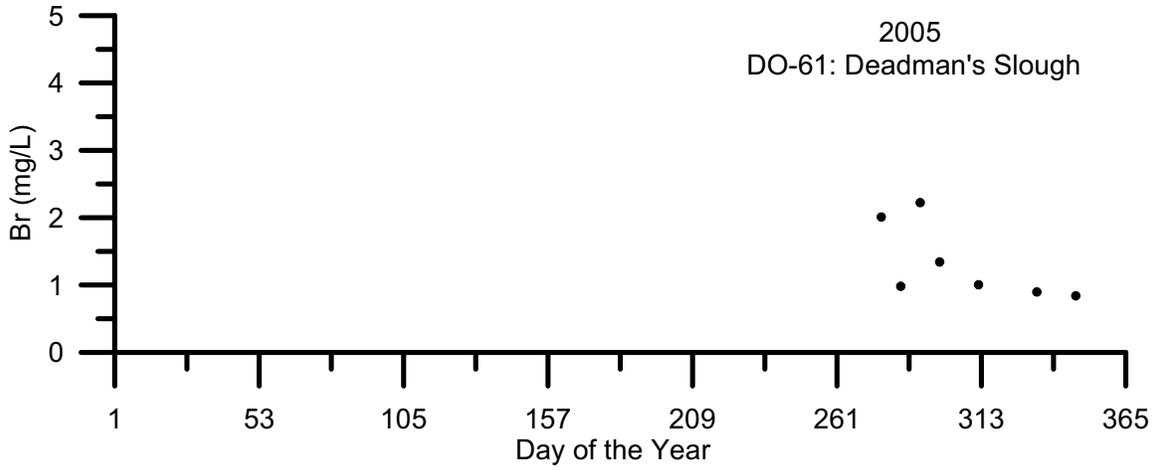


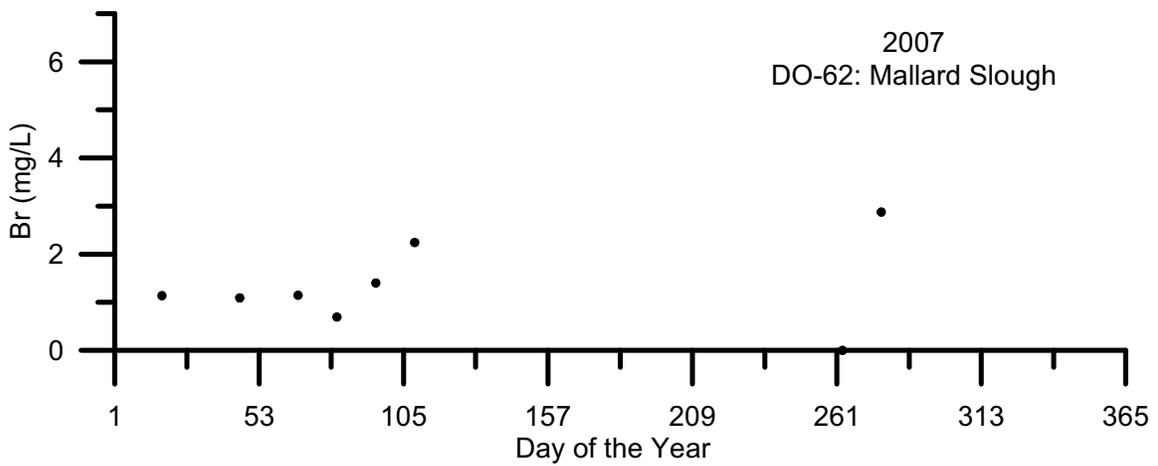
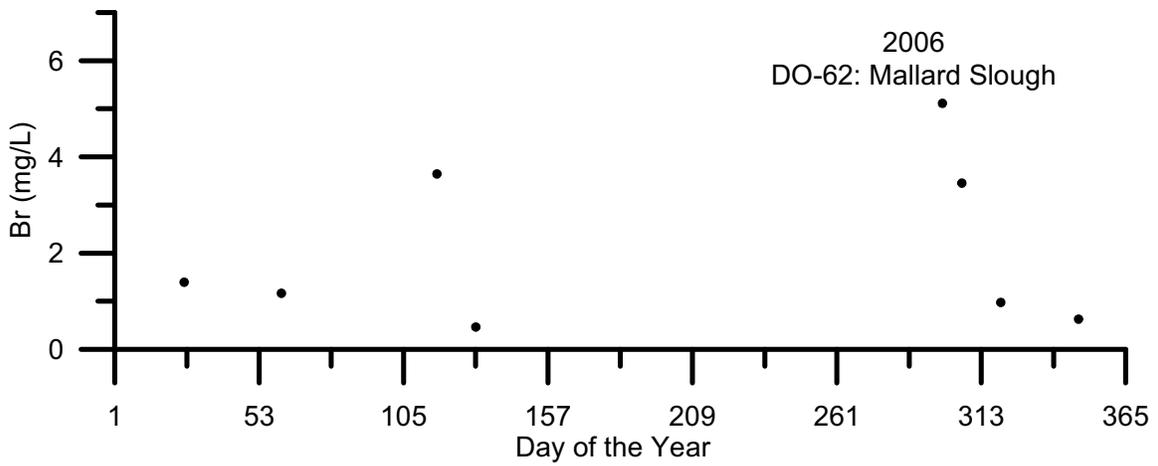
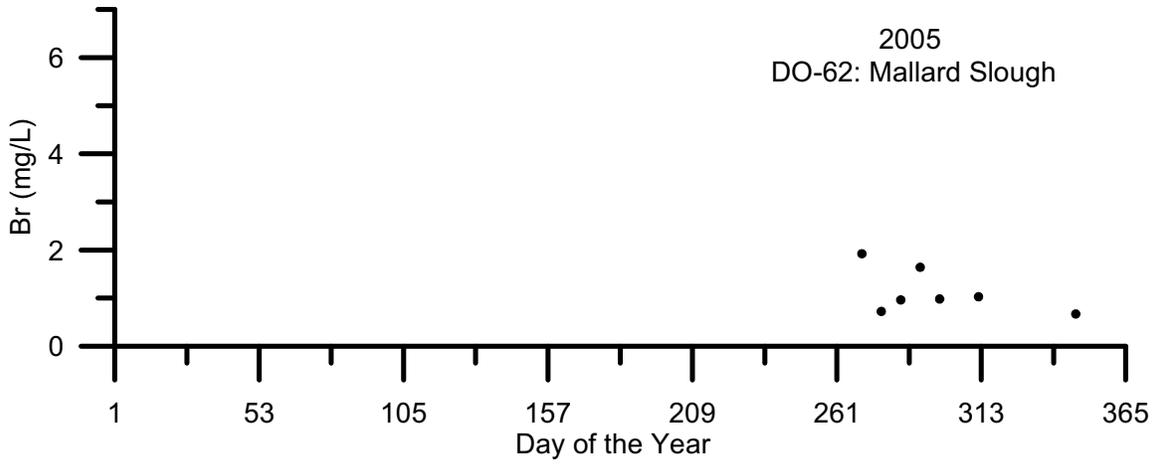


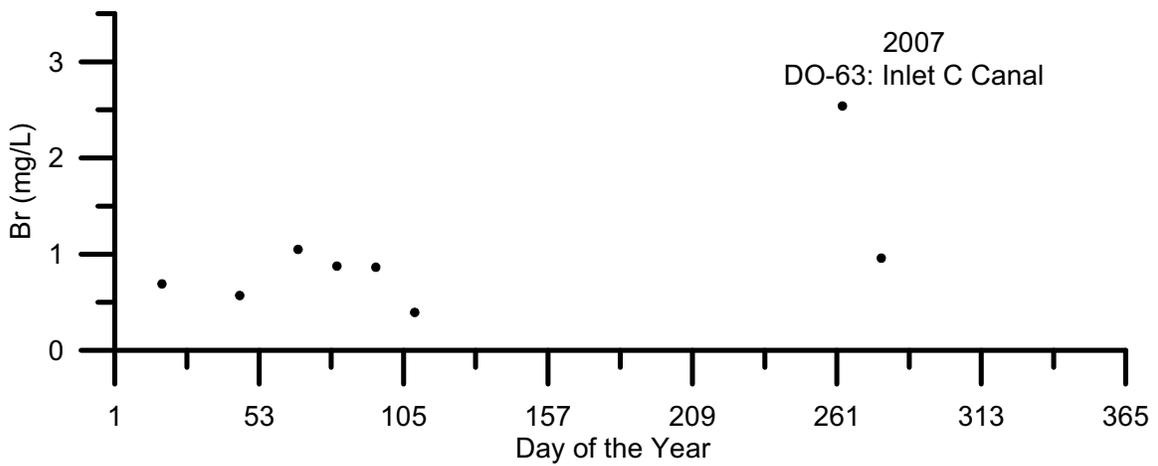
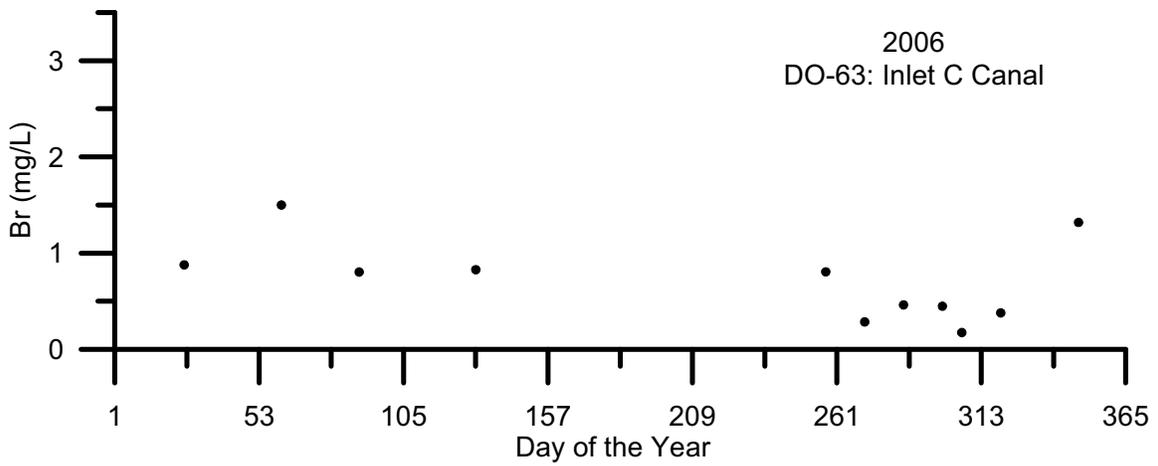
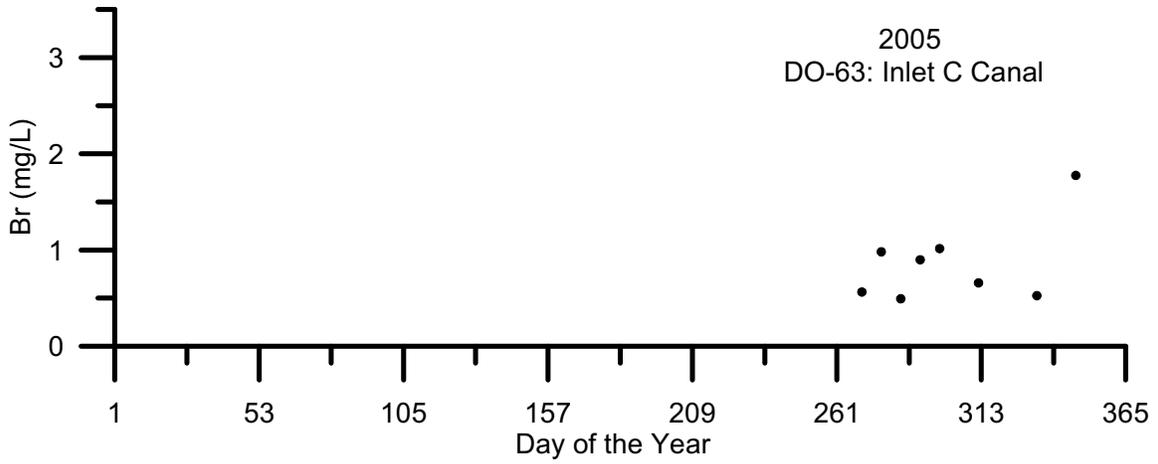


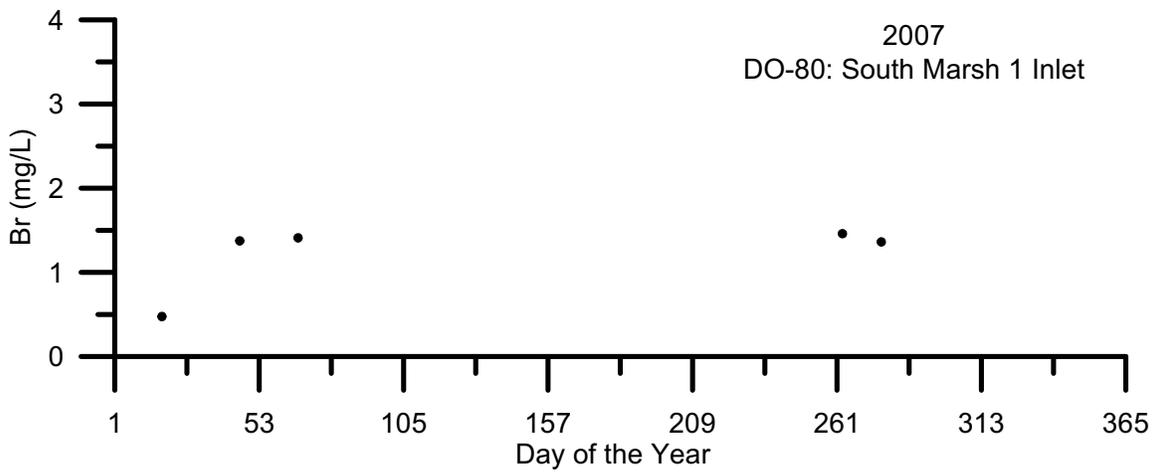
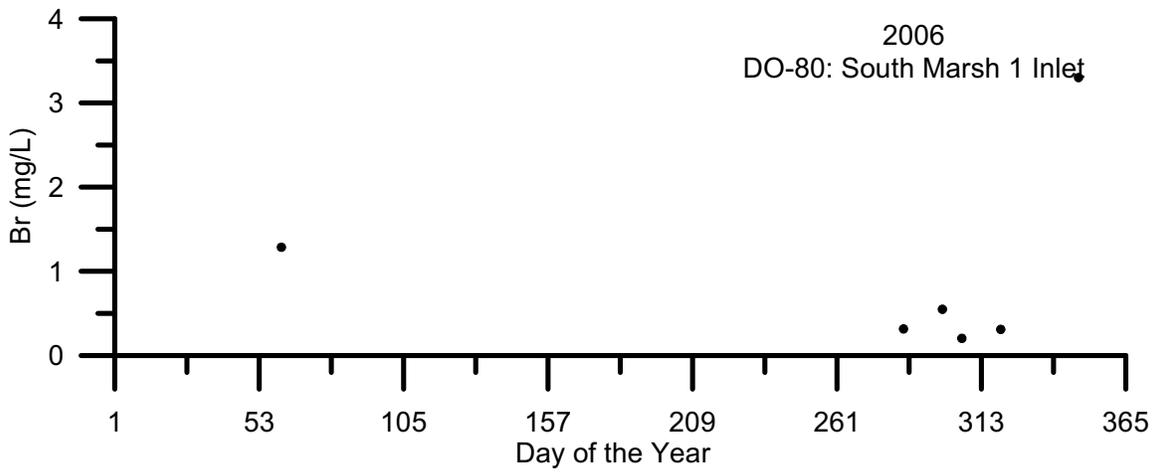
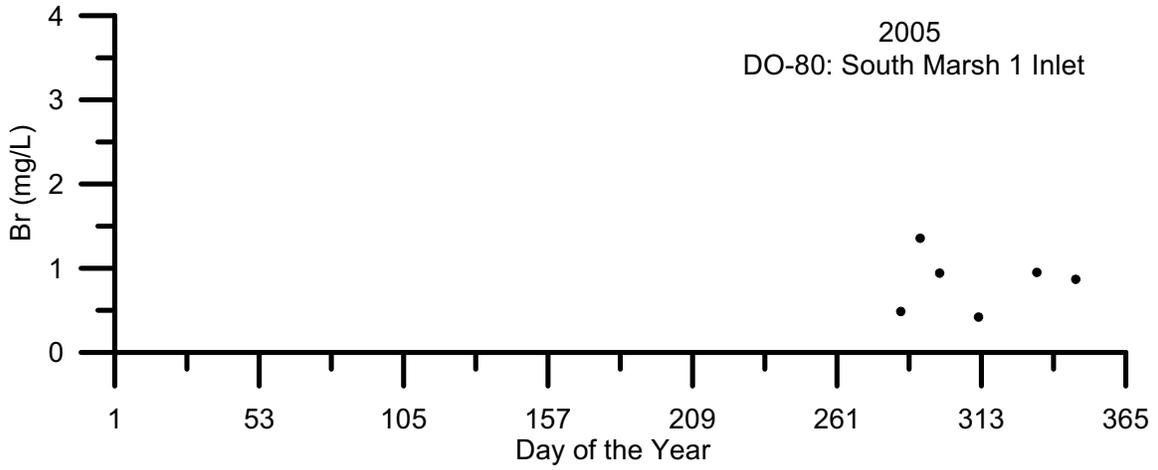


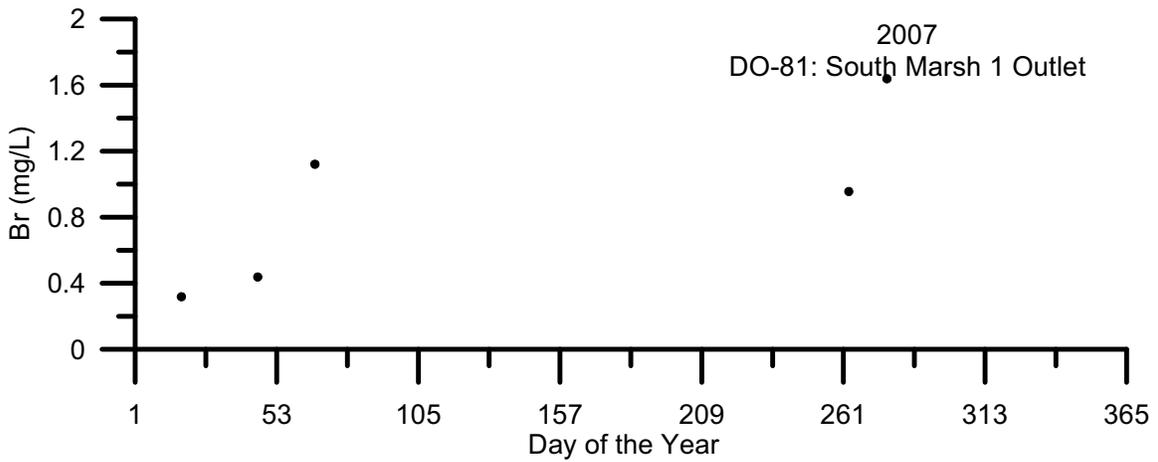
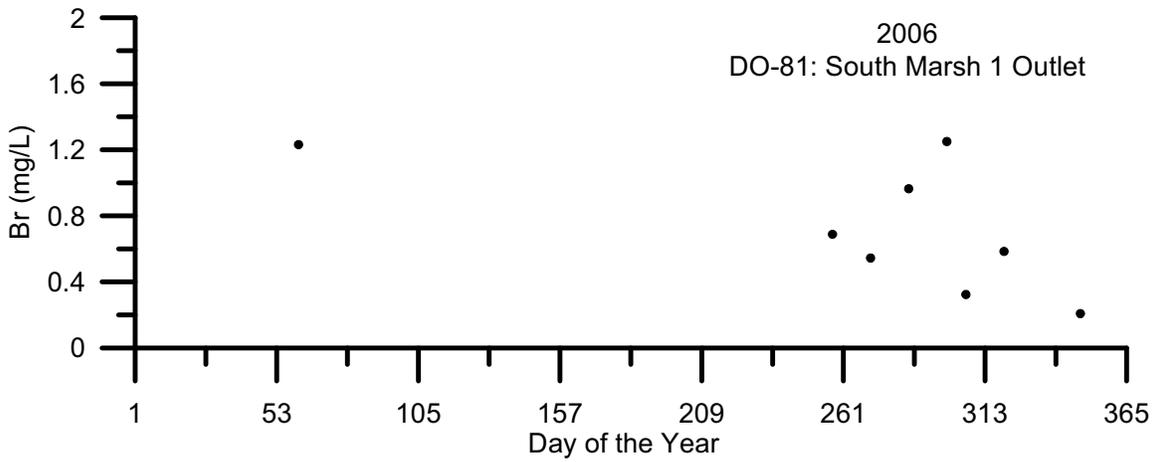
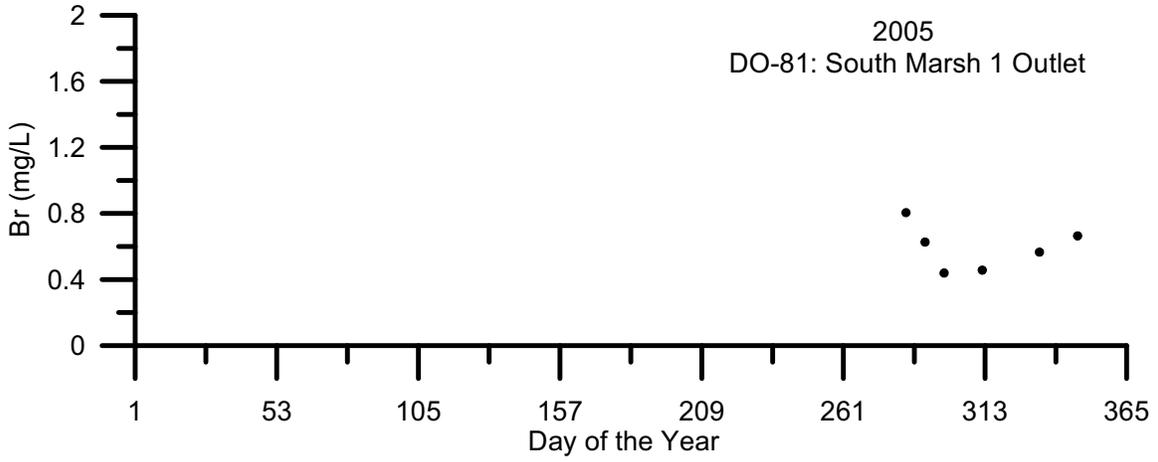


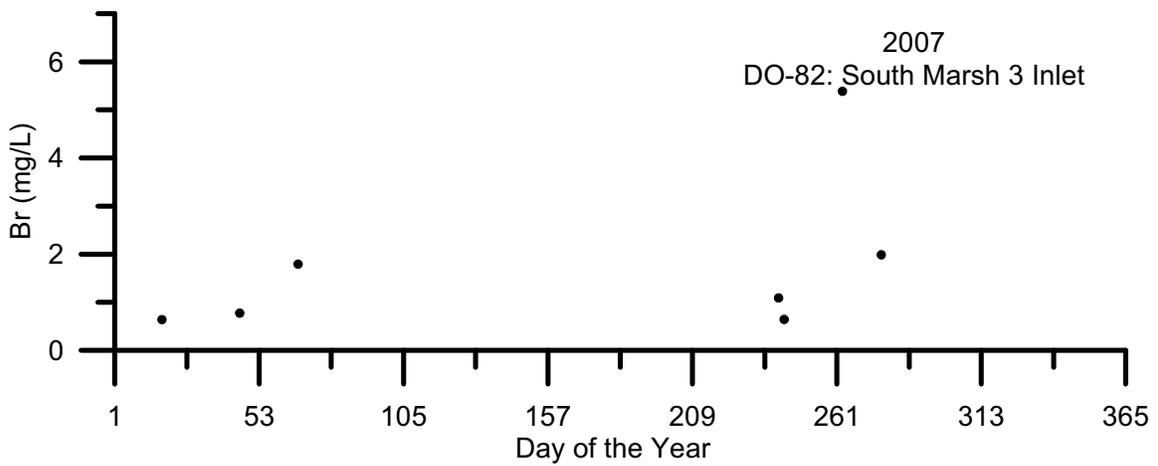
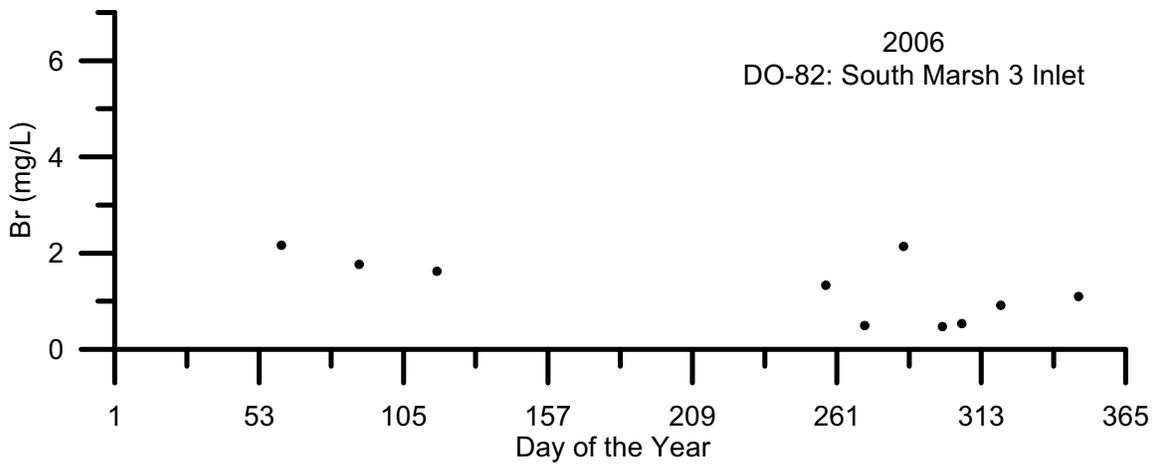
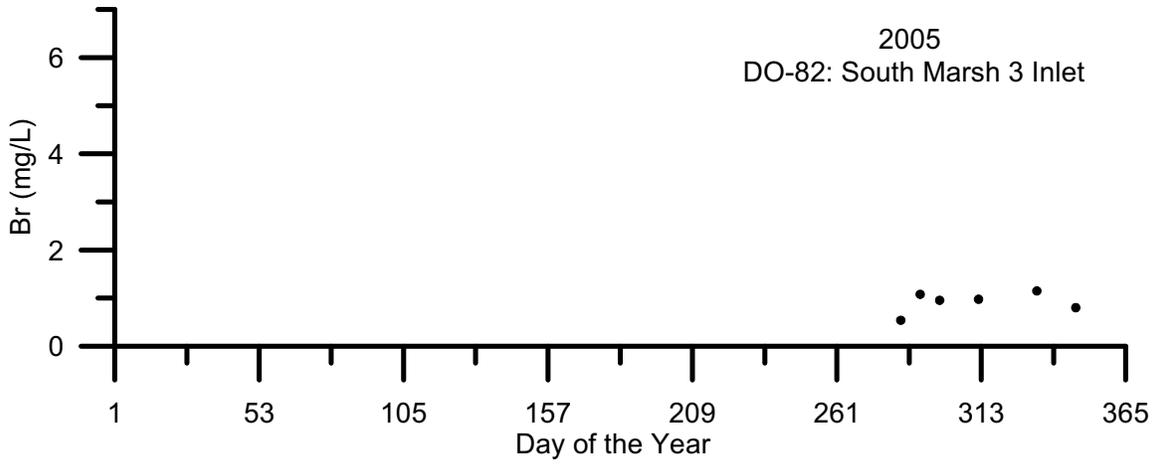


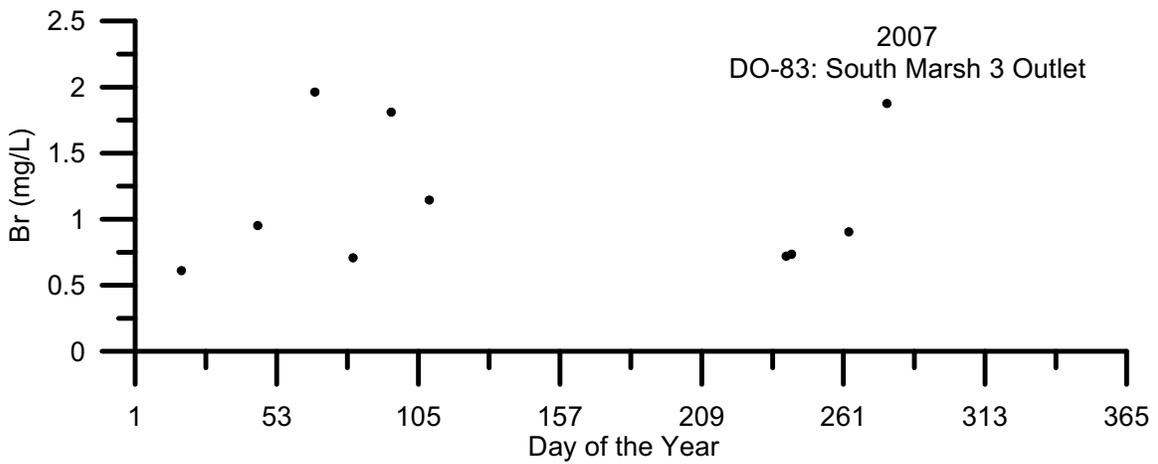
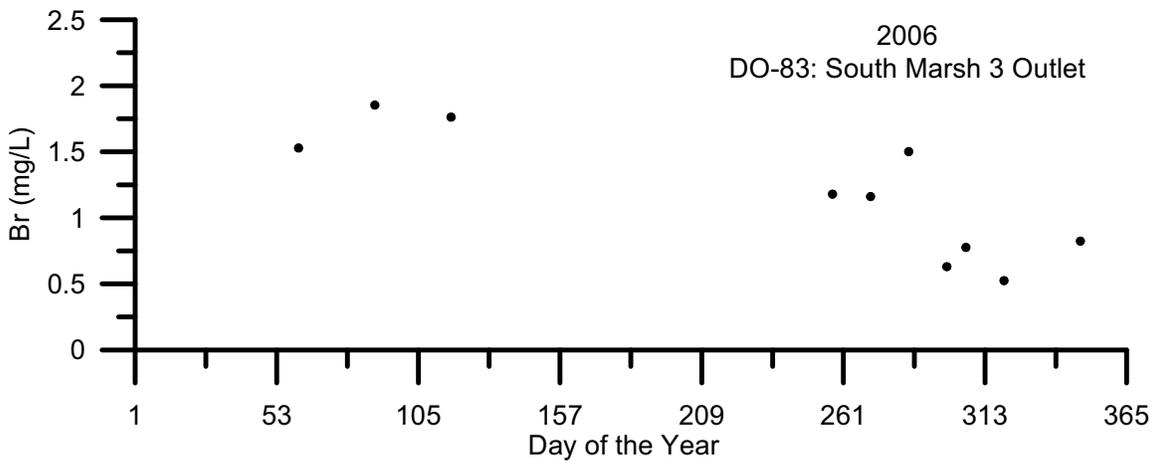
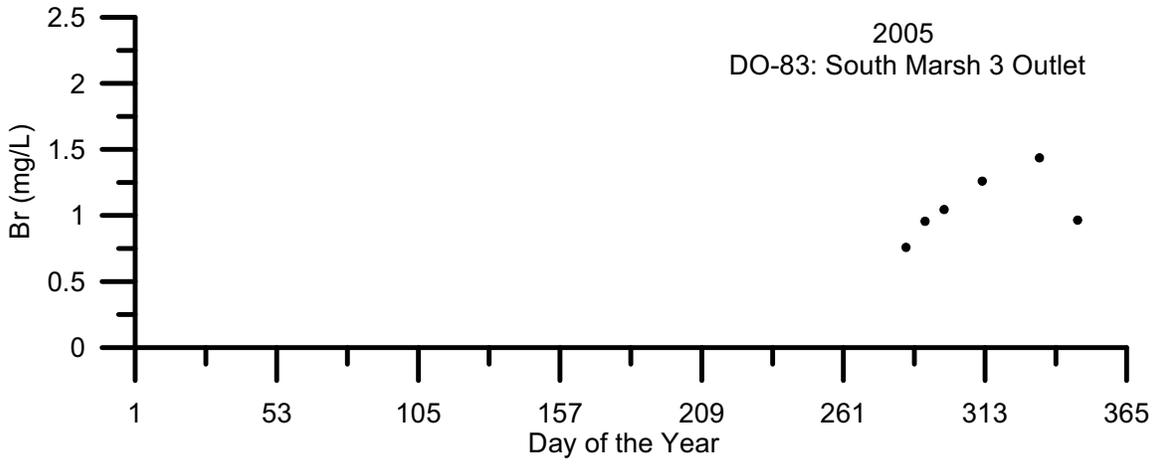


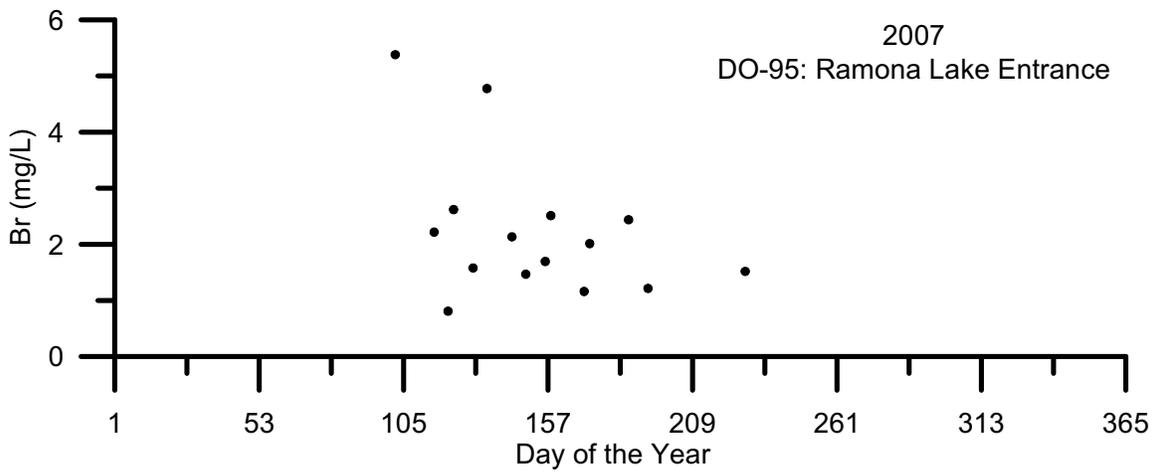
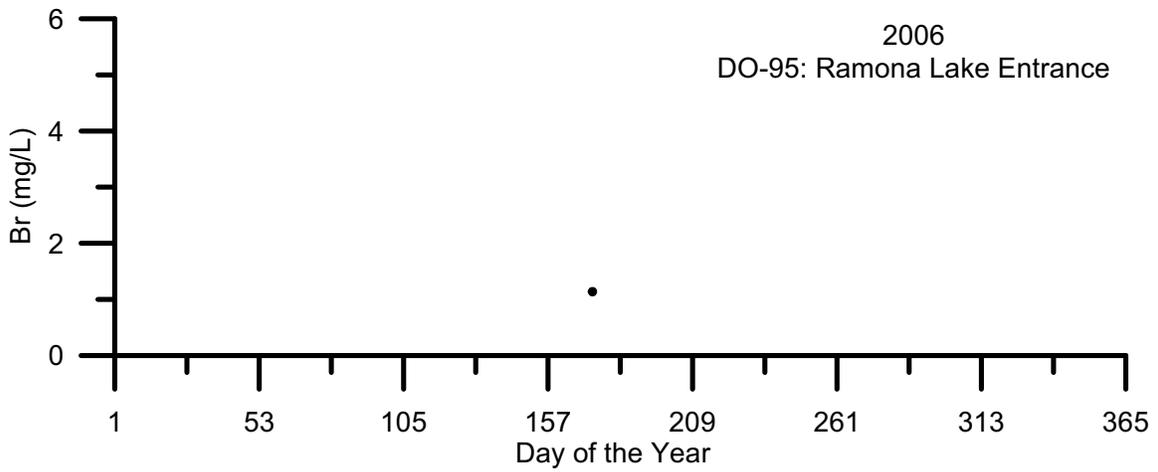
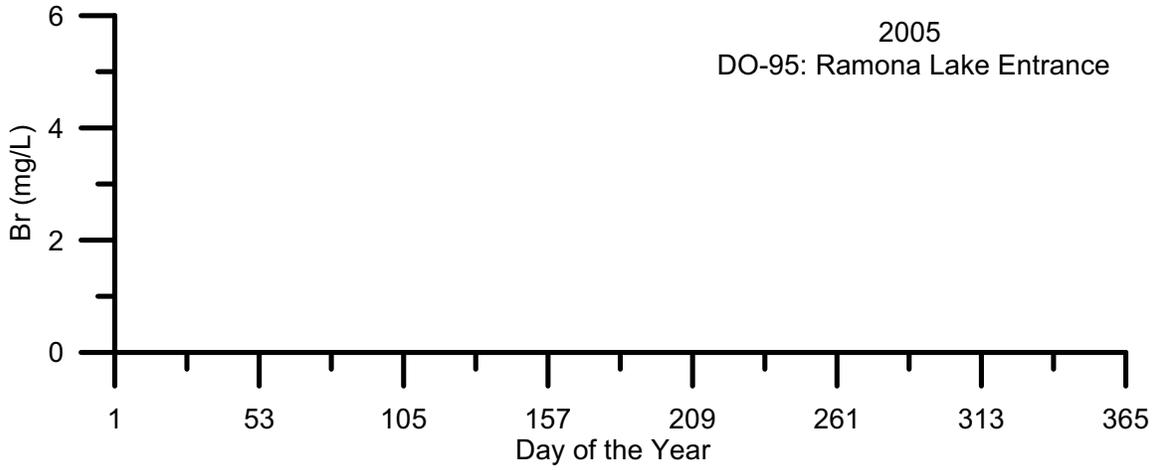














Temporal Plots of 2005-2007 Total Phosphorus Data from the Upstream San Joaquin River

Chelsea Spier¹
Remie Burks¹
Sharon Borglin¹
Randy Dahlgren²
Jeremy Hanlon¹
Justin Graham¹
William Stringfellow¹

February 2008

¹Environmental Engineering Research Program
School of Engineering & Computer Sciences
University of the Pacific
3601 Pacific Avenue, Sears Hall
Stockton, CA 95211

²UC Davis, Davis, CA

Introduction

The San Joaquin River (SJR) supports one of the most productive agricultural regions in the world and its productivity is heavily dependant on irrigated agriculture. A consequence of irrigated agriculture is the production of return flows conveyed down gradient drains that eventually discharge to surface waters. Agricultural drainage may have significant nutrient load and can impact algae growth and general water quality in the SJR. Individual farmers and agricultural organizations, such as drainage authorities, are in need of tools to manage the environmental impacts of agricultural activities (Stringfellow, 2008).

For the years 2005 through 2007, sites throughout the San Joaquin Valley watershed were sampled to assess the overall water quality in the region. One thousand nine hundred and ninety-six (1996) individual surface water samples were collected and analyzed and WQ was assessed at 113 locations in the SJR basin (Borglin et al., 2008). Samples were processed and analyzed by the Environmental Engineering Research Program (EERP) laboratory at the University of the Pacific as well as at the University of California, Davis, Dahlgren Lab. This report includes temporal plots of total phosphorus (Tot-P) data analyzed by the EERP laboratory starting in late 2006.

Methods

Depth integrated field samples were collected during 2005-2007 in the upper San Joaquin River in accordance with EERP Field Standard Operating Procedures Protocol Book (Graham 2008). Water samples were collected in glass 1000 mL bottles (Wheaton Science Products, Millville, NJ), 1000 mL HDPE Trace-Clean narrow mouth plastic bottles (VWR International), 250 mL HDPE Trace-Clean wide mouth plastic bottles (VWR International) as well as 40 mL trace clean vials with PTFE septa (IChem, Rockwood, TN). Bottles were labeled with the appropriate sample number, site name and sampling date. All bottles were rinsed with sample water prior to sample collection. Some sites required a bucket to collect sample water because of accessibility from a high bridge or platform. For these sites, the bucket was pre-rinsed with sample water and sample bottles were filled using a rinsed funnel. Care was taken to distribute water simultaneously to all sample bottles (rather than sequentially). Samples were immediately stored at 4°C after sampling (cooler temperature was recorded in the lab upon delivery) and transported to the EERP lab on the day of sampling.

Within 24 hours of collection samples were aliquotted and stored at 4°C until Tot-P analysis could be completed.

Tot-P was determined on 5.0 mL of unfiltered sample by persulfate digestion and colorimetric determination by the ascorbic acid method adapted from SM 4500-P B, E (APHA, 2005). To digest samples equal parts of sample and digestion reagent (10 g potassium persulfate, 6 g boric acid, and 3 g NaOH in 1000mL Millipore water) were combined and samples were autoclaved in a Tuttnauer Brinkman autoclave (Westbury, NY). After digestion and sample cooling, the total phosphorus concentrations were determined spectrophotometrically using Hach PhosVer3 packets (Loveland, CO) on a PerkinElmer Lambda 35 UV/VIS Spectrometer (Shelton, CT). The limit of detection for

this analysis was 0.06 mg/L Tot-P.

Results/Discussion

With each set of Tot-P field samples analyzed in the EERP laboratory, quality assurance samples including a lab duplicate, field duplicate, matrix spike, matrix spike duplicate, calibration check standards, laboratory control standard, trip blank, and lab blanks were also analyzed. Between 2005 and 2007, 100% of all quality assurance samples were within a passing range (Borglin et al, 2008). Proficiency check samples, standards with unknown concentration to the laboratory analyst, were run approximately twice a year. Two proficiency check samples were analyzed for Tot-P in the EERP laboratory during 2006 and 2007, and both of these samples were found to be within the acceptable range. Samples were measured ranging from 0.0-5.06 mg/L Tot-P. The average concentration of Tot-P in samples collected was 0.32 mg/L Tot-P. Tot-P was also analyzed at UC Davis on all of the same water samples and has a high correlation to values measured by EERP. When all data points measured by the two labs are compared they have $r^2=0.951$ (Figure 2). EERP measured 89.1% as much Tot-P as UCD (Figure 2).

One problem that occurred with this analysis is dissolved ortho-phosphate-phosphorus ($PO_4\text{-P}$) values were sometimes found to be slightly higher than the measured total phosphorus (Total P) values. When this happened one or both of these analyses were re-run. This problem occurred most often in samples with high $PO_4\text{-P}$ and Tot-P which required dilutions. Because of this problem all $PO_4\text{-P}$ and TP dilutions are now run in triplicate to reduce the potential for dilution errors. These temporal plots (Figures 3-104) as well as plotting EERP data against UCD's data (Figure 2) created an easy visual way to find outliers and double check data entry for possible mistakes. For the purpose of these plots any data points that were slightly negative were changed to 0.

References

- American Public Health Association (APHA). 2005. Standard Methods for the Examination of Water and Wastewater, 21st Edition. American Public Health Association, Washington, DC.
- Borglin, S., W. Stringfellow, J. Hanlon. 2005. Standard Operating Procedures for the Up-Stream Dissolved Oxygen TMDL Project. LBNL/Pub-937.
- Borglin, S., Burks, R., Hanlon, J., Graham, J., Spier, C., Stringfellow, W., and Dahlgren, R., (2008) Methods overview, quality assurance, and quality control, University of the Pacific, Stockton, CA
- Borglin, S.E., Burks, R.D., Hanlon, J.S., Stringfellow, W.T. (2008) EERP Lab Protocol Book, University of the Pacific, Stockton, CA.
- Graham, J., Hanlon, J.S., Stringfellow, W.T., (2008) EERP Field Protocol Book, University of the Pacific, Stockton, CA.
- Stringfellow, W.T., et al., (2008) Evaluation of Vegetated Ditches, Ponds, and Wetlands as BMPs for Mitigating the Water Quality Impact of Irrigated Agriculture in the San Joaquin Valley, University of the Pacific, Stockton, CA

YSI Environmental Operations Manual (2005) 6-Series Environmental Monitoring Systems, Yellow Springs, OH.

Table 1: EERP Sampling Site List

DO Number	Site Name	Type
4	SJR at Mossdale	Core sites
5	SJR at Vernalis-McCune Station (River Club)	Core sites, BMP
6	SJR at Maze	Core sites, BMP
7	SJR at Patterson	Core sites, BMP
8	SJR at Crows Landing	Core sites, BMP
10	SJR at Lander Avenue	Core sites
12	Stanislaus River at Caswell Park	Core sites
14	Tuolumne River at Shiloh Bridge	Core sites
16	Merced River at River Road	Core sites
18	Mud Slough near Gustine	Core sites, Wetland
19	Salt Slough at Lander Avenue	Core sites, Wetland
20	Los Banos Creek Flow Station	Core sites, Wetland
21	Orestimba Creek at River Road	Core sites, BMP
23	Modesto ID Lateral 5 to Tuolumne	Core sites
25	Modesto ID Main Drain to Stan. R. via Miller Lake	Core sites
28	Turlock ID Westport Drain Flow station	Core sites
29	Turlock ID Harding Drain	Core sites
30	Turlock ID Lateral 6 & 7 at Levee	Core sites
33	Hospital Creek	Intermittent, BMP
34	Ingram Creek	Core sites, BMP
36	Del Puerto Creek Flow Station	Core sites, BMP
44	San Luis Drain End	Core sites
53	Salt Slough at Wolfsen Road	Wetland
57	Ramona Lake Drain	Core sites, BMP
59	SJR Laird Park	Core sites
60	Moffit 1 South	Wetland
61	Deadman's Slough	Wetland
62	Mallard Slough	Wetland
63	Inlet C Canal	Wetland
80	South Marsh-1-Inlet	Wetland
81	South Marsh-1-Outlet	Wetland
82	South Marsh-3-Inlet	Wetland
83	South Marsh-3-Outlet	Wetland
95	Ramona drain at Ramona Lake	BMP, Intermittent

Figure 1: EERP Sampling Site Map of SJR Watershed and Tributaries

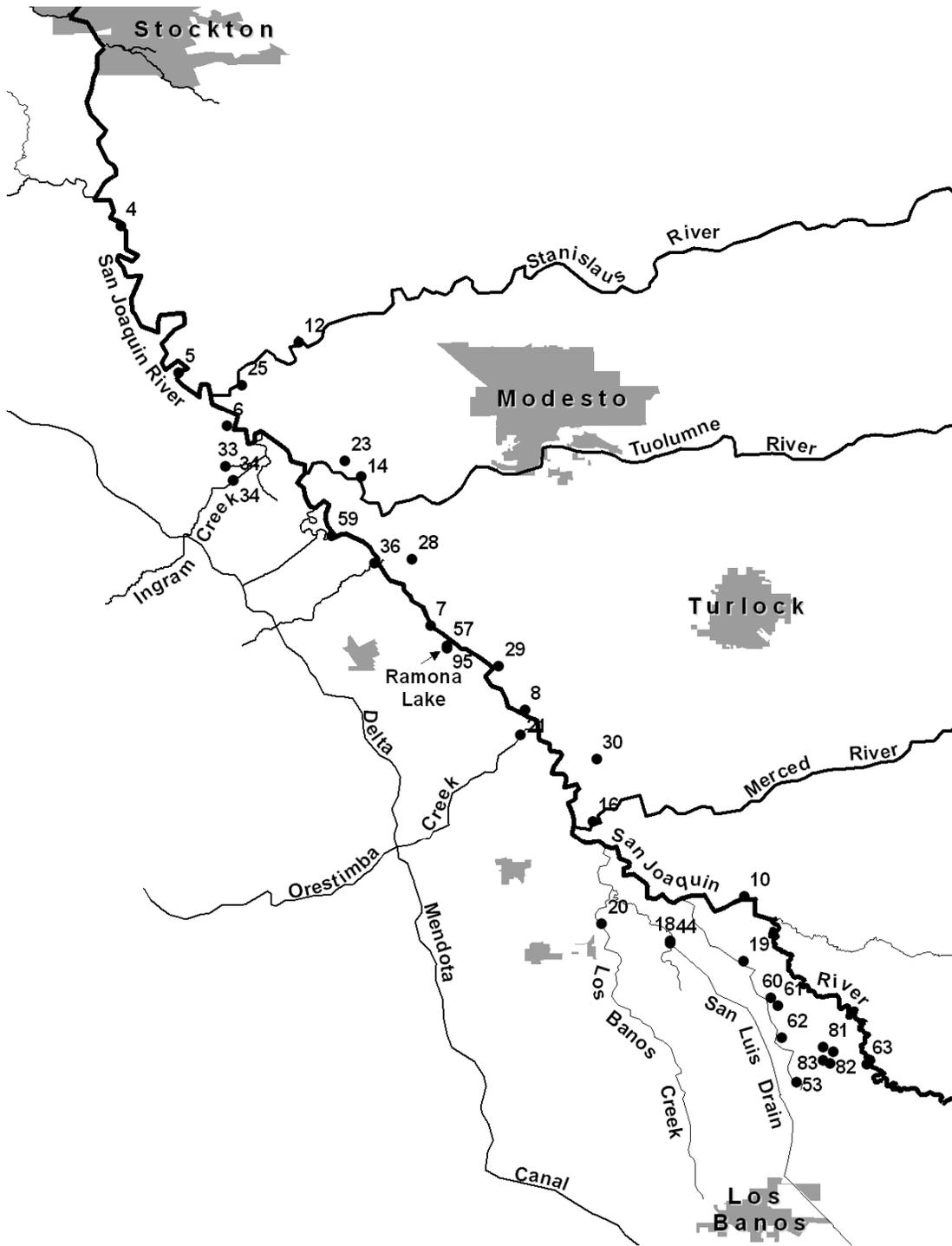
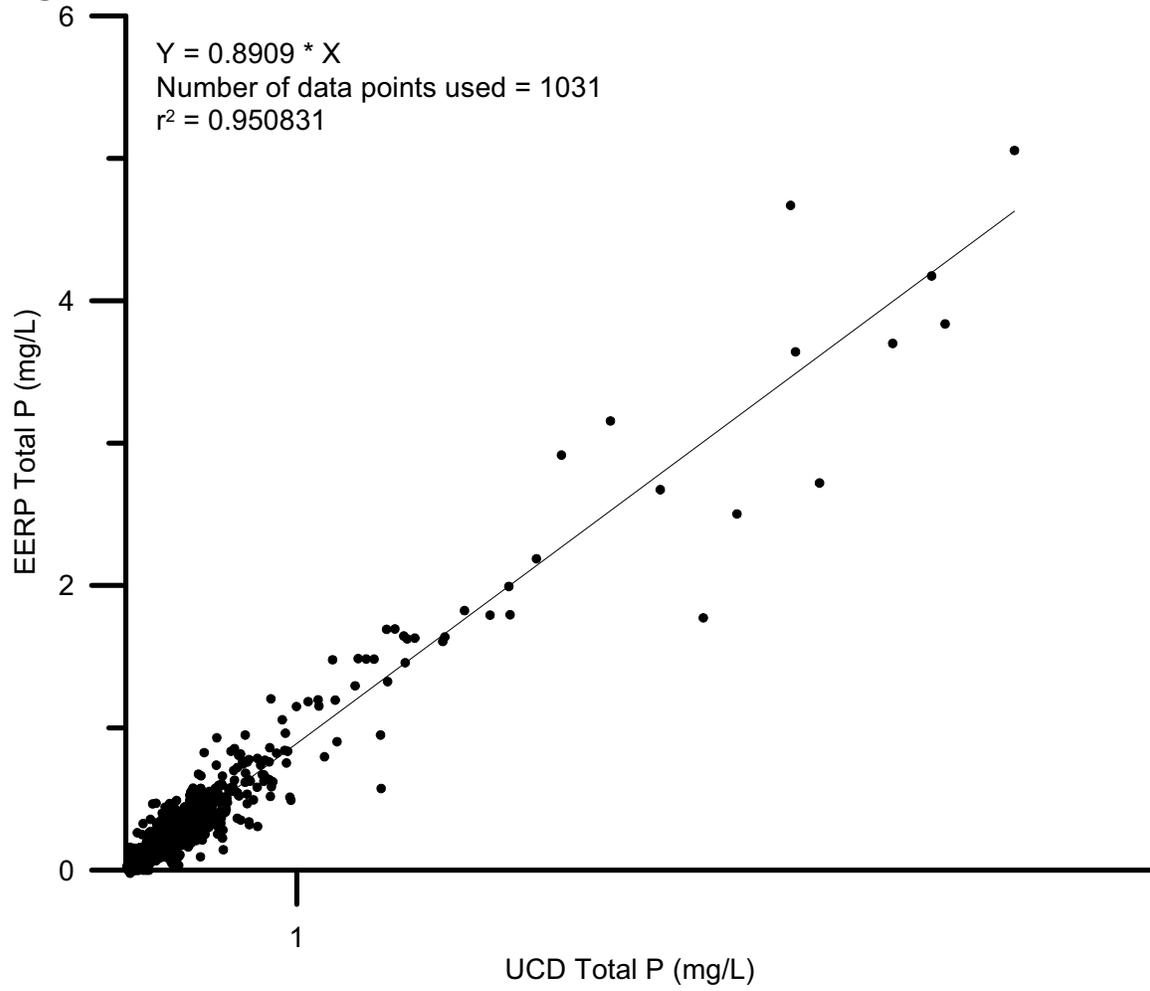
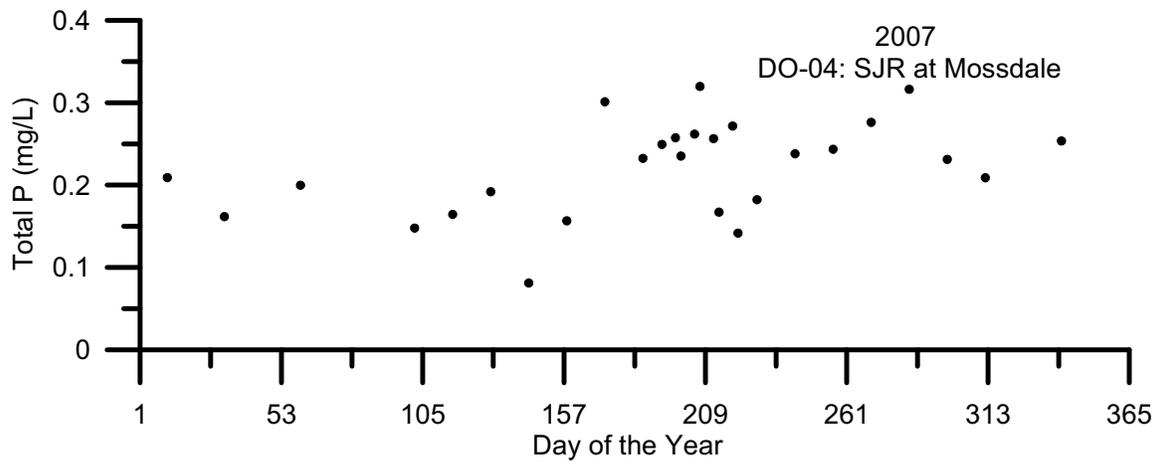
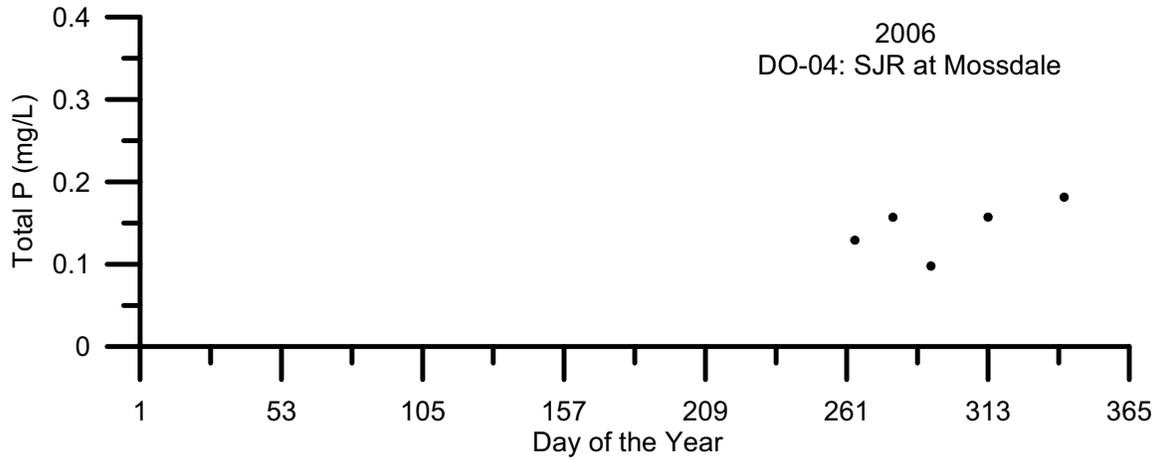
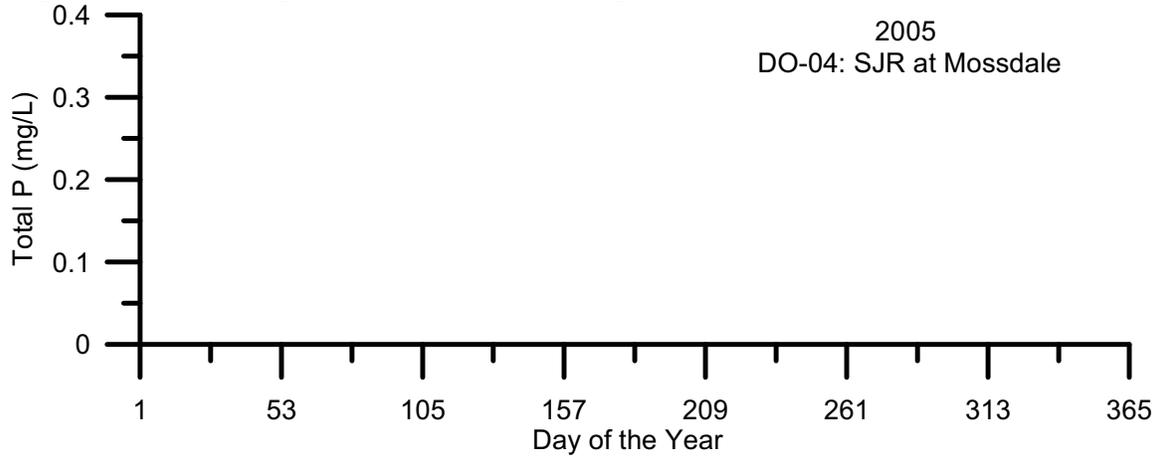
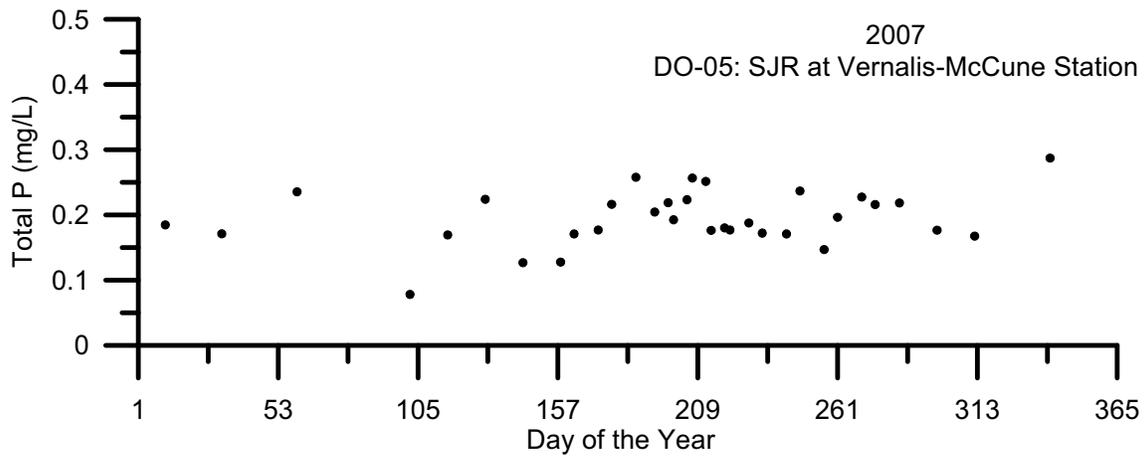
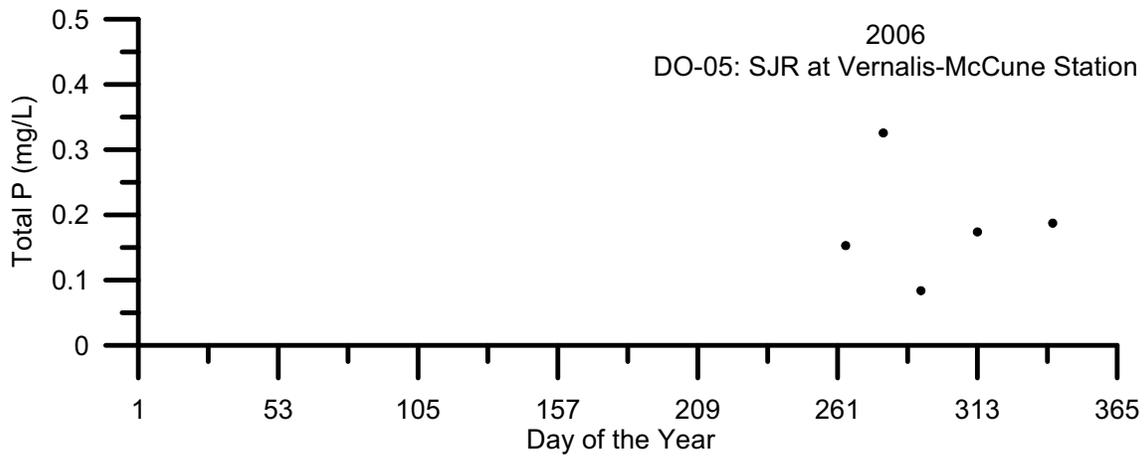
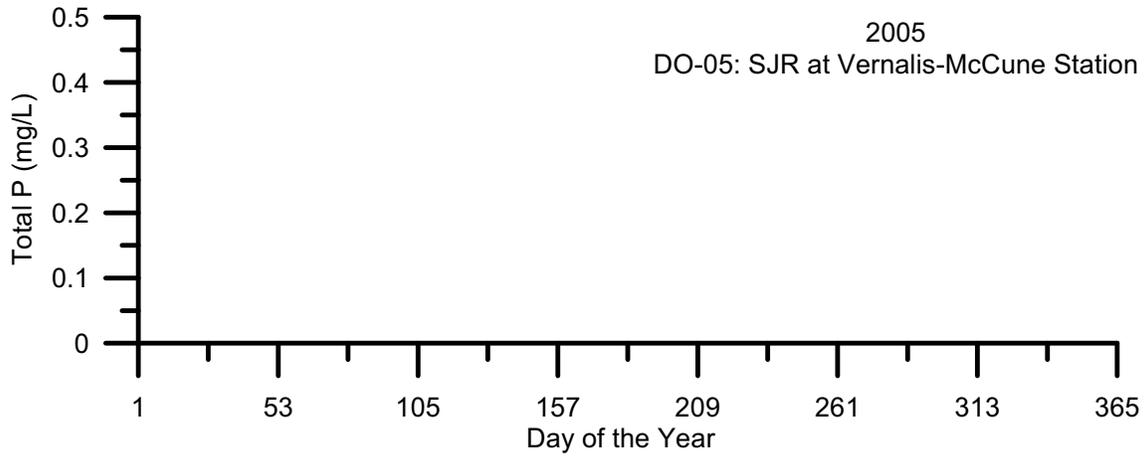


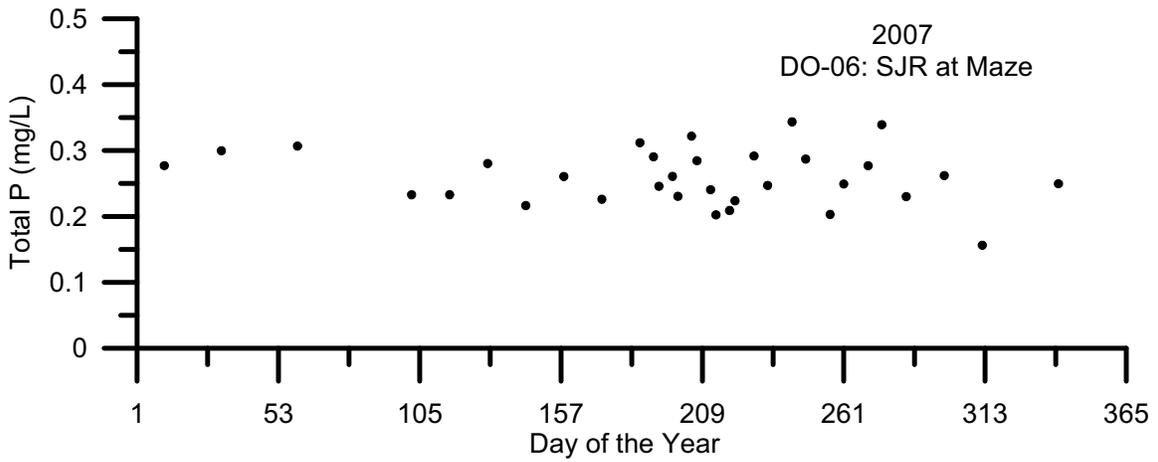
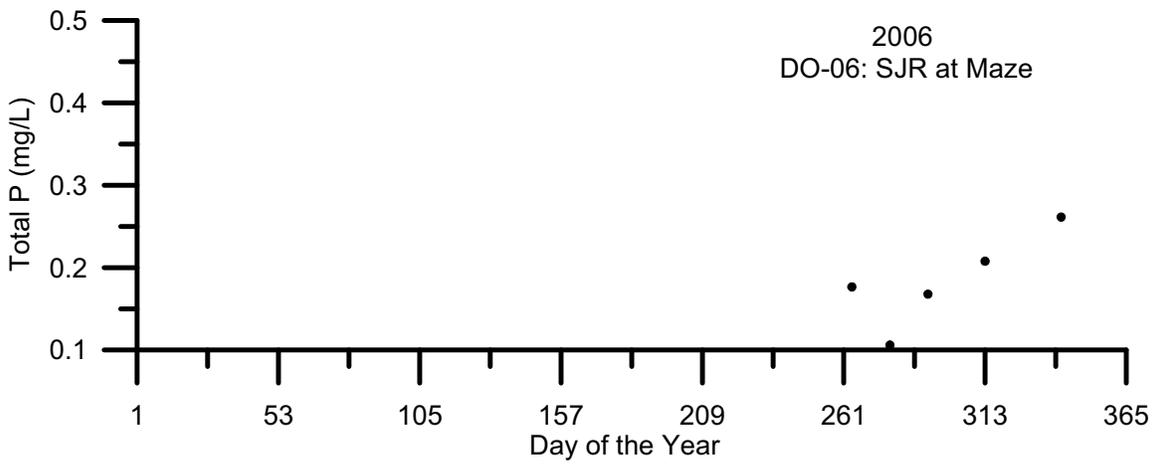
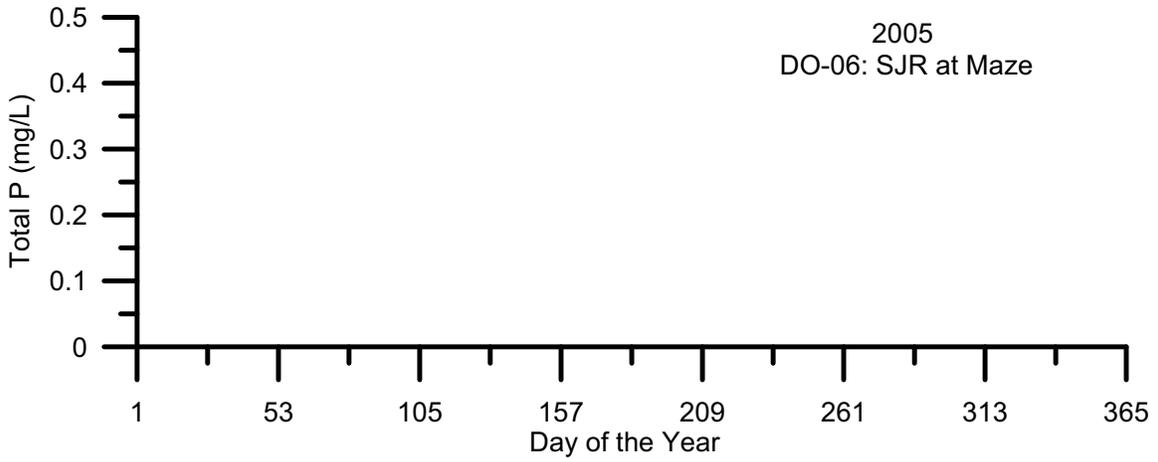
Figure 2: EERP vs UCD Total P Data 2006-2007

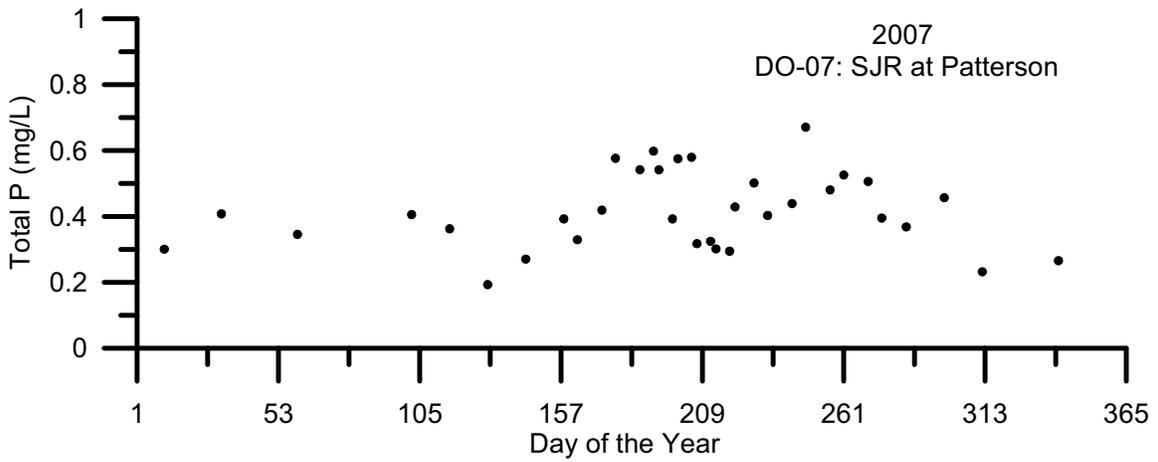
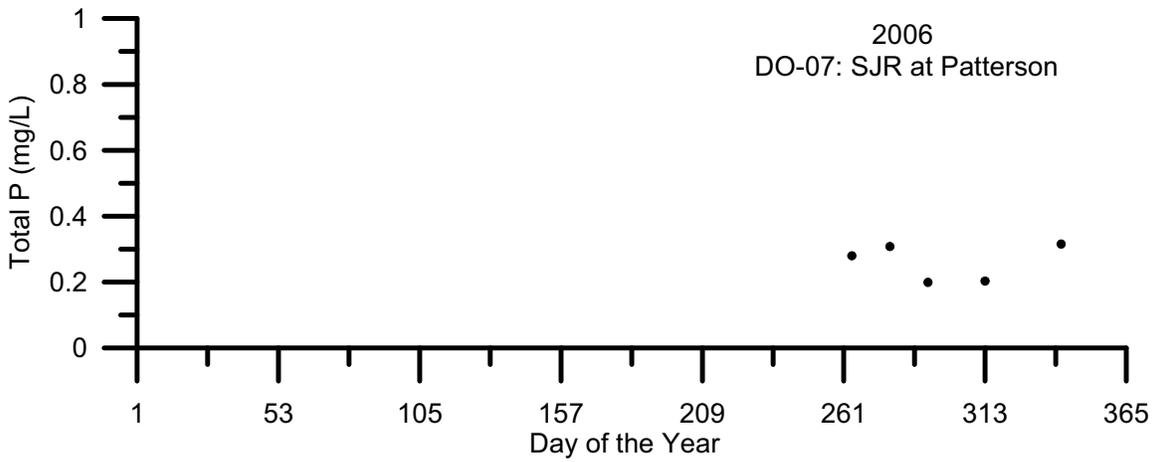
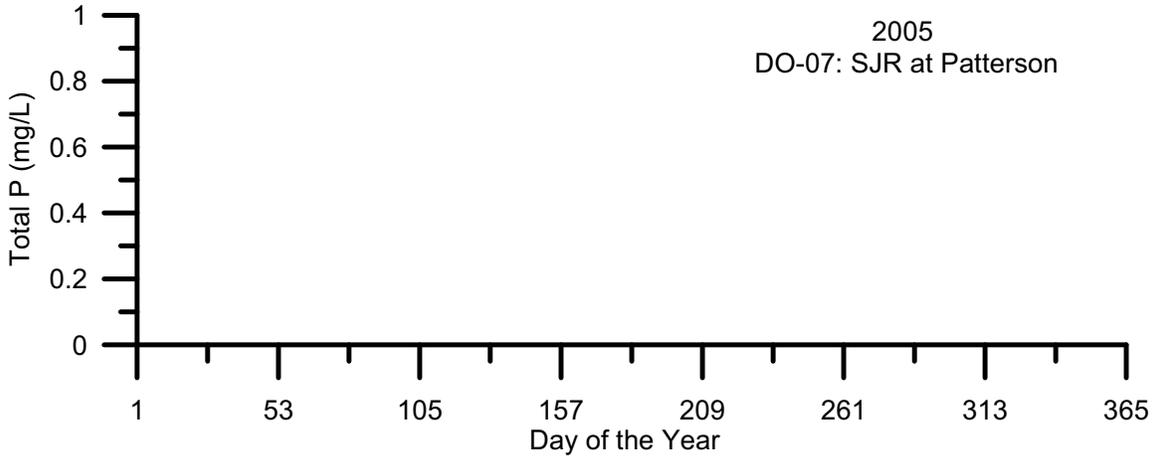


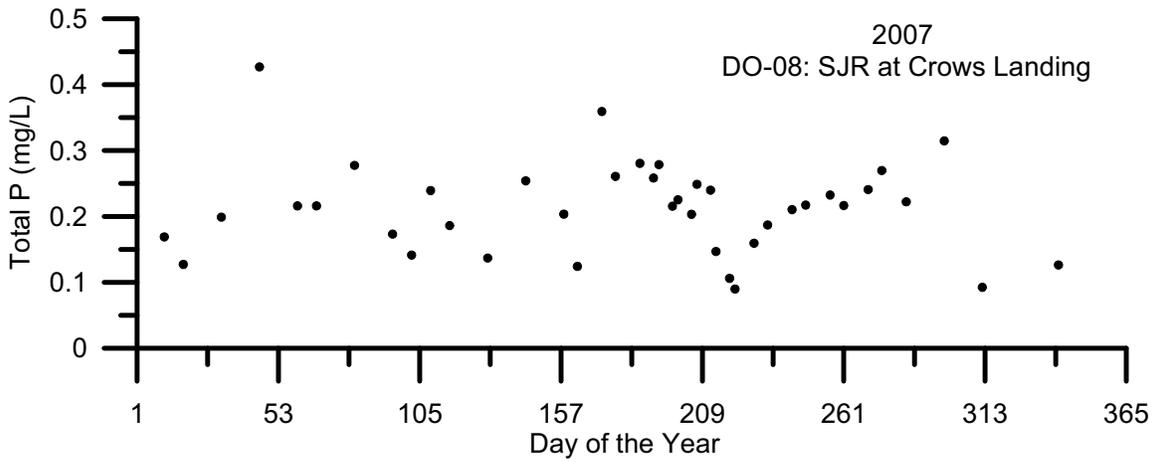
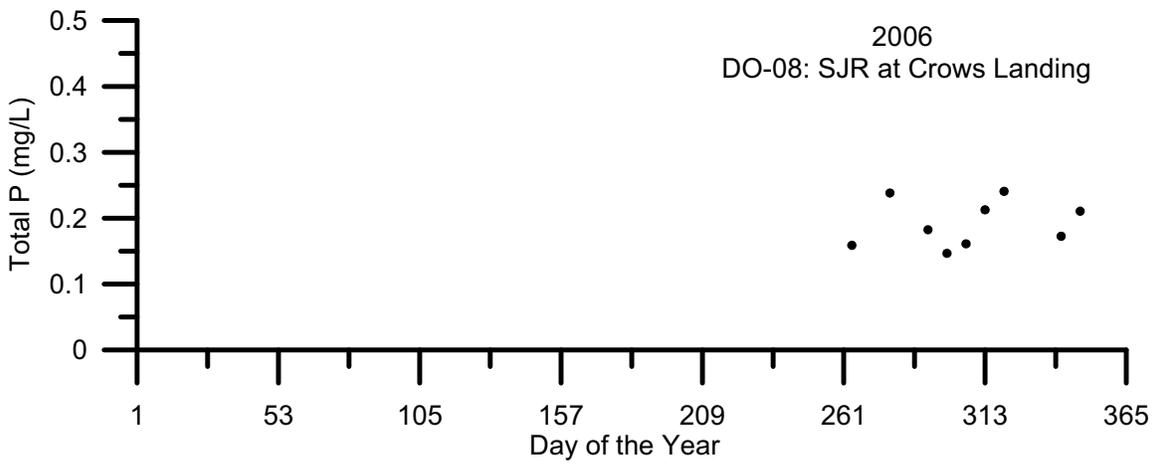
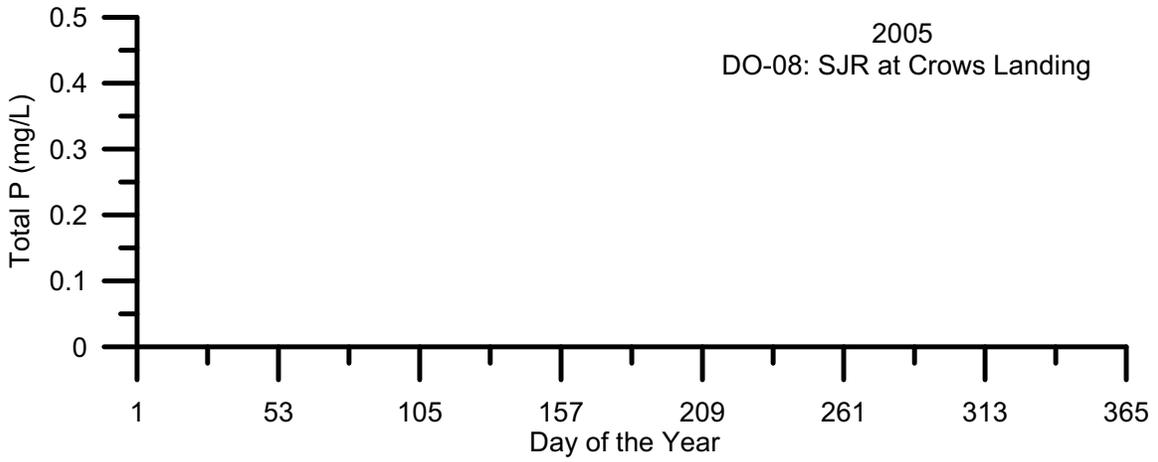
Figures 3 -104: Temporal Plots of Total Phosphorus By Site ID

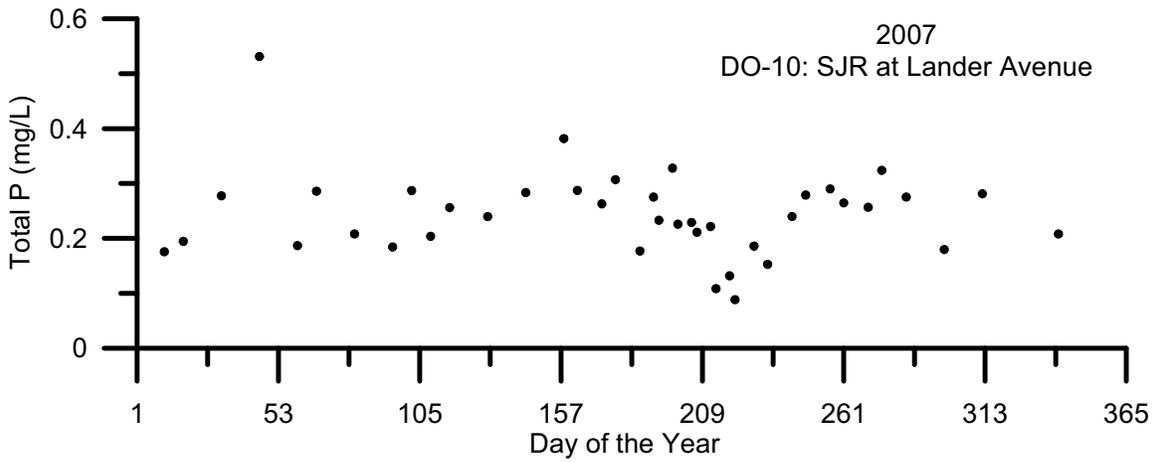
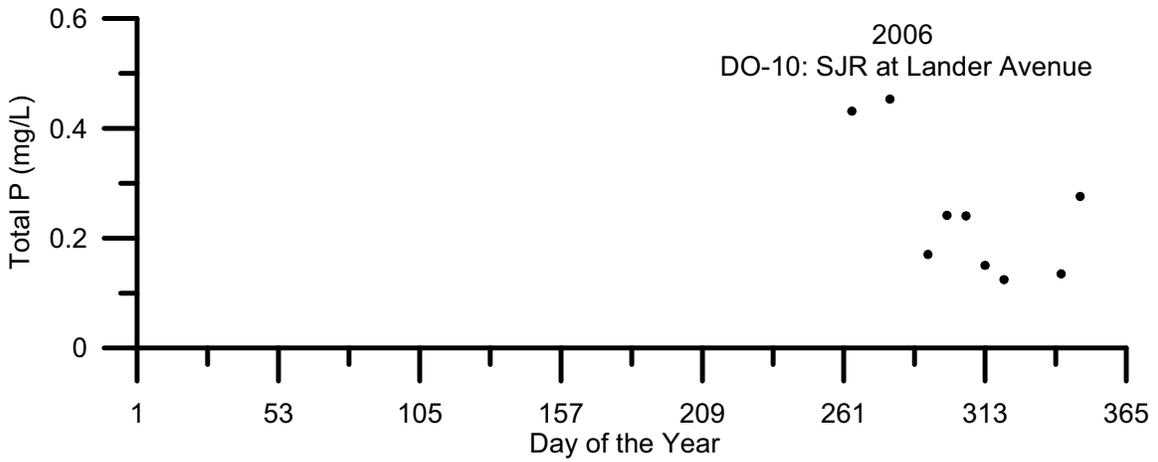
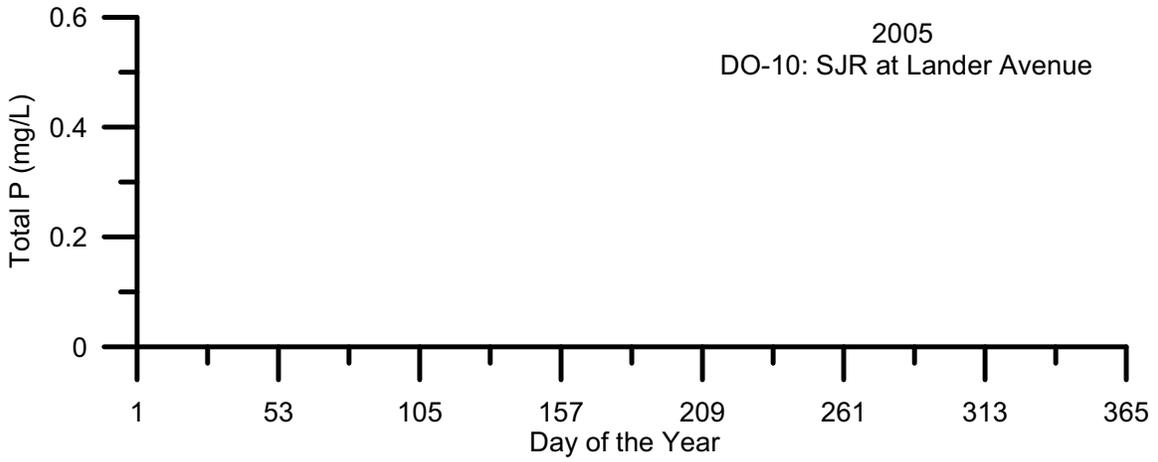


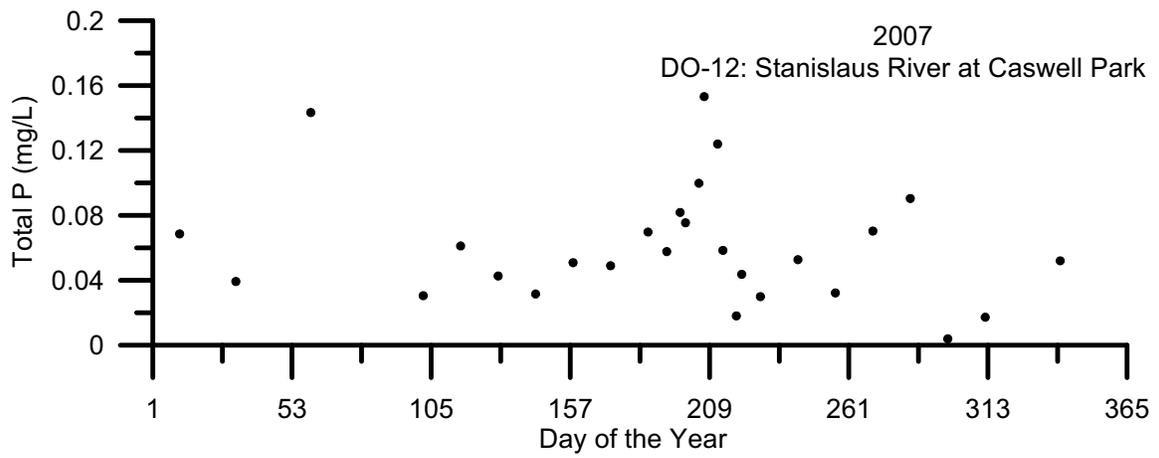
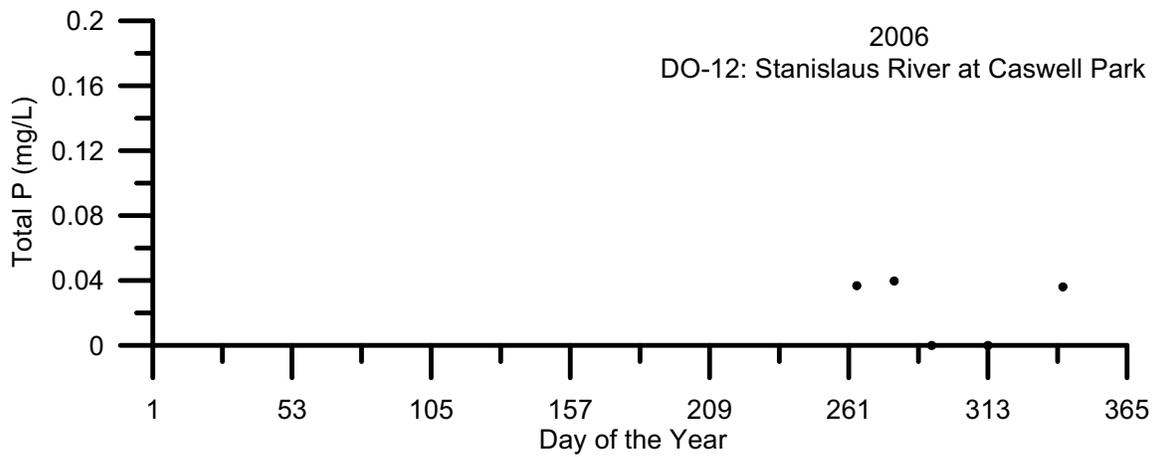
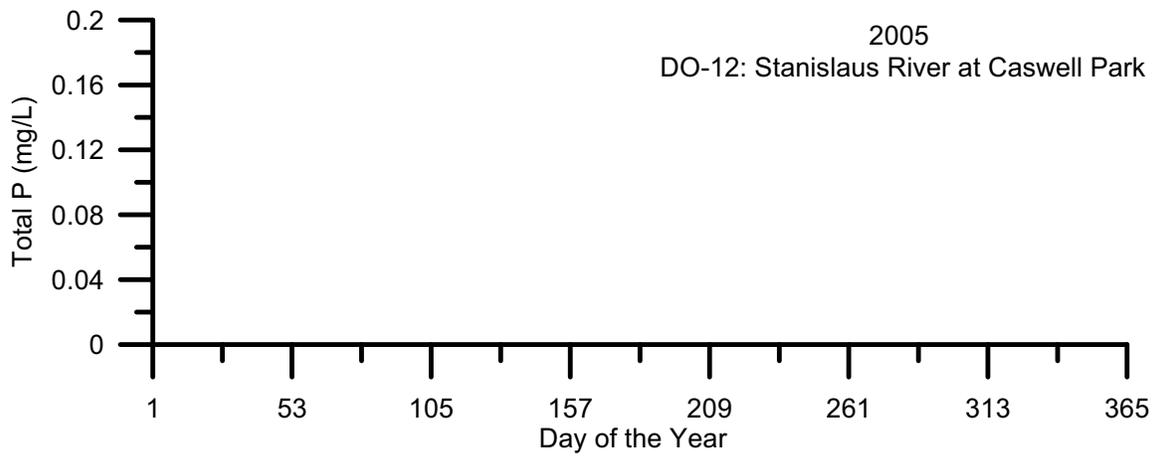


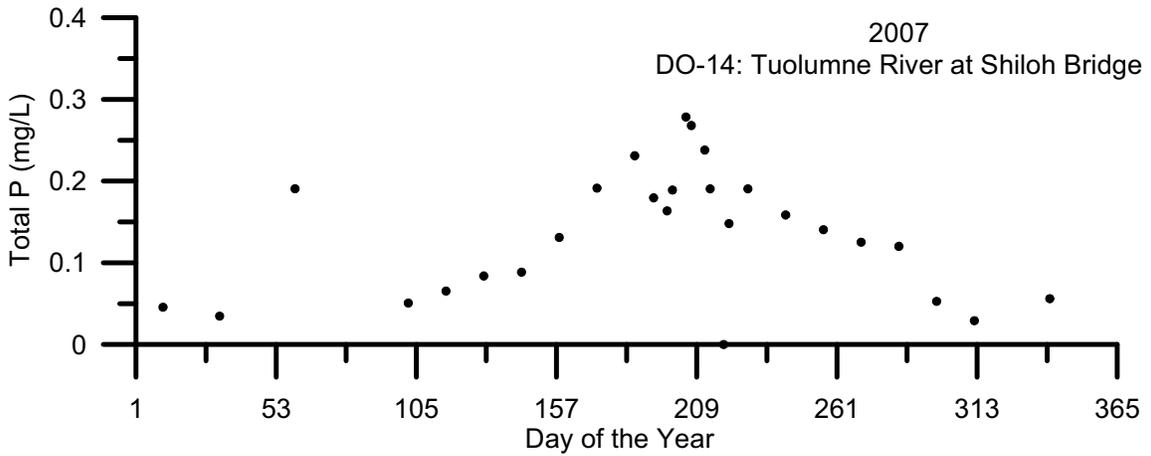
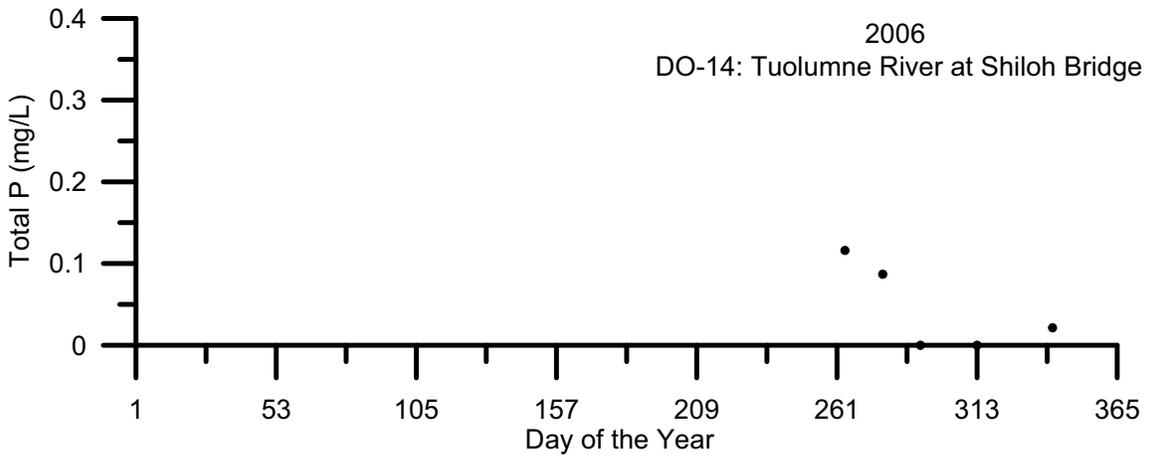
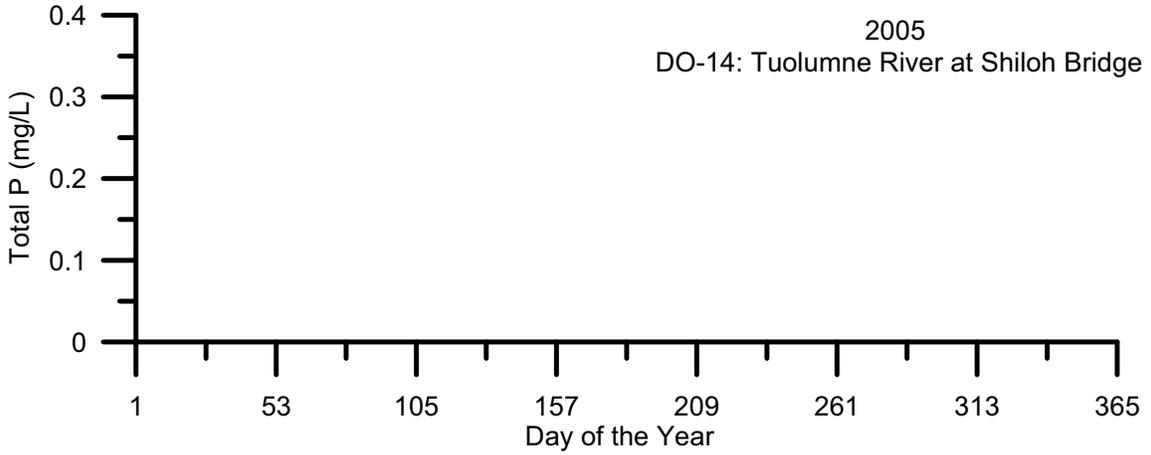


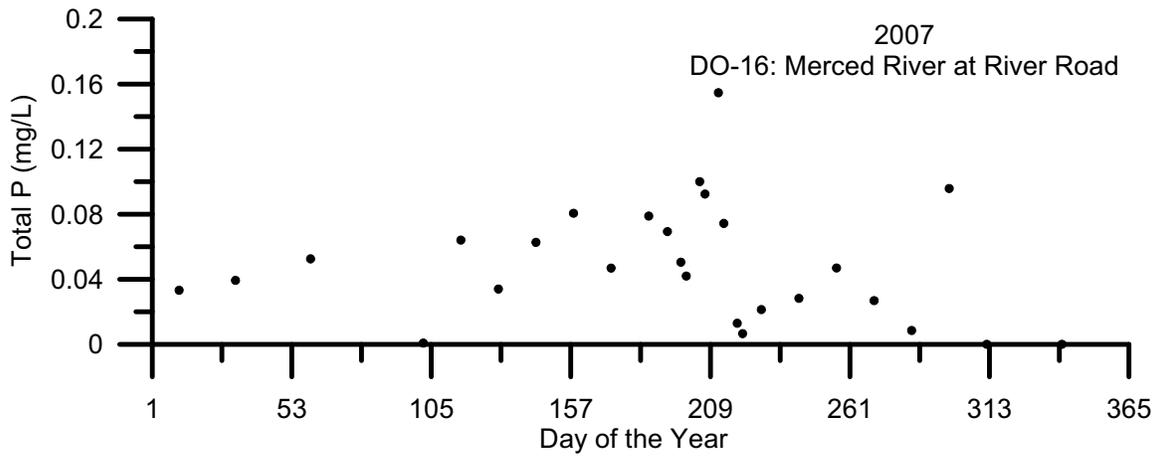
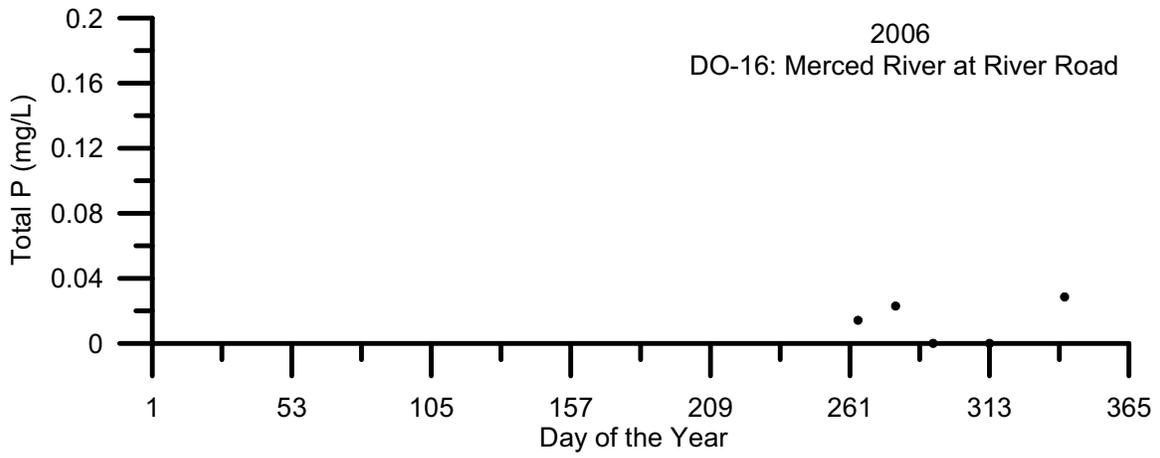
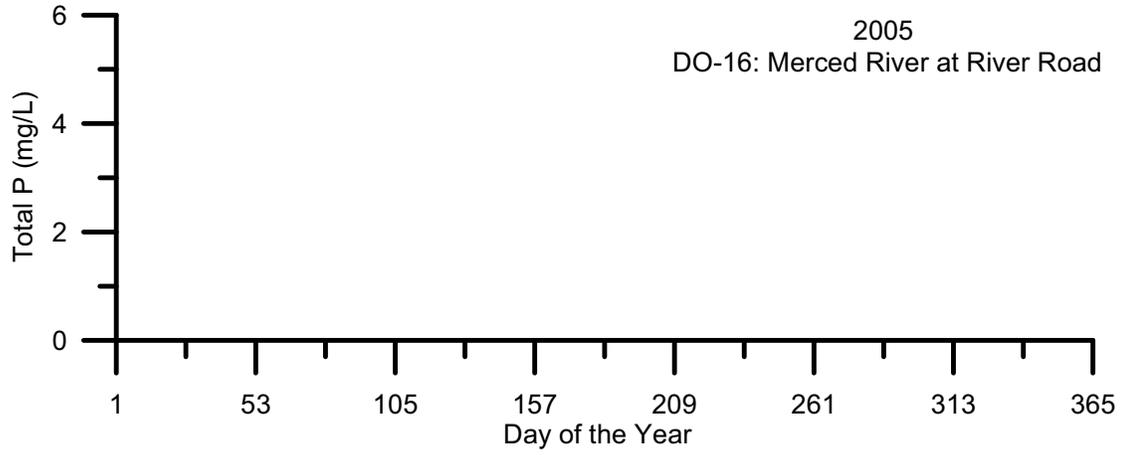


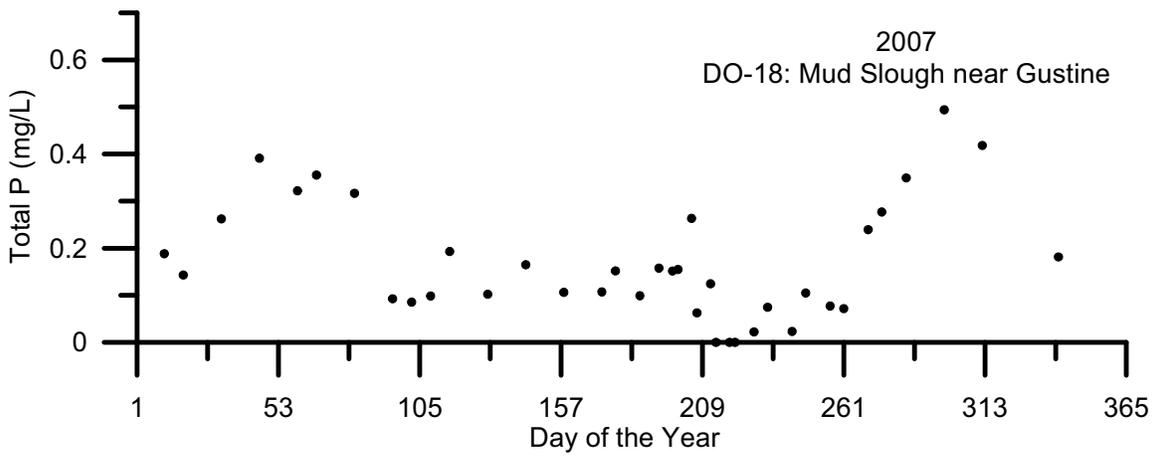
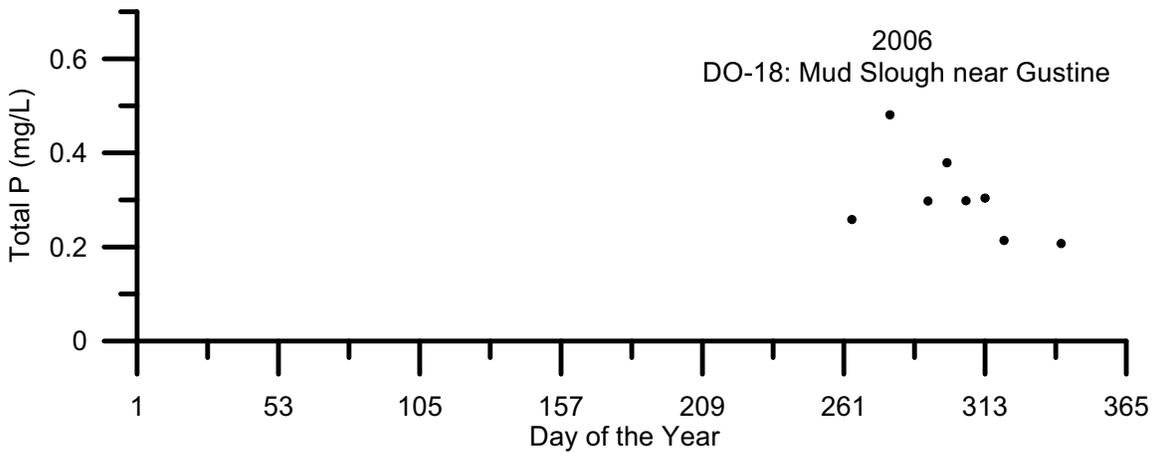
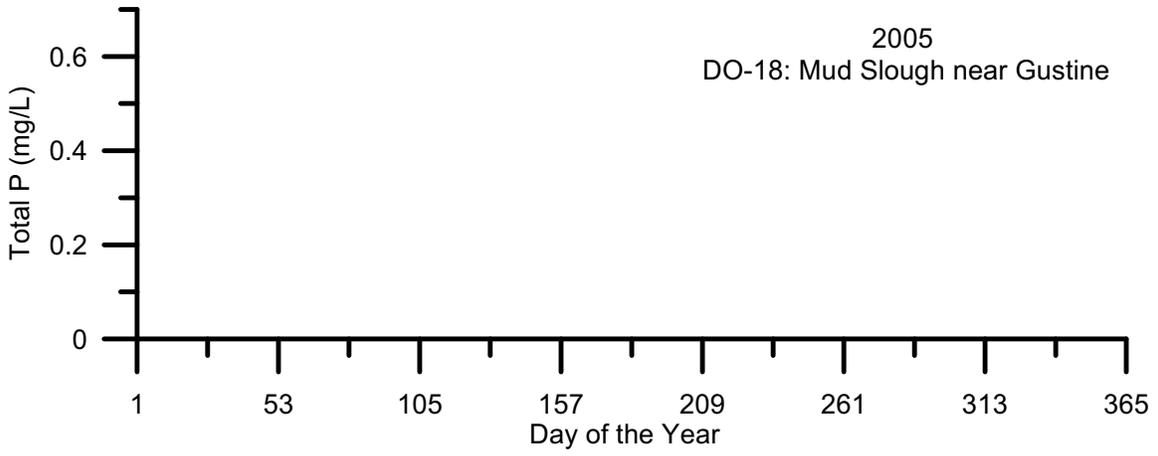


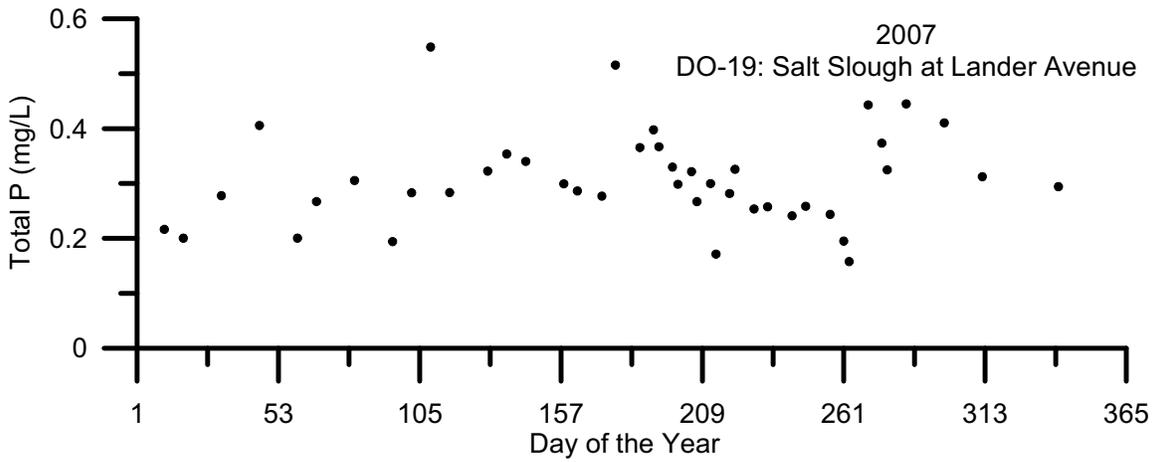
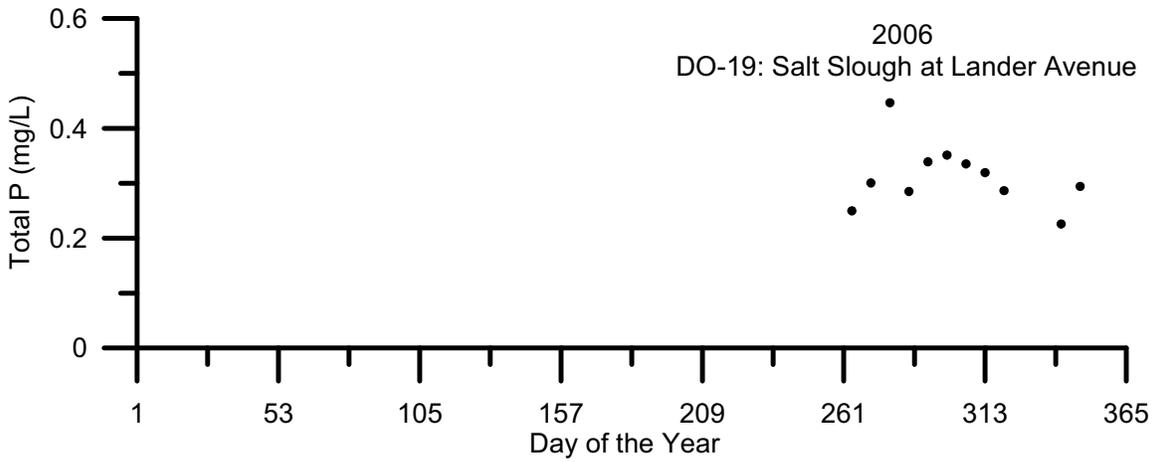
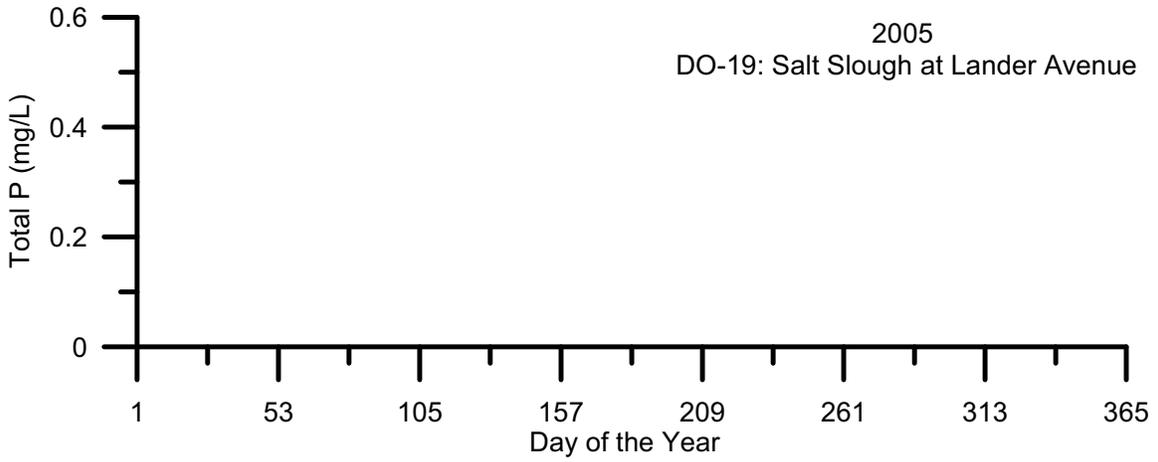


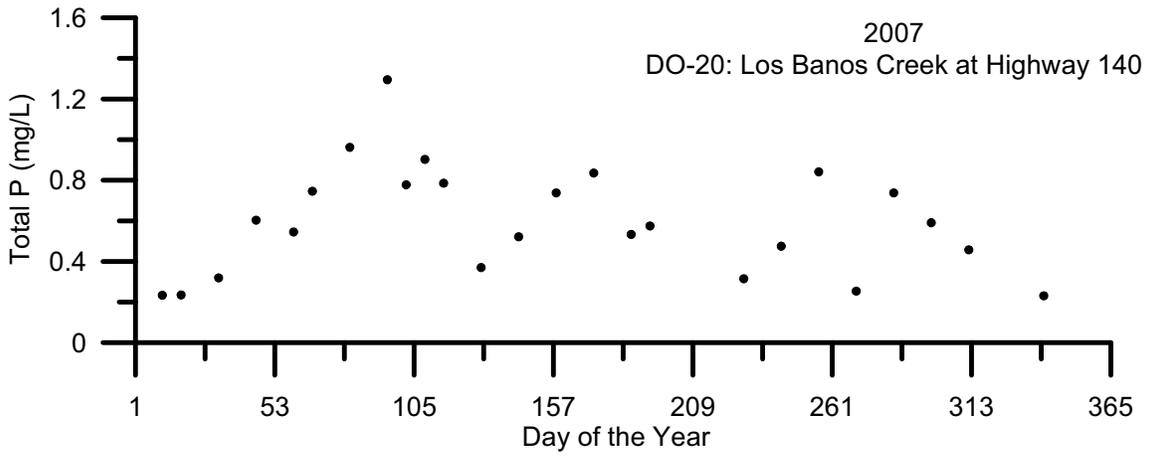
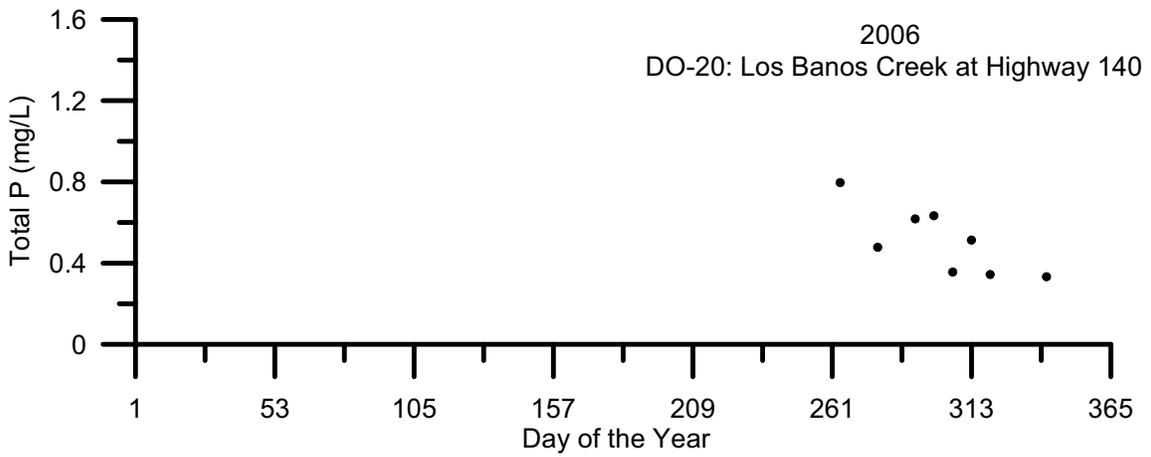
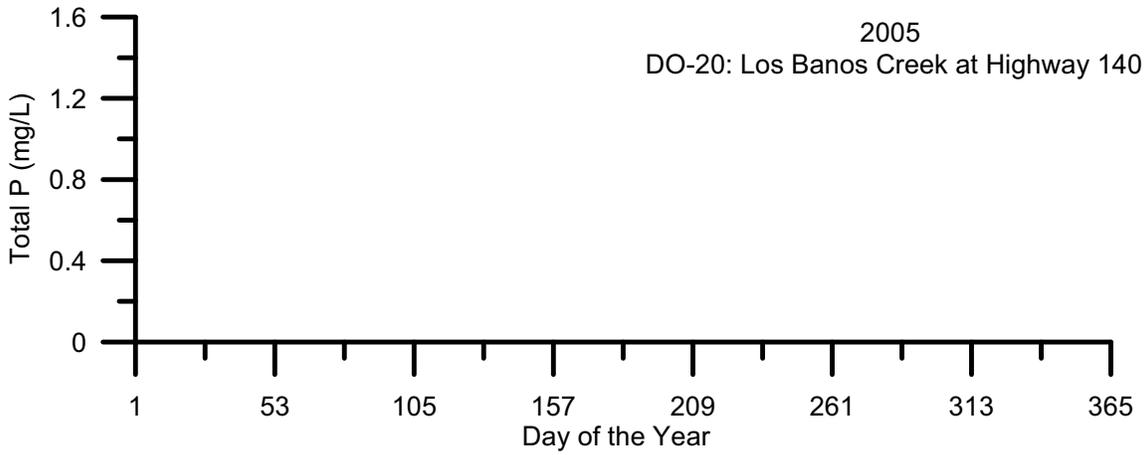


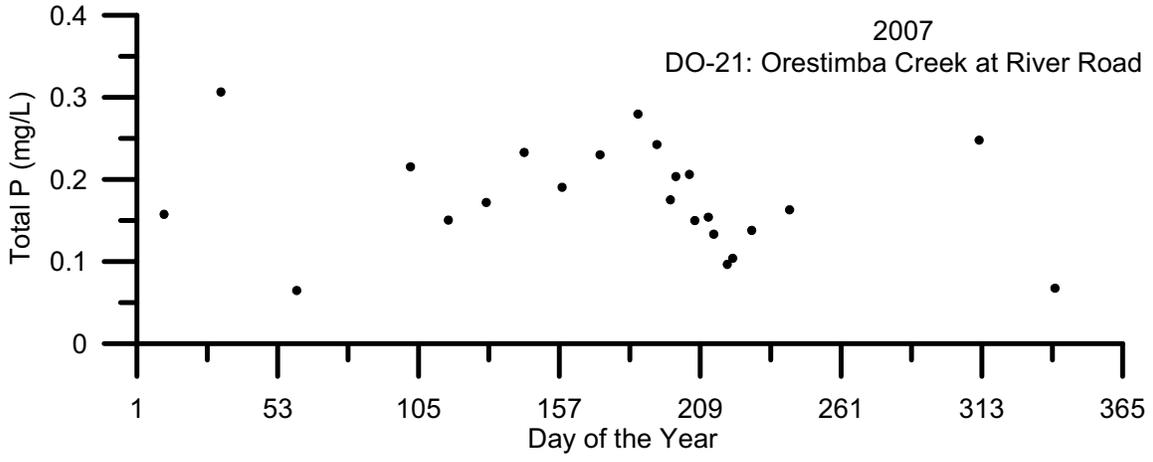
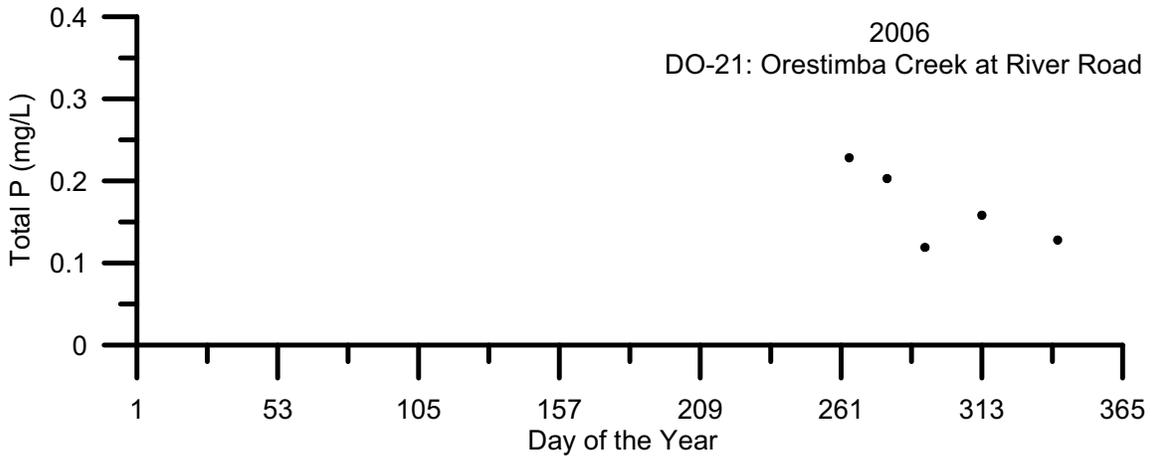
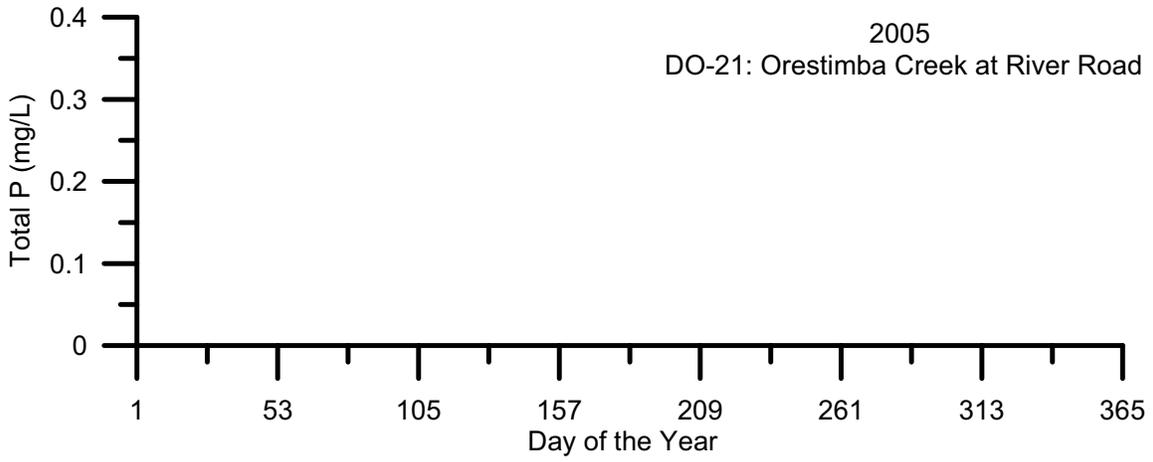


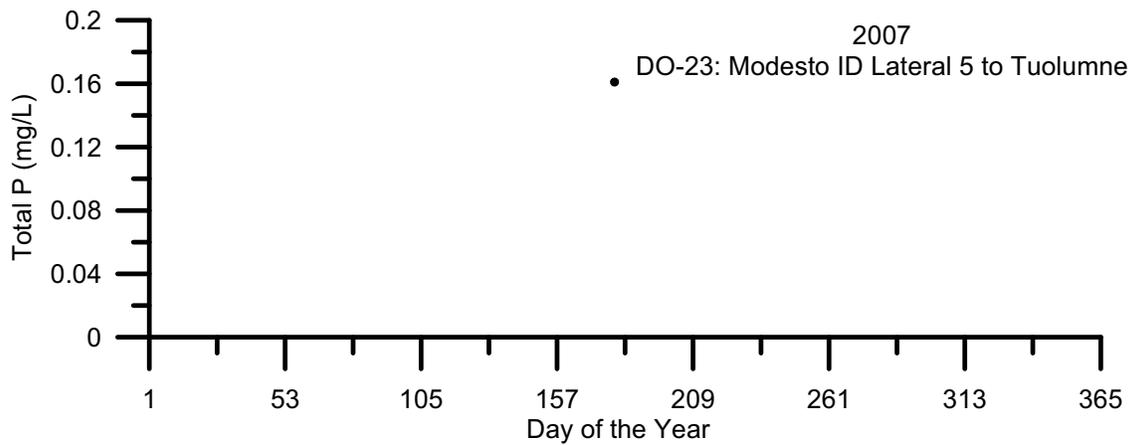
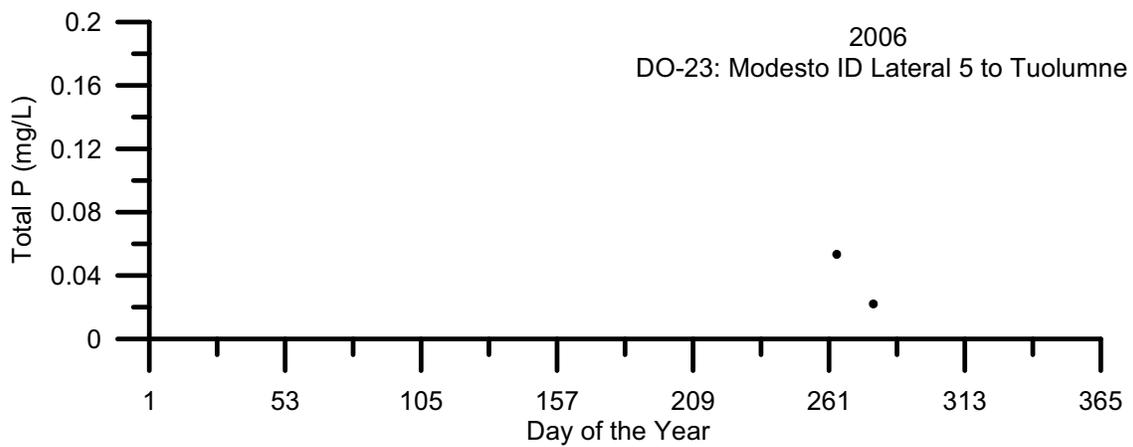
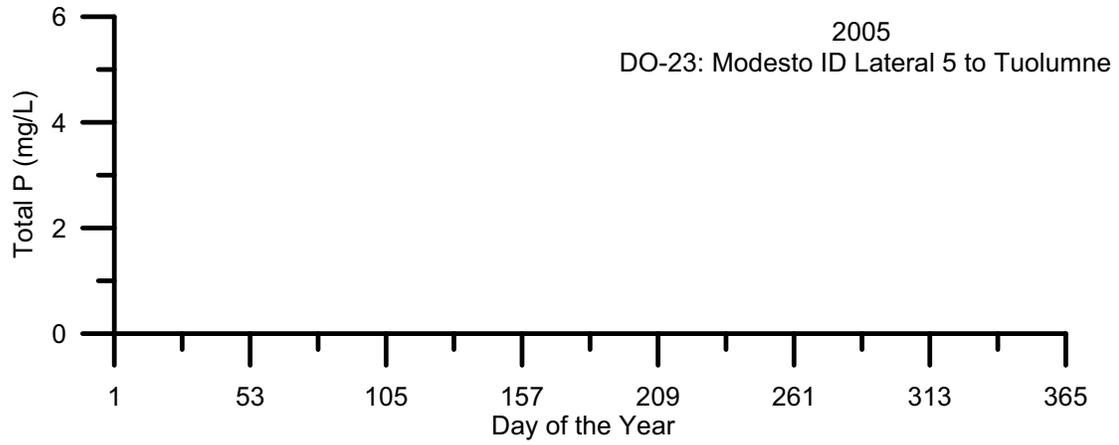


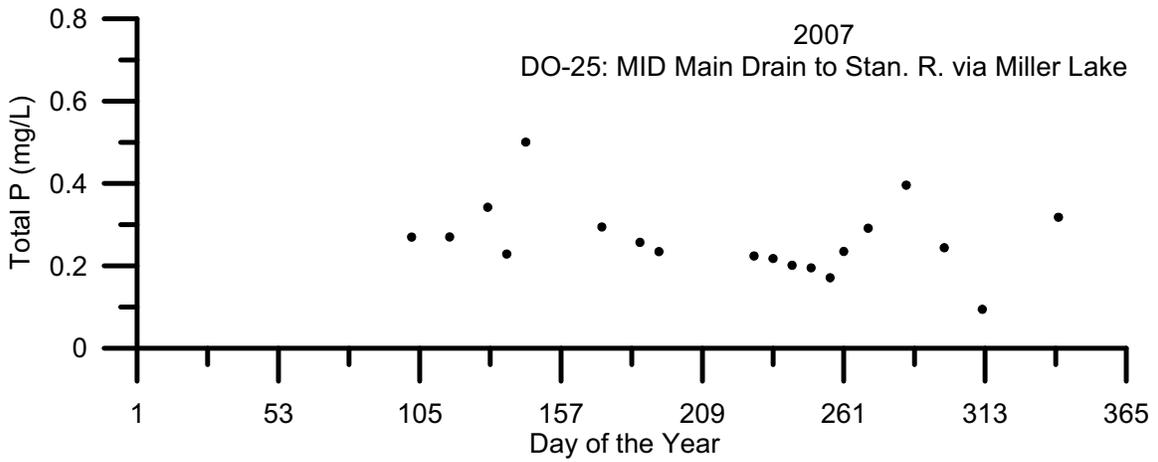
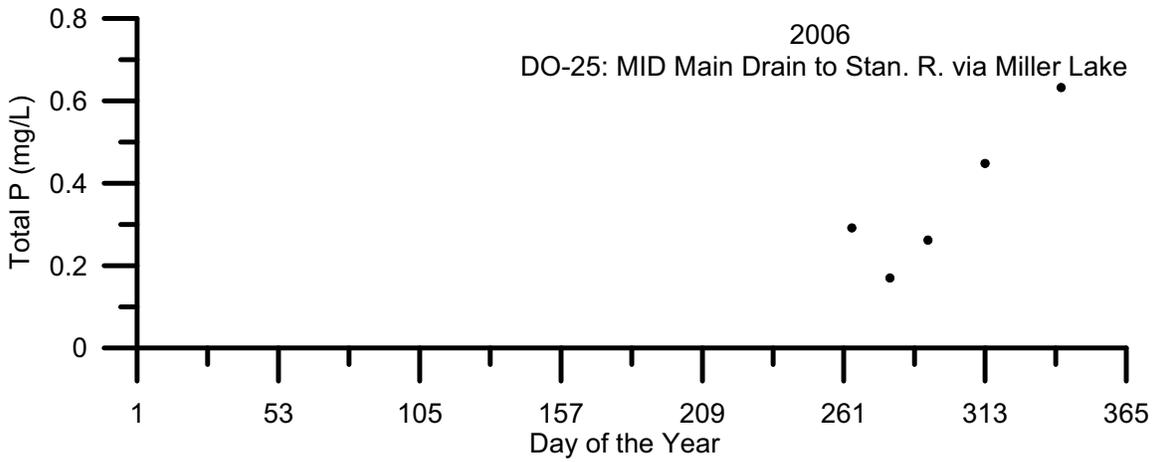
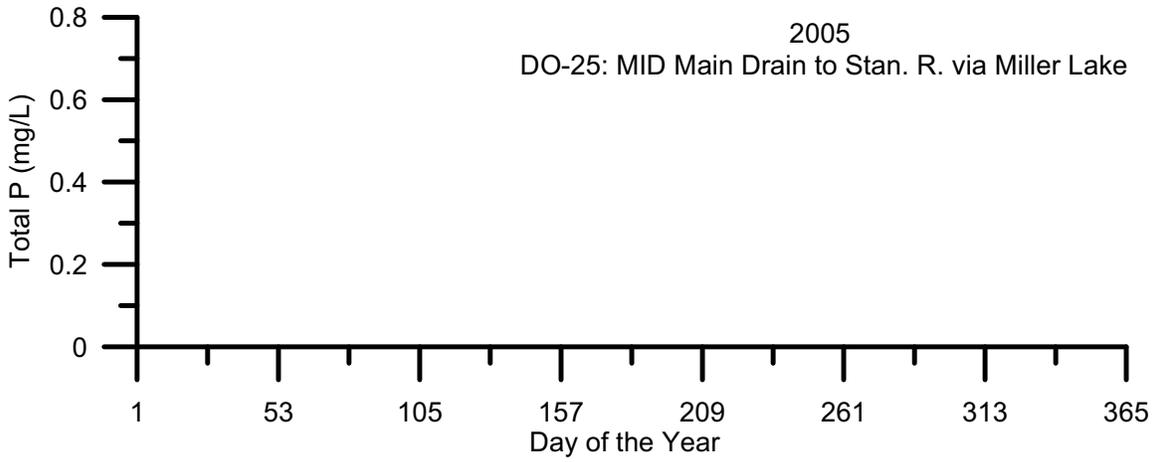


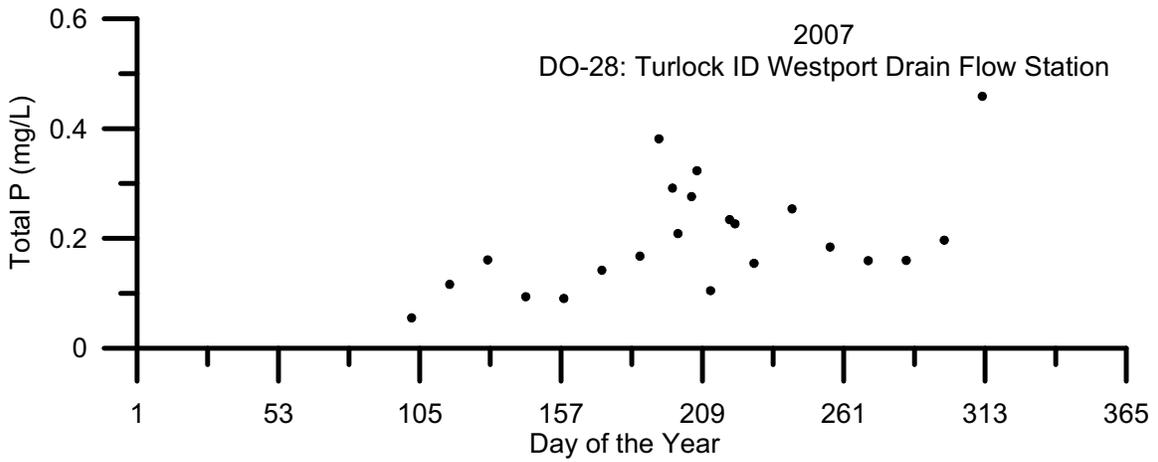
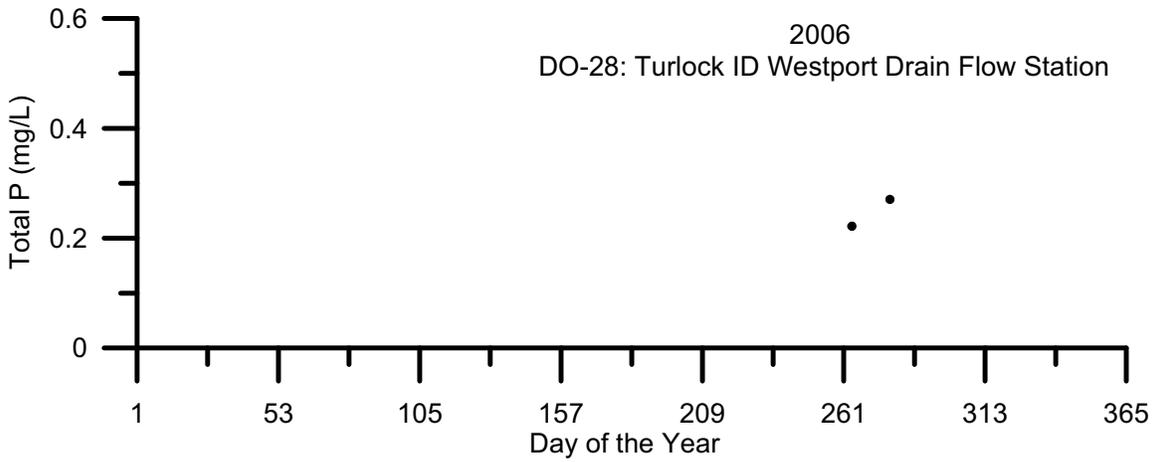
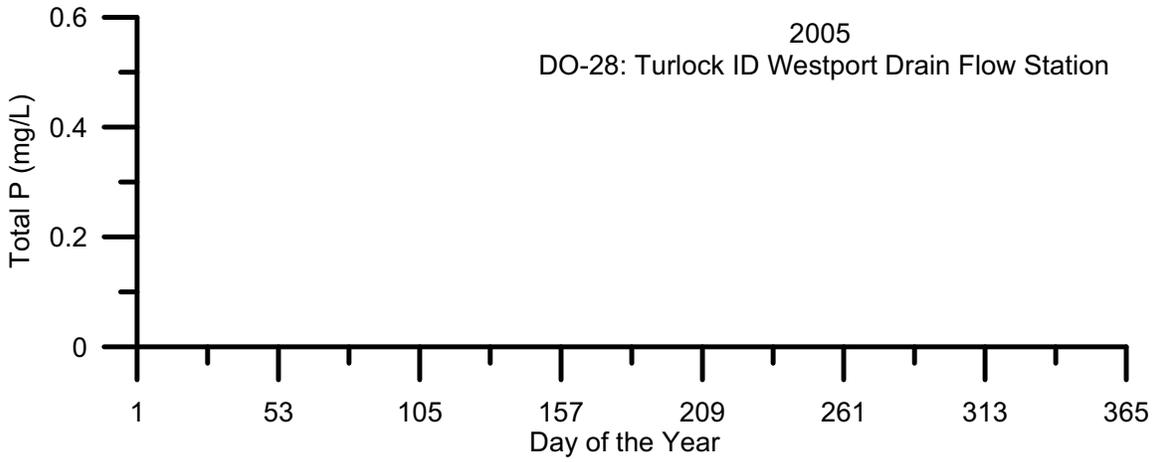


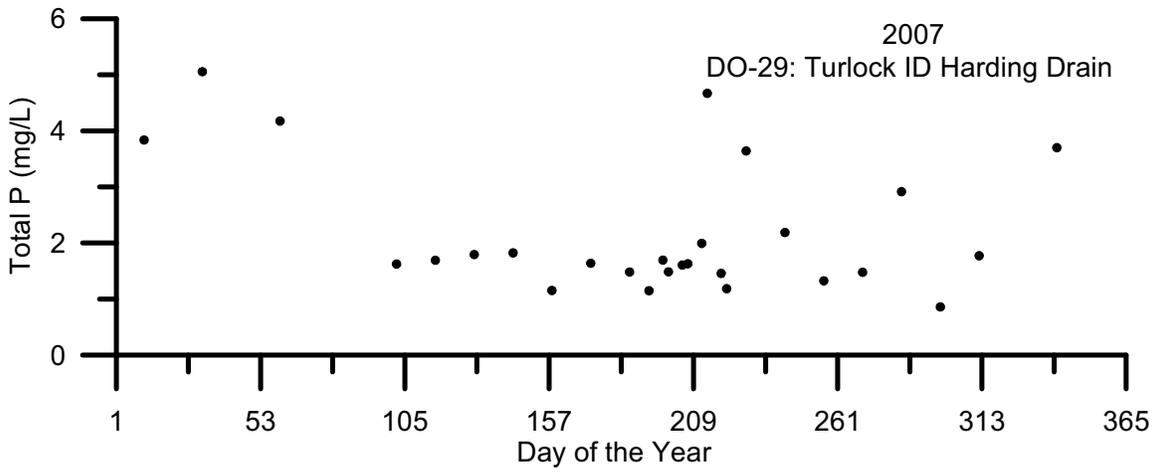
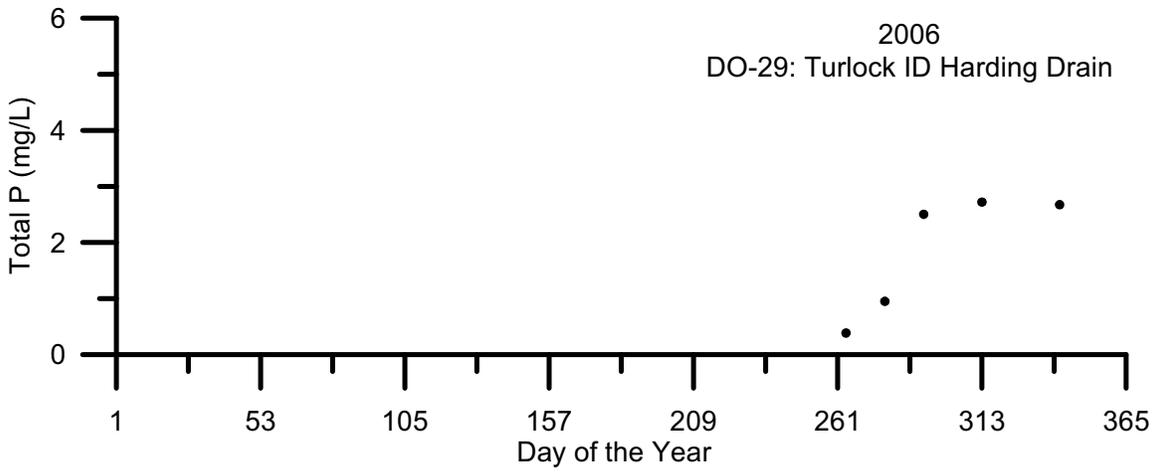
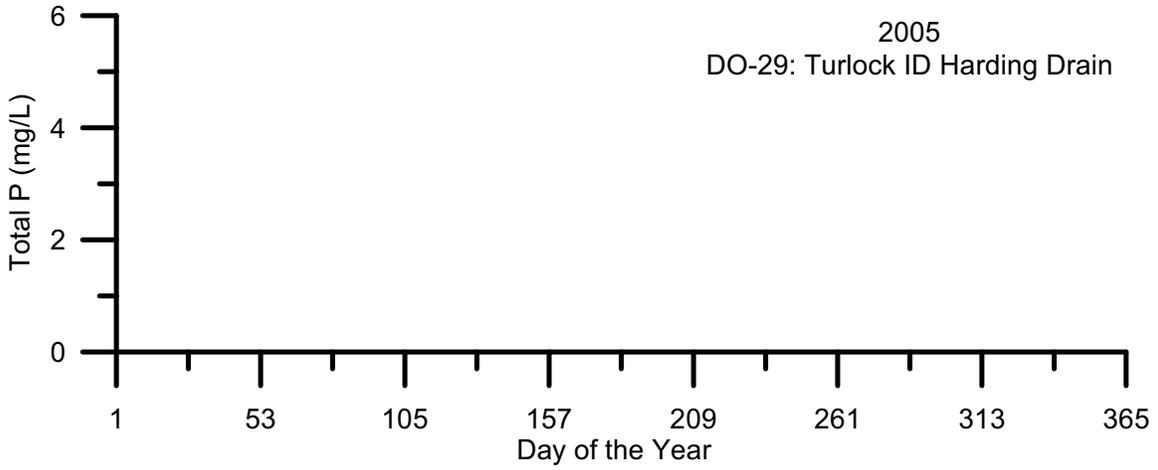


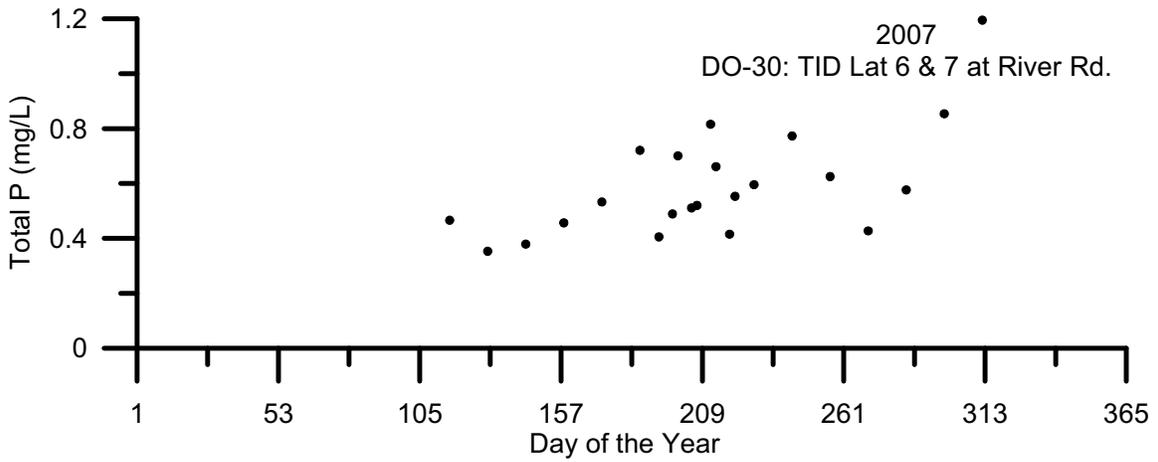
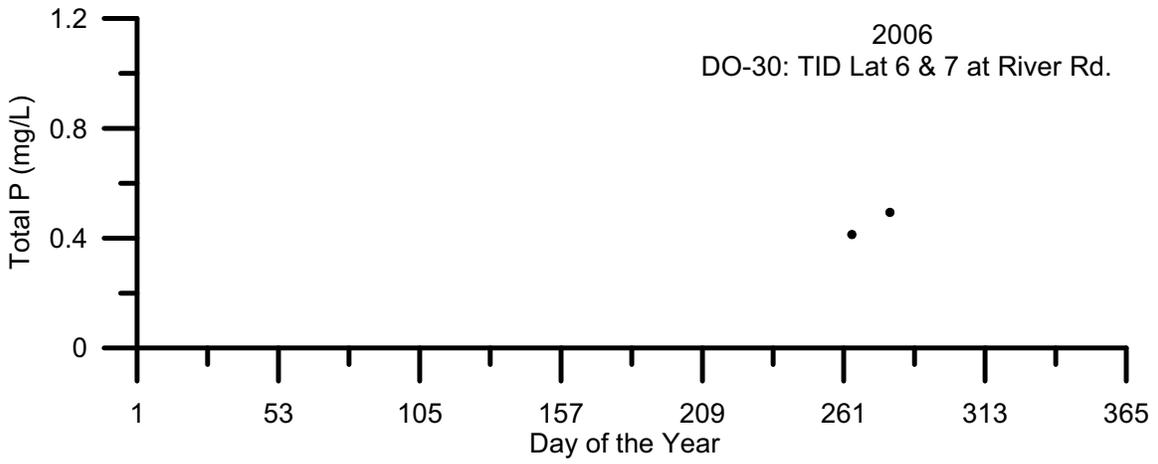
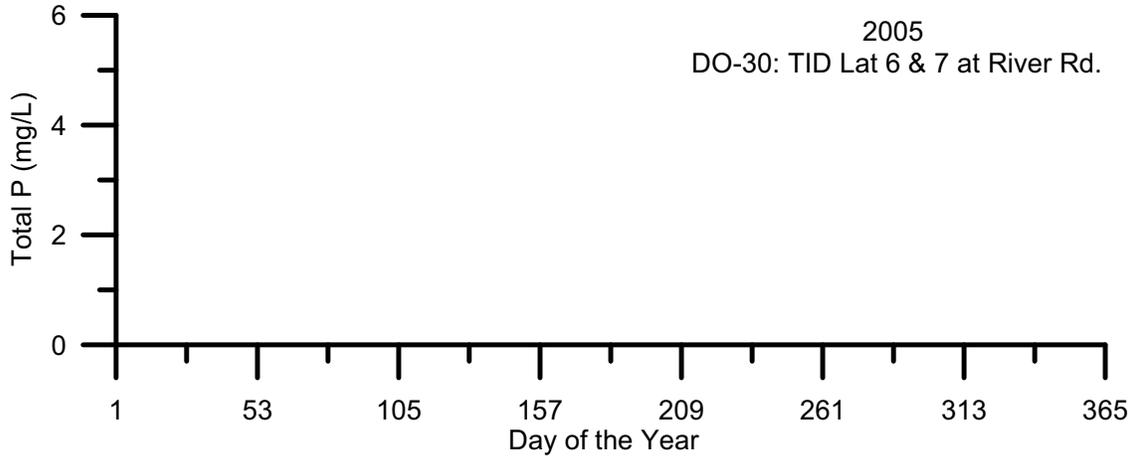


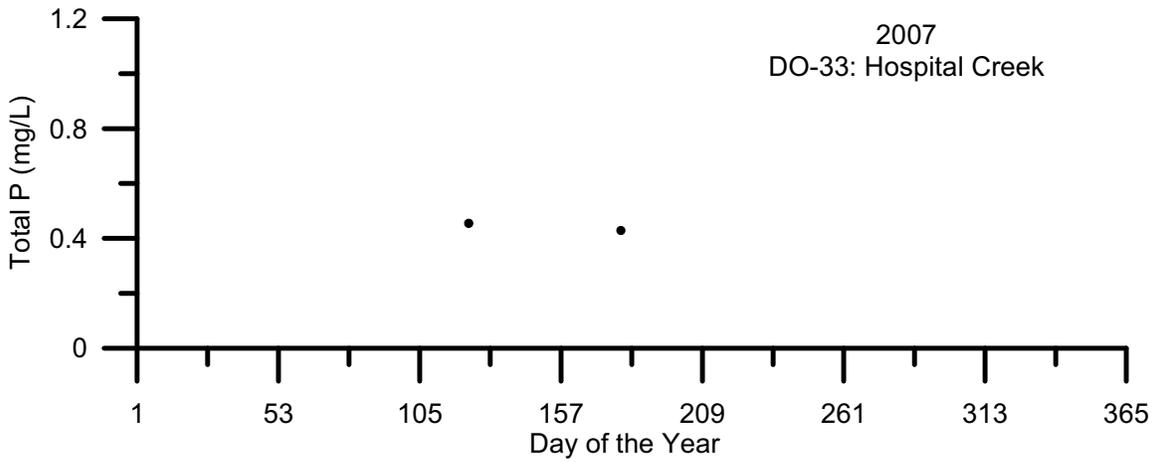
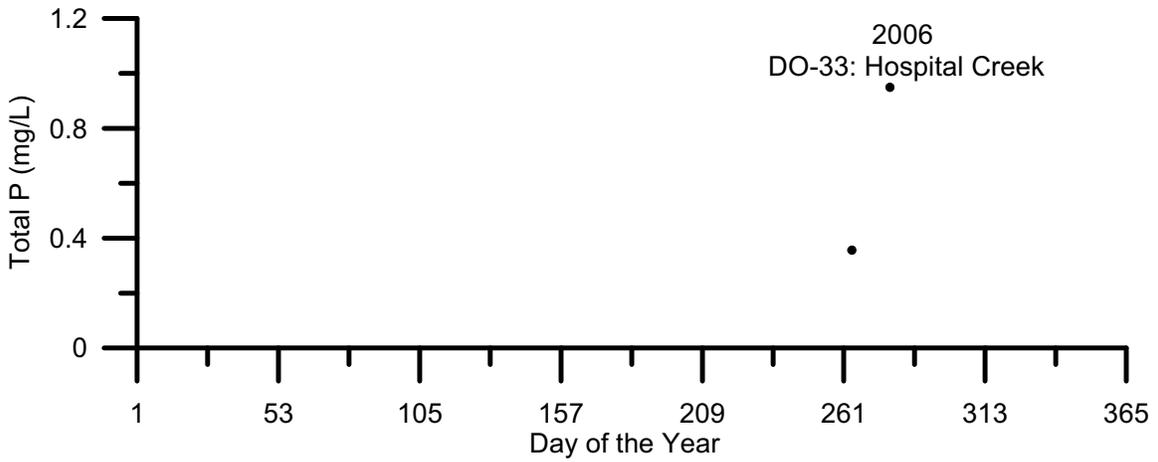
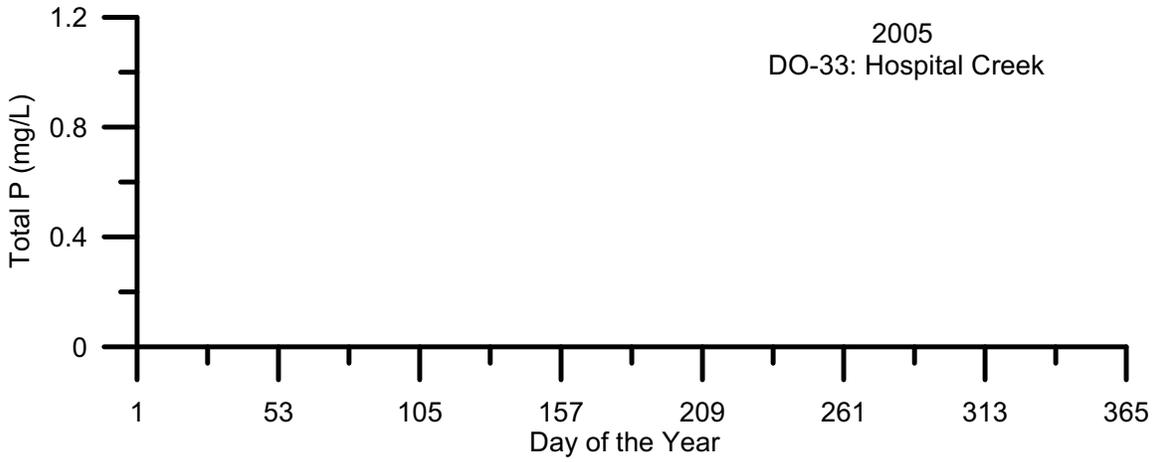


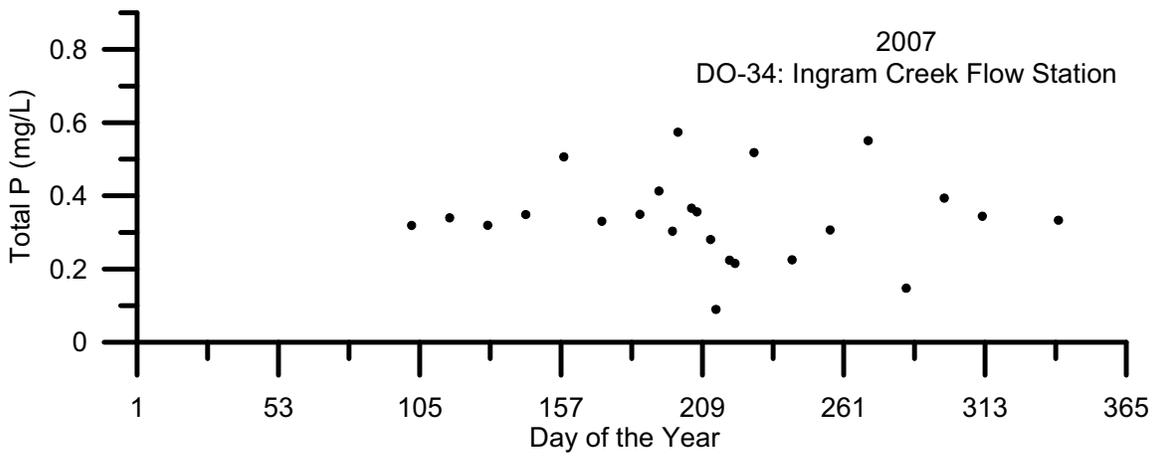
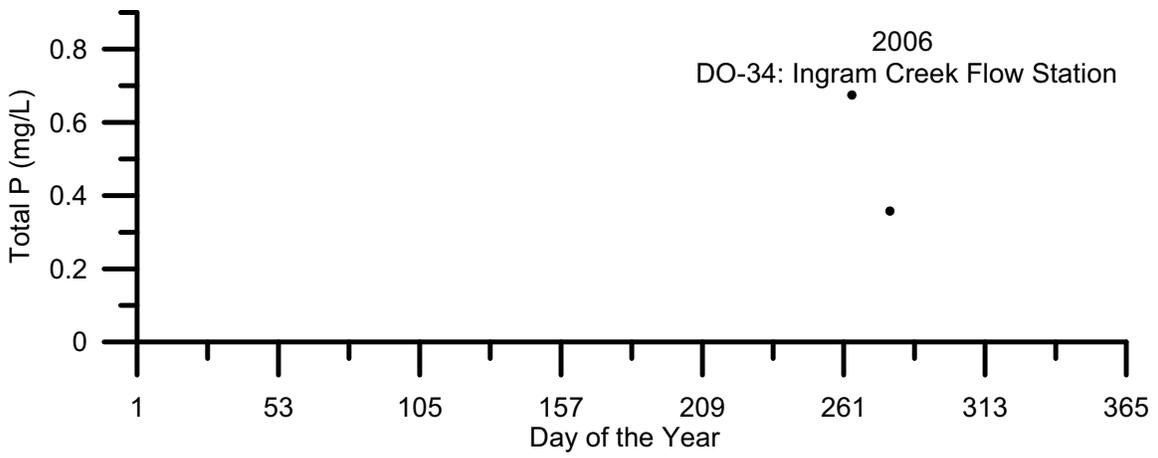
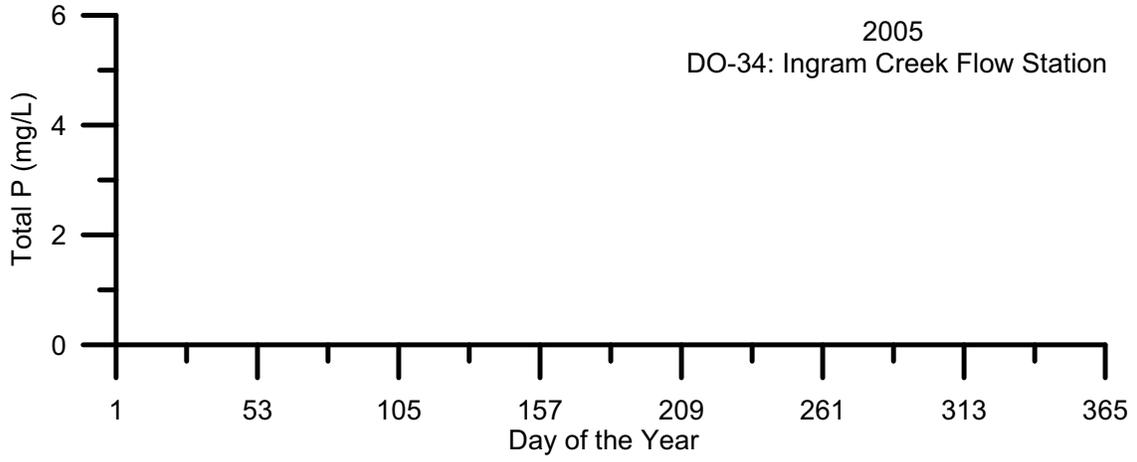


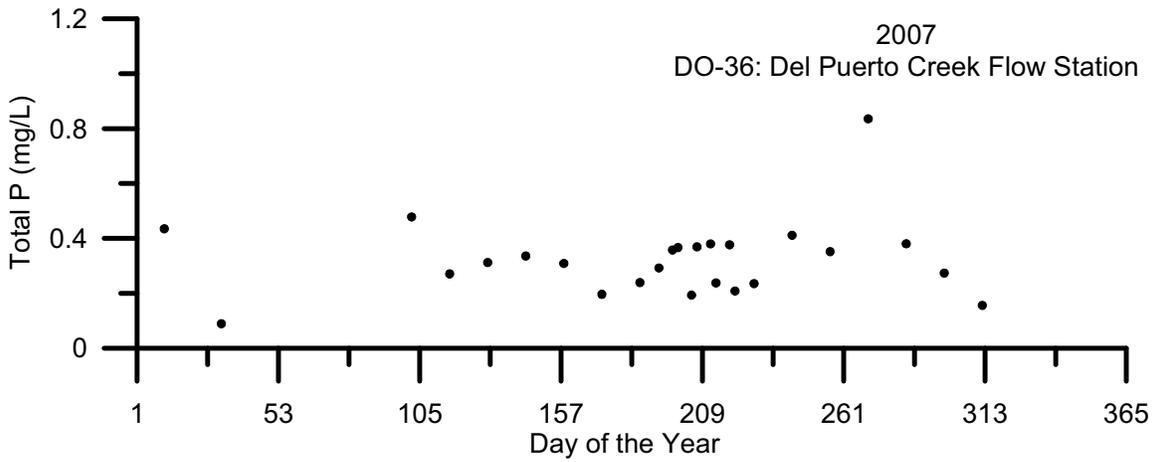
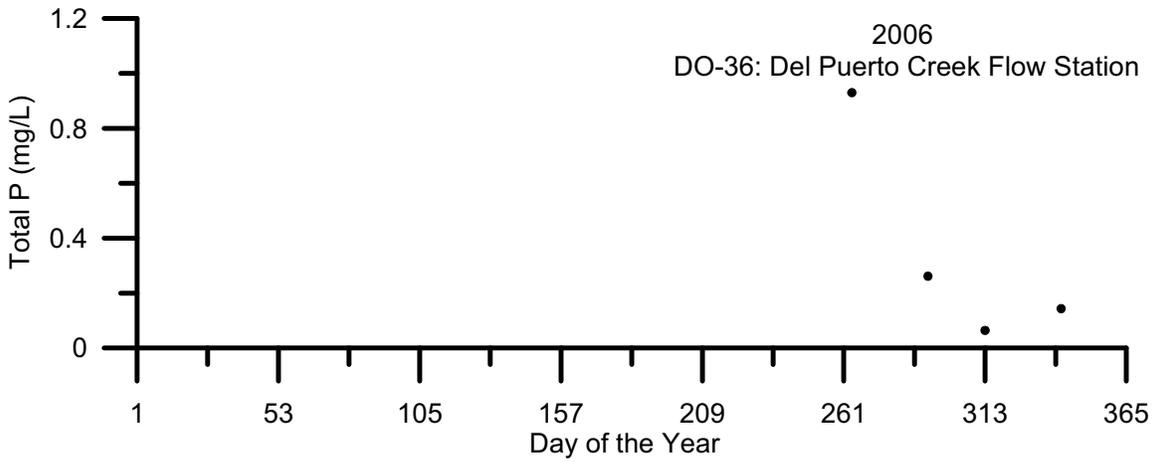
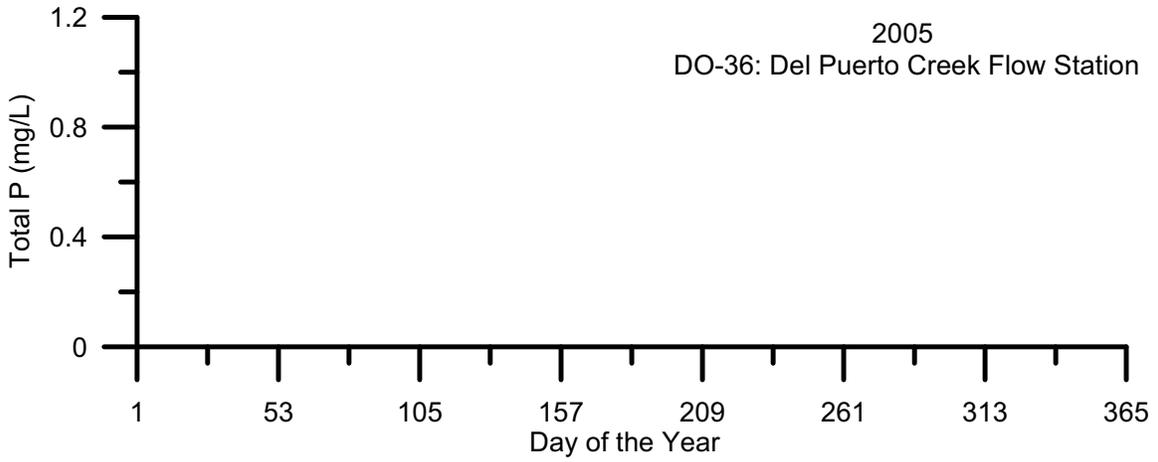


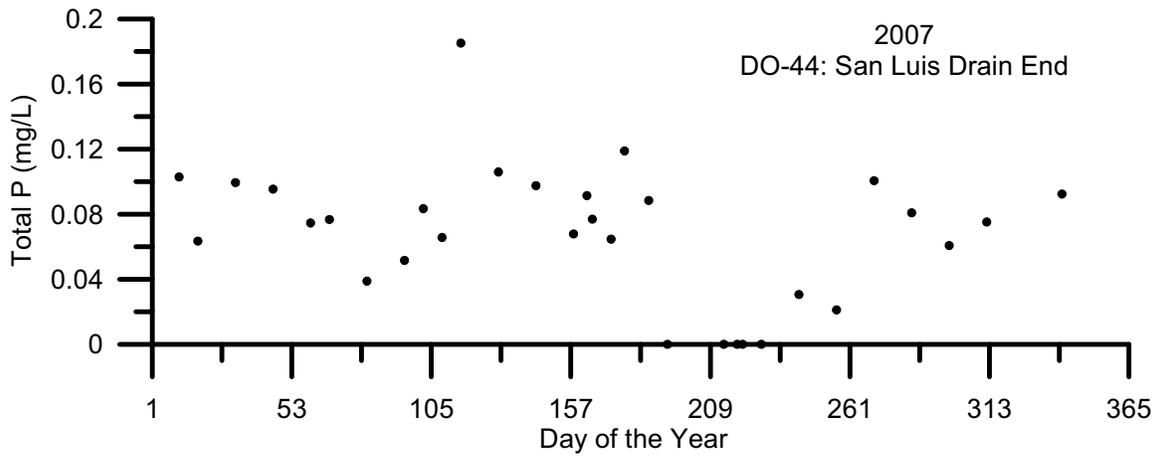
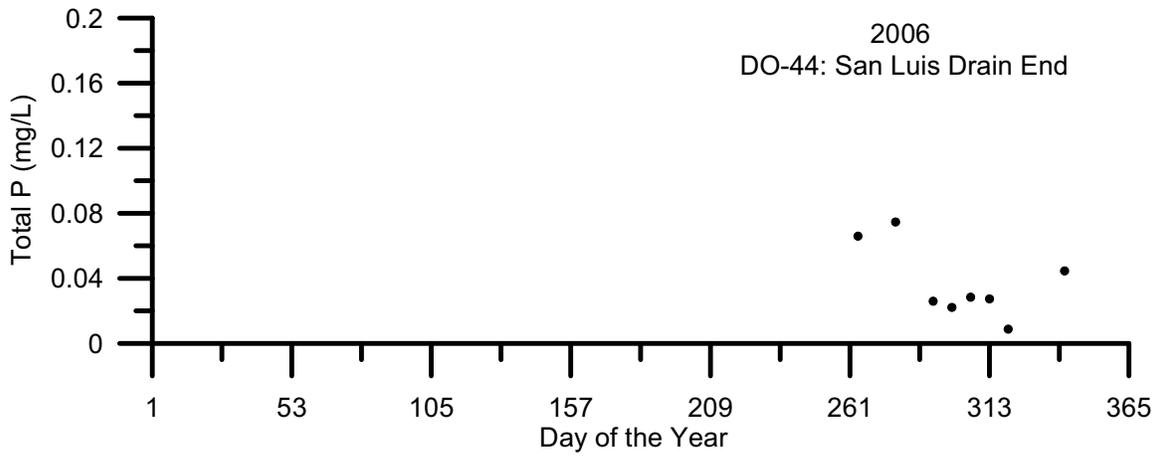
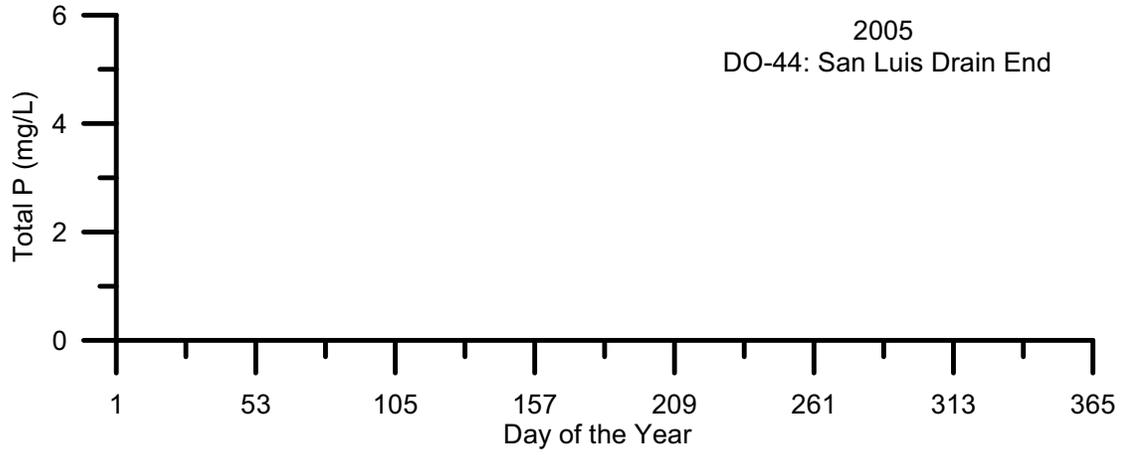


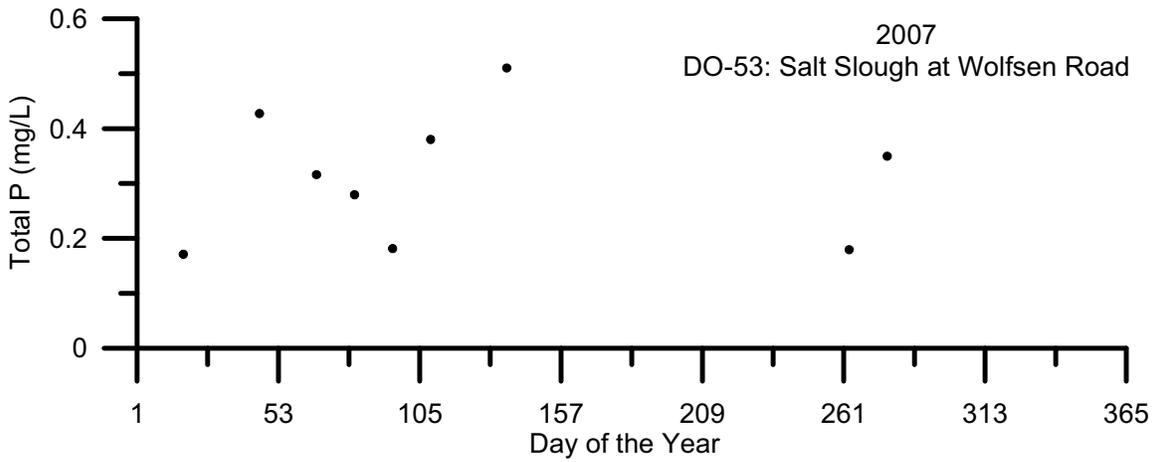
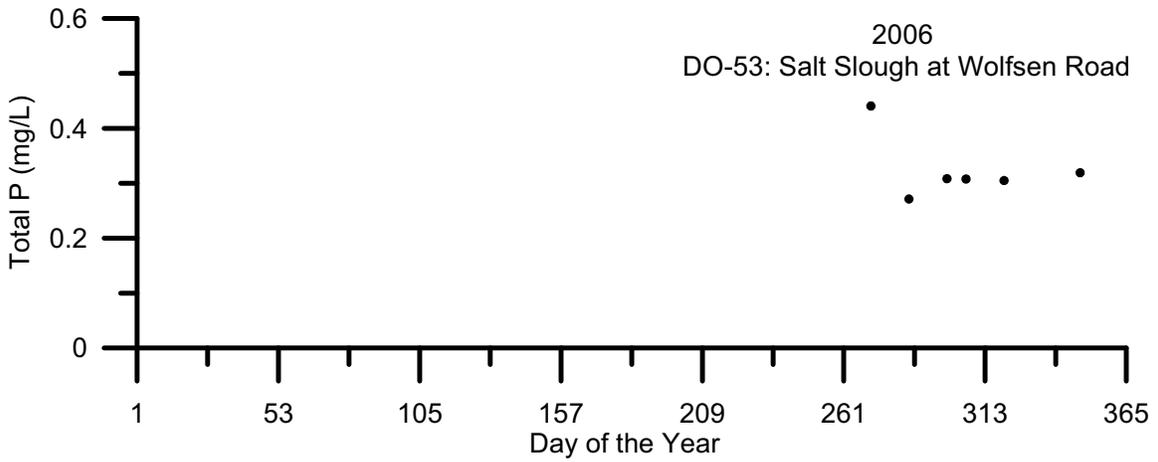
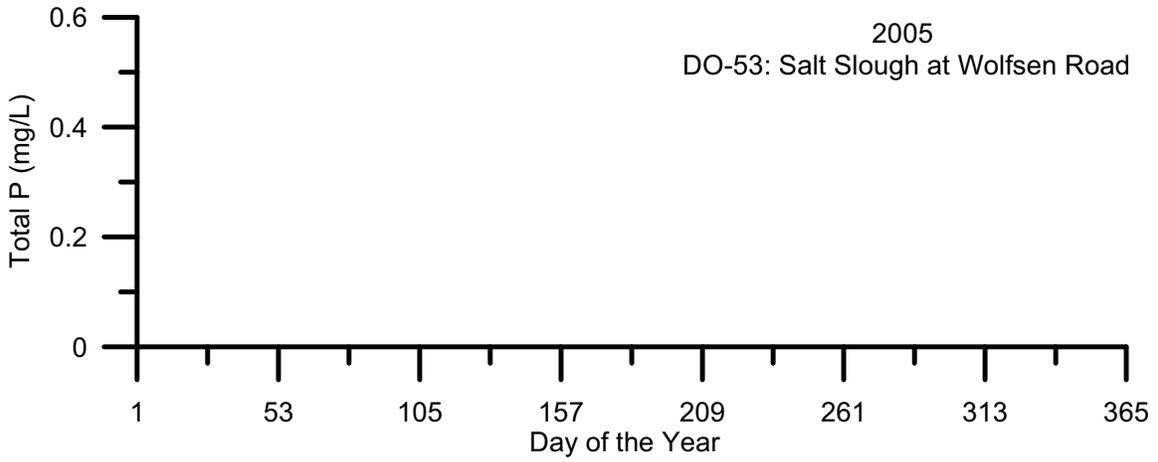


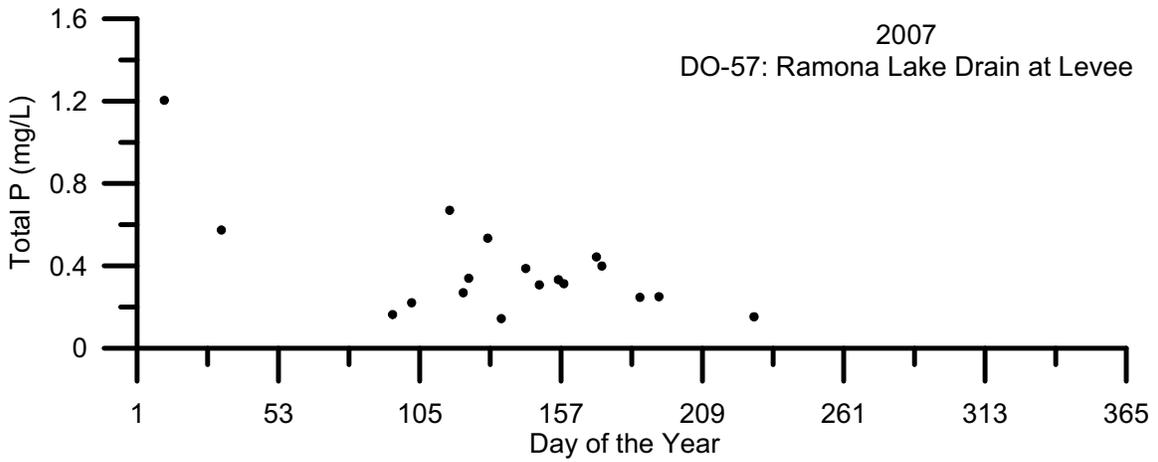
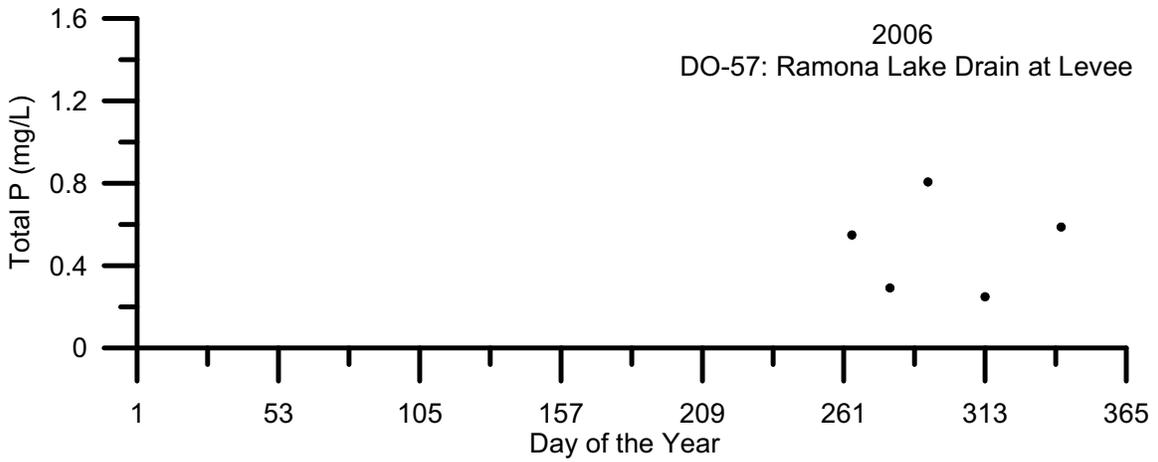
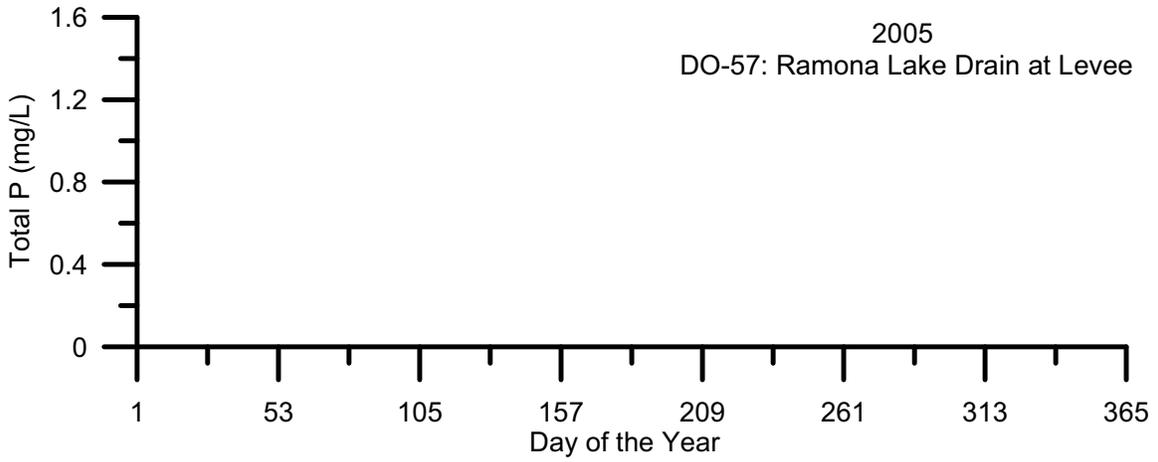


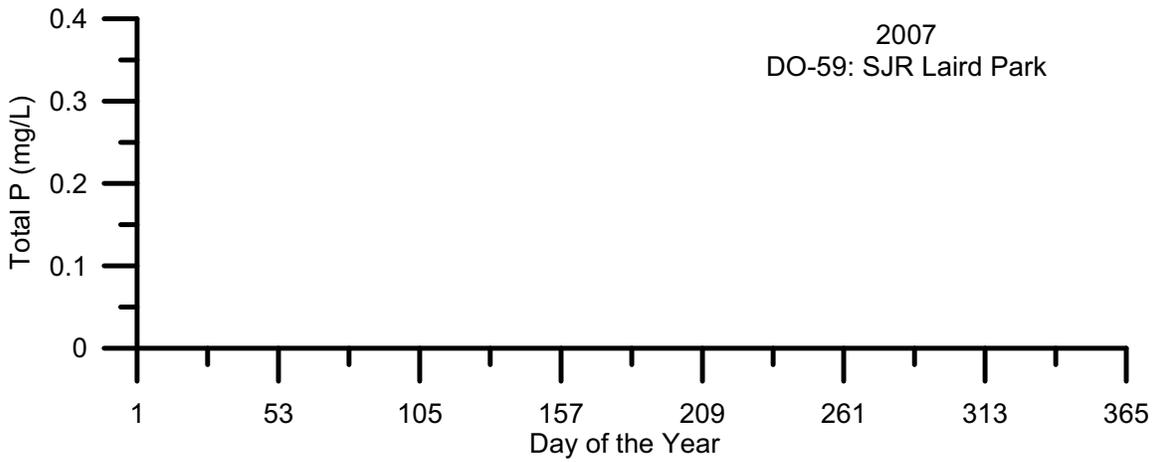
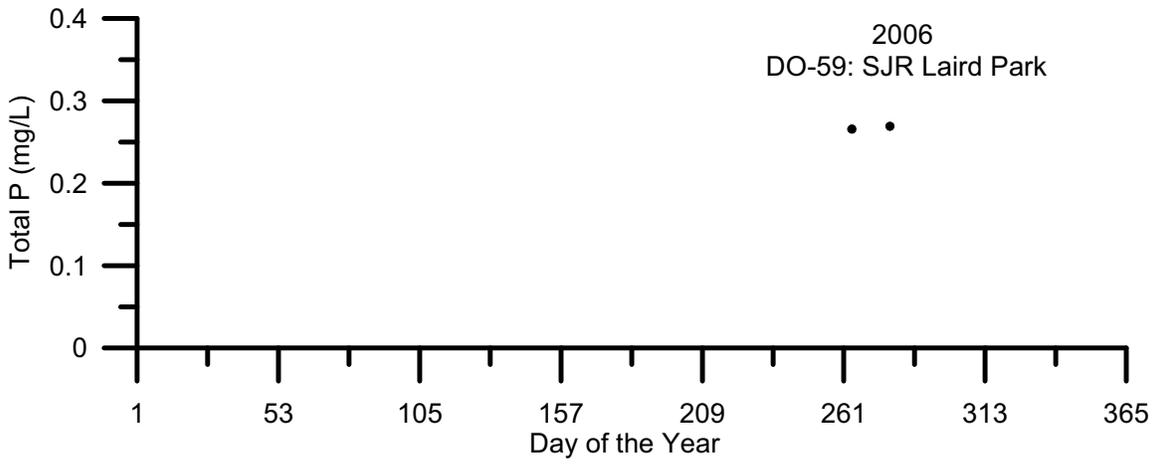
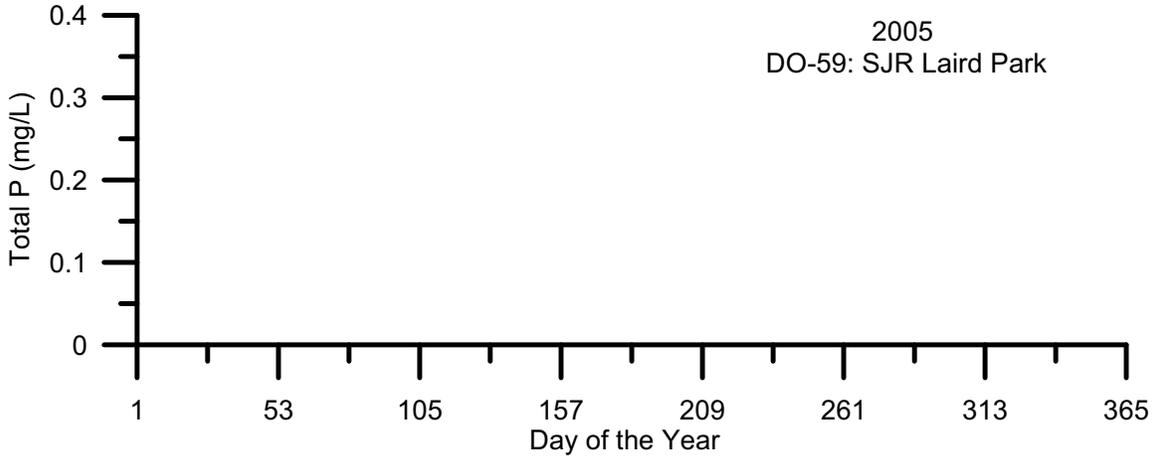


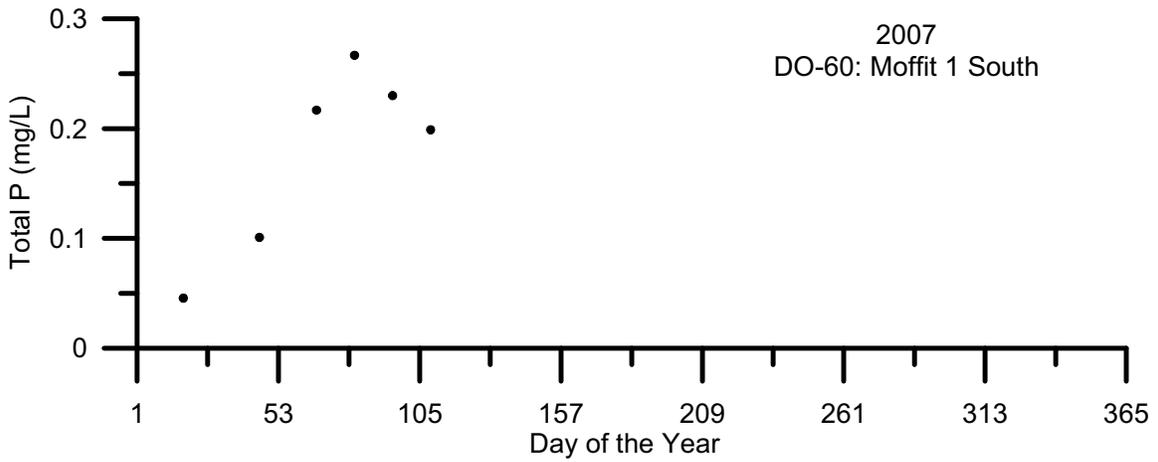
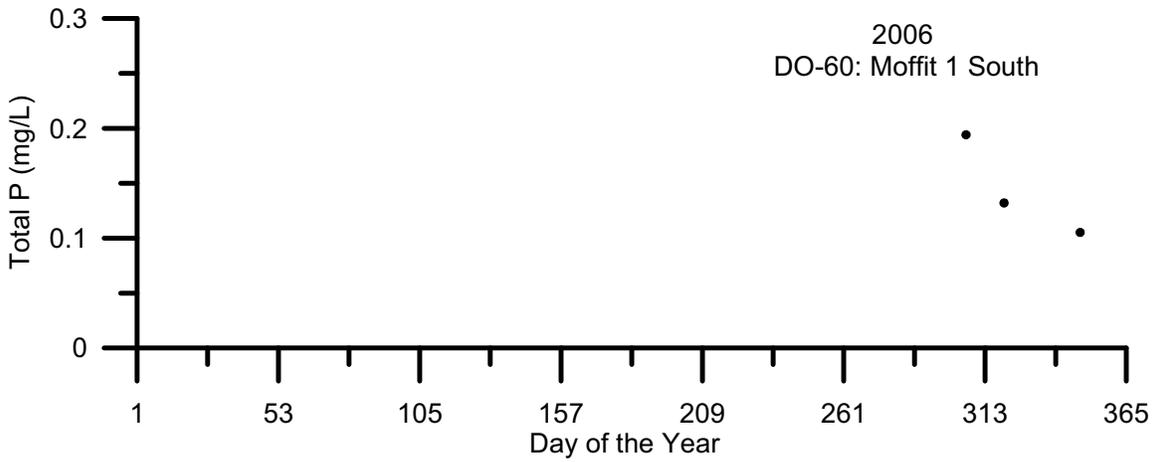
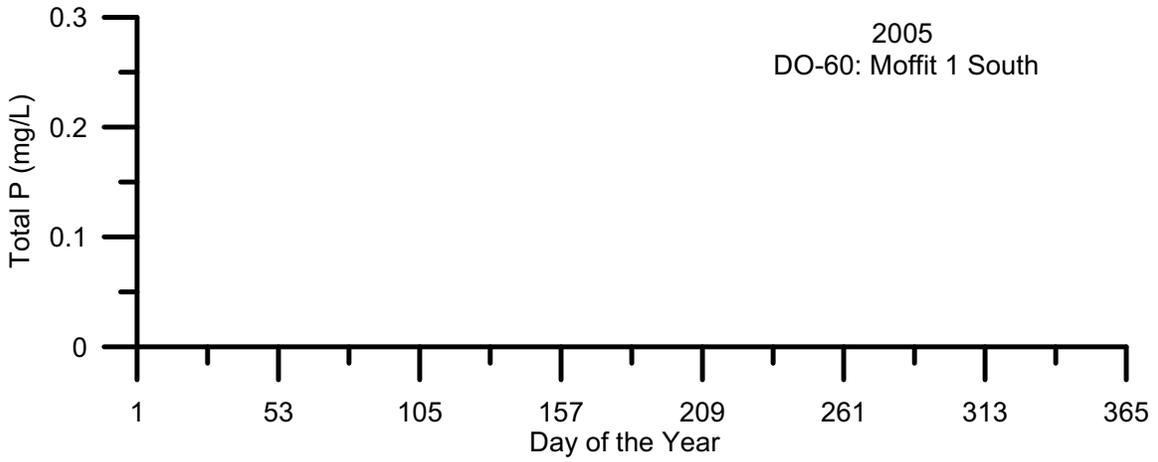


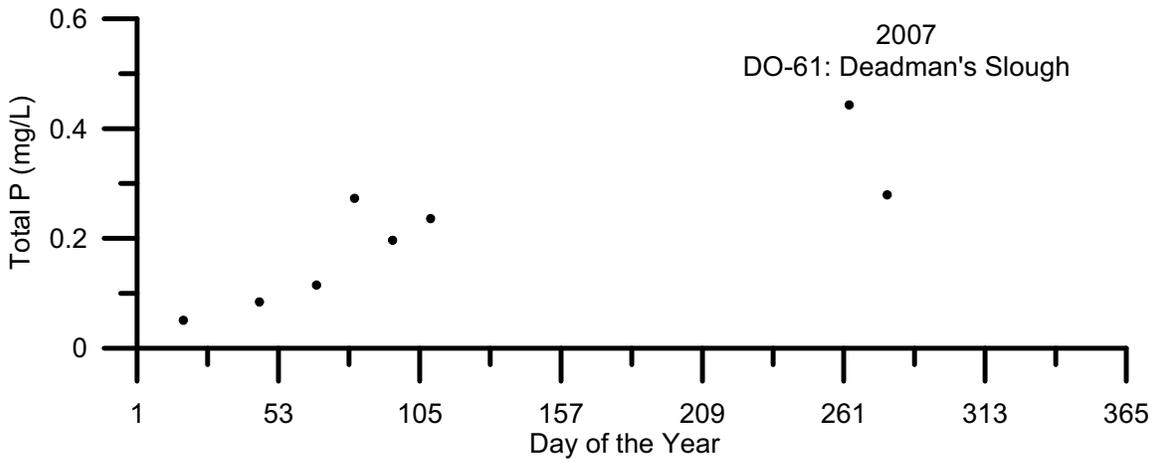
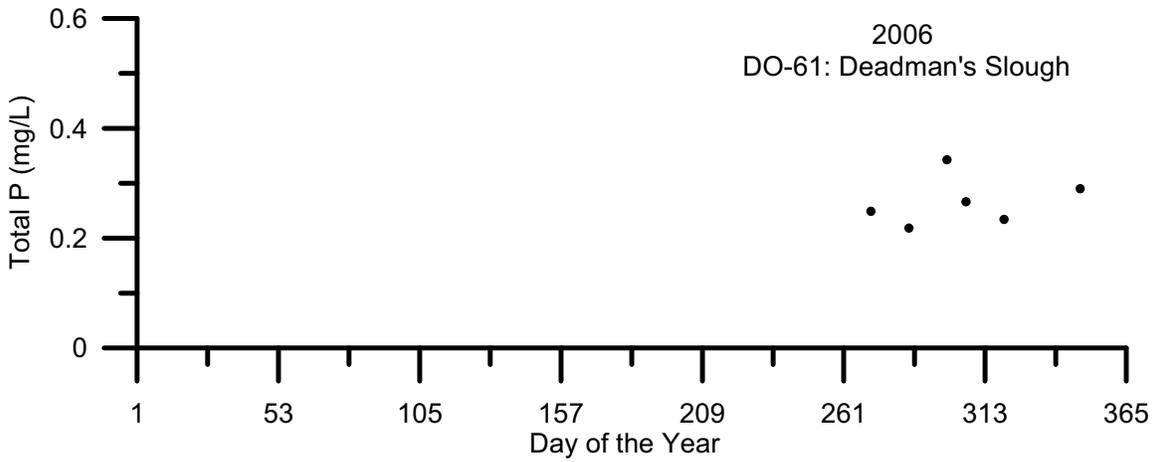
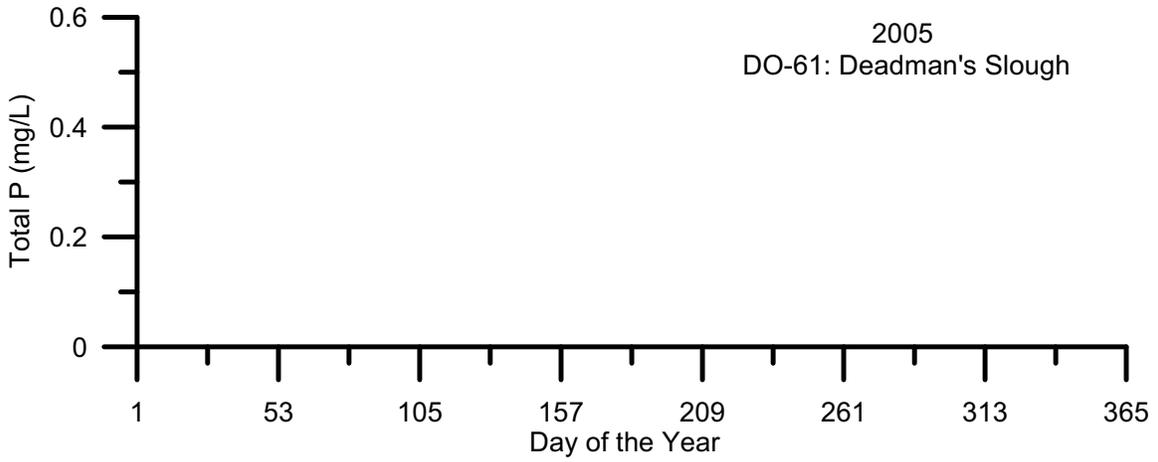


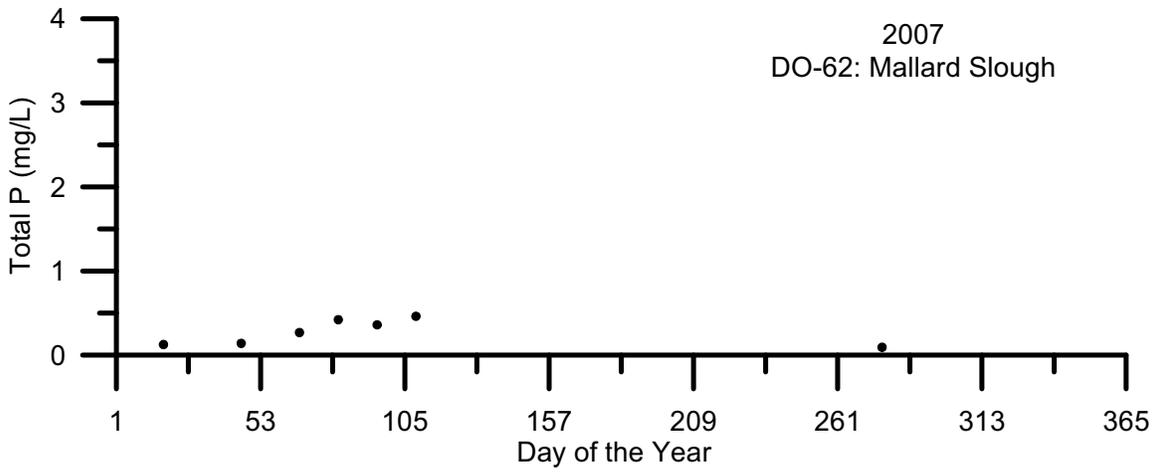
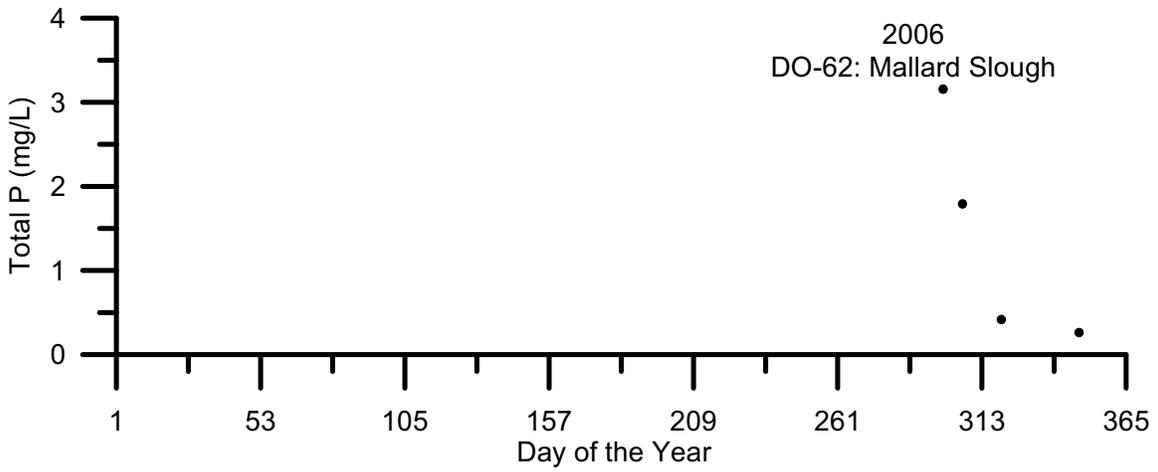
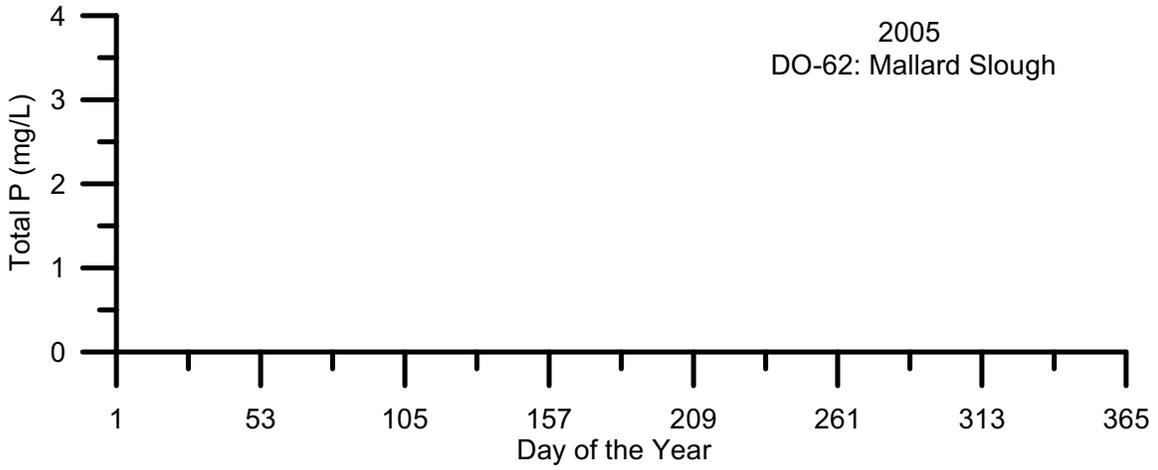


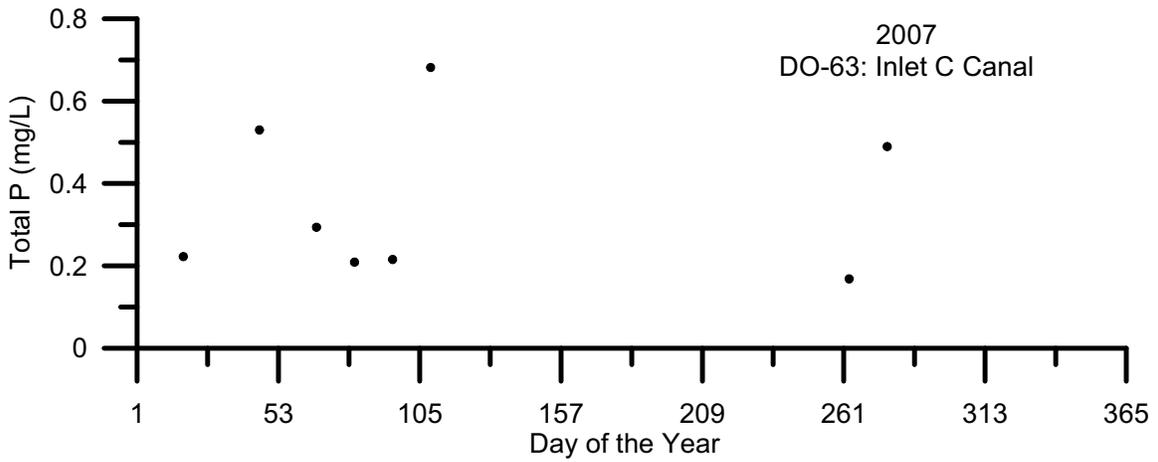
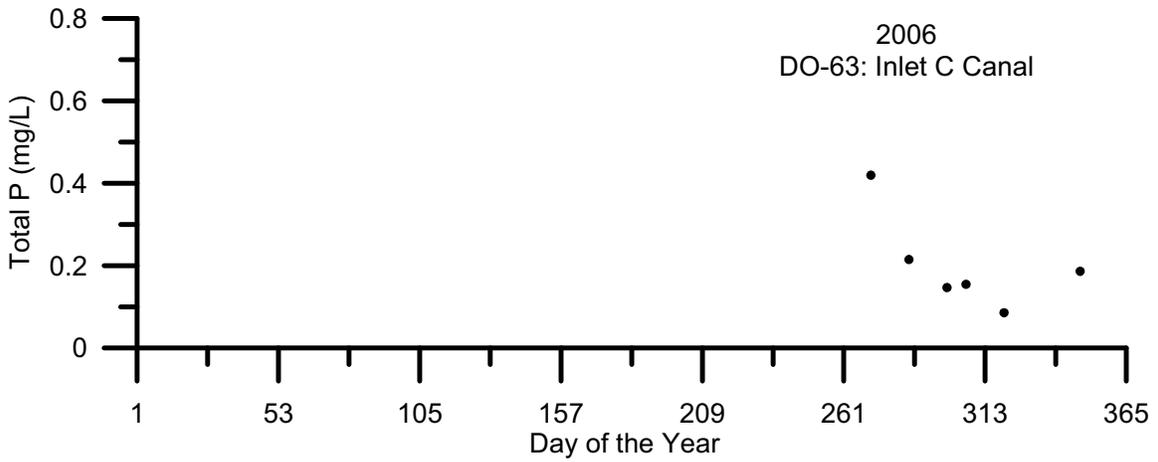
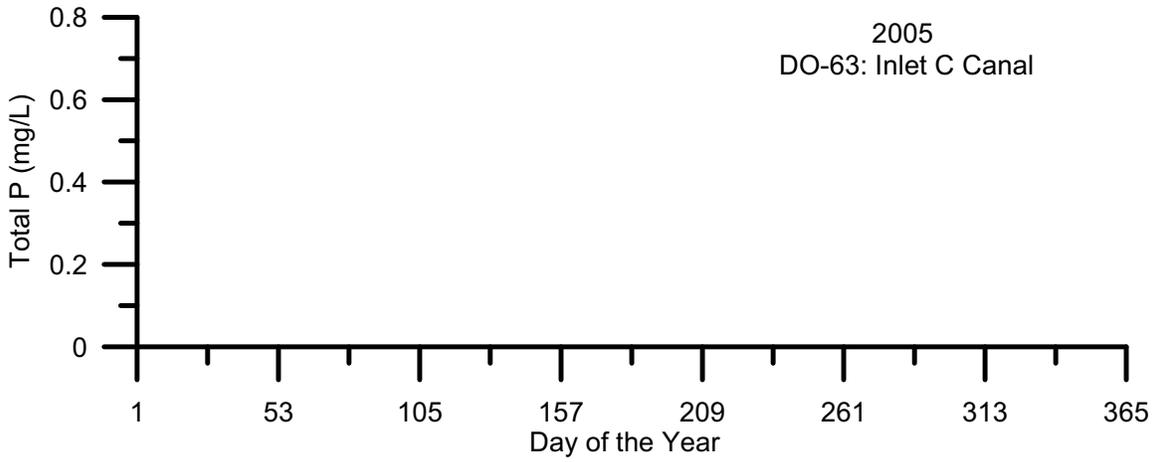


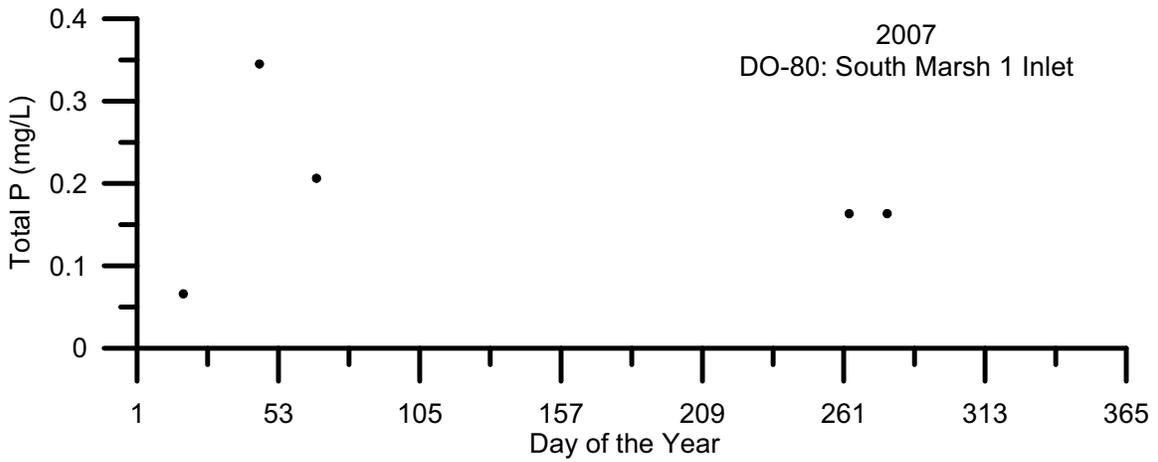
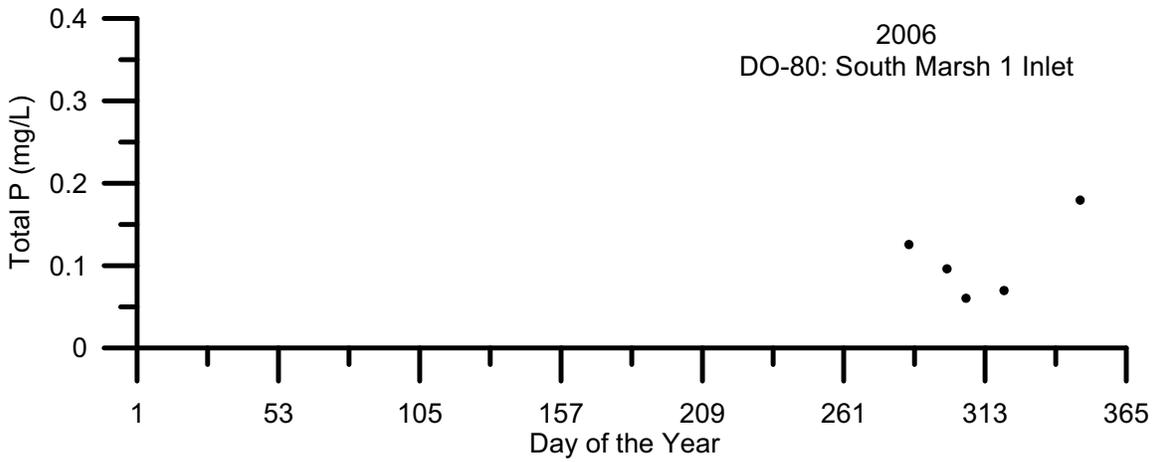
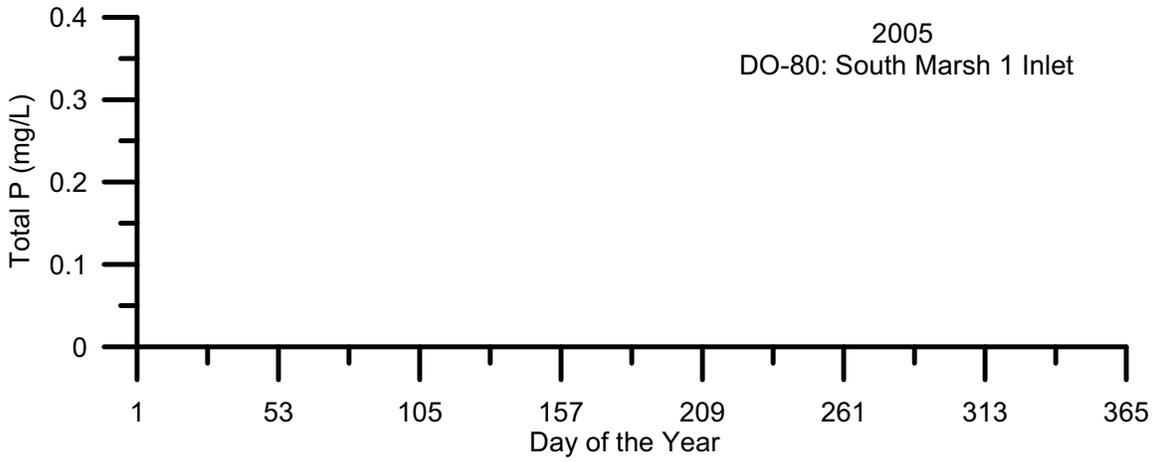


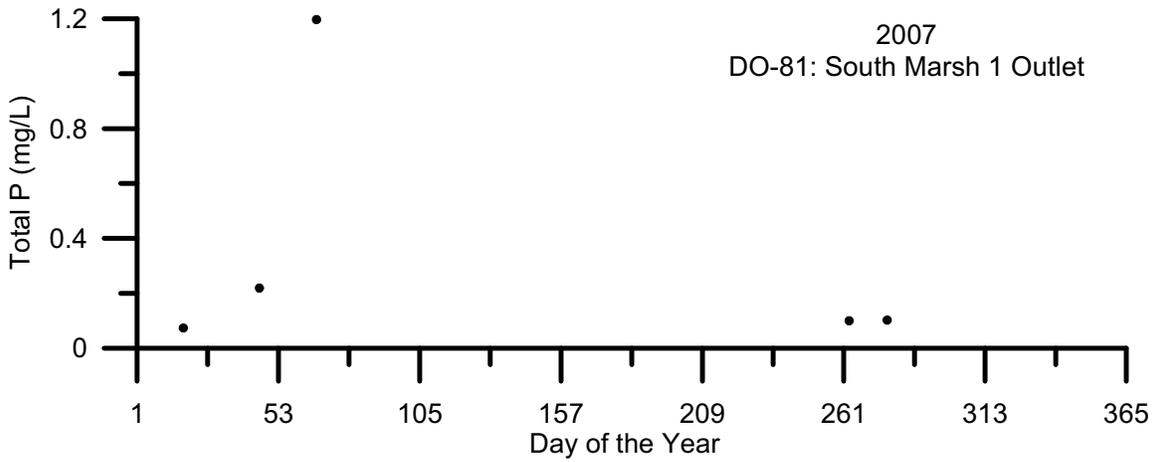
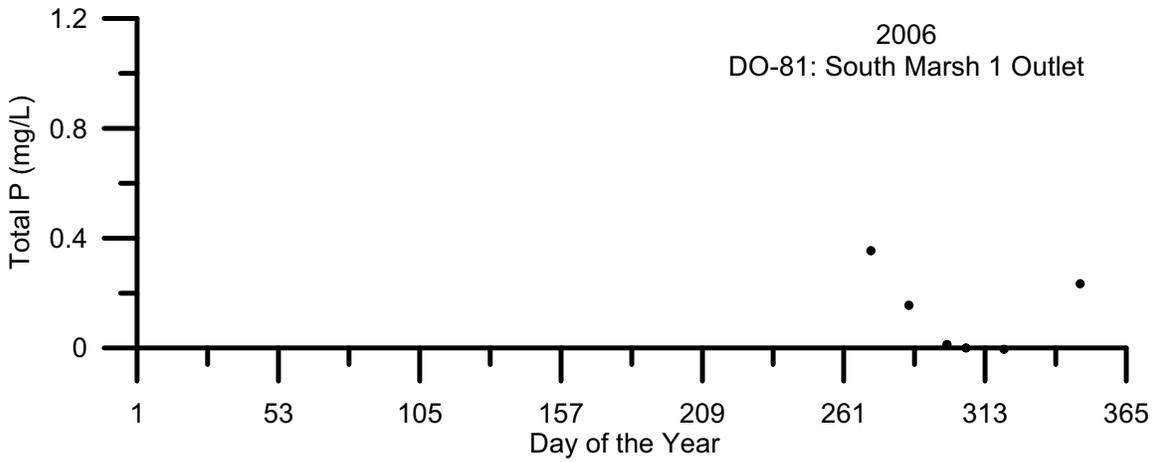
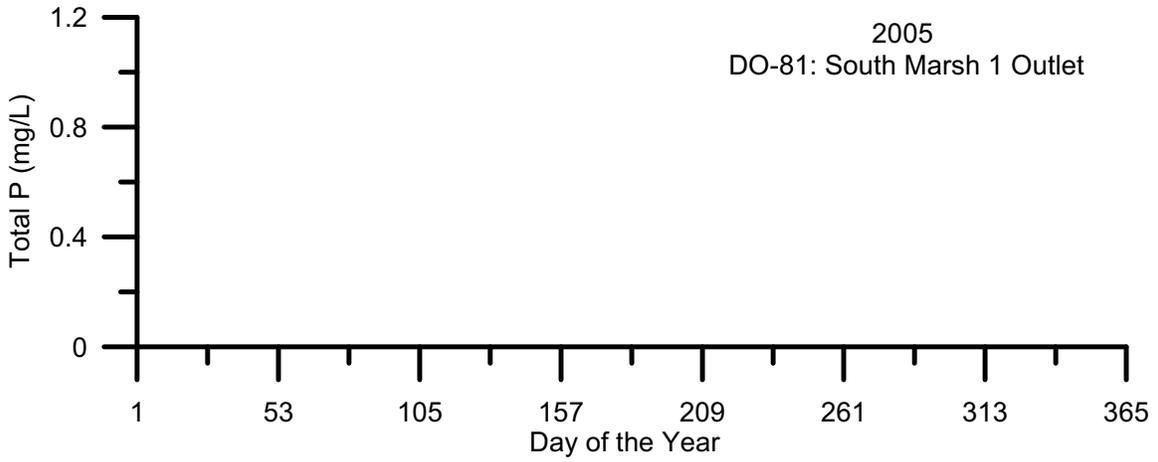


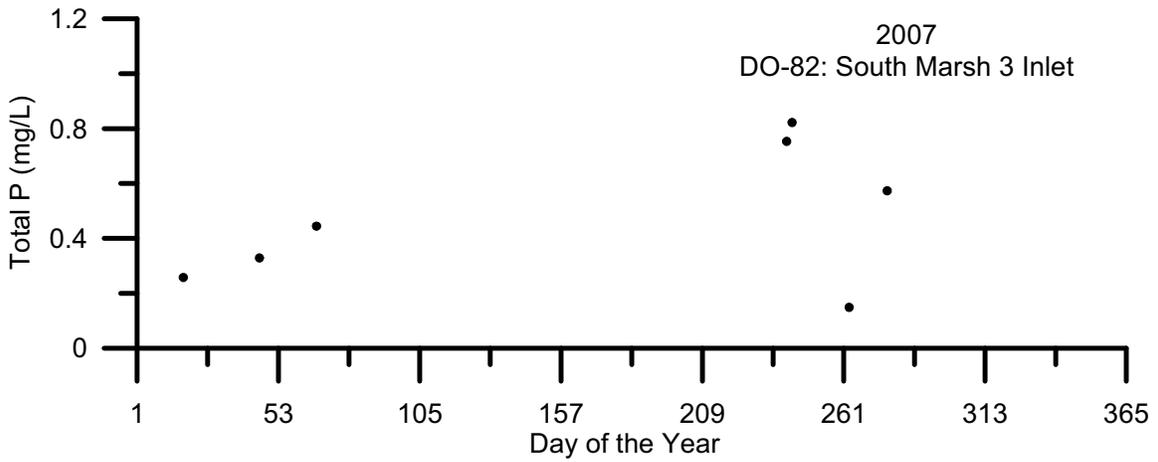
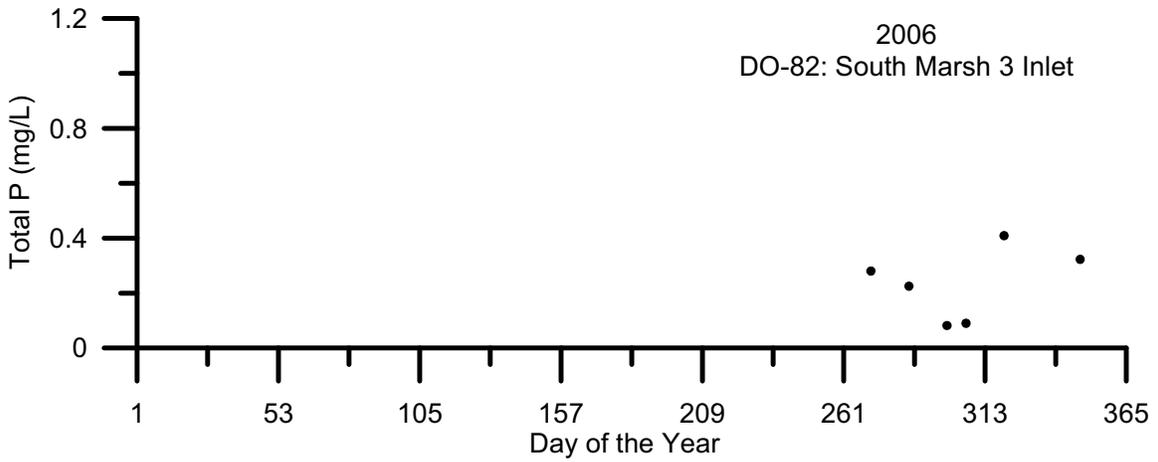
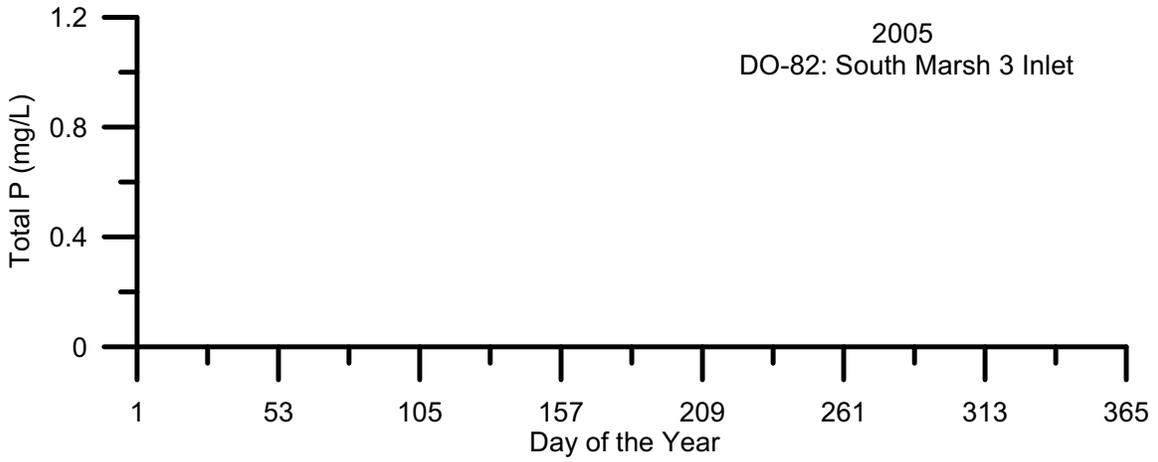


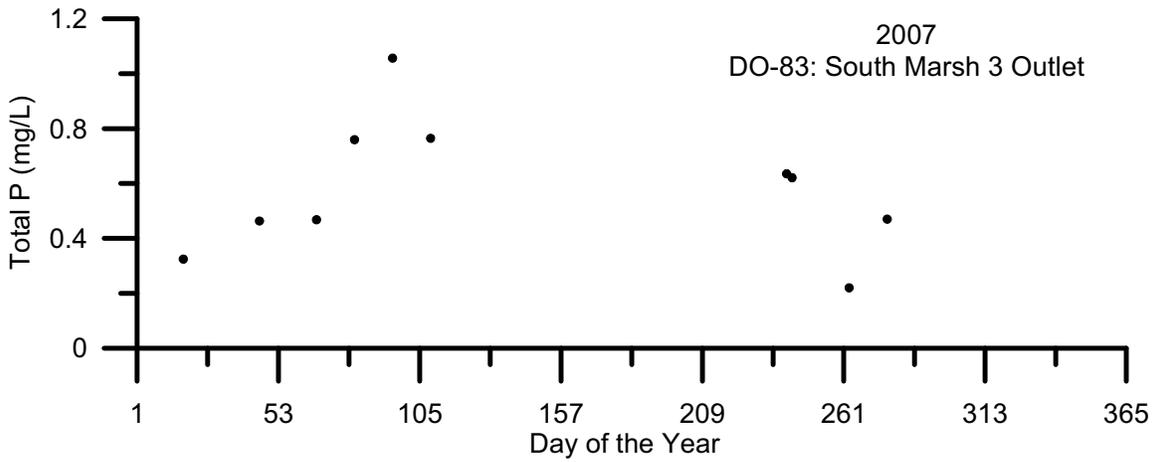
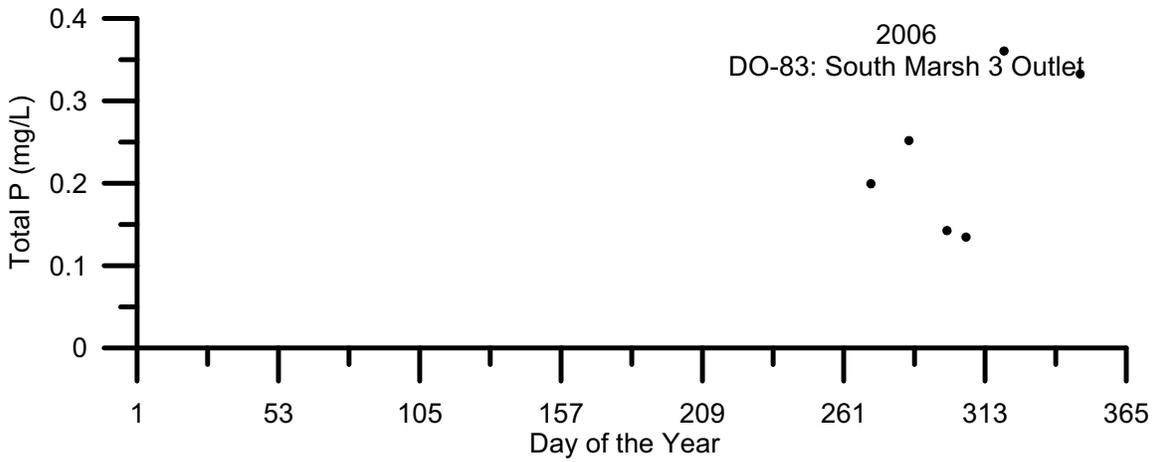
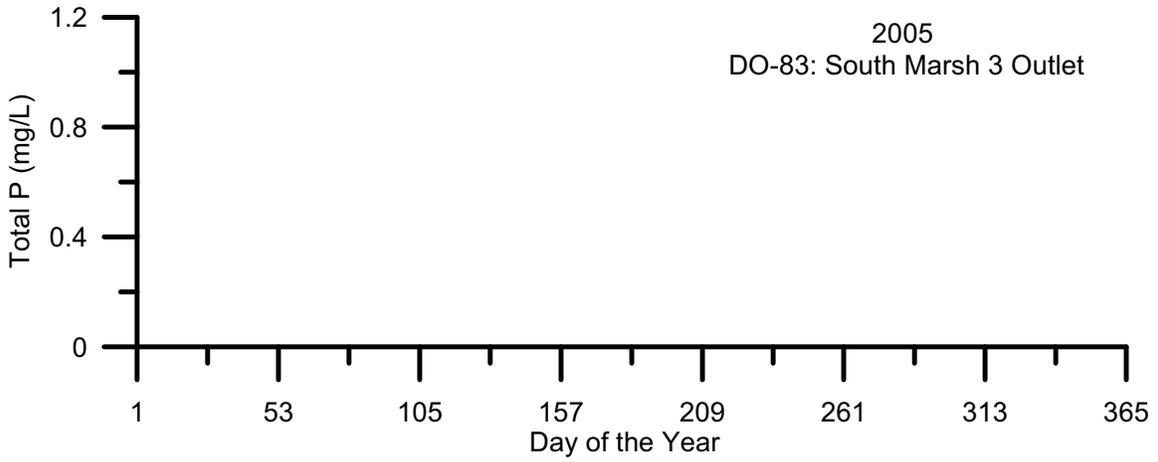


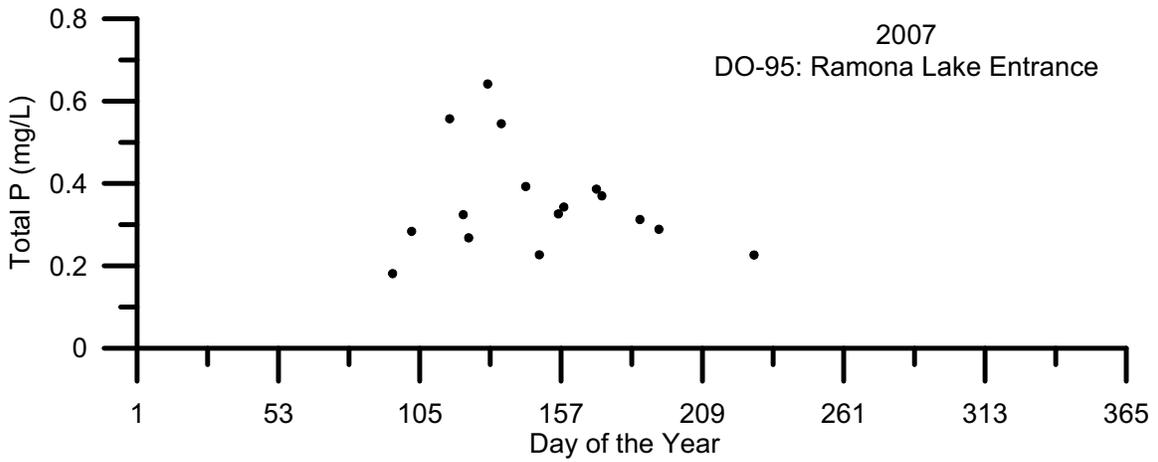
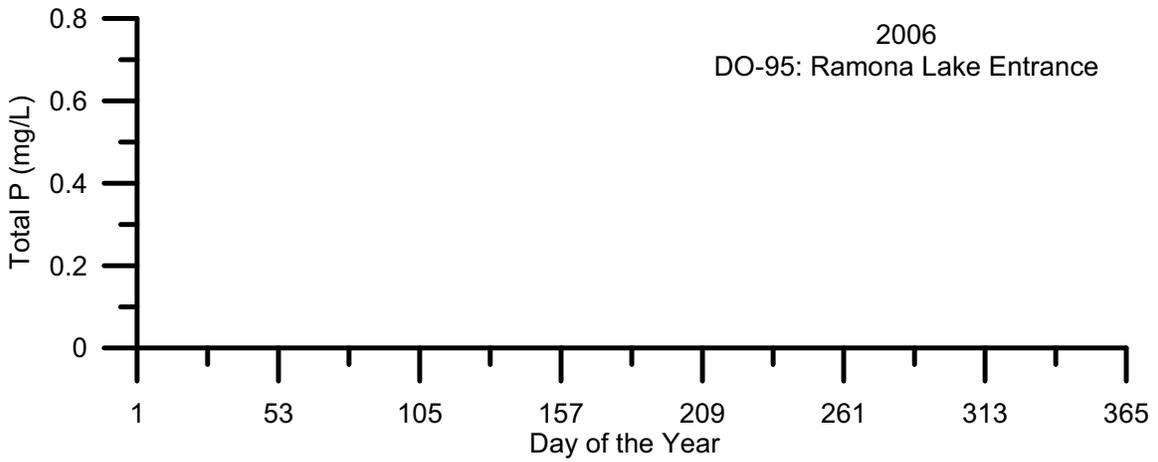
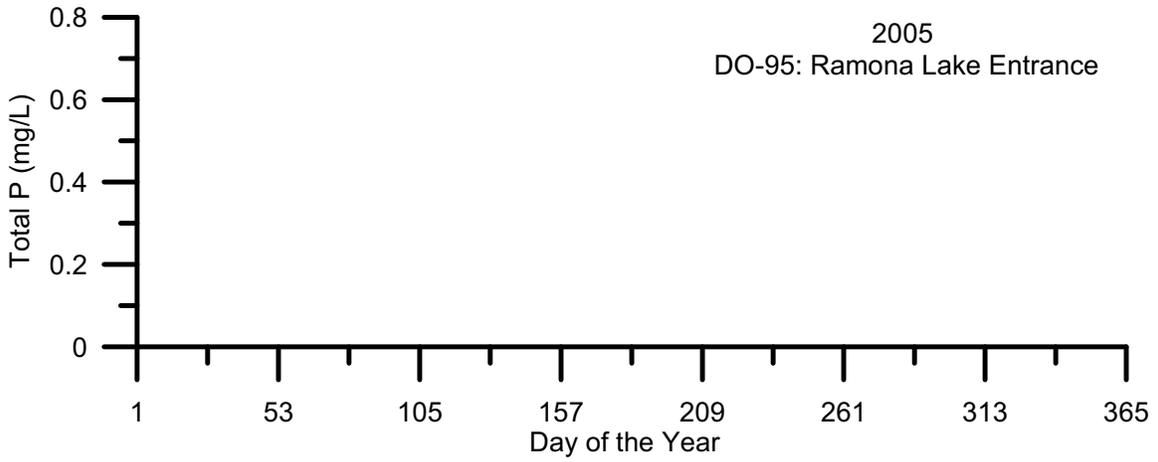














Temporal Plots of 2005-2007 Total Nitrogen Data from the Upstream San Joaquin River

Chelsea Spier¹
Remie Burks¹
Sharon Borglin¹
Randy Dahlgren²
Jeremy Hanlon¹
Justin Graham¹
William Stringfellow¹

February 2008

¹Environmental Engineering Research Program
School of Engineering & Computer Sciences
University of the Pacific
3601 Pacific Avenue, Sears Hall
Stockton, CA 95211

²UC Davis, Davis, CA

Introduction

The San Joaquin River (SJR) supports one of the most productive agricultural regions in the world and its productivity is heavily dependant on irrigated agriculture. A consequence of irrigated agriculture is the production of return flows conveyed down gradient drains that eventually discharge to surface waters. Agricultural drainage may have significant nutrient load and can impact algae growth and general water quality in the SJR. Individual farmers and agricultural organizations, such as drainage authorities, are in need of tools to manage the environmental impacts of agricultural activities (Stringfellow, 2008).

For the years 2005 through 2007, sites throughout the San Joaquin Valley watershed were sampled to assess the overall water quality in the region. One thousand nine hundred and ninety-six (1996) individual surface water samples were collected and analyzed and WQ was assessed at 113 locations in the SJR basin (Borglin et al., 2008). Samples were processed and analyzed by the Environmental Engineering Research Program (EERP) laboratory at the University of the Pacific as well as at the University of California, Davis, Dahlgren Lab. This report includes temporal plots of total nitrogen (Tot-N) data analyzed by the EERP laboratory starting in the summer of 2007.

Methods

Depth integrated field samples were collected during 2005-2007 in the upper San Joaquin River in accordance with EERP Field Standard Operating Procedures Protocol Book (Graham 2008). Water samples were collected in glass 1000 mL bottles (Wheaton Science Products, Millville, NJ), 1000 mL HDPE Trace-Clean narrow mouth plastic bottles (VWR International), 250 mL HDPE Trace-Clean wide mouth plastic bottles (VWR International) as well as 40 mL trace clean vials with PTFE septa (ICChem, Rockwood, TN). Bottles were labeled with the appropriate sample number, site name and sampling date. All bottles were rinsed with sample water prior to sample collection. Some sites required a bucket to collect sample water because of accessibility from a high bridge or platform. For these sites, the bucket was pre-rinsed with sample water and sample bottles were filled using a rinsed funnel. Care was taken to distribute water simultaneously to all sample bottles (rather than sequentially). Samples were immediately stored at 4 °C after sampling (cooler temperature was recorded in the lab upon delivery) and transported to the EERP lab on the day of sampling.

Within 24 hours of collection samples were aliquotted and stored at -20°C until Tot-N analysis could be completed.

Starting in 2007 Tot-N was quantified using the TL-2800 ammonia analyzer made by Timberline Instruments (Boulder, CO). All N in a water sample was converted to NO₃-N by the addition of equal parts of unfiltered sample and digestion reagent under high heat and pressure. NO₃-N in the digested sample was mixed with caustic solution (to pH 11-13) and was then passed through a reducing zinc cartridge to convert all NO₃-N to NH₃-N. NH₃ (g) diffused across a gas-permeable membrane and was dissolved in a buffer solution. The dissolution of NH₃ (g) in the buffer causes a change in conductivity which can be correlated to the concentration of NH₃. The Timberline instrument automates the

mixing of the caustic solution with the sample, reduction of NO₃ to NH₃ by passing the sample through the zinc cartridge, and pumping of the buffer solution through the gas permeable membrane (Borglin et al, 2008) and (Carlson, 1990). The reportable limit for this method is 0.15 mg/L Tot-N.

Results/Discussion

With each set of Tot-N field samples analyzed in the EERP laboratory, quality assurance samples including a lab duplicate, field duplicate, matrix spike, matrix spike duplicate, calibration check standards, laboratory control standard, trip blank, and lab blanks were also analyzed. 100% of all quality assurance samples were within a passing range (Spier, 2008). Proficiency check samples, standards with unknown concentration to the laboratory analyst, were run approximately twice a year. Four proficiency check samples were analyzed for Tot-N in the EERP laboratory during 2007, and all of these samples were found to be within the acceptable range. Samples were measured ranging from 0.03-42.27 mg/L Tot-N. The average concentration of Tot-N in samples collected was 5.09 mg/L Tot-N. Tot-N was also analyzed at UC Davis on all of the same water samples and has a high correlation to values measured by EERP. When all data points measured by the two labs are compared they have $r^2=0.9896$ (Spier, 2008). Tot-N samples measured by EERP have about 101% as much Tot-N as the same samples measured by UCD (Figure 2).

EERP used to measure Tot-N Using a Tekmar Teledyne Apollo 9000 TOC Combustion Analyzer. It was found that this machine did not measure all forms of N, and had a very low correlation to Tot-N measured at UCD so it is no longer used by the EERP laboratory to measure Tot-N. These temporal plots (Figures 3-103) as well as the plot comparing EERP's laboratory data to UCD's data (Figure 2) created an easy visual way to find outliers and double check data entry for possible mistakes. For the purpose of these plots any data points that were slightly negative were changed to 0.

References

- American Public Health Association (APHA). 2005. Standard Methods for the Examination of Water and Wastewater, 21st Edition. American Public Health Association, Washington, DC.
- Borglin, S., W. Stringfellow, J. Hanlon. 2005. Standard Operating Procedures for the Up-Stream Dissolved Oxygen TMDL Project. LBNL/Pub-937.
- Borglin, S., Burks, R., Hanlon, J., Graham, J., Spier, C., Stringfellow, W., and Dahlgren, R., (2008) Methods overview, quality assurance, and quality control, University of the Pacific, Stockton, CA
- Borglin, S.E., Burks, R.D., Hanlon, J.S., Stringfellow, W.T. (2008) EERP Lab Protocol Book, University of the Pacific, Stockton, CA.

- Carlson, R.M., R. Cabrera, J. Paul, J. Quick and R.Y. Evans. 1990. Rapid direct determination of ammonium and nitrate in soil and plant tissue extracts. *Comm. Soil Sci. Plant Anal.* 21:1519-1530.
- Graham, J., Hanlon, J.S., Stringfellow, W.T., (2008) *EERP Field Protocol Book*, University of the Pacific, Stockton, CA.
- Spier, C., Burks, R., Borglin, S.E. (2008) *Comparison of Methods for Analyzing Ammonia, Nitrate and Total Nitrogen*, University of the Pacific, Stockton, CA.
- Stringfellow, W.T., et al., (2008) *Evaluation of Vegetated Ditches, Ponds, and Wetlands as BMPs for Mitigating the Water Quality Impact of Irrigated Agriculture in the San Joaquin Valley*, University of the Pacific, Stockton, CA
- YSI Environmental Operations Manual (2005) *6-Series Environmental Monitoring Systems*, Yellow Springs, OH.

Table 1: EERP Sampling Site List

DO Number	Site Name	Type
4	SJR at Mossdale	Core sites
5	SJR at Vernalis-McCune Station (River Club)	Core sites, BMP
6	SJR at Maze	Core sites, BMP
7	SJR at Patterson	Core sites, BMP
8	SJR at Crows Landing	Core sites, BMP
10	SJR at Lander Avenue	Core sites
12	Stanislaus River at Caswell Park	Core sites
14	Tuolumne River at Shiloh Bridge	Core sites
16	Merced River at River Road	Core sites
18	Mud Slough near Gustine	Core sites, Wetland
19	Salt Slough at Lander Avenue	Core sites, Wetland
20	Los Banos Creek Flow Station	Core sites, Wetland
21	Orestimba Creek at River Road	Core sites, BMP
23	Modesto ID Lateral 5 to Tuolumne	Core sites
25	Modesto ID Main Drain to Stan. R. via Miller Lake	Core sites
28	Turlock ID Westport Drain Flow station	Core sites
29	Turlock ID Harding Drain	Core sites
30	Turlock ID Lateral 6 & 7 at Levee	Core sites
33	Hospital Creek	Intermittent, BMP
34	Ingram Creek	Core sites, BMP
36	Del Puerto Creek Flow Station	Core sites, BMP
44	San Luis Drain End	Core sites
53	Salt Slough at Wolfsen Road	Wetland
57	Ramona Lake Drain	Core sites, BMP
59	SJR Laird Park	Core sites
60	Moffit 1 South	Wetland
61	Deadman's Slough	Wetland
62	Mallard Slough	Wetland
63	Inlet C Canal	Wetland
80	South Marsh-1-Inlet	Wetland
81	South Marsh-1-Outlet	Wetland
82	South Marsh-3-Inlet	Wetland
83	South Marsh-3-Outlet	Wetland
95	Ramona drain at Ramona Lake	BMP, Intermittent

Figure 1: EERP Sampling Site Map of SJR Watershed and Tributaries

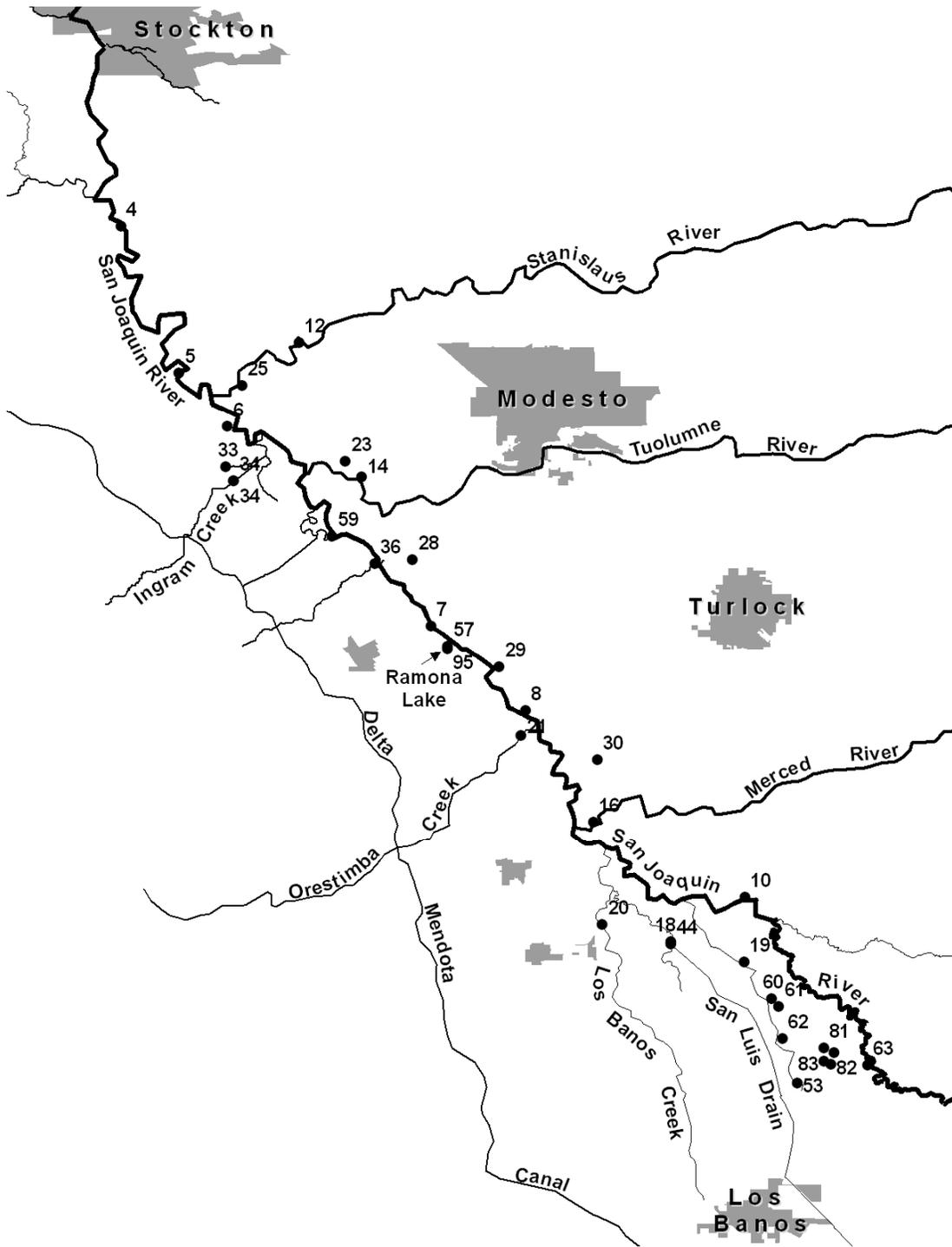
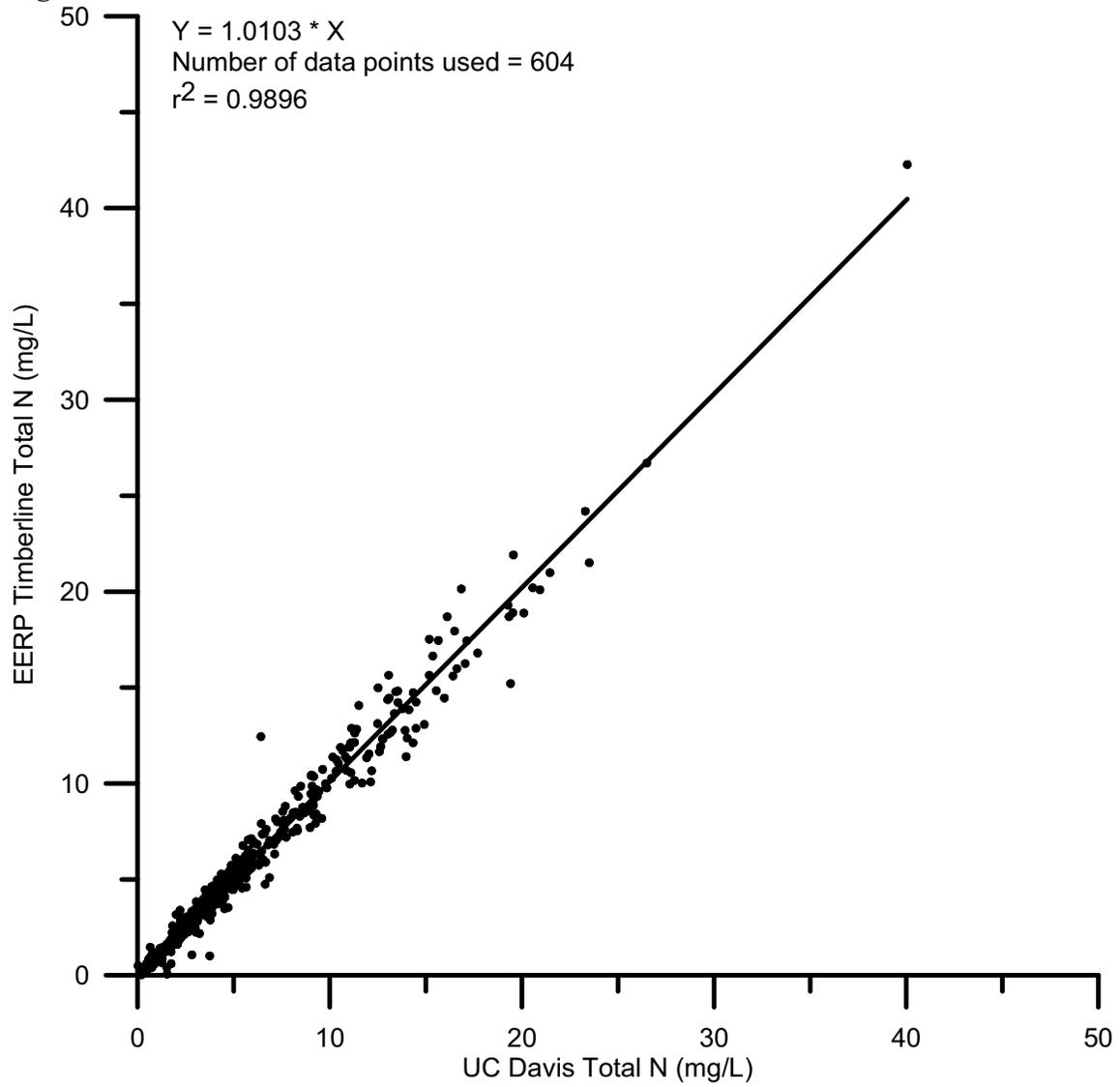
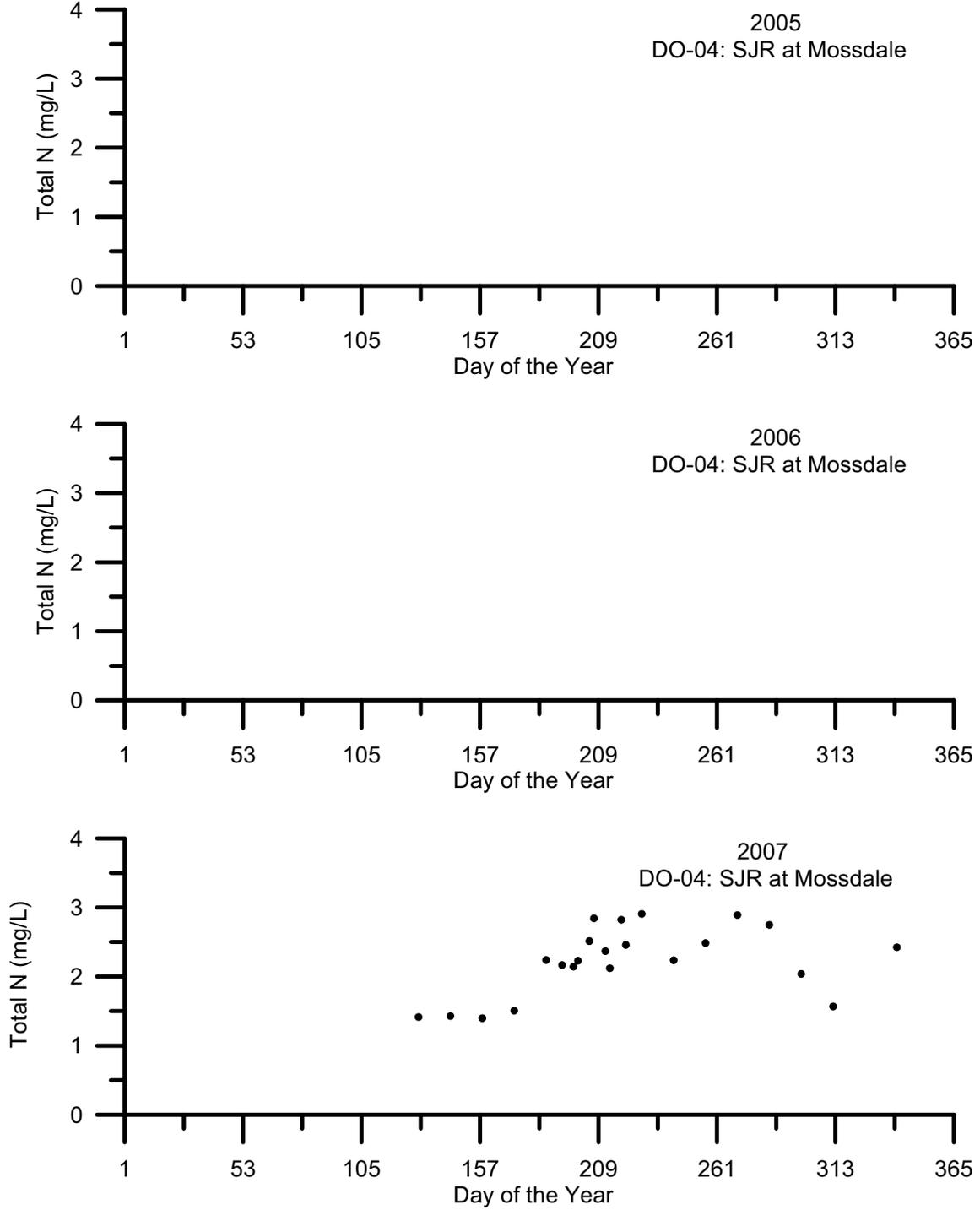
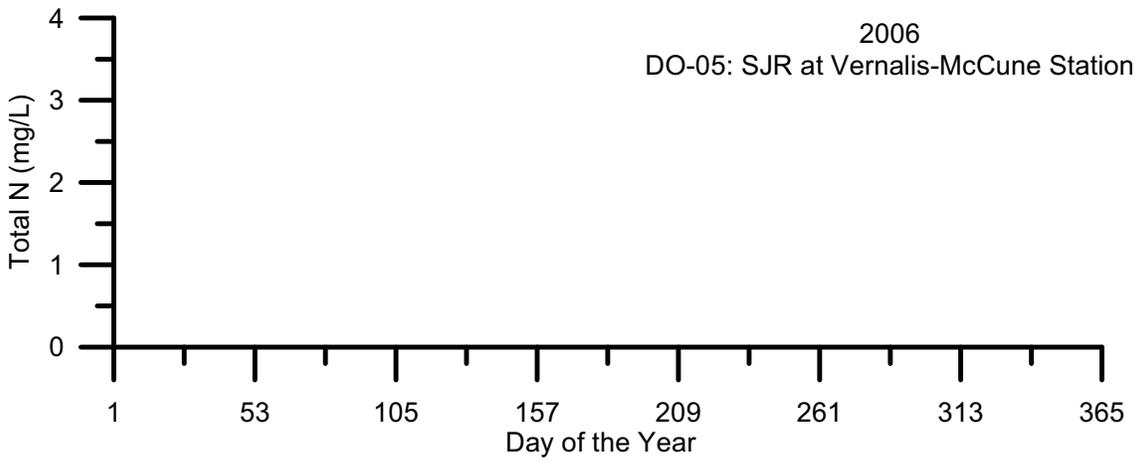
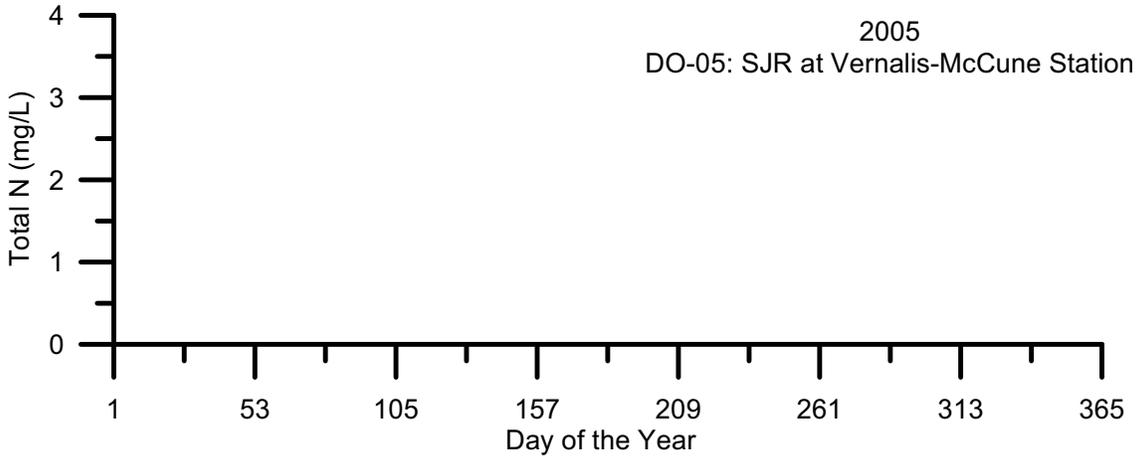


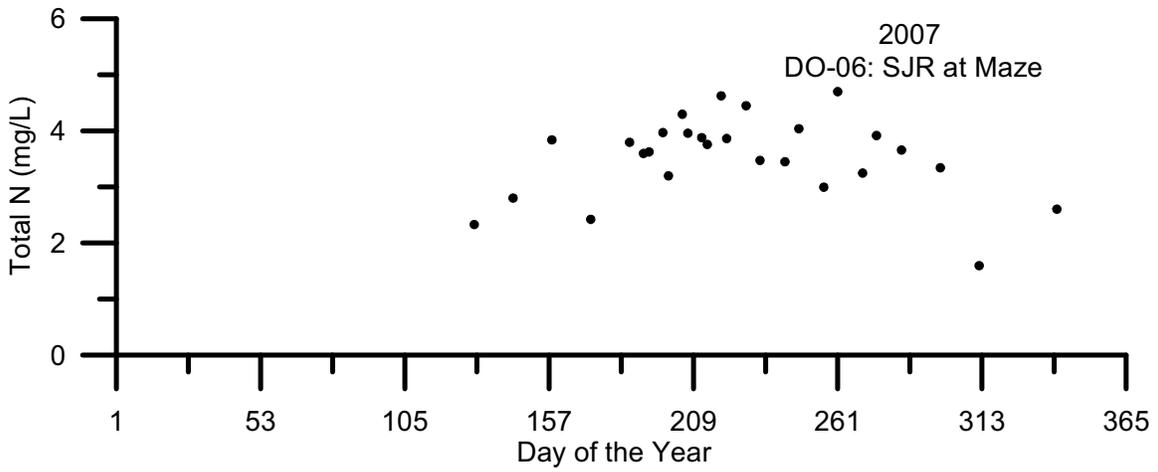
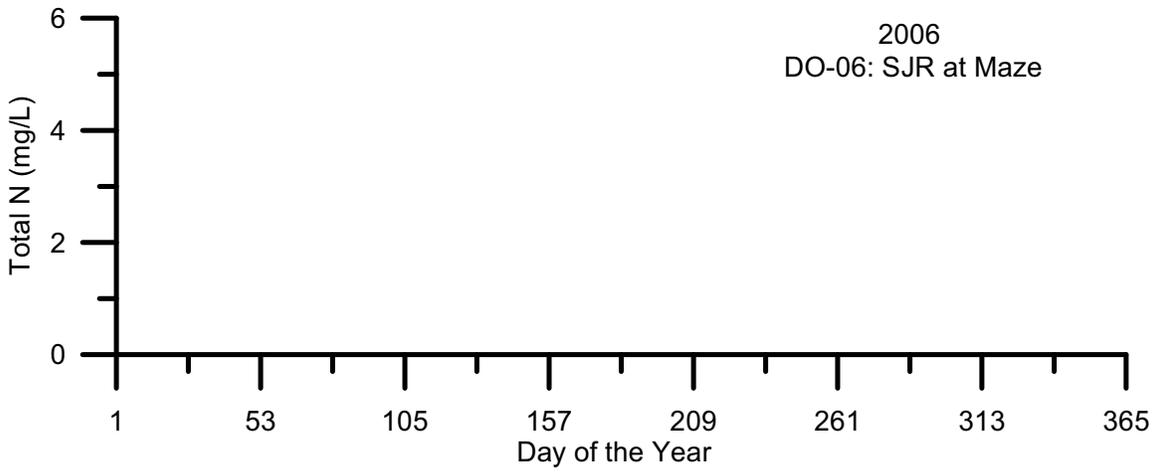
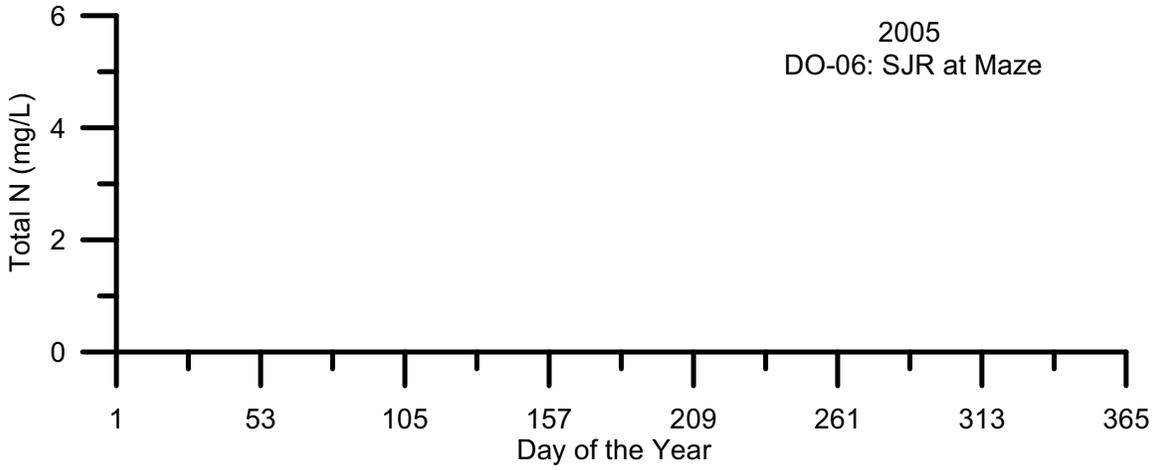
Figure 2: UCD vs EERP Timberline Plot of Tot-N.

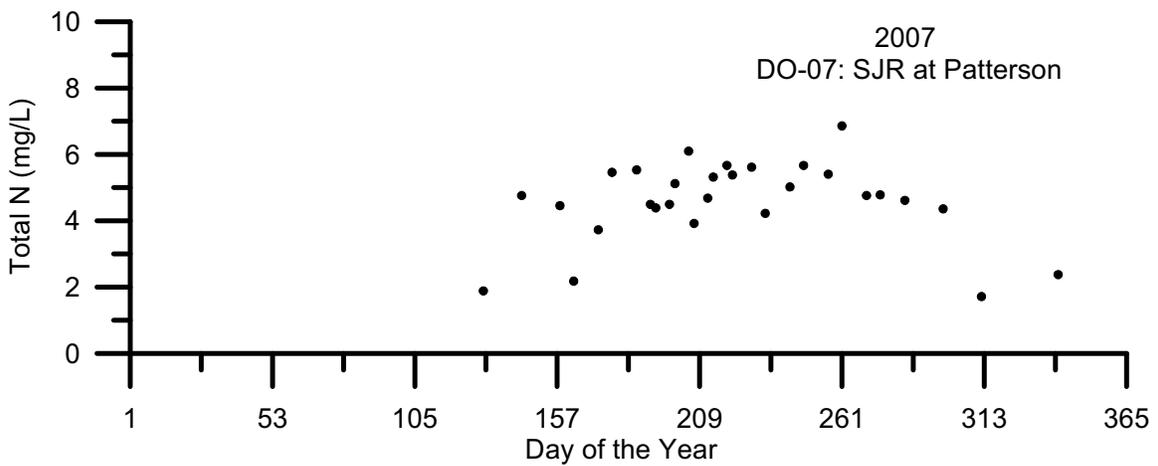
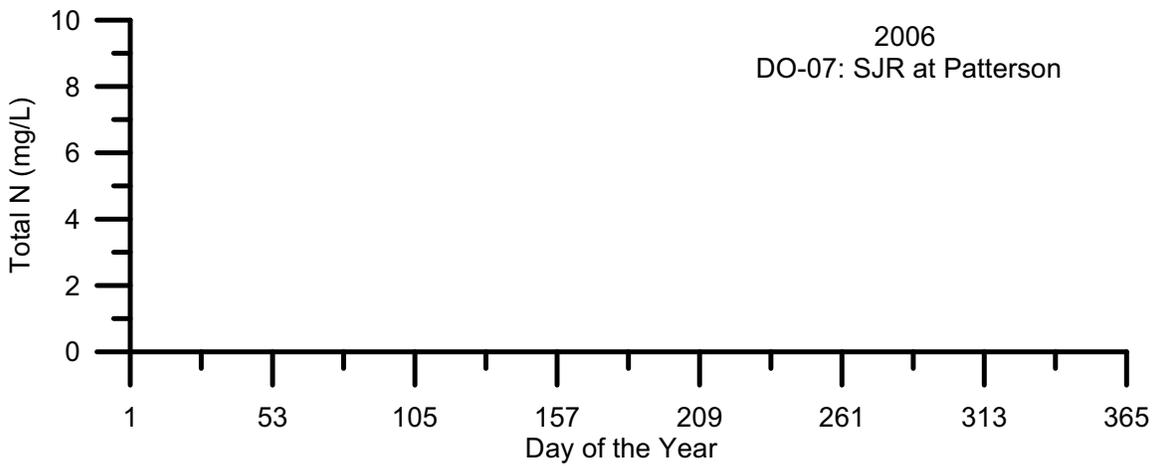
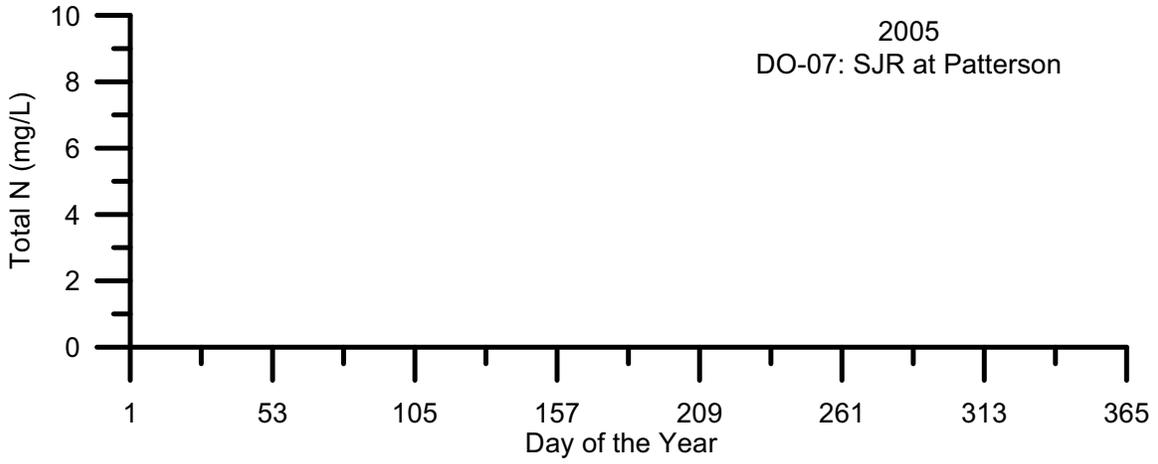


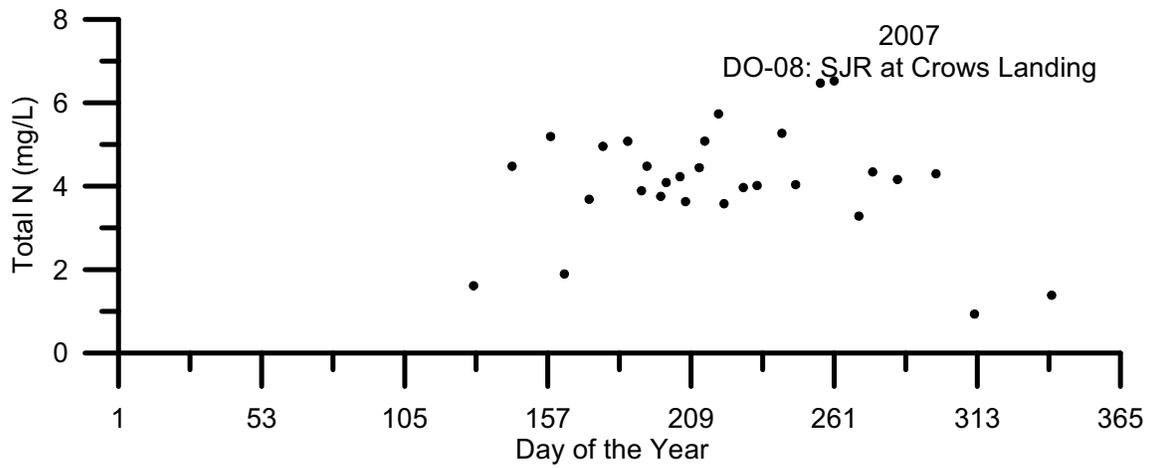
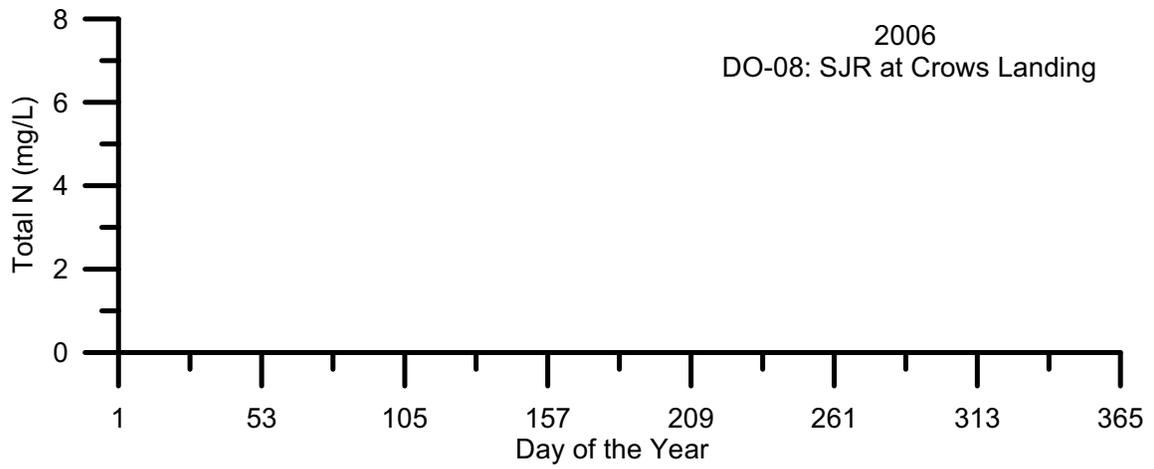
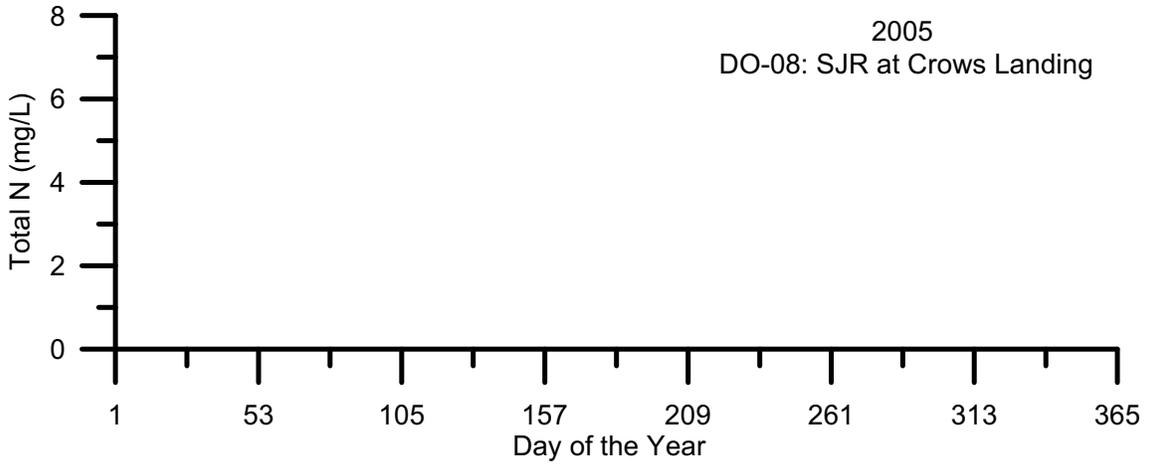
Figures 3 -104: Temporal Plots of Tot-N By Site ID

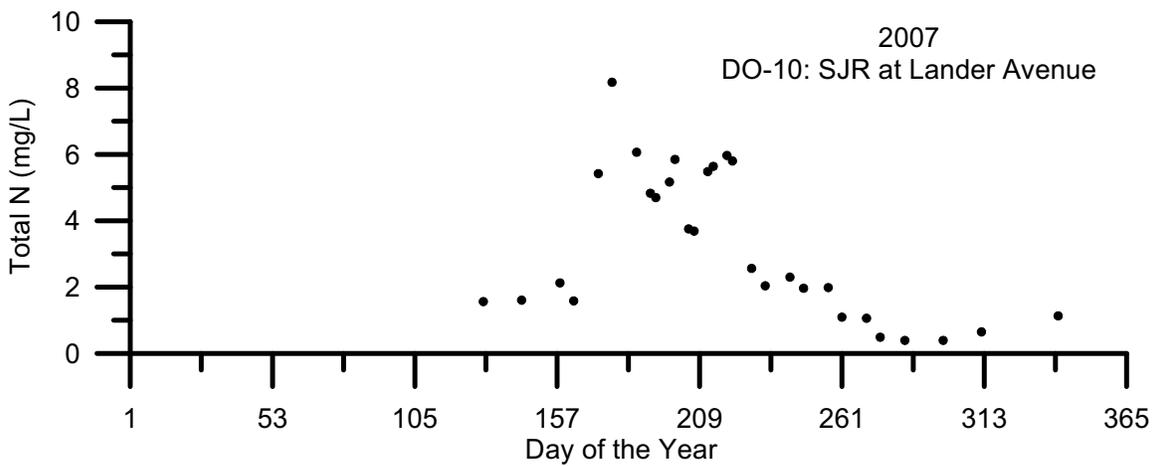
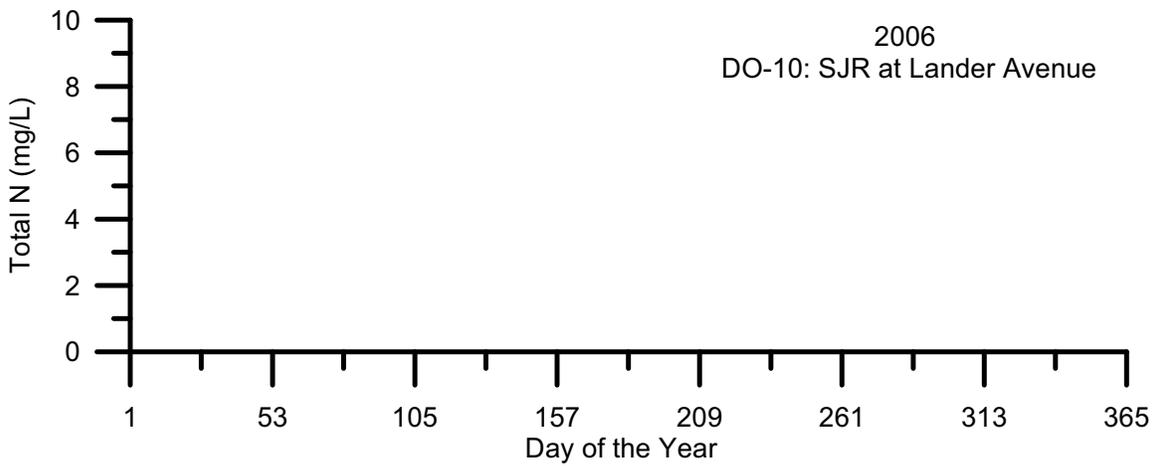
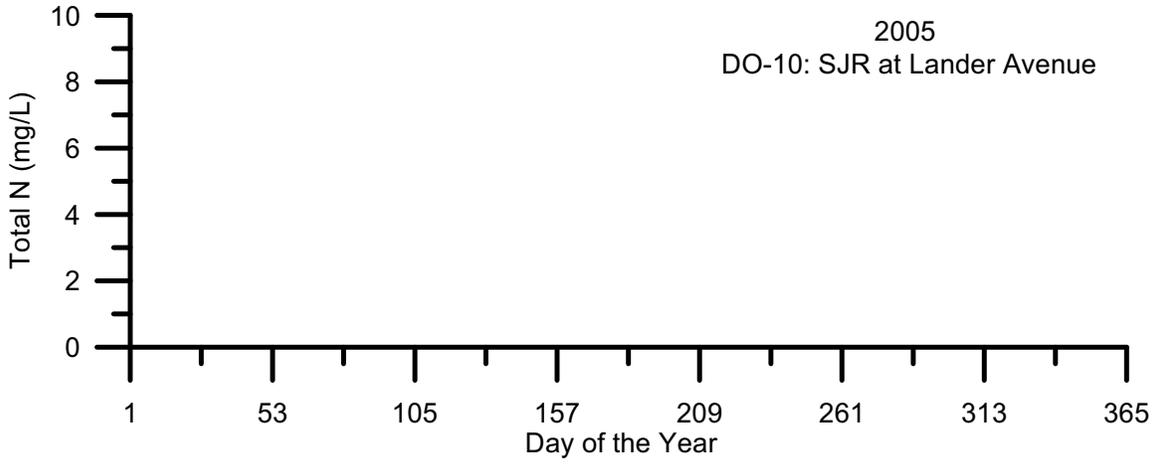


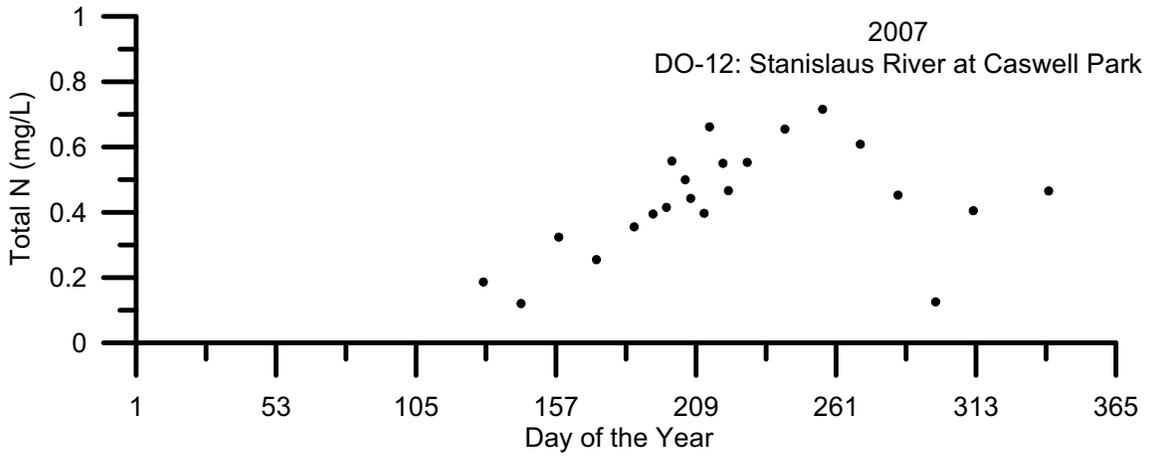
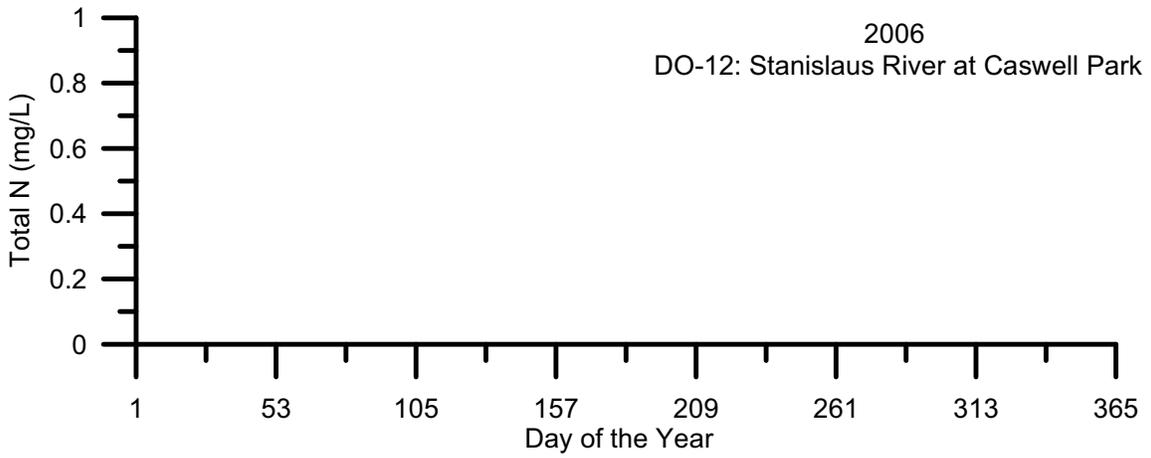
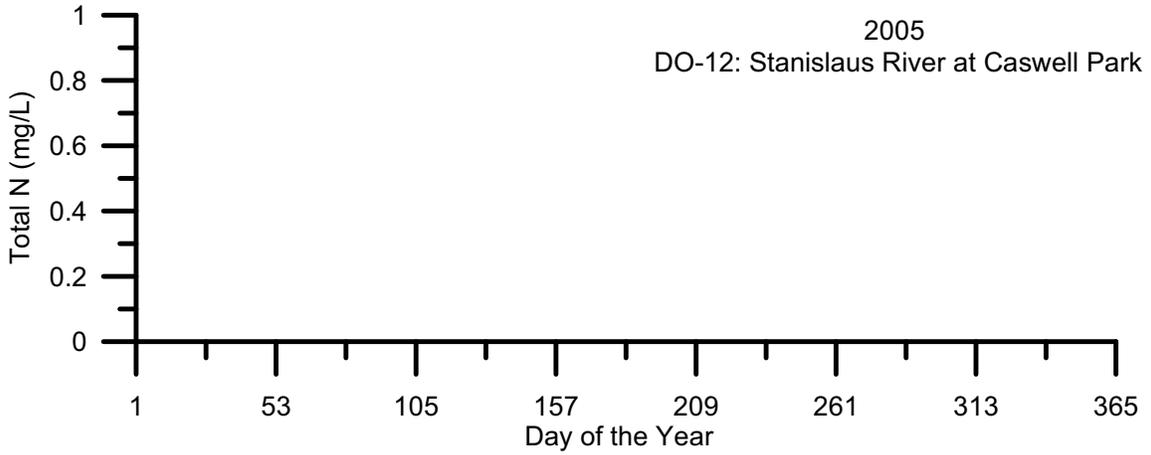


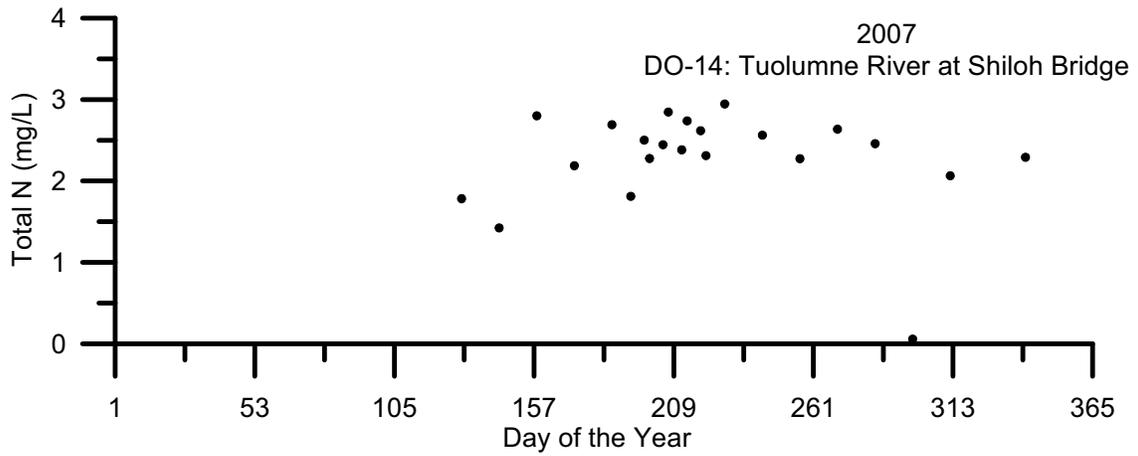
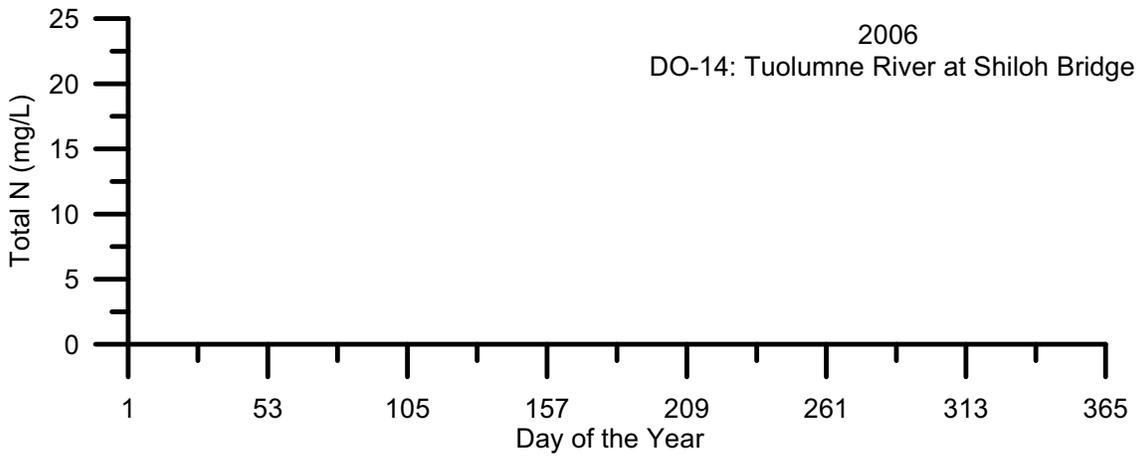
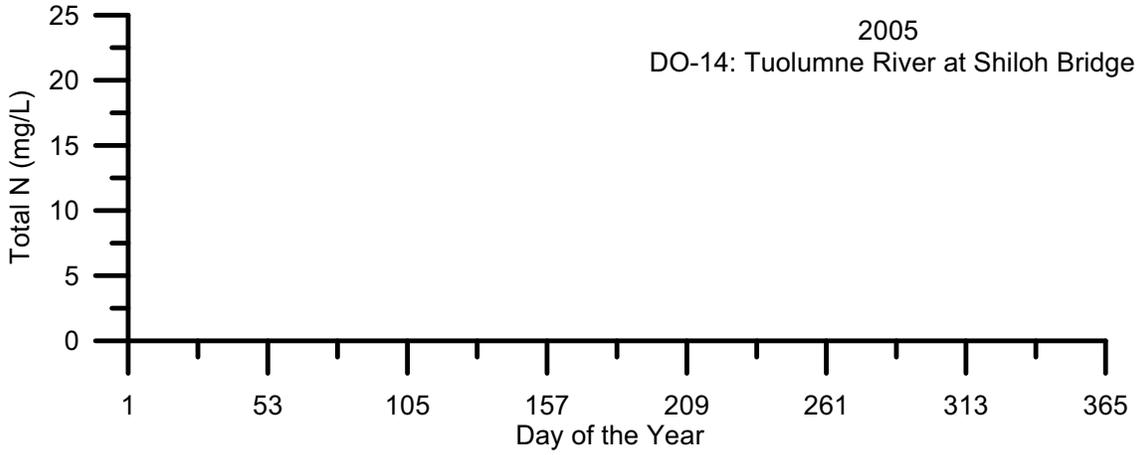


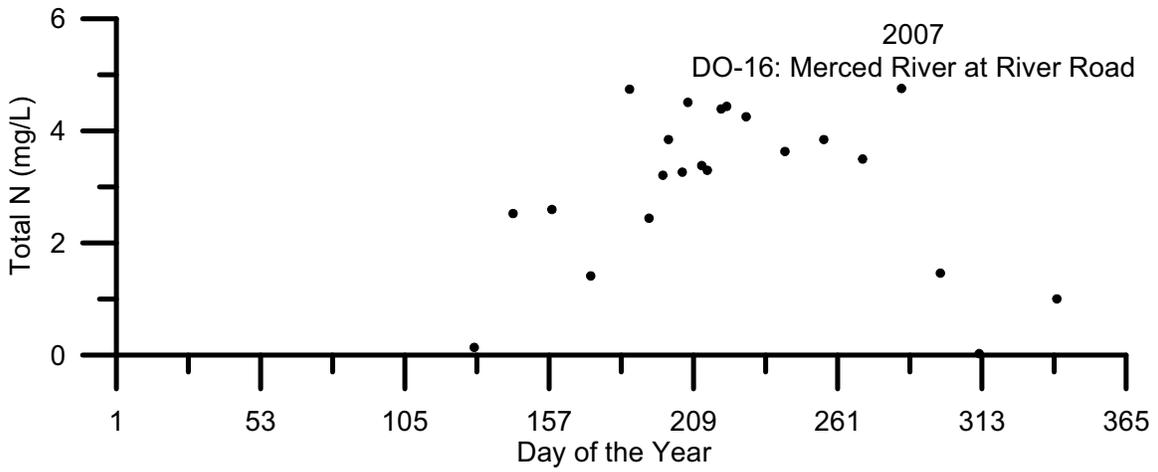
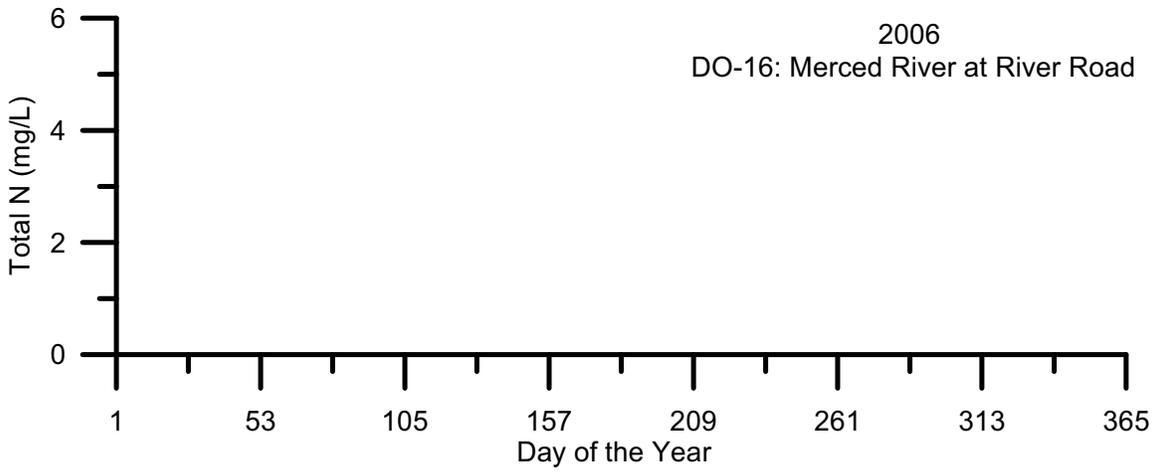
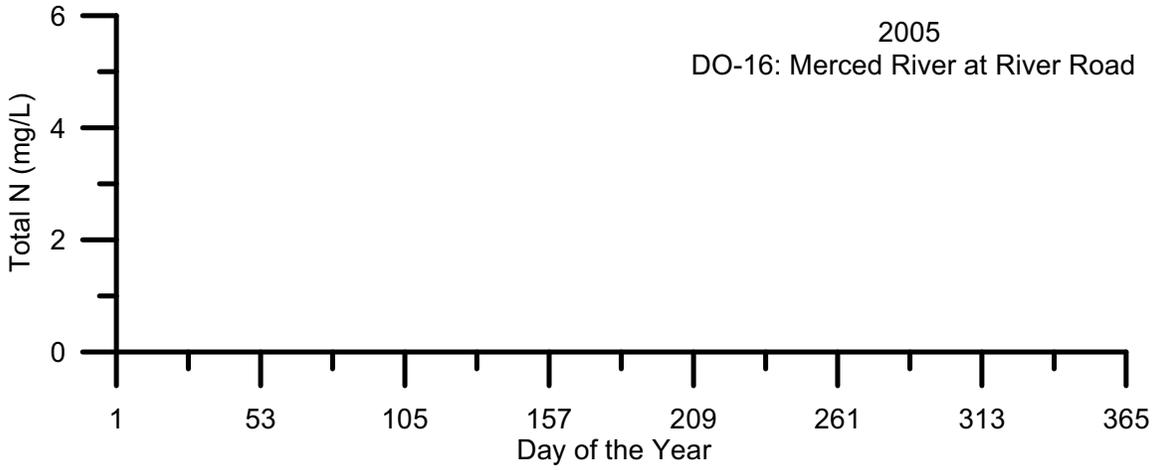


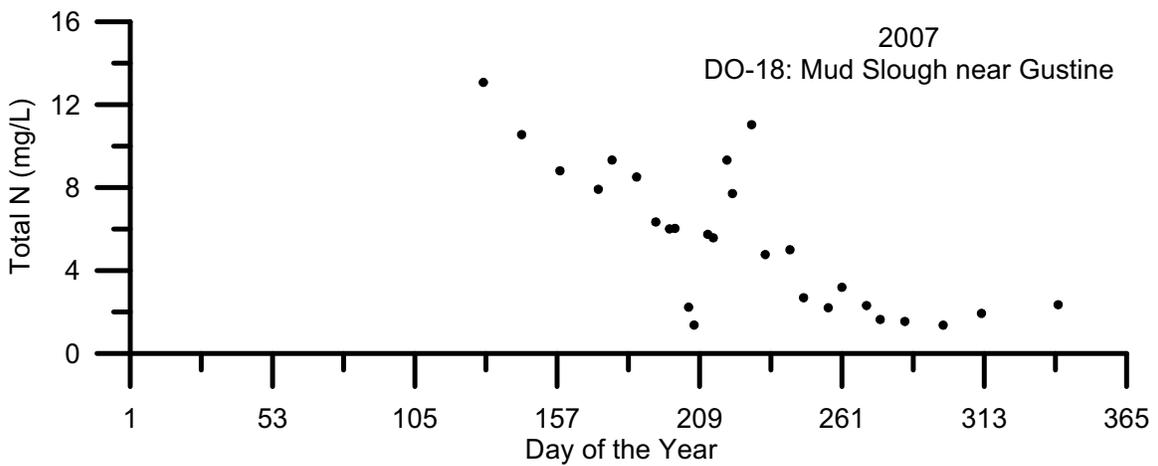
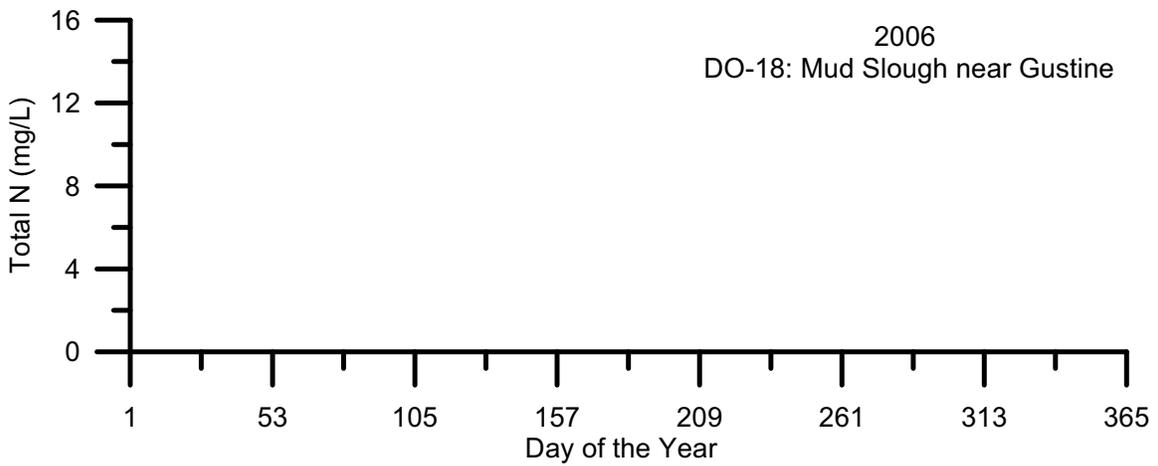
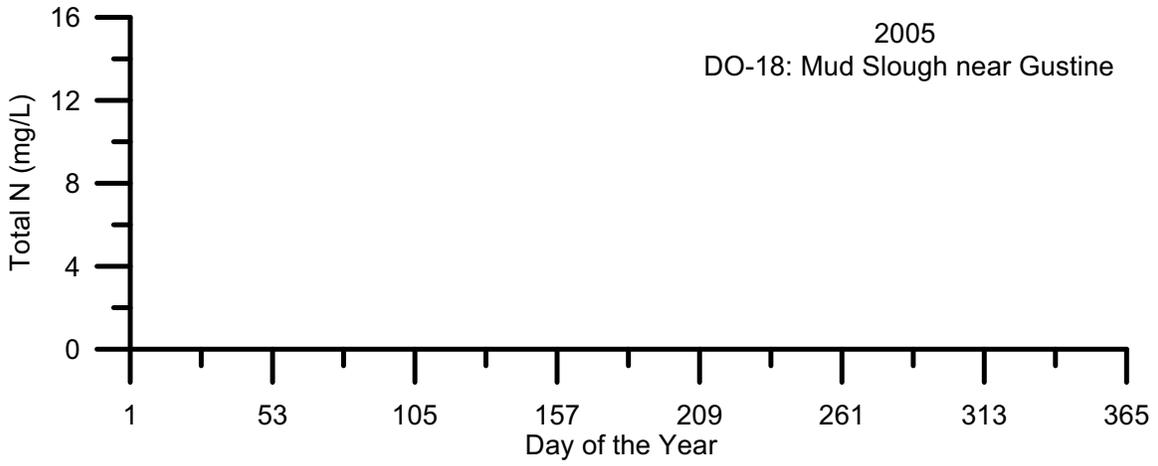


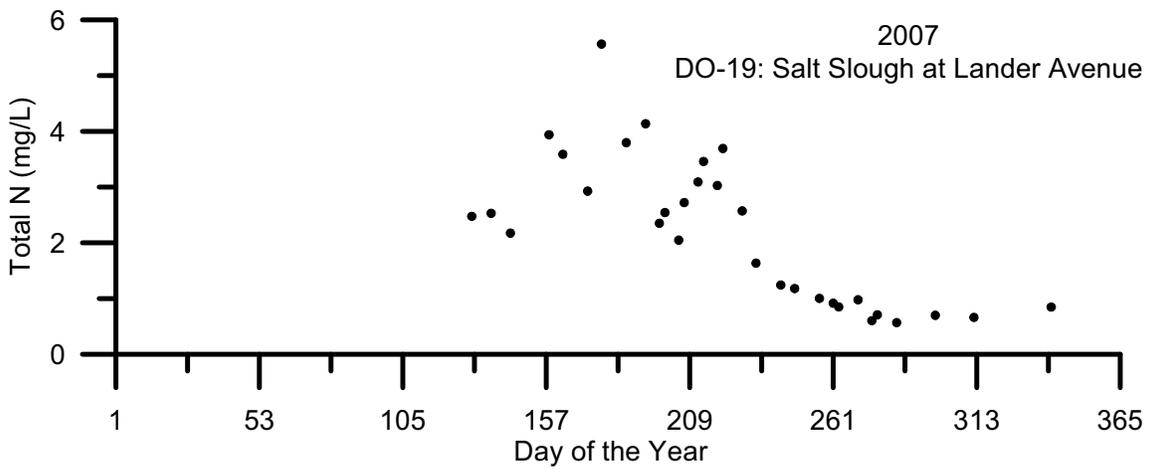
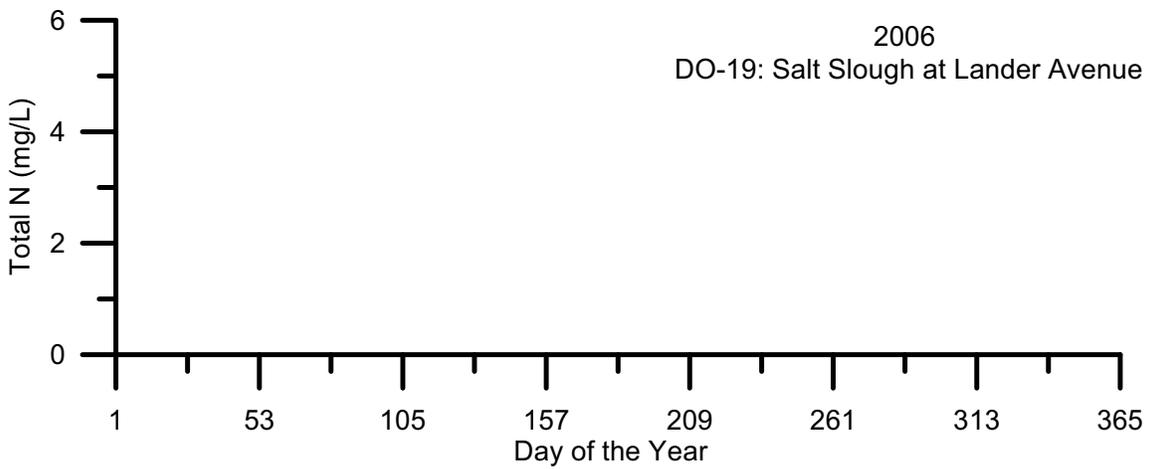
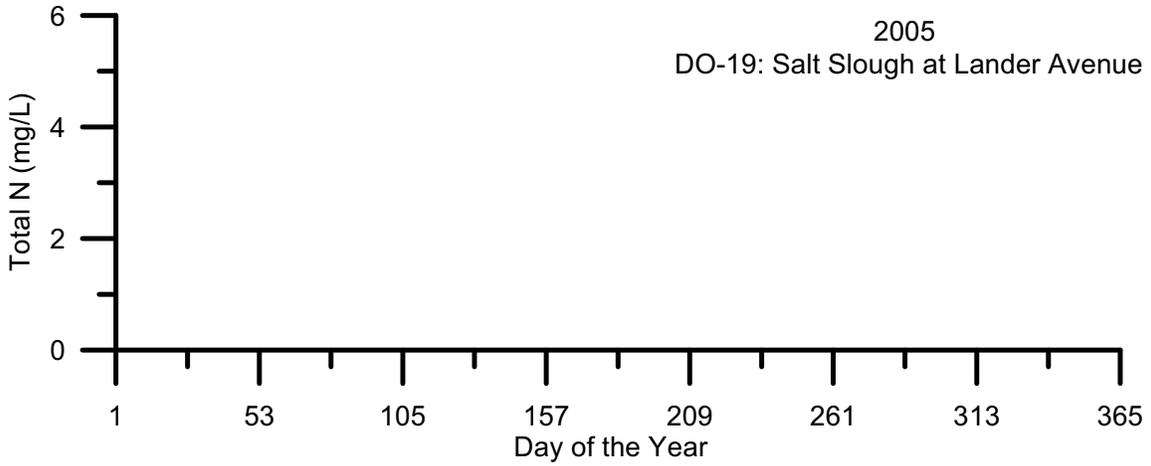


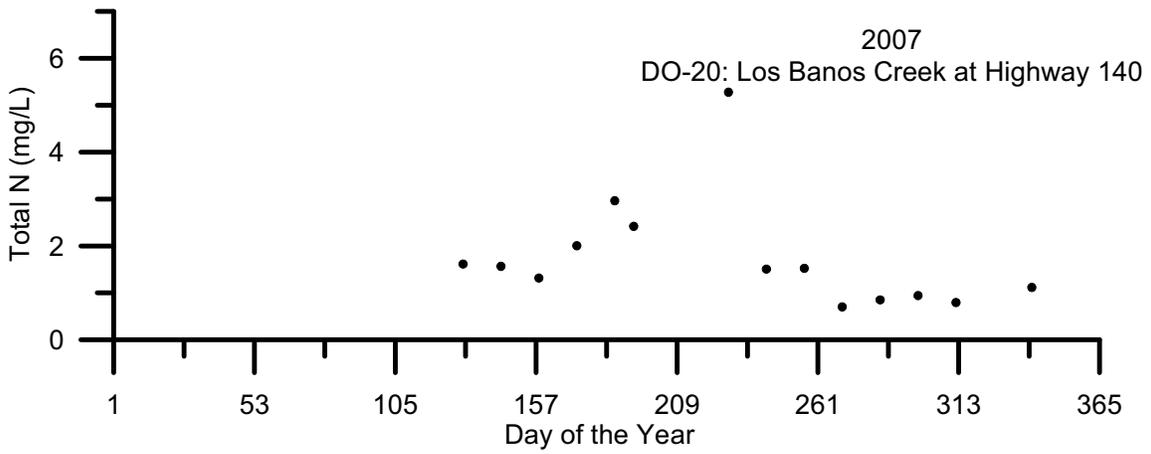
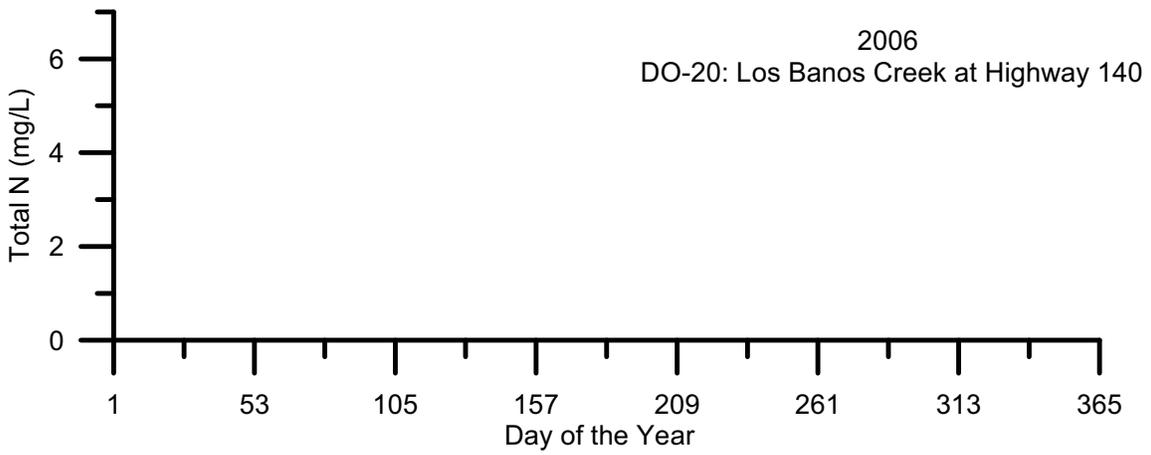
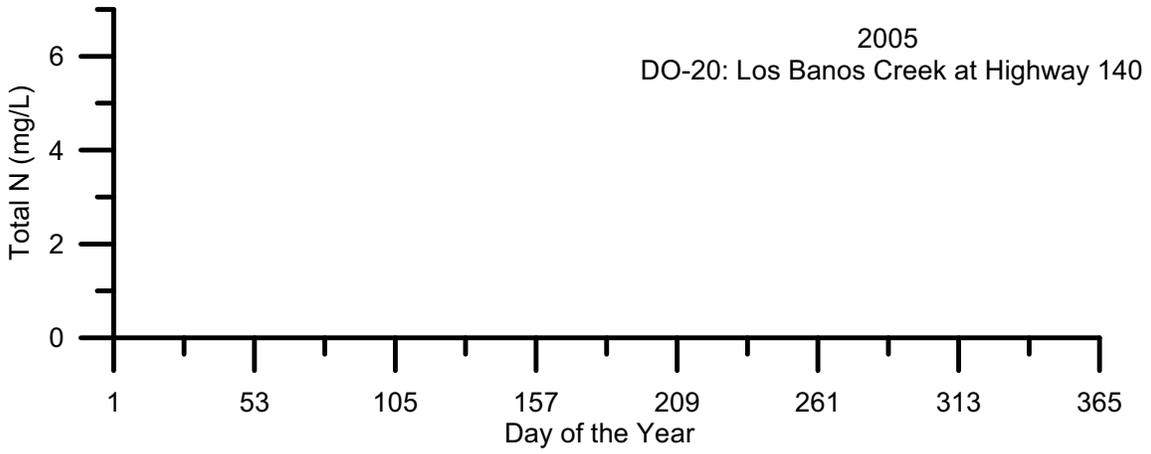


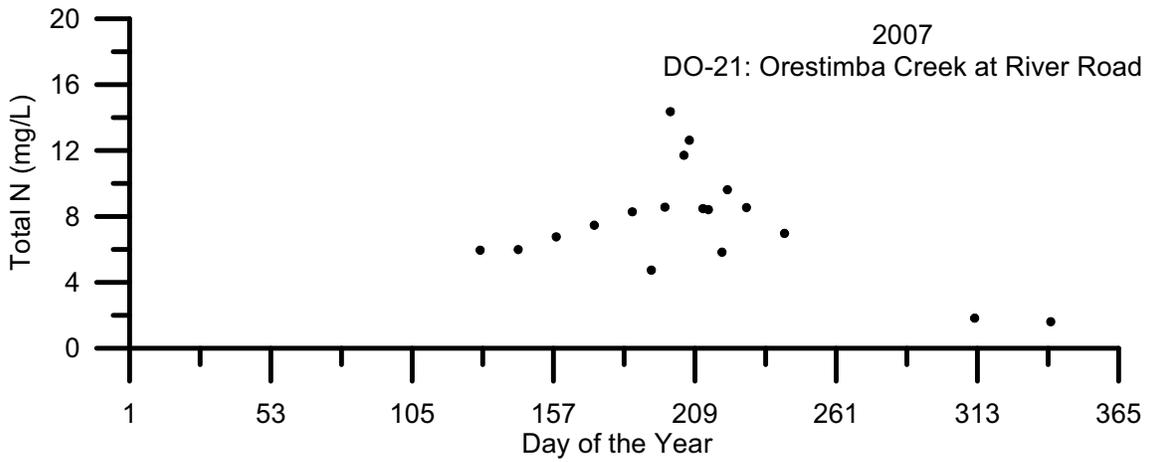
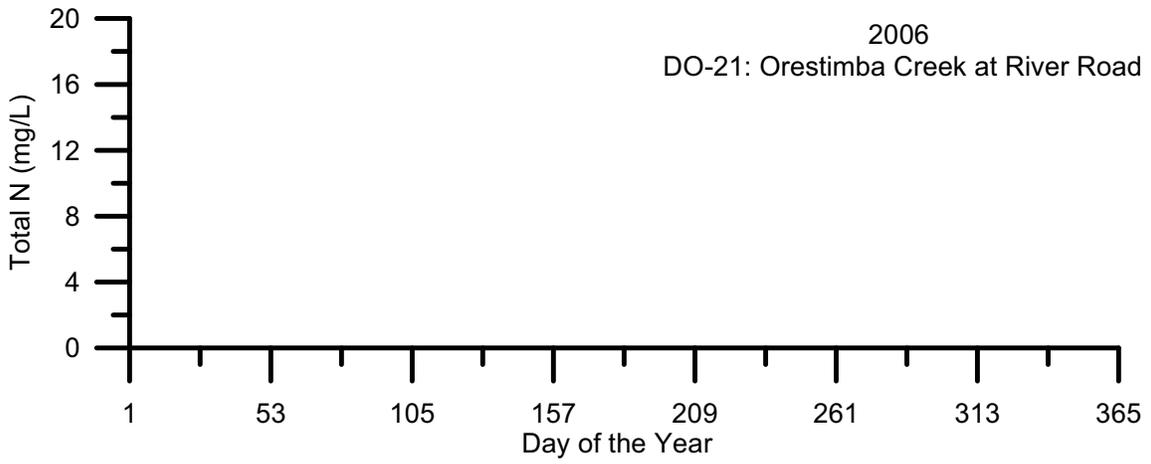
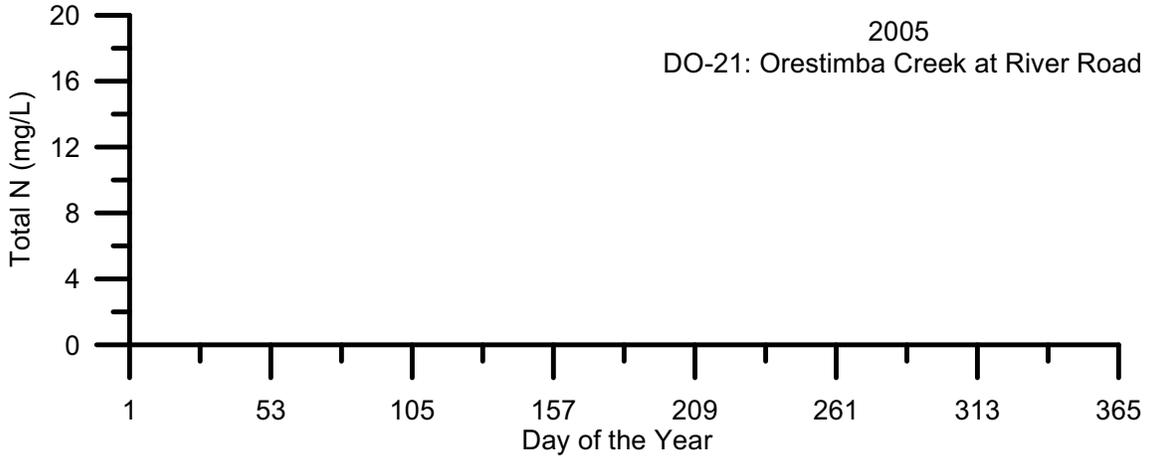


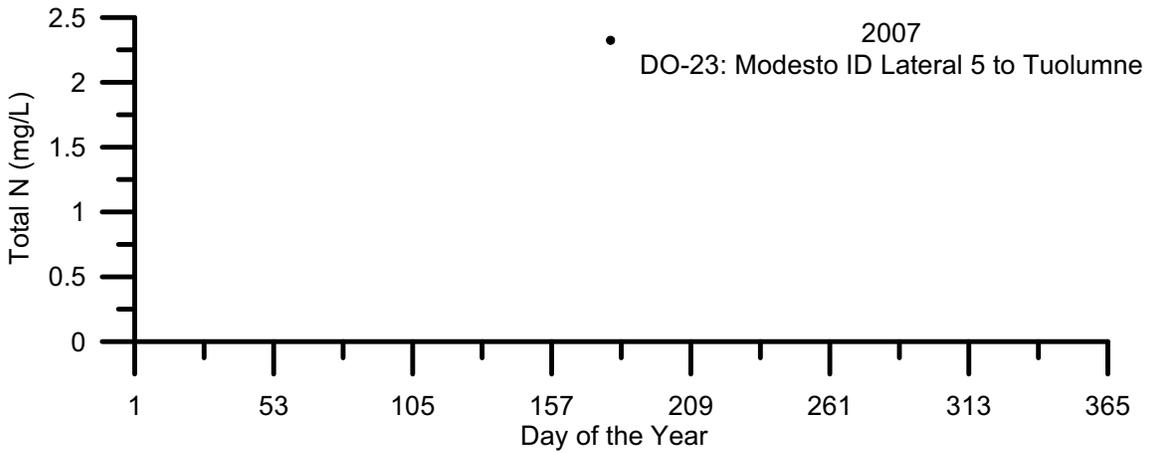
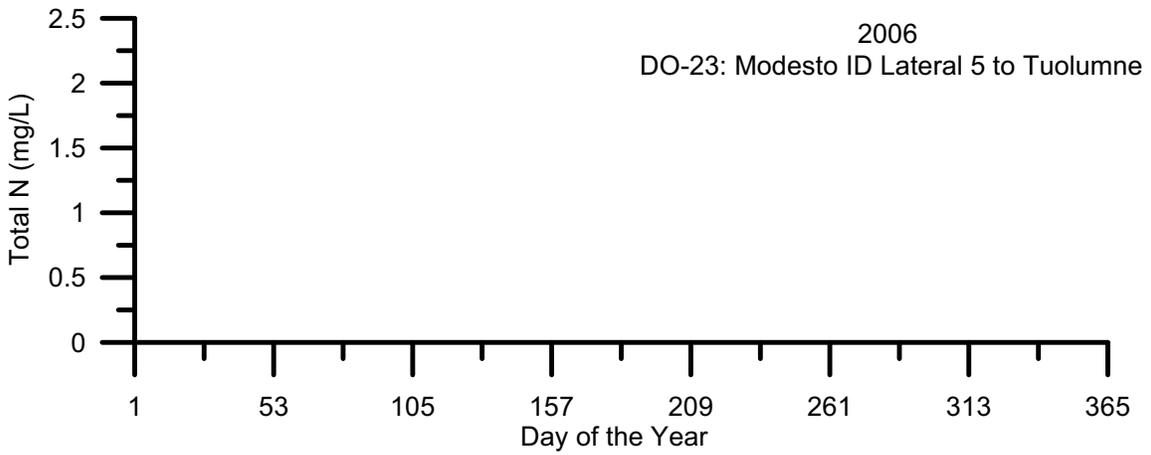
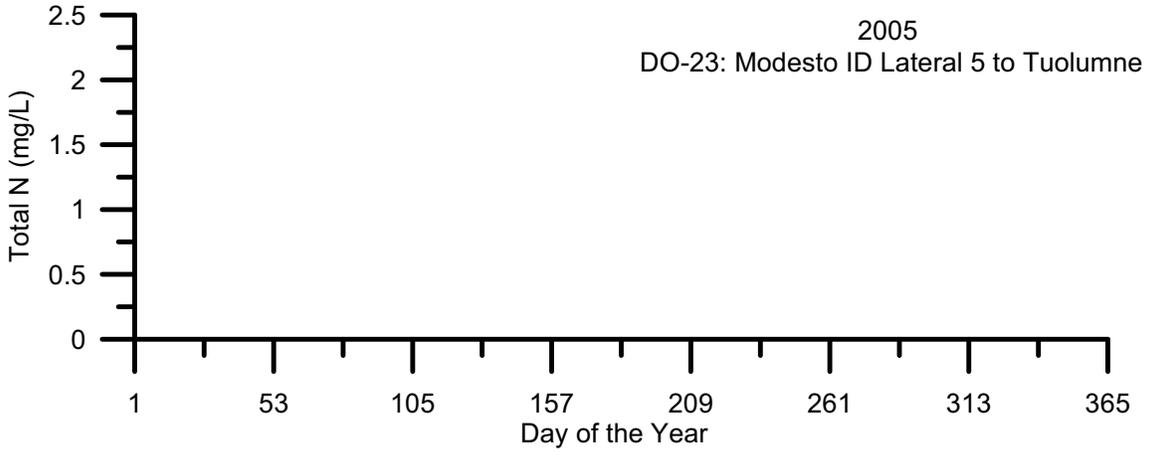


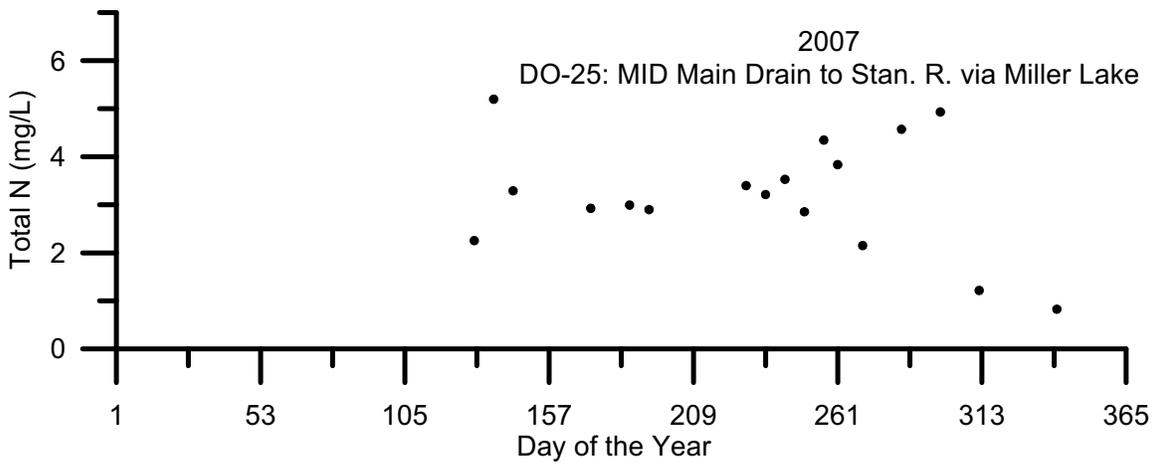
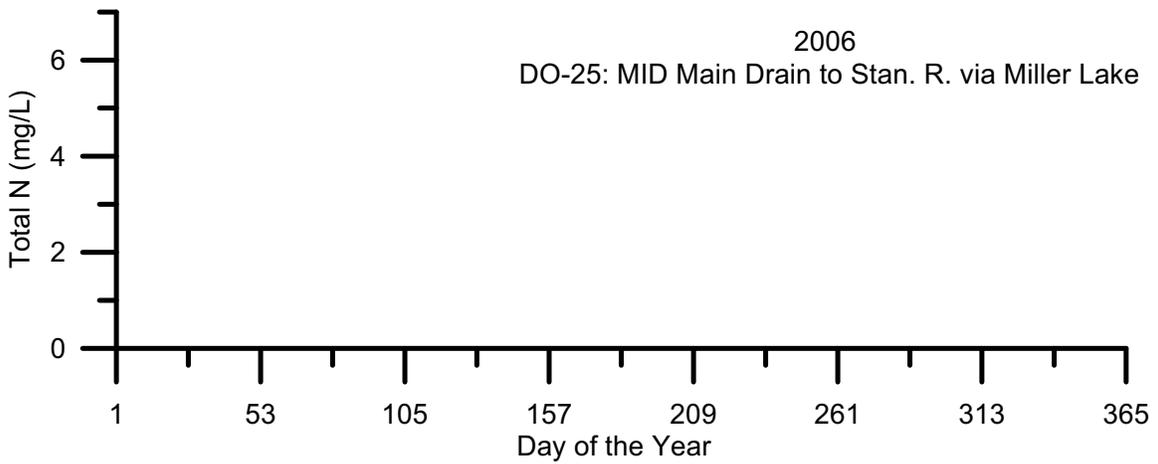
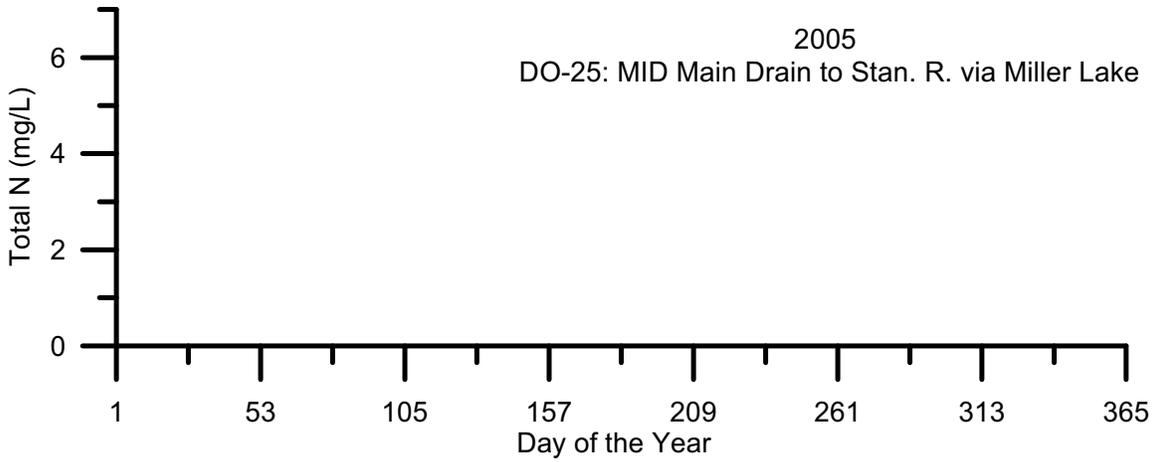


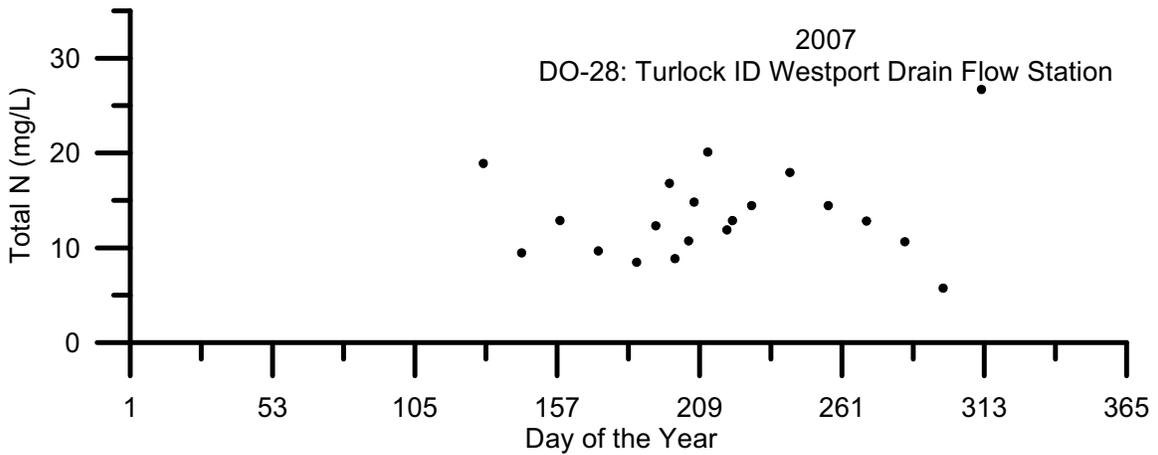
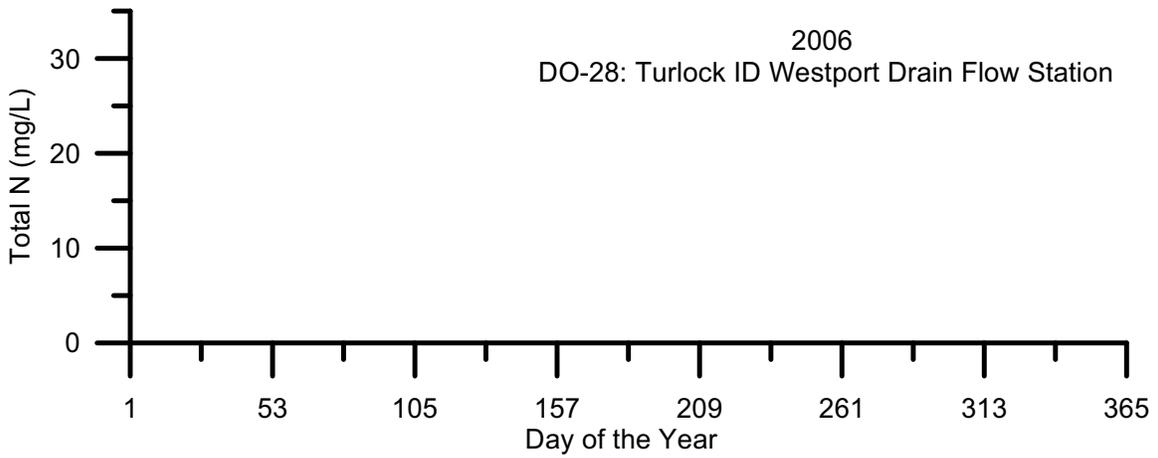
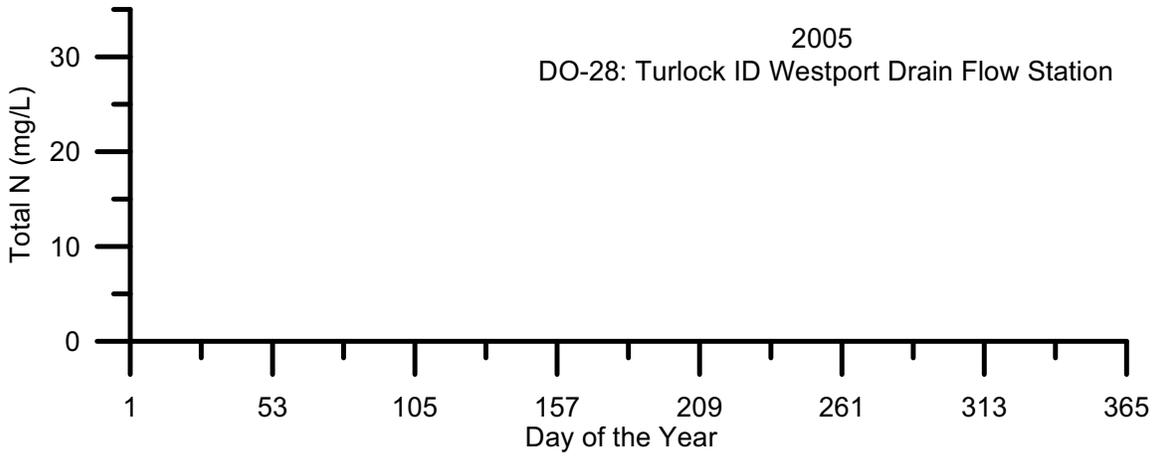


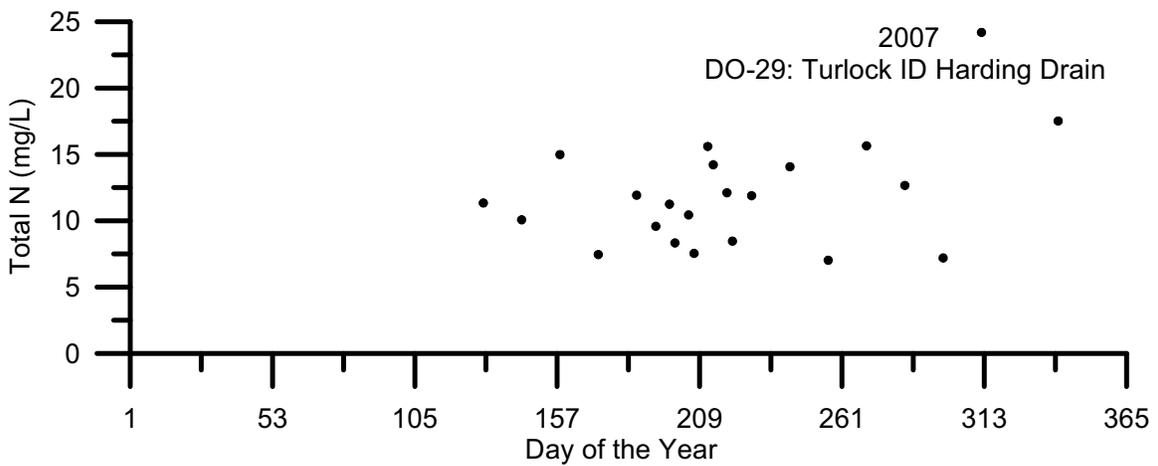
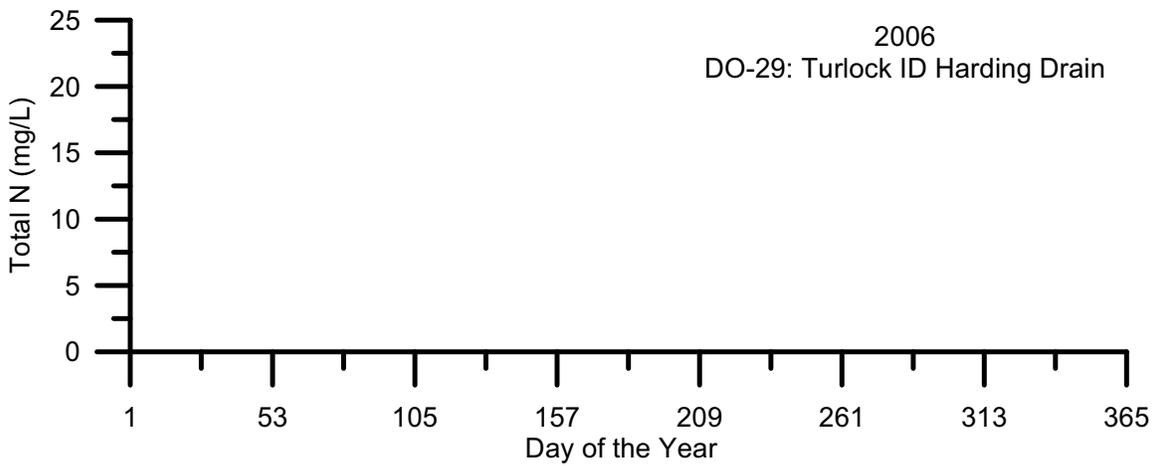
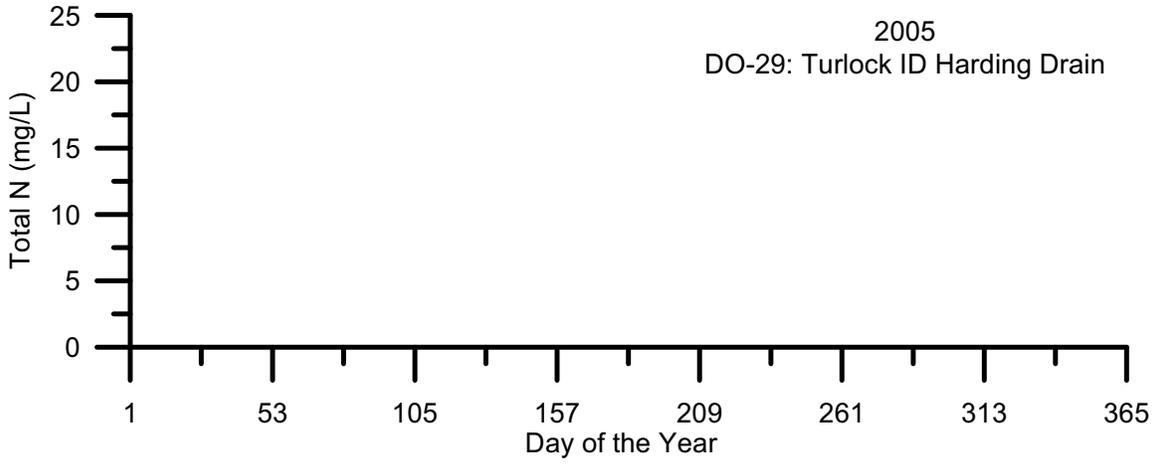


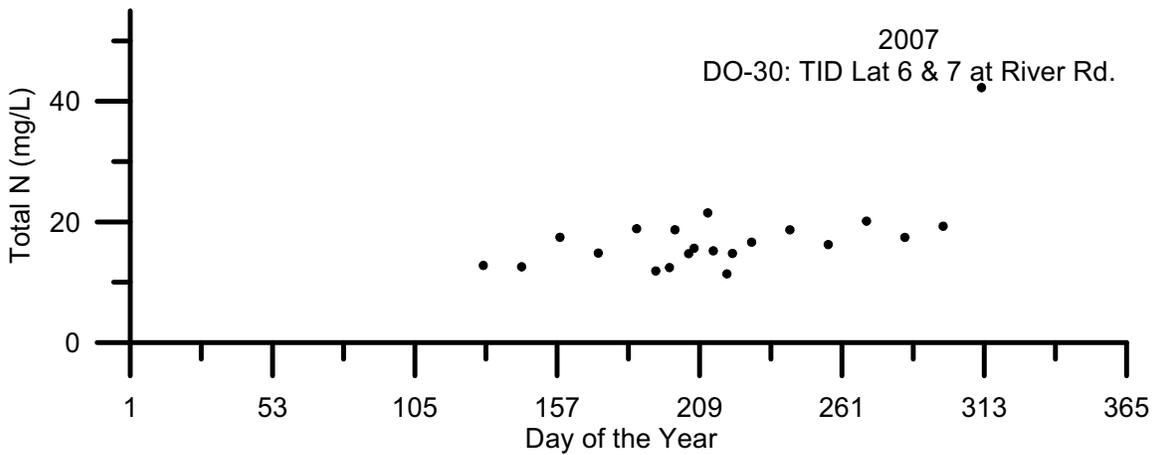
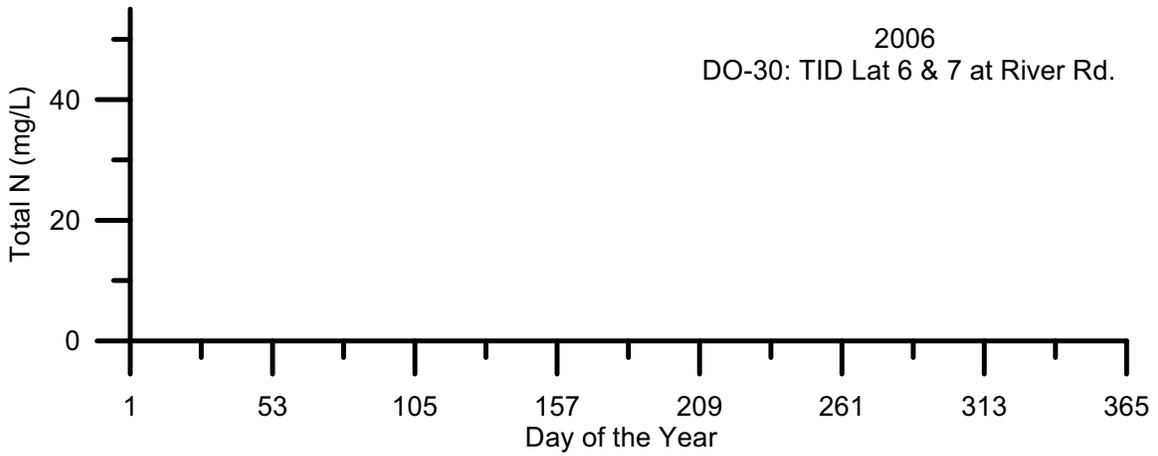
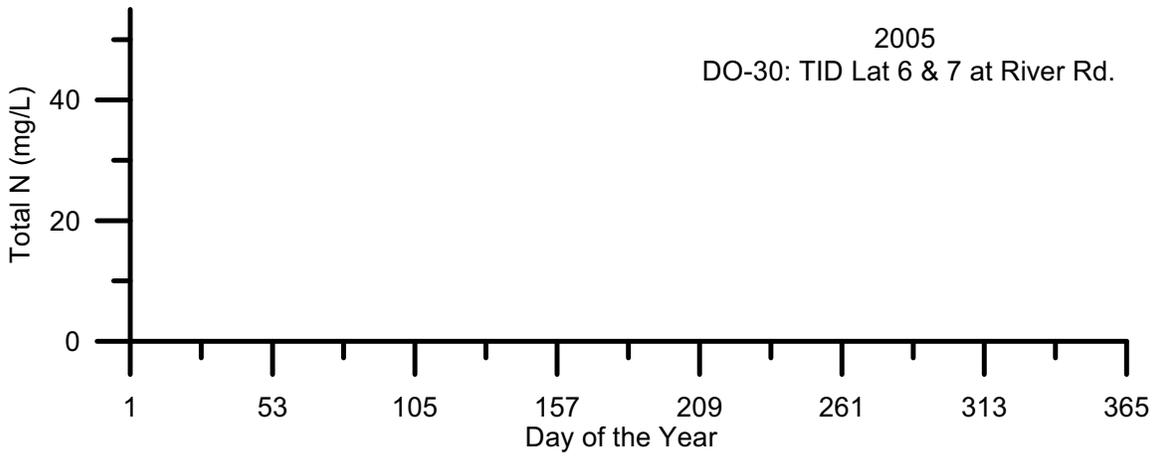


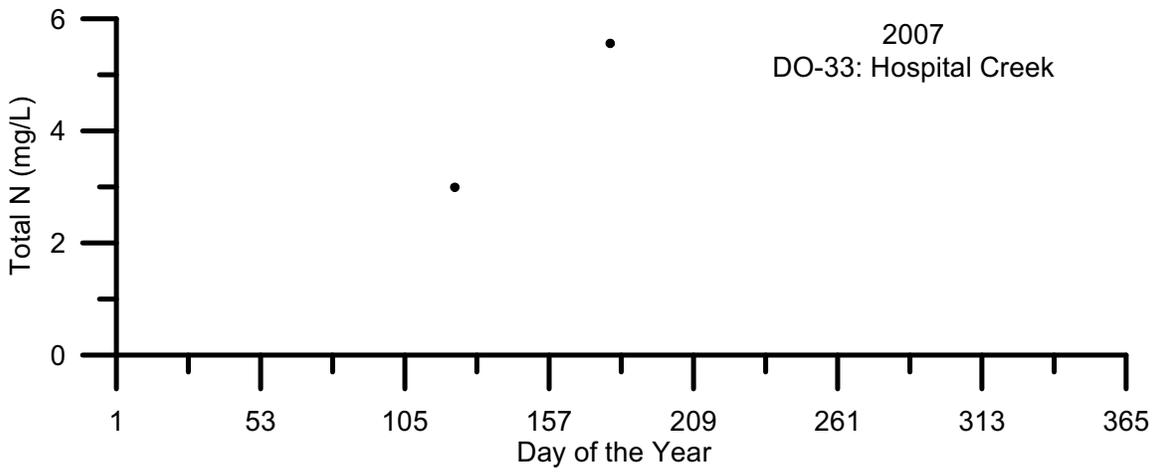
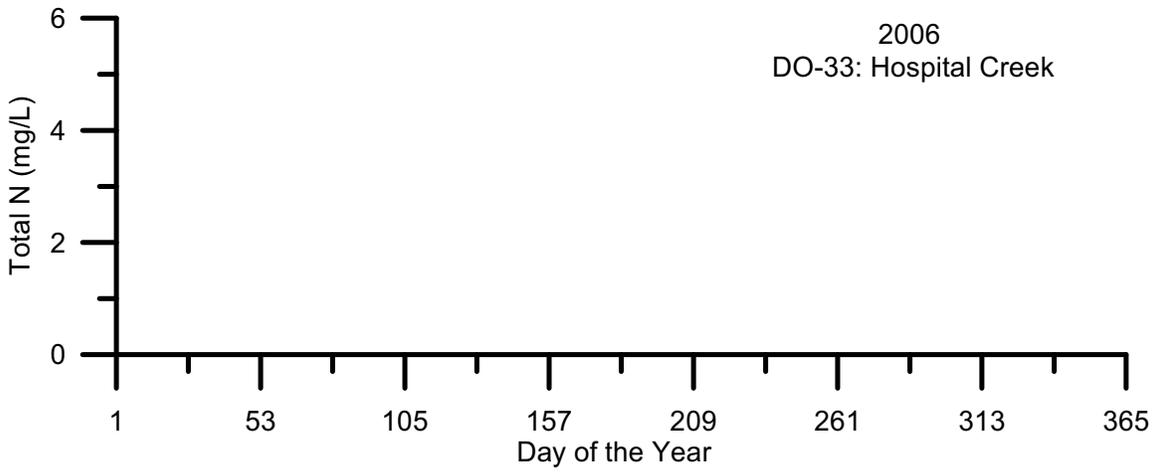
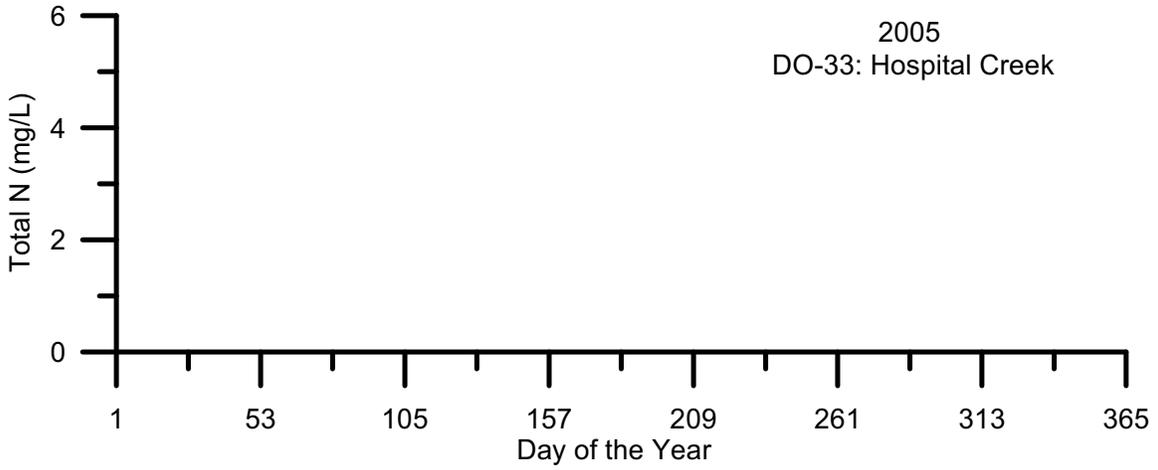


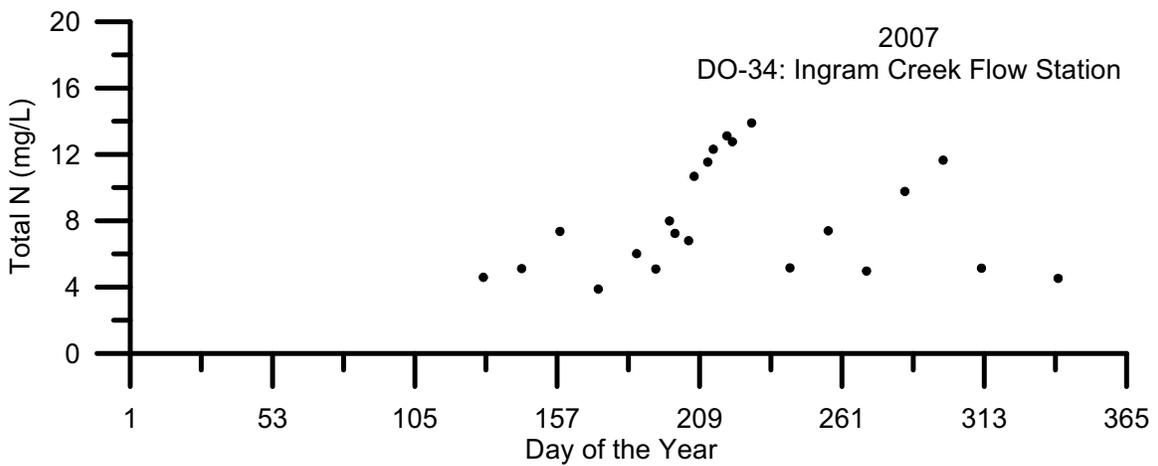
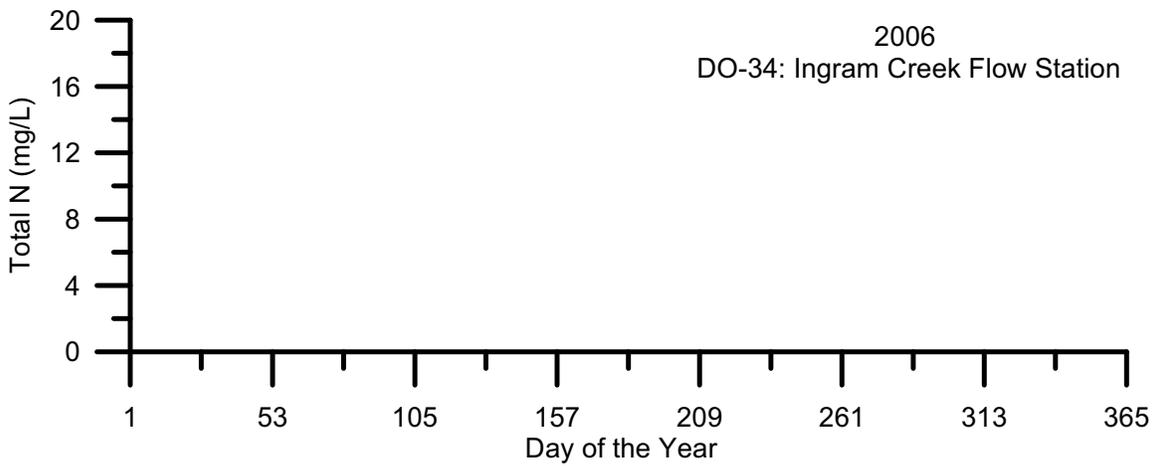
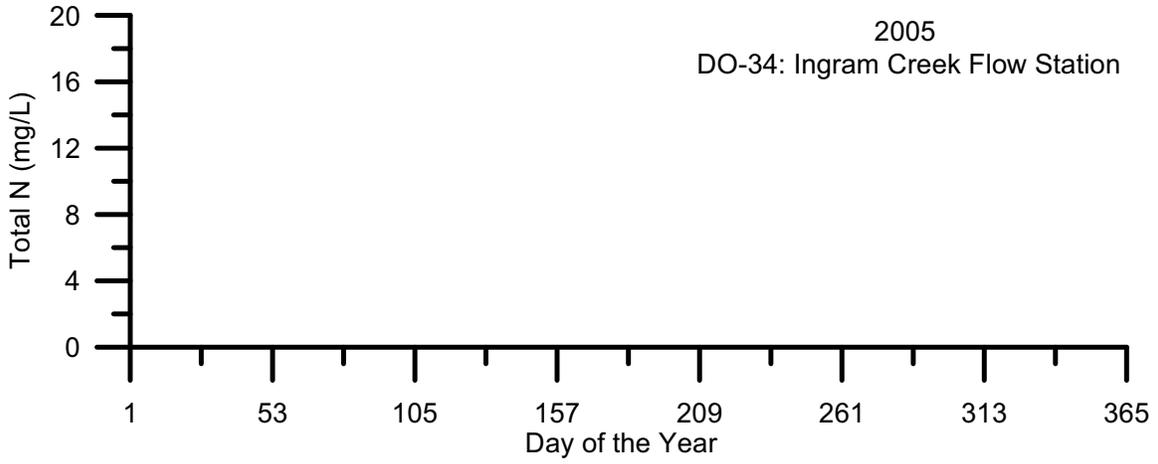


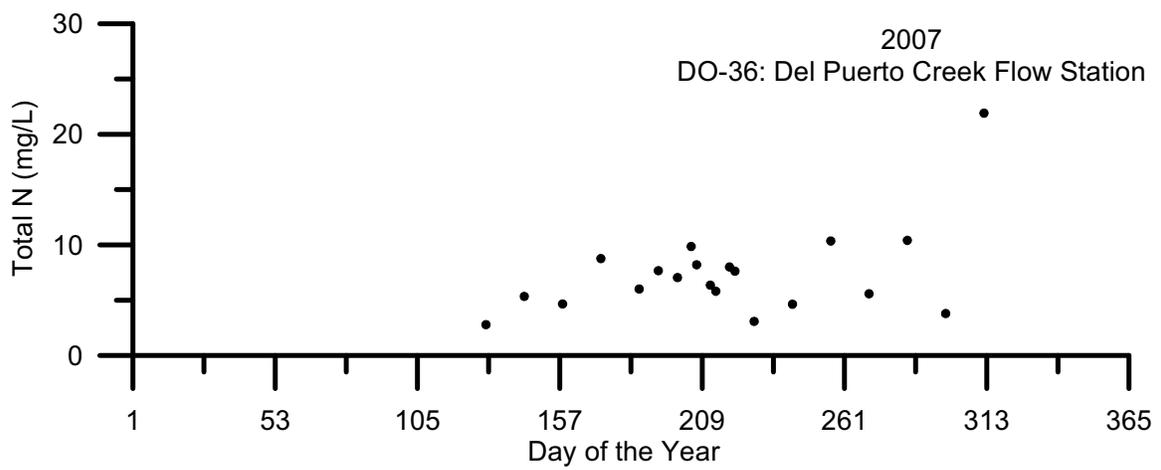
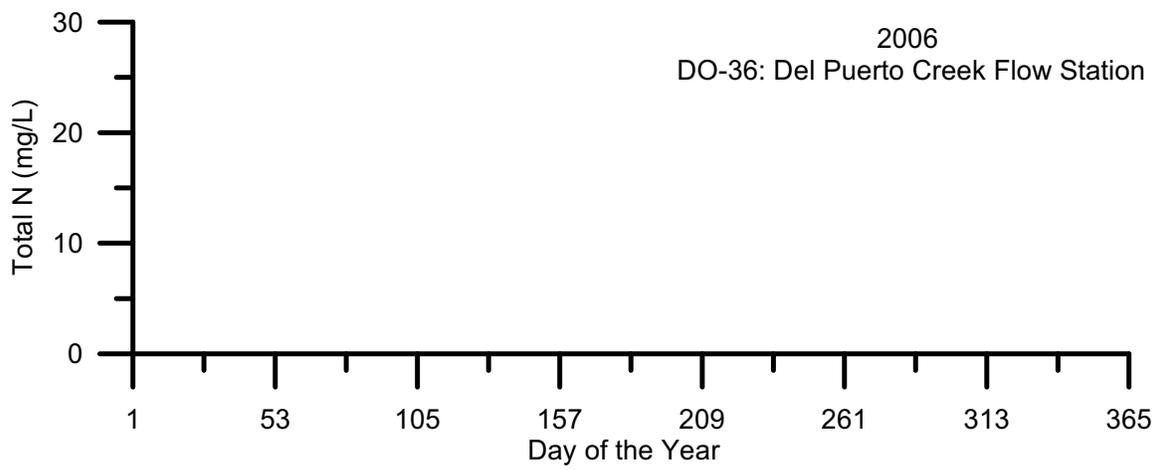
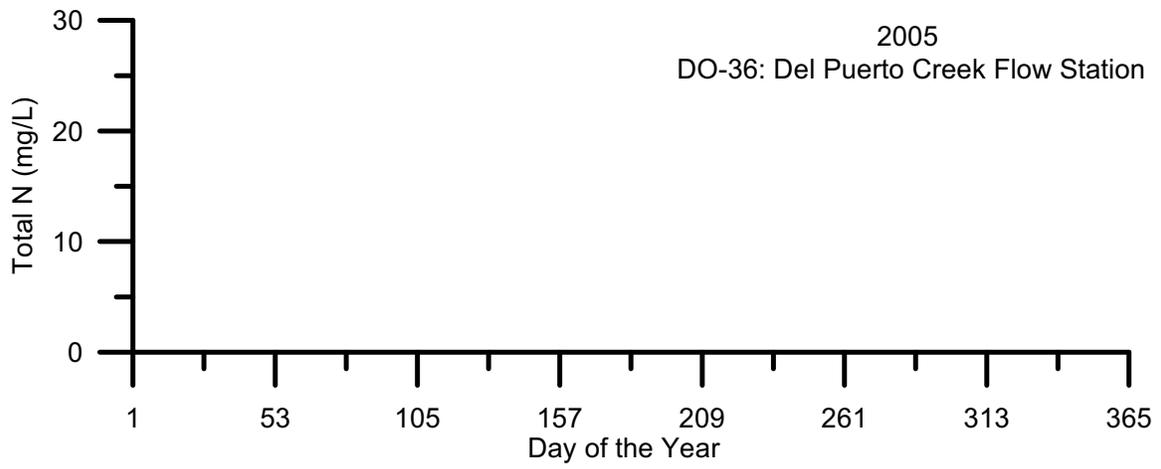


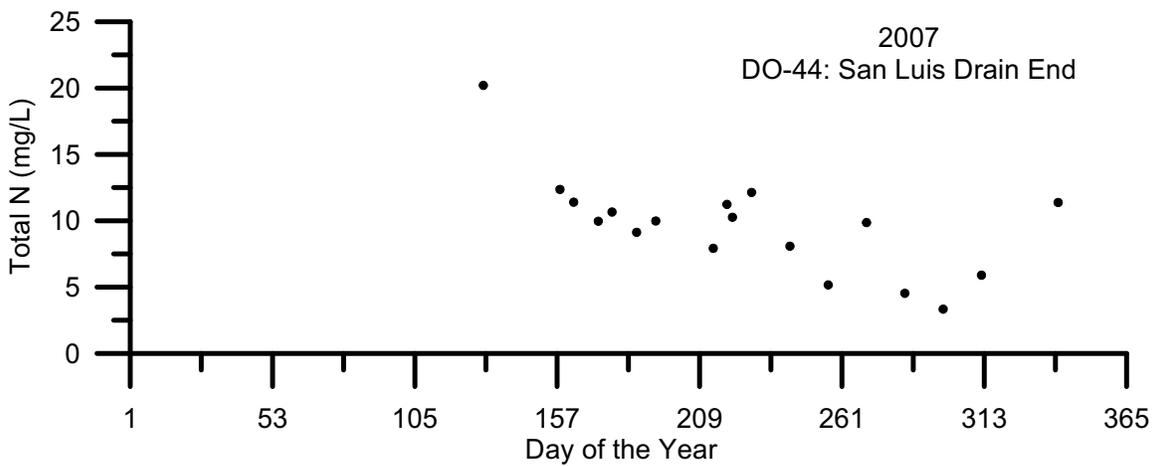
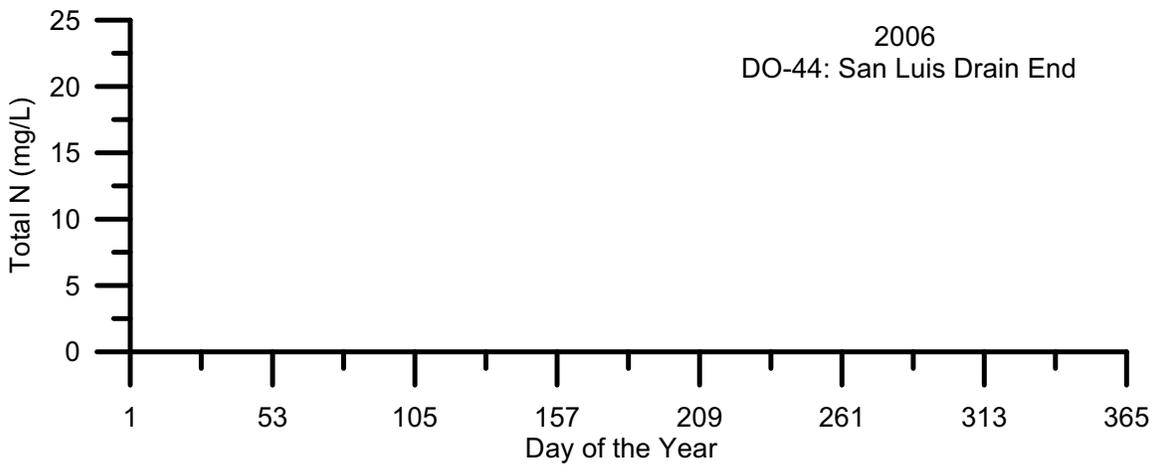
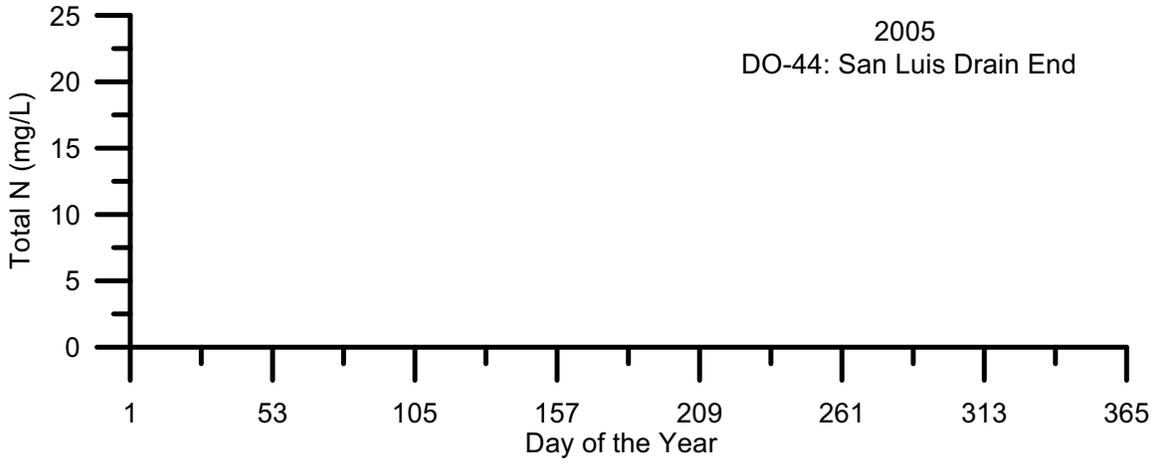


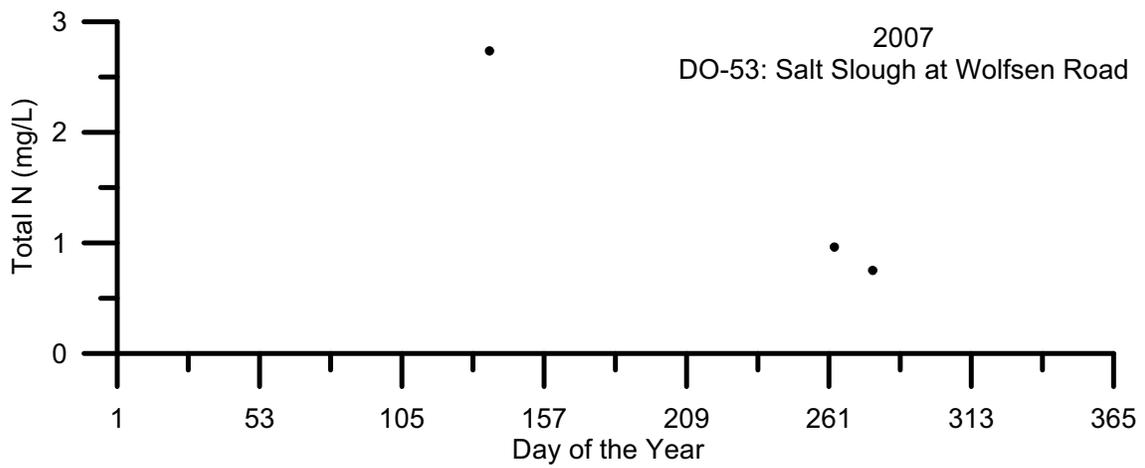
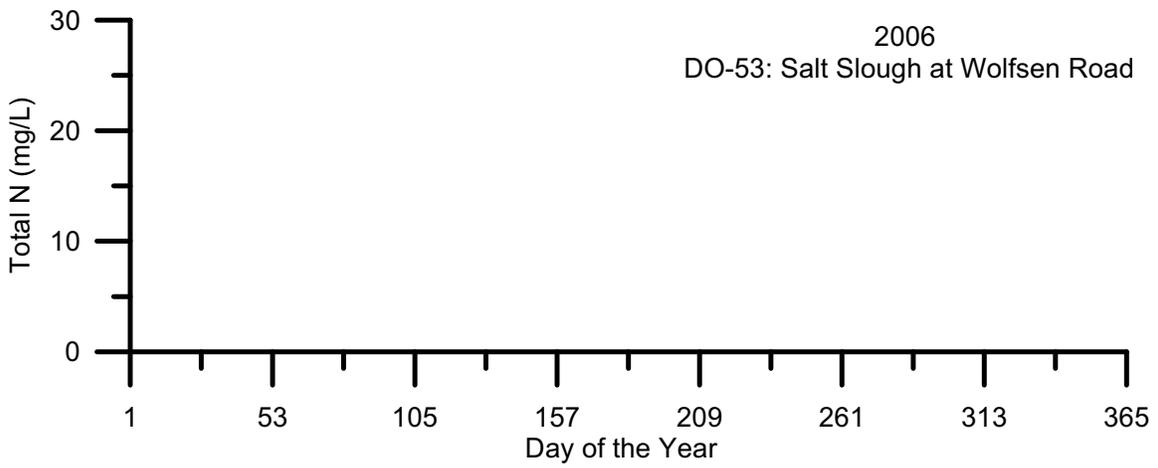
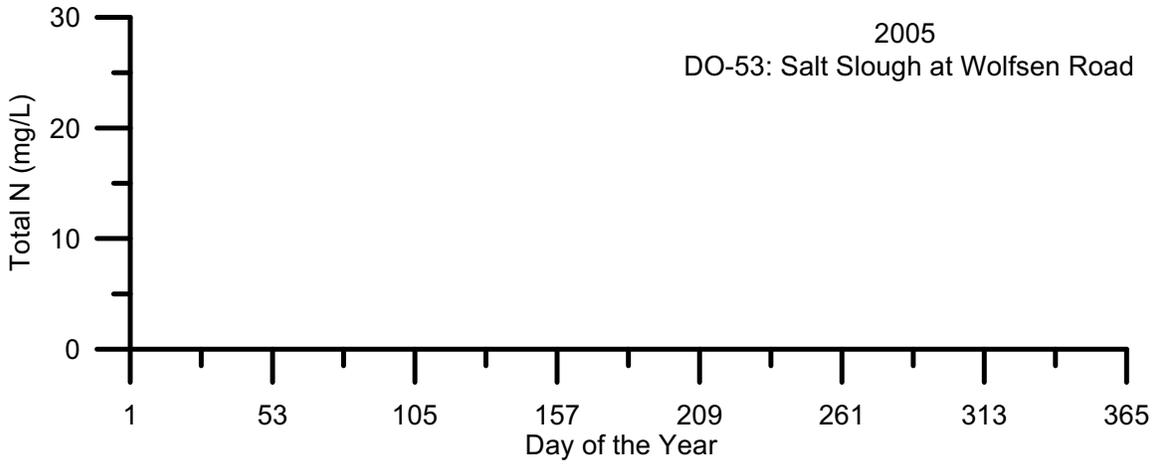


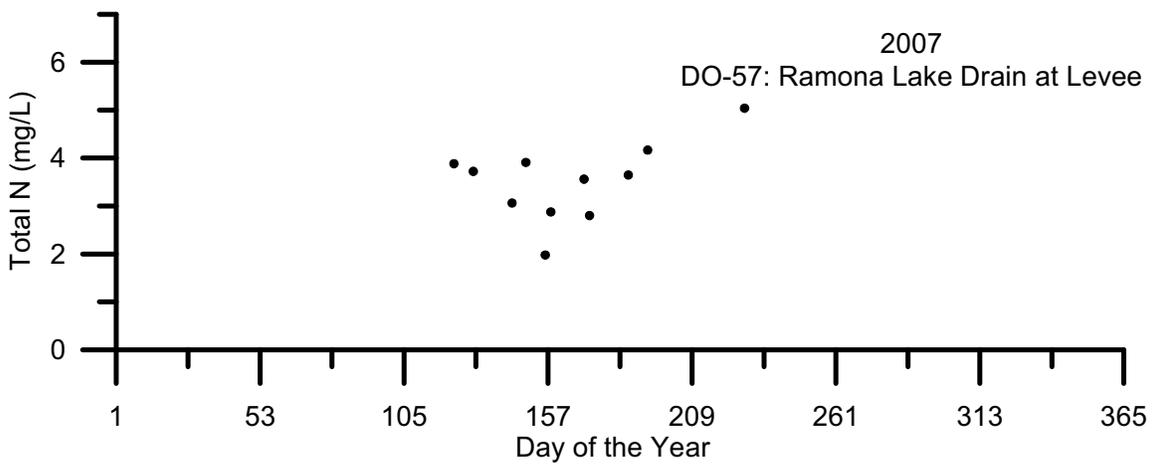
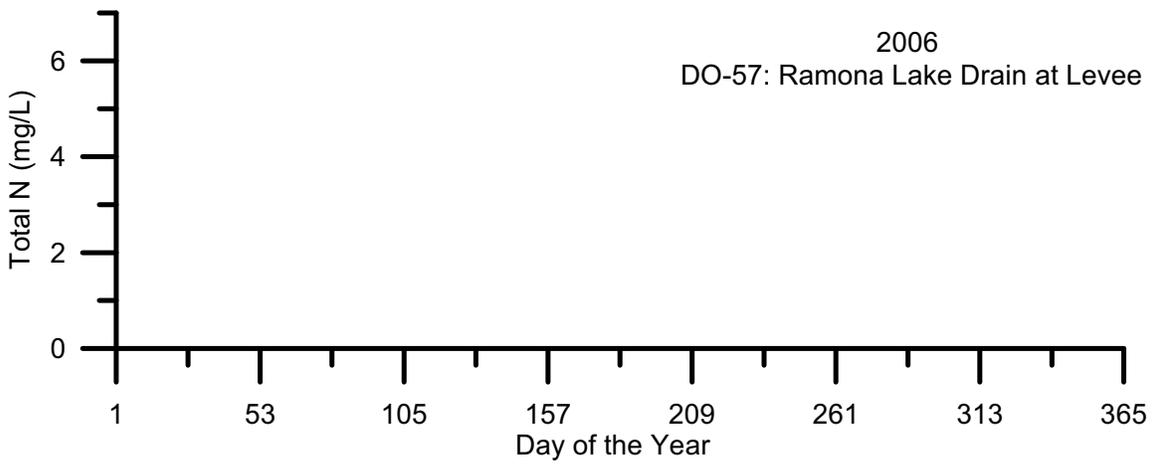
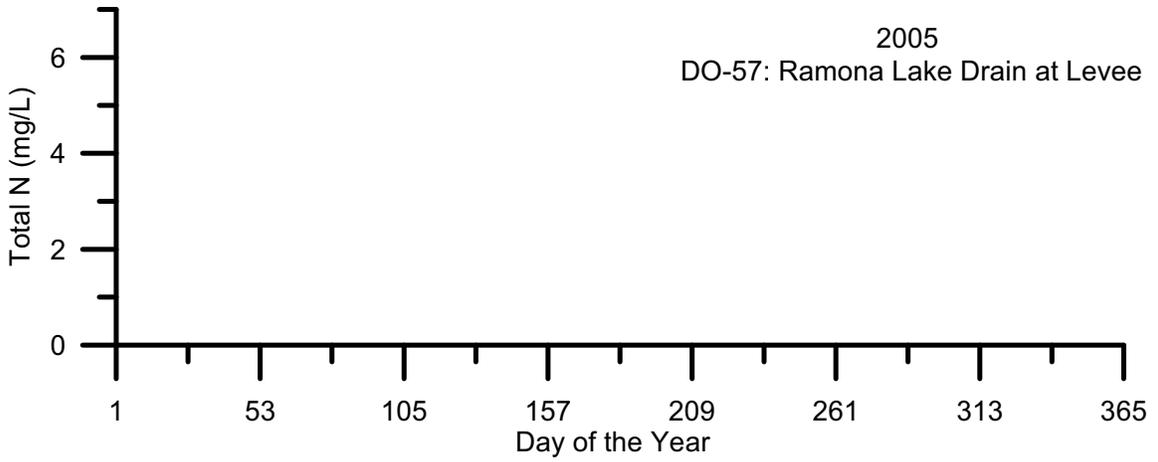


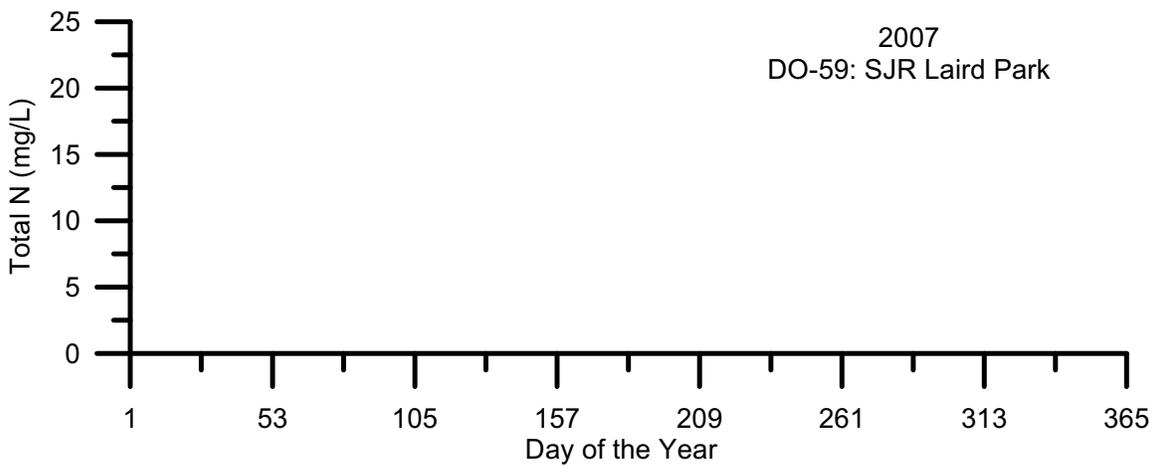
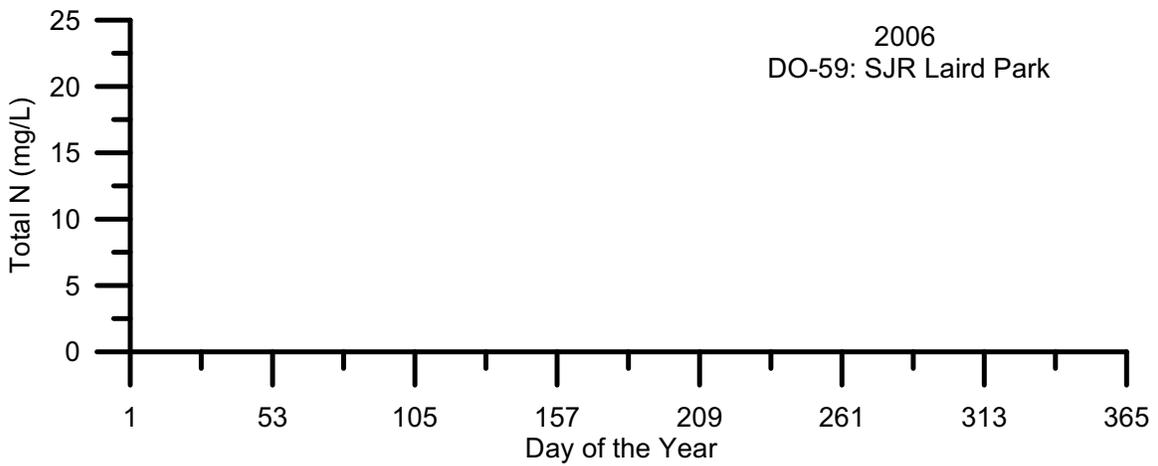
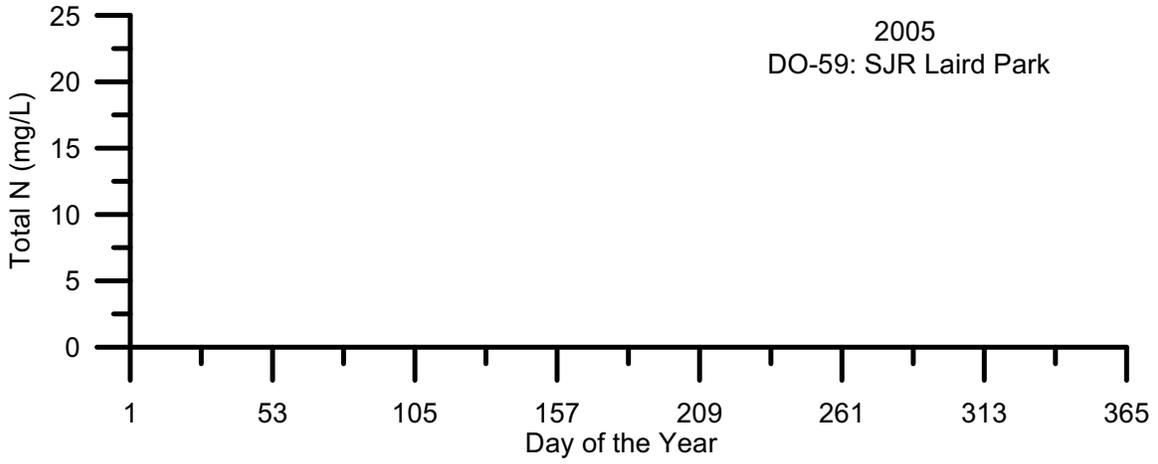


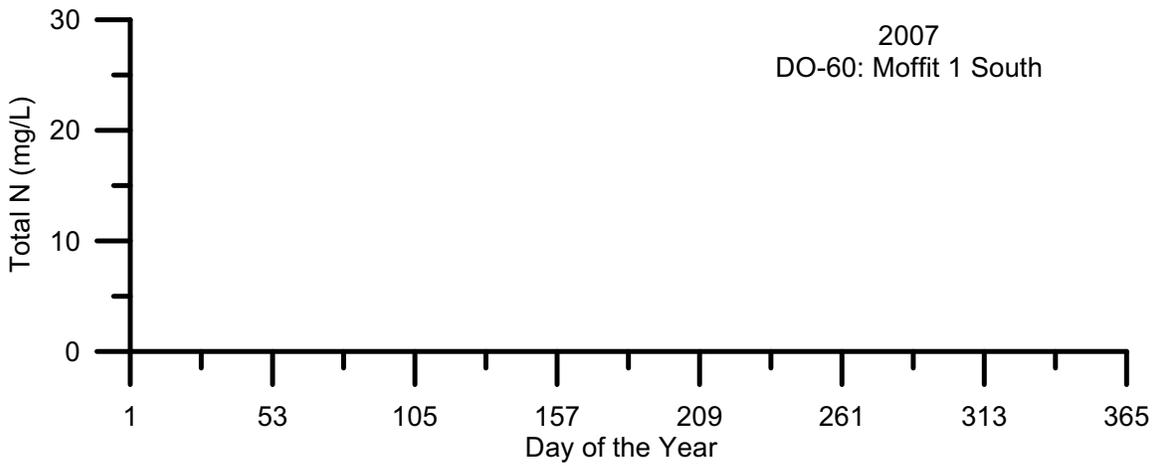
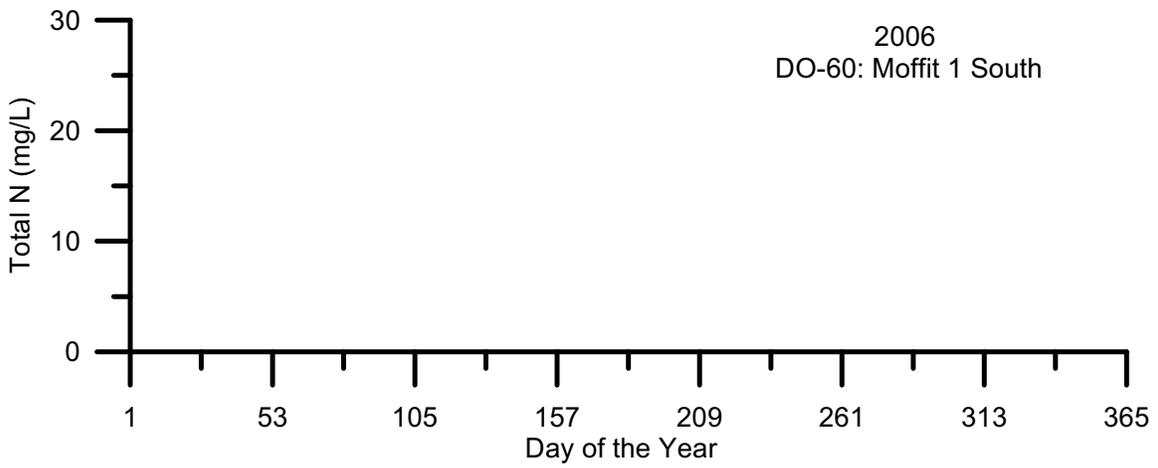
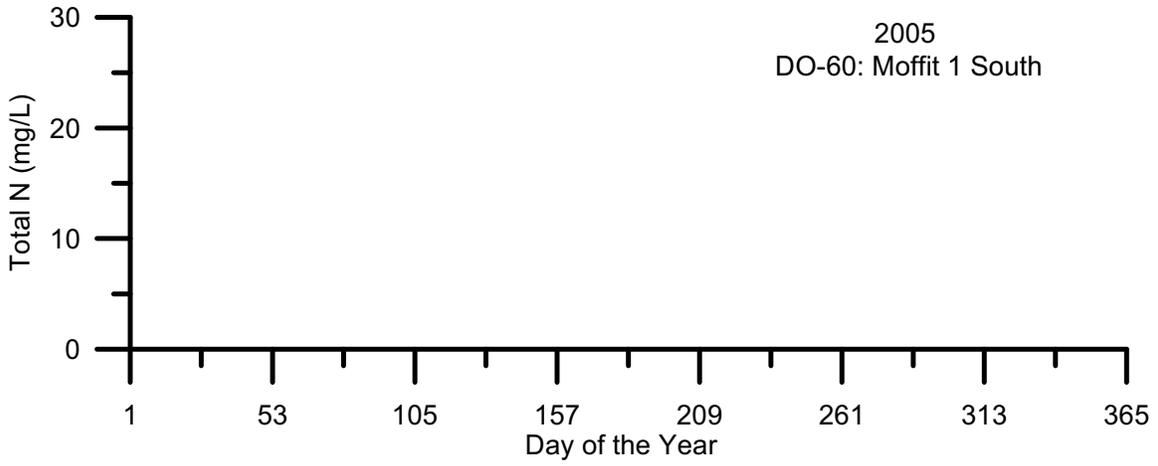


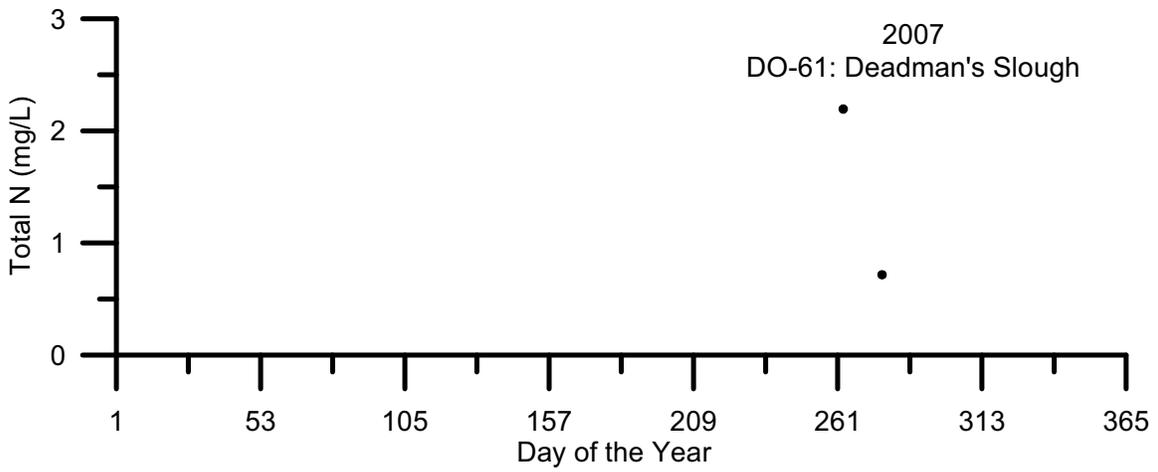
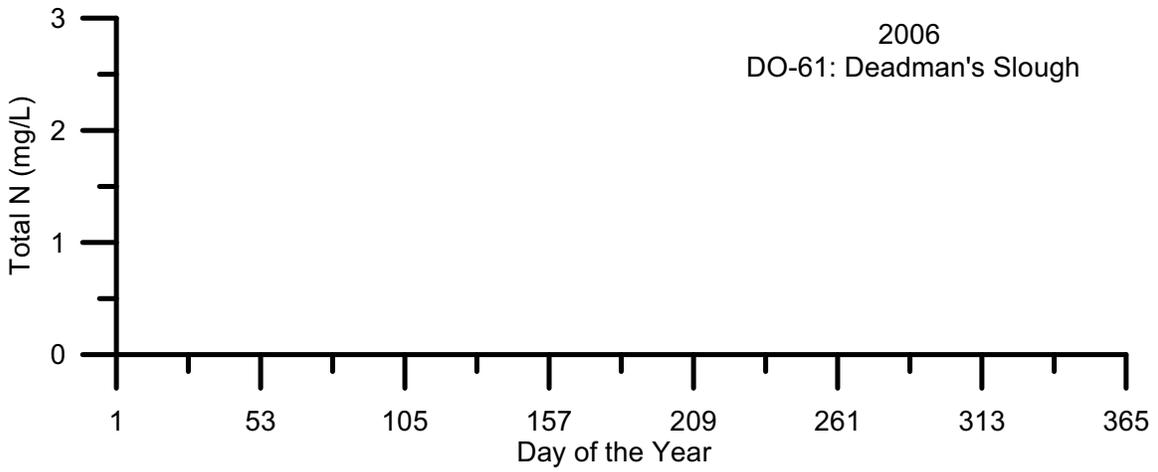
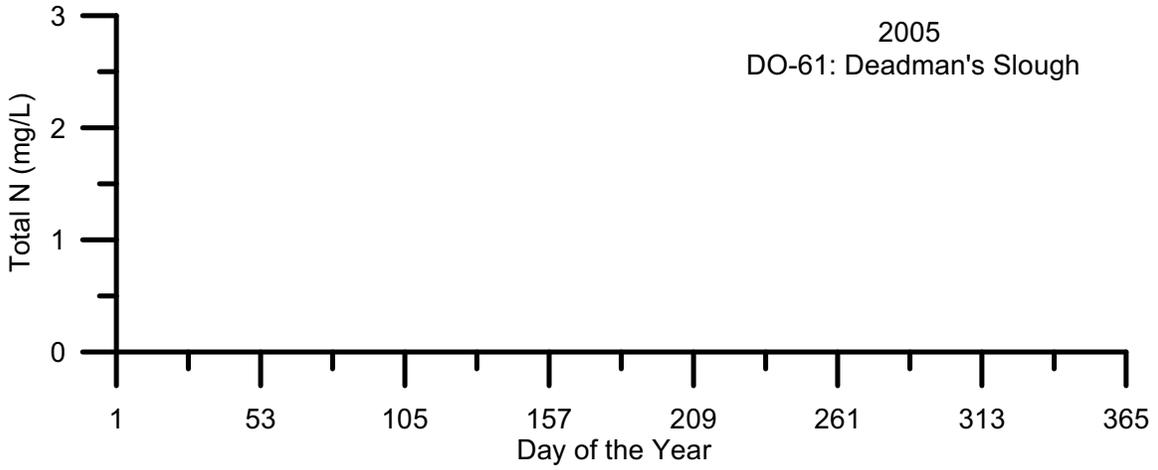


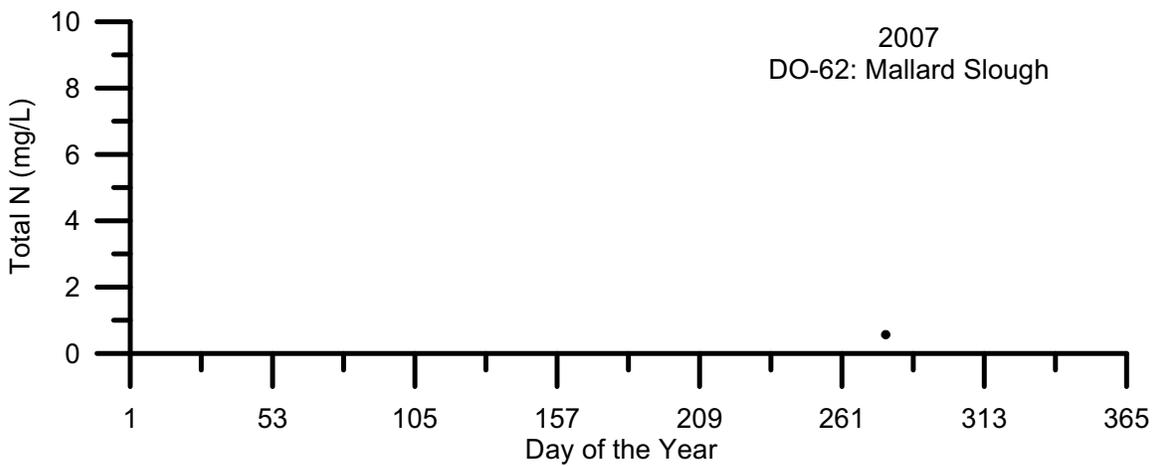
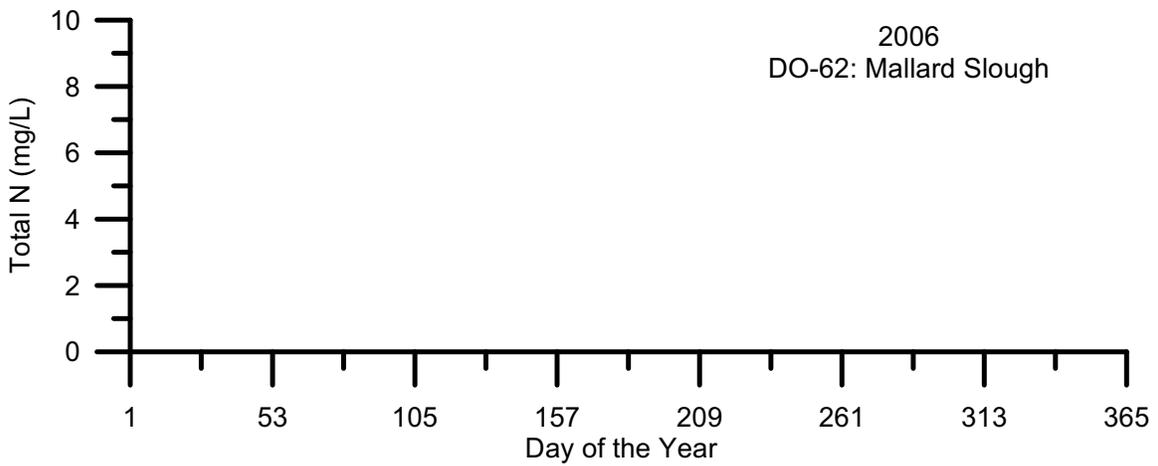
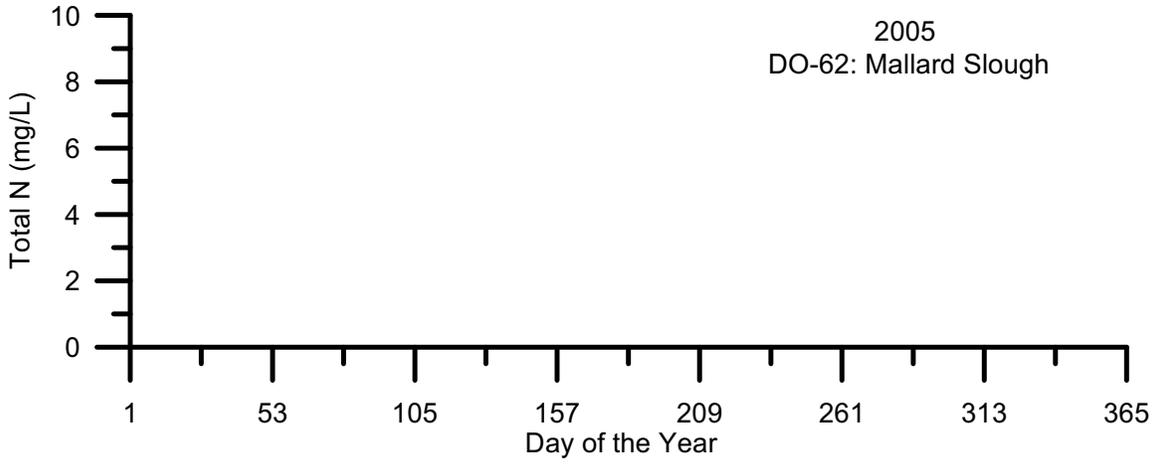


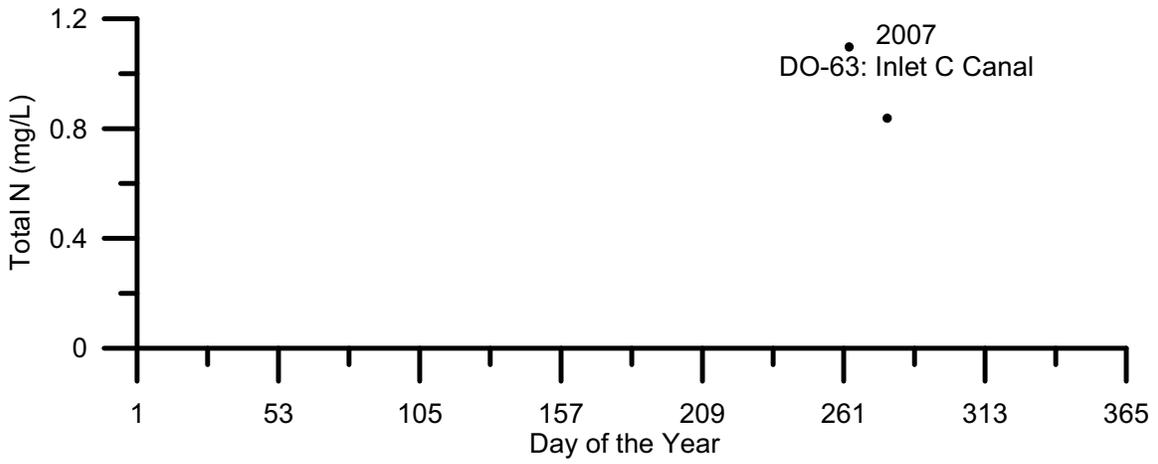
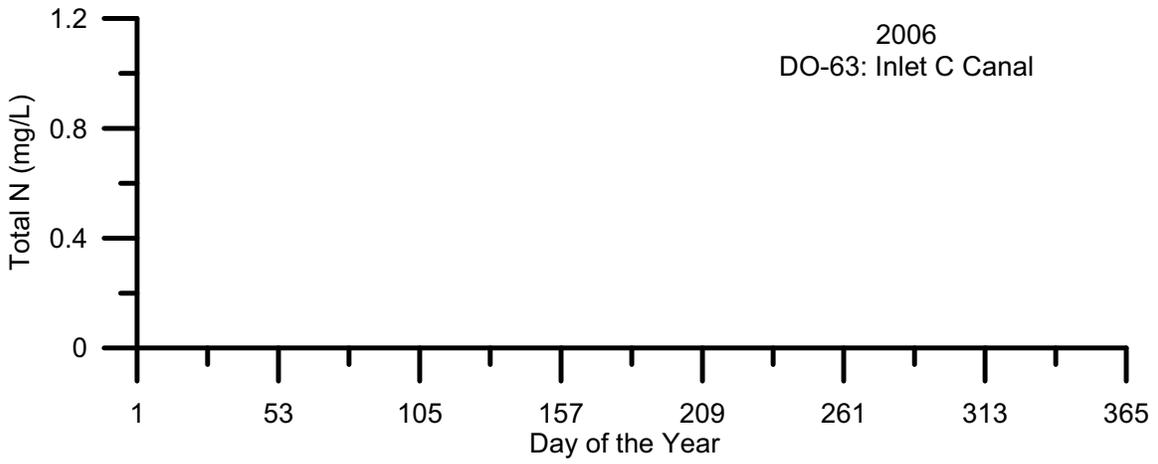
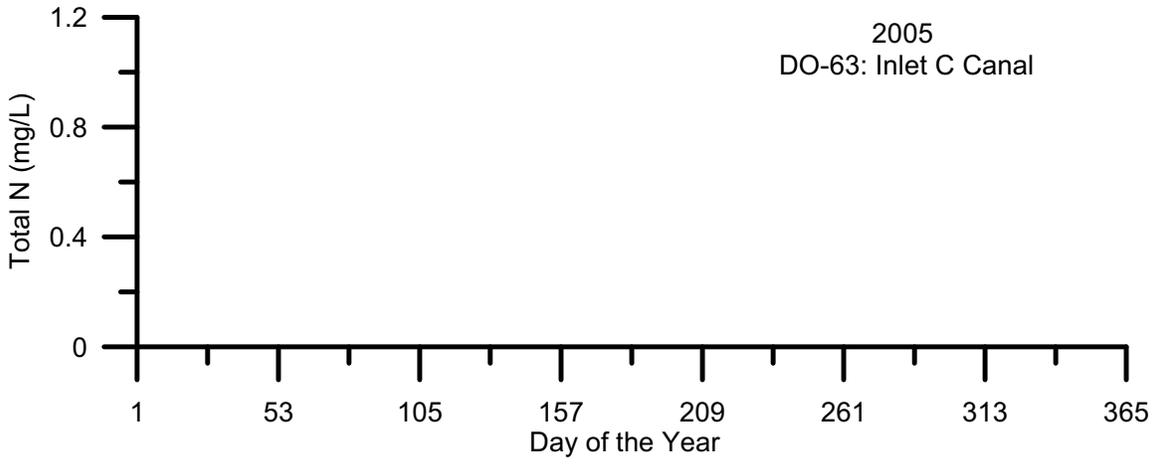


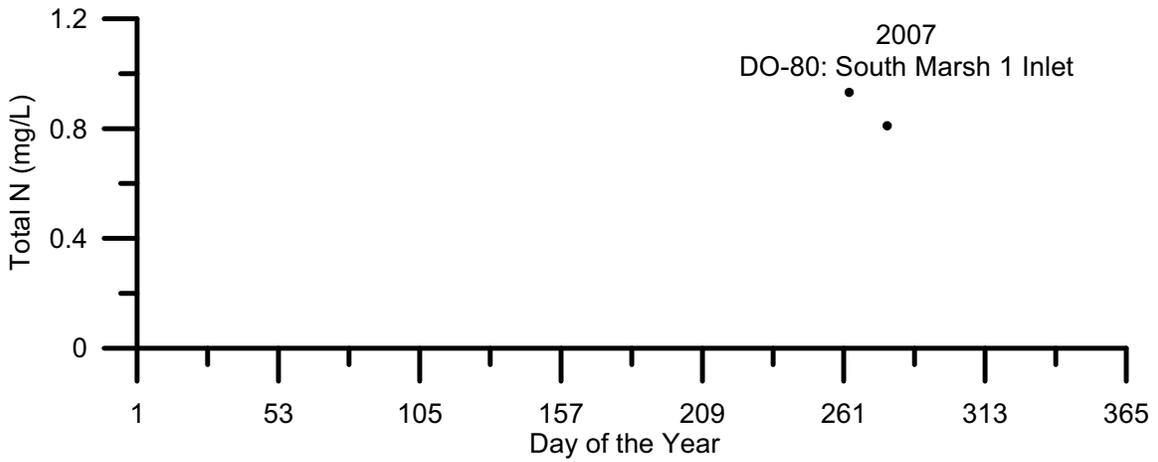
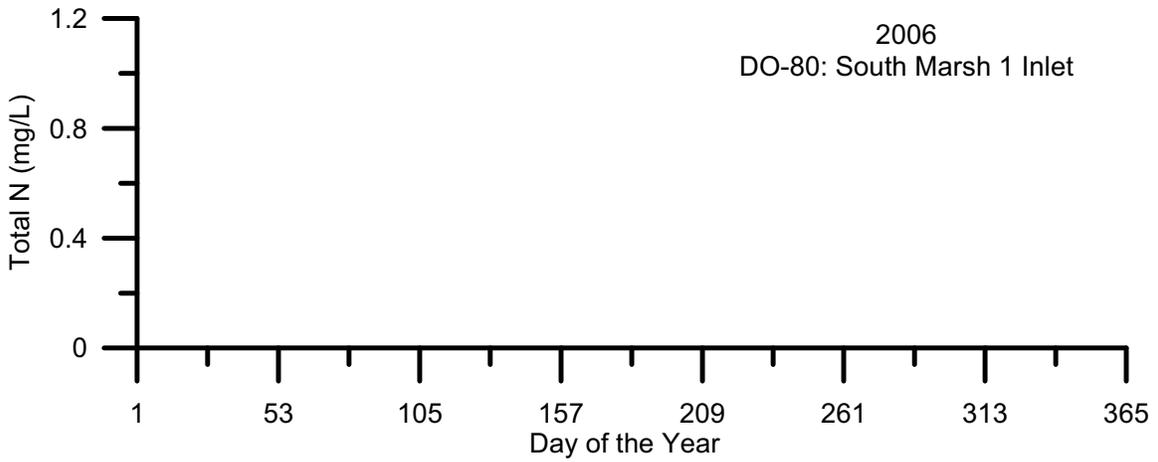
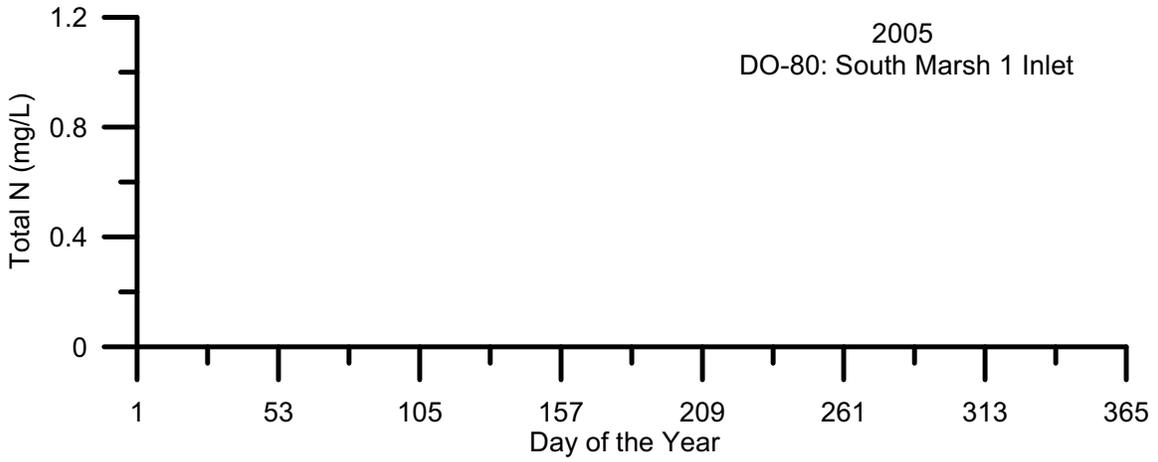


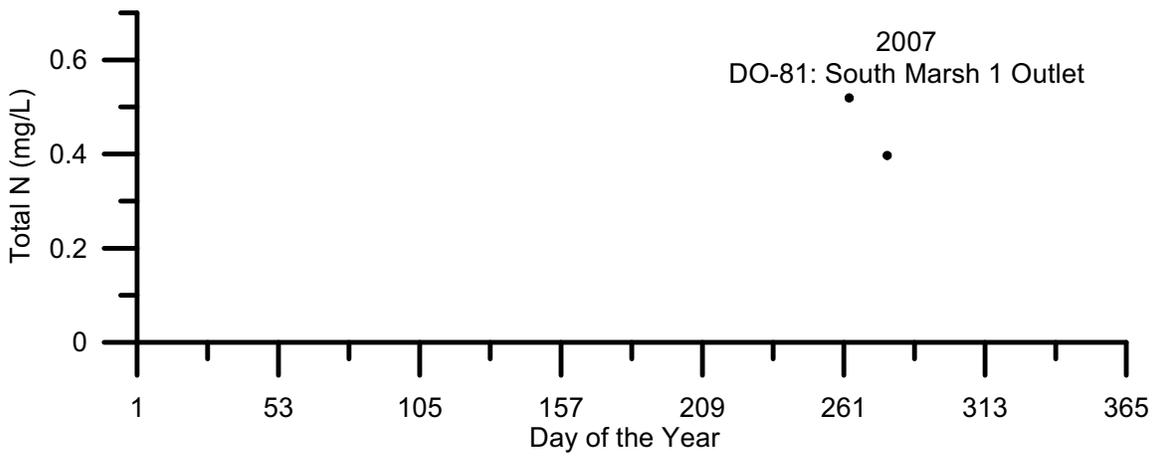
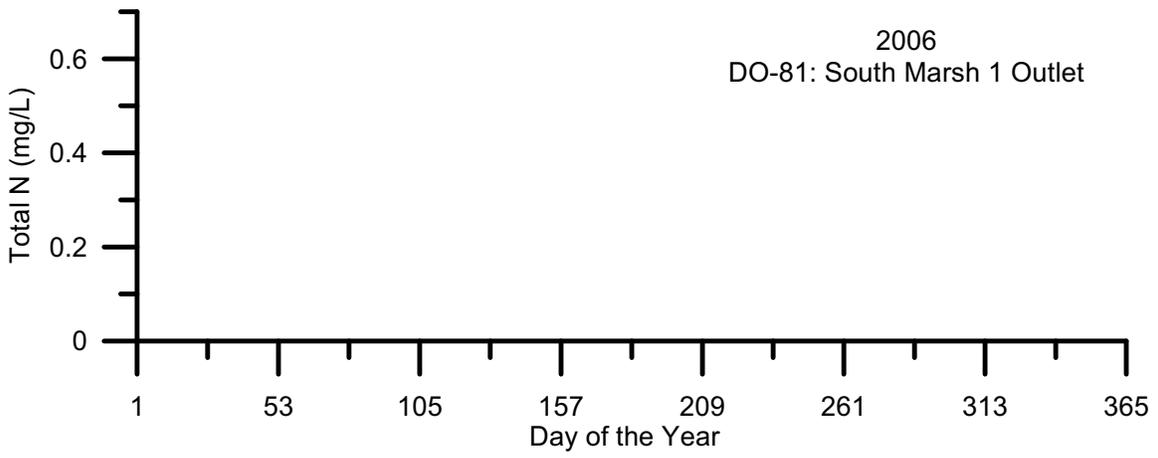
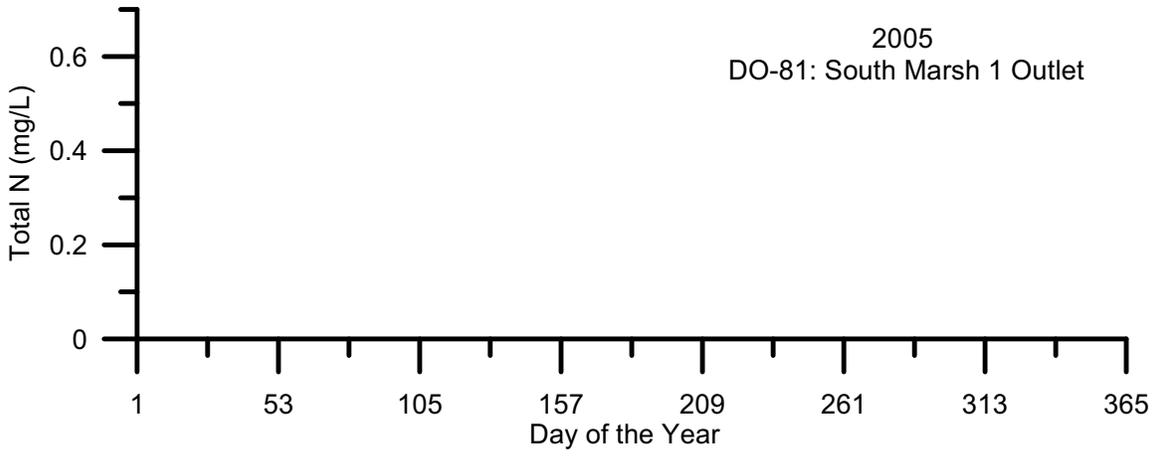


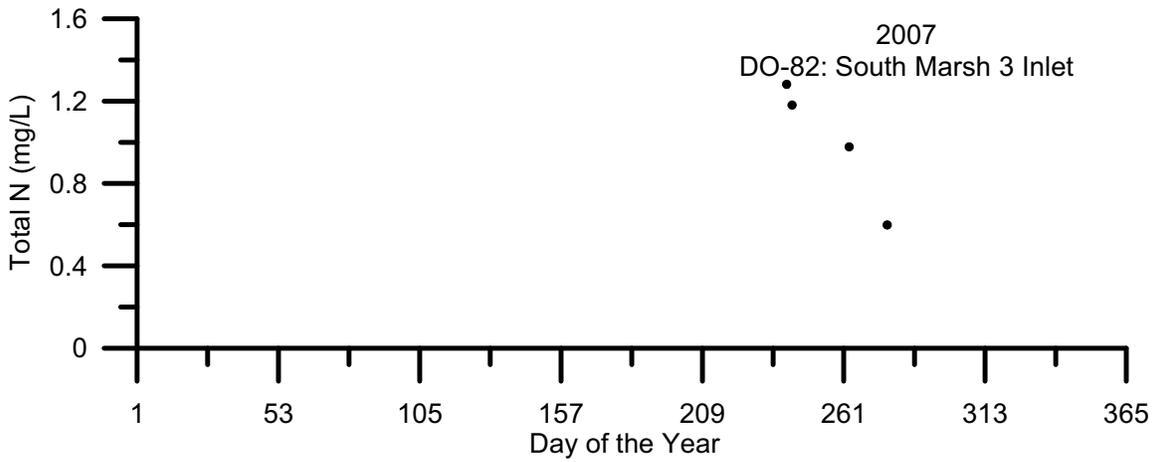
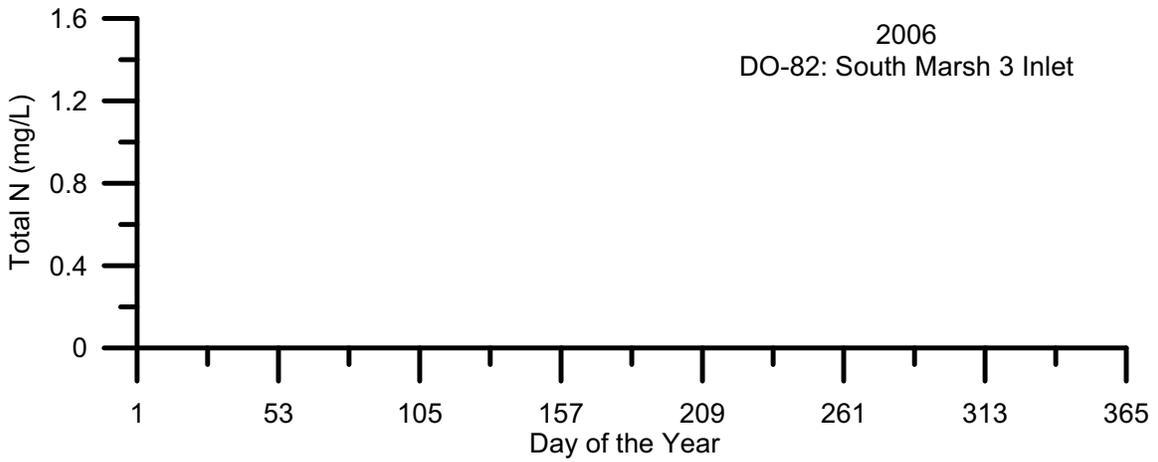
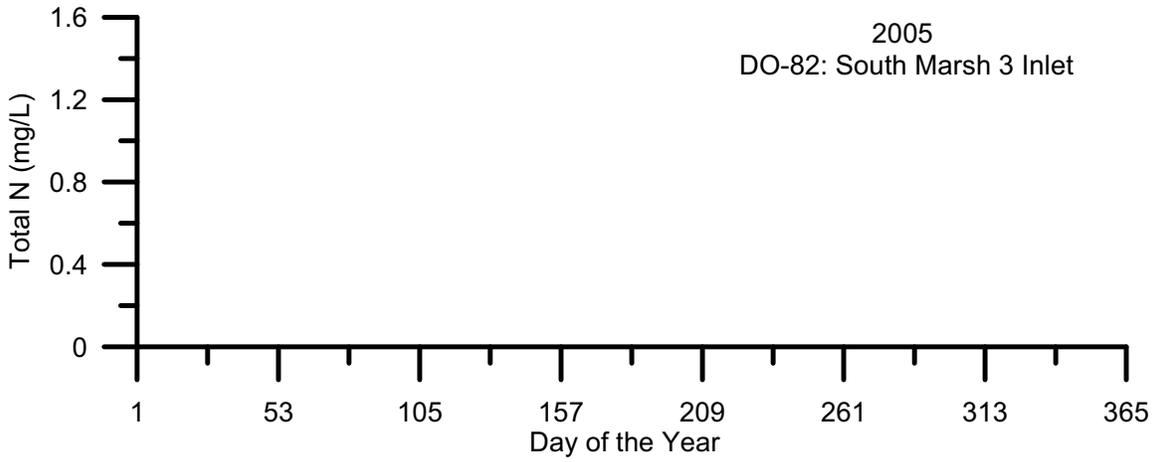


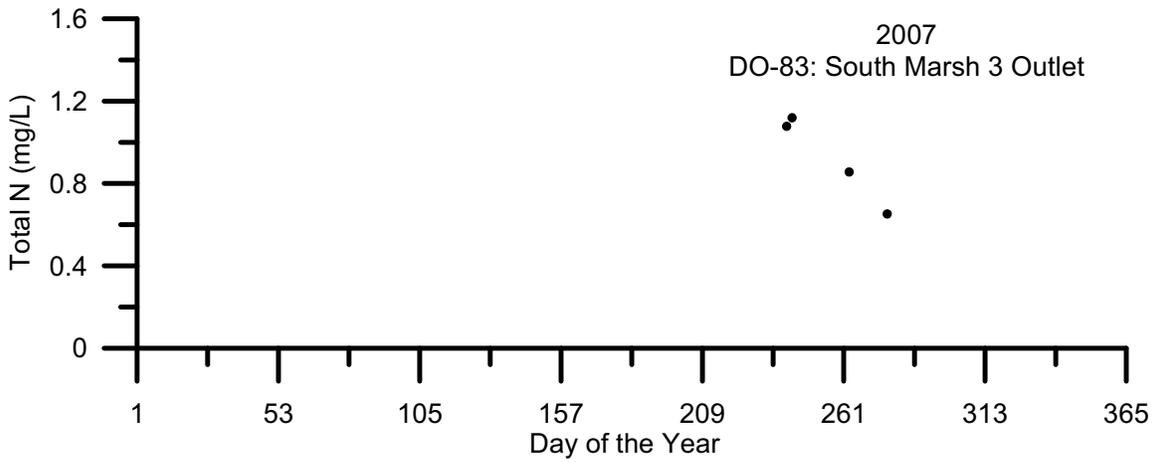
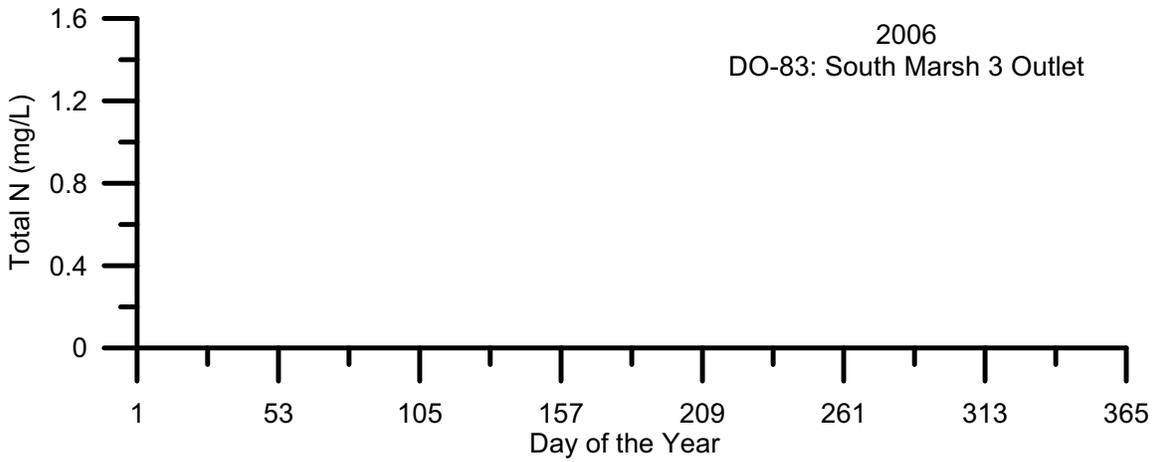
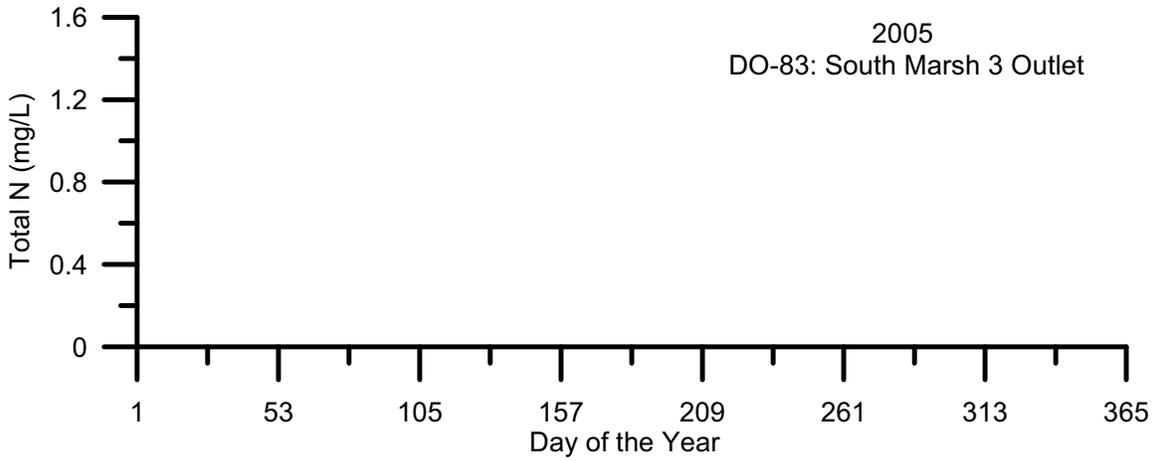


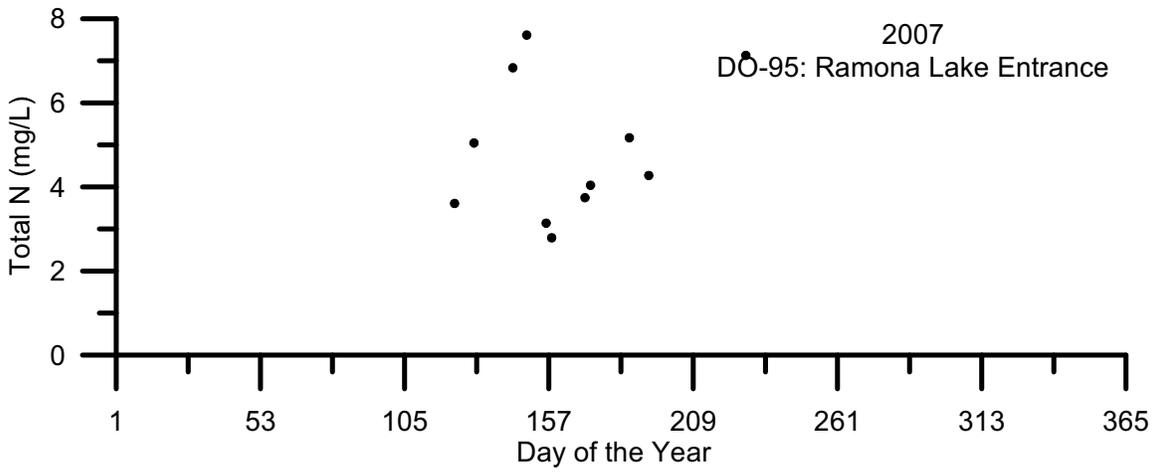
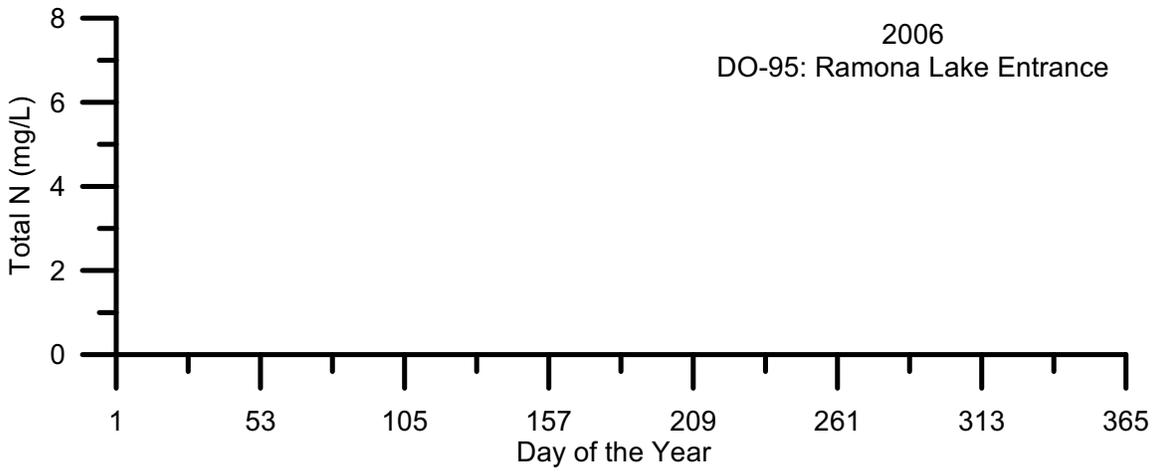
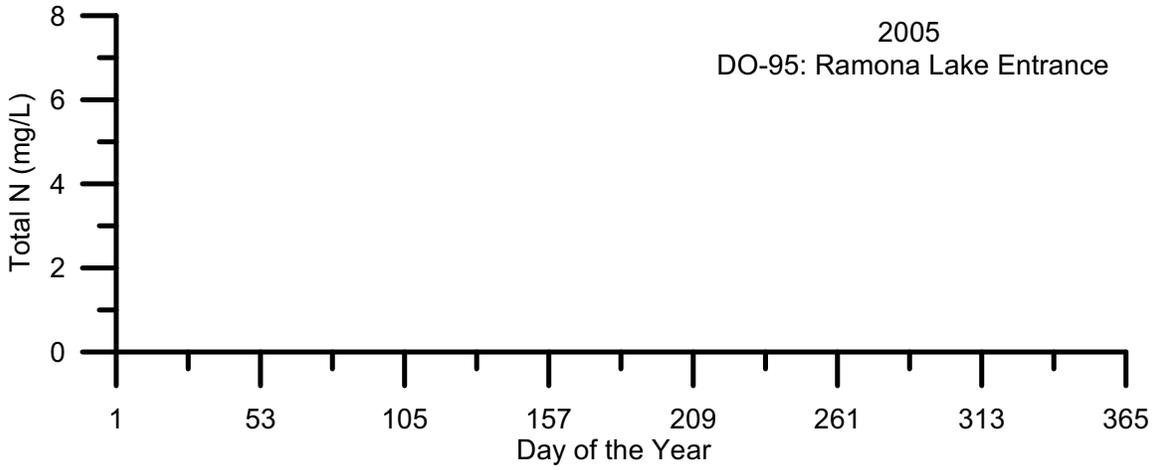














Temporal Plots of 2005-2007 Total Iron Data from the Upstream San Joaquin River

Chelsea Spier¹
Remie Burks¹
Sharon Borglin¹
Jeremy Hanlon¹
Justin Graham¹
William Stringfellow¹

February 2008

¹Environmental Engineering Research Program
School of Engineering & Computer Sciences
University of the Pacific
3601 Pacific Avenue, Sears Hall
Stockton, CA 95211

Introduction

The San Joaquin River (SJR) supports one of the most productive agricultural regions in the world and its productivity is heavily dependant on irrigated agriculture. A consequence of irrigated agriculture is the production of return flows conveyed down gradient drains that eventually discharge to surface waters. Agricultural drainage may have significant nutrient load and can impact algae growth and general water quality in the SJR. Individual farmers and agricultural organizations, such as drainage authorities, are in need of tools to manage the environmental impacts of agricultural activities (Stringfellow, 2008).

For the years 2005 through 2007, sites throughout the San Joaquin Valley watershed were sampled to assess the overall water quality in the region. One thousand nine hundred and ninety-six (1996) individual surface water samples were collected and analyzed and WQ was assessed at 113 locations in the SJR basin (Borglin et al., 2008). Samples were processed and analyzed by the Environmental Engineering Research Program (EERP) laboratory at the University of the Pacific. This report includes temporal plots of total iron (Fe) data analyzed by the EERP laboratory between 2005 and 2007.

Methods

Depth integrated field samples were collected during 2005-2007 in the upper San Joaquin River in accordance with EERP Field Standard Operating Procedures Protocol Book (Graham 2008). Water samples were collected in glass 1000 mL bottles (Wheaton Science Products, Millville, NJ), 1000 mL HDPE Trace-Clean narrow mouth plastic bottles (VWR International), 250 mL HDPE Trace-Clean wide mouth plastic bottles (VWR International) as well as 40 mL trace clean vials with PTFE septa (ICChem, Rockwood, TN). Bottles were labeled with the appropriate sample number, site name and sampling date. All bottles were rinsed with sample water prior to sample collection. Some sites required a bucket to collect sample water because of accessibility from a high bridge or platform. For these sites, the bucket was pre-rinsed with sample water and sample bottles were filled using a rinsed funnel. Care was taken to distribute water simultaneously to all sample bottles (rather than sequentially). Samples were immediately stored at 4°C after sampling (cooler temperature was recorded in the lab upon delivery) and transported to the EERP lab on the day of sampling.

Within twenty-four hours of sample collection, 6 mL aliquots of unfiltered sample were placed in 15 mL disposal centrifuge tubes and stored at -20°C for later quantification of Total Fe.

Fe was measured using a reaction with phenanthroline according to SM 3500-Fe B (APHA, 2005) using FerroVer reagents purchased from Hach (Loveland, CO). Prior to analysis, the samples were defrosted and 1 mL of sample was removed and used to measure the background absorbance of the water sample at 510 nm on the PerkinElmer Lambda 35 UV/VIS Spectrometer (Shelton, CT). Total Fe was measured on the remaining 5 mL of unfiltered sample by the addition of pre-made Hach FerroVer phenanthroline reagent and measurement at 510 nm. The background sample absorbance was subtracted from the sample absorbance with reagent added. The limit of detection for this method is 0.02mg/L Fe.

Results/Discussion

With each set of Fe field samples analyzed in the EERP laboratory, quality assurance samples including a lab duplicate, field duplicate, matrix spike, matrix spike duplicate, calibration check standards, laboratory control standard, trip blank, and lab blanks were also analyzed. Between 2005 and 2007, 100% of all quality assurance samples were within a passing range (Borglin et al, 2008). Samples were measured ranging from 0.0-60.3 mg/L Fe. The average concentration of Fe in samples collected was 0.58 mg/L Fe. For many sites Fe peaks towards the end of winter and also in early summer.

These temporal plots (Figures 2-103) created an easy visual way to find outliers and double check data entry for possible mistakes. For the purpose of these plots any data points that were slightly negative were changed to 0.

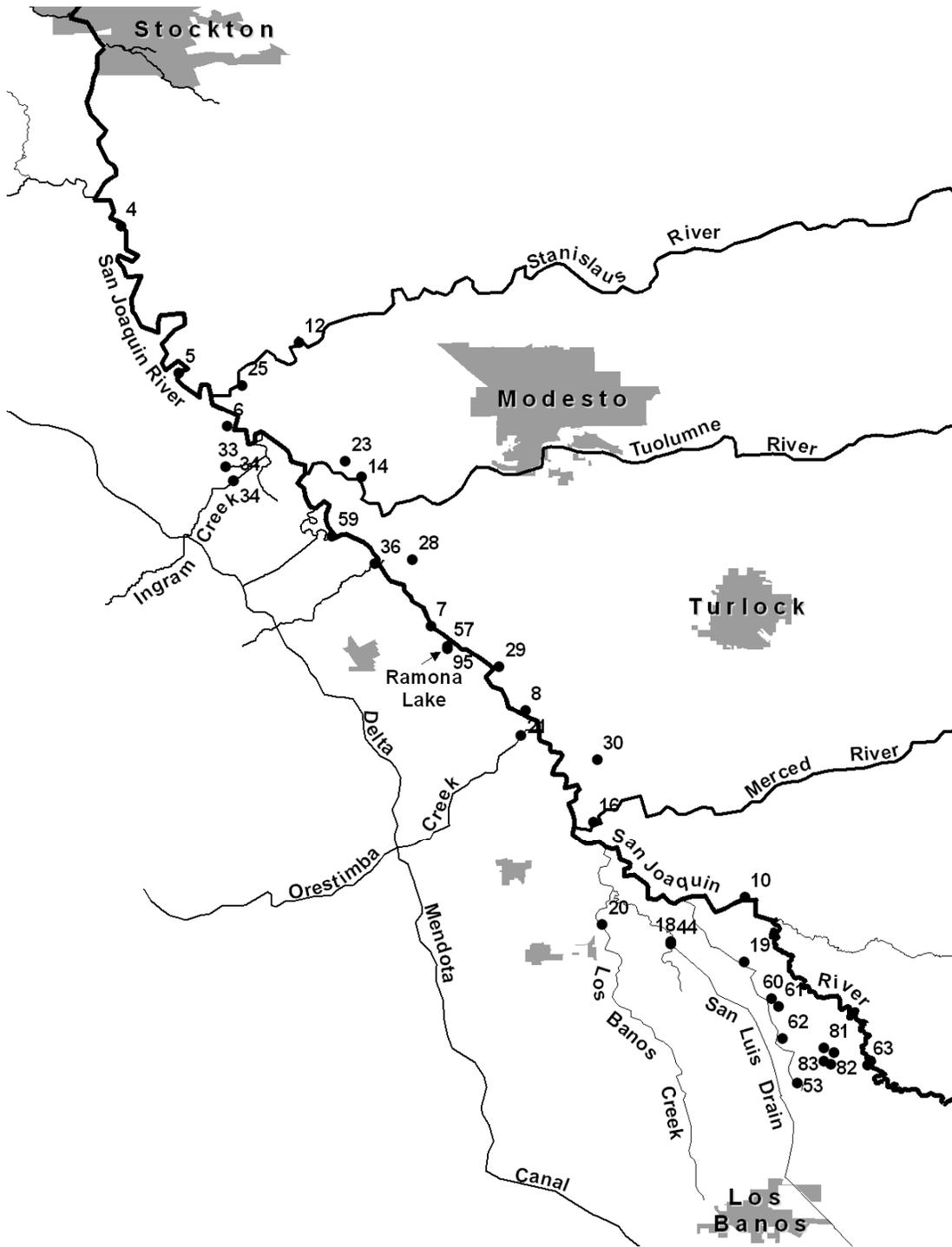
References

- American Public Health Association (APHA). 2005. Standard Methods for the Examination of Water and Wastewater, 21st Edition. American Public Health Association, Washington, DC.
- Borglin, S., W. Stringfellow, J. Hanlon. 2005. Standard Operating Procedures for the Up-Stream Dissolved Oxygen TMDL Project. LBNL/Pub-937.
- Borglin, S., Burks, R., Hanlon, J., Graham, J., Spier, C., Stringfellow, W., and Dahlgren, R., (2008) Methods overview, quality assurance, and quality control, University of the Pacific, Stockton, CA
- Borglin, S.E., Burks, R.D., Hanlon, J.S., Stringfellow, W.T. (2008) EERP Lab Protocol Book, University of the Pacific, Stockton, CA.
- Graham, J., Hanlon, J.S., Stringfellow, W.T., (2008) EERP Field Protocol Book, University of the Pacific, Stockton, CA.
- Stringfellow, W.T., et al., (2008) Evaluation of Vegetated Ditches, Ponds, and Wetlands as BMPs for Mitigating the Water Quality Impact of Irrigated Agriculture in the San Joaquin Valley, University of the Pacific, Stockton, CA
- YSI Environmental Operations Manual (2005) 6-Series Environmental Monitoring Systems, Yellow Springs, OH.

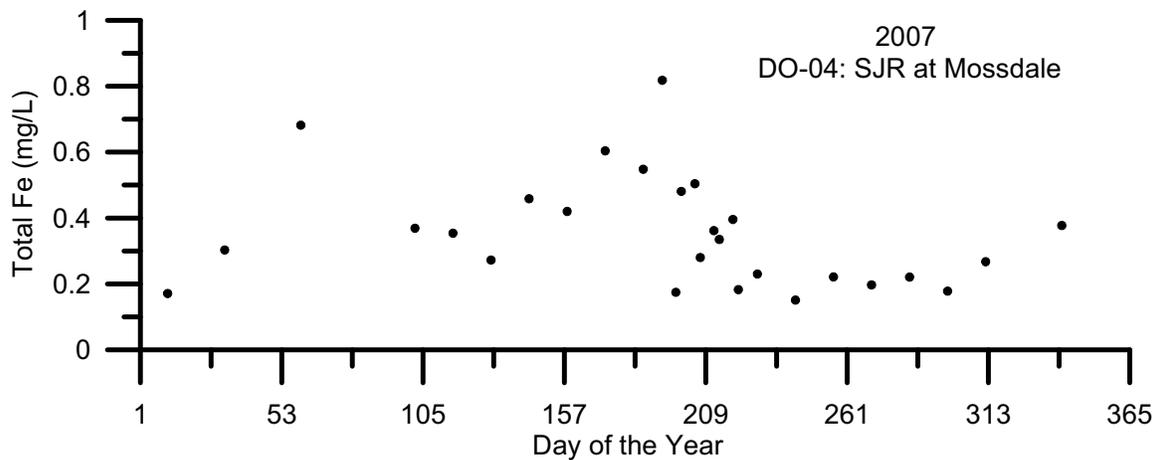
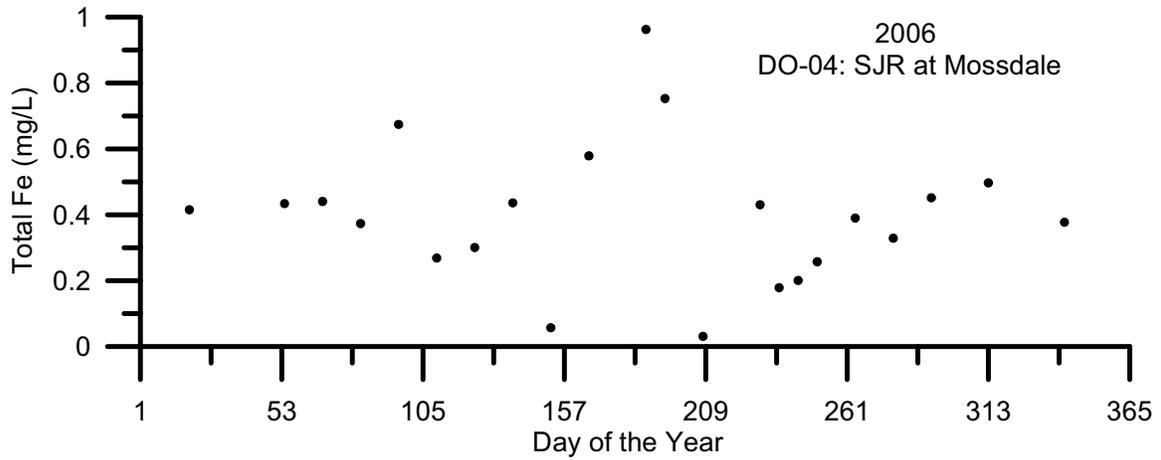
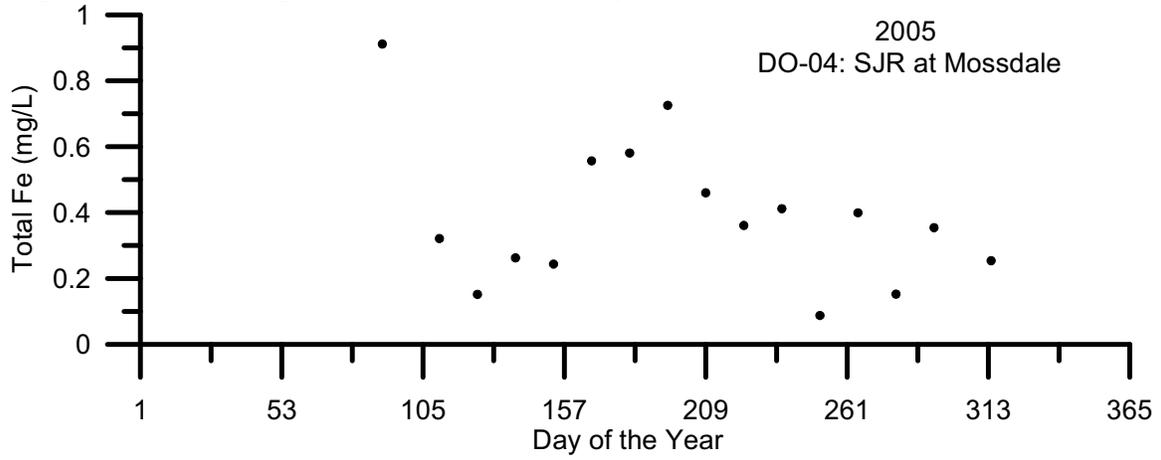
Table 1: EERP Sampling Site List

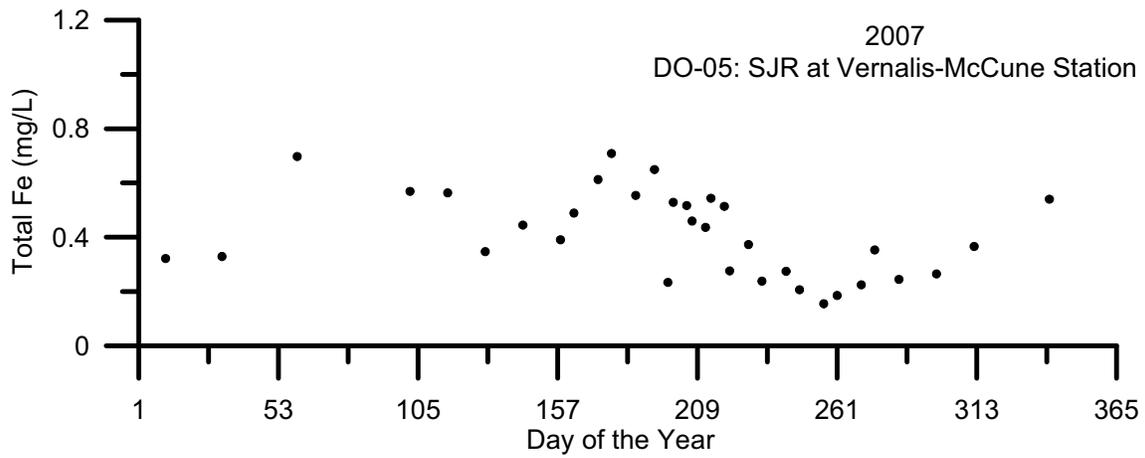
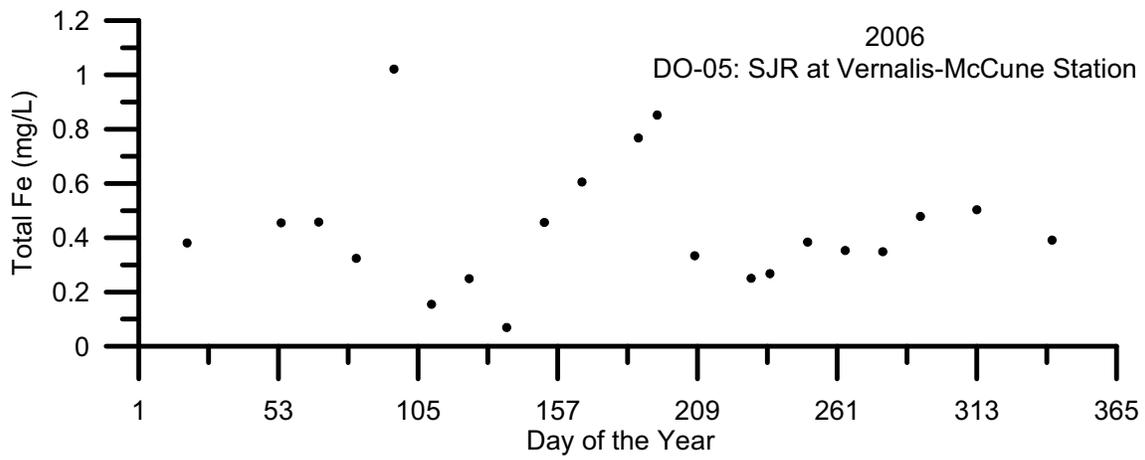
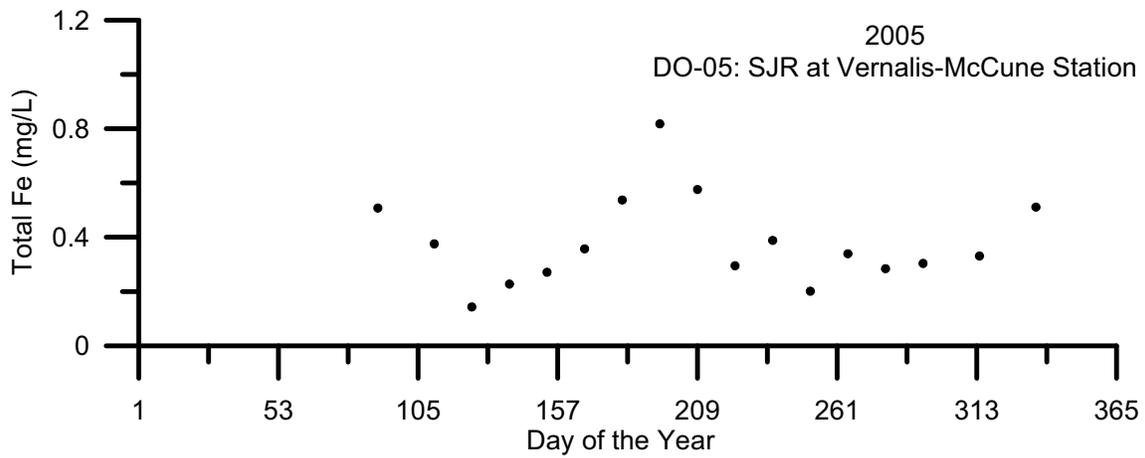
DO Number	Site Name	Type
4	SJR at Mossdale	Core sites
5	SJR at Vernalis-McCune Station (River Club)	Core sites, BMP
6	SJR at Maze	Core sites, BMP
7	SJR at Patterson	Core sites, BMP
8	SJR at Crows Landing	Core sites, BMP
10	SJR at Lander Avenue	Core sites
12	Stanislaus River at Caswell Park	Core sites
14	Tuolumne River at Shiloh Bridge	Core sites
16	Merced River at River Road	Core sites
18	Mud Slough near Gustine	Core sites, Wetland
19	Salt Slough at Lander Avenue	Core sites, Wetland
20	Los Banos Creek Flow Station	Core sites, Wetland
21	Orestimba Creek at River Road	Core sites, BMP
23	Modesto ID Lateral 5 to Tuolumne	Core sites
25	Modesto ID Main Drain to Stan. R. via Miller Lake	Core sites
28	Turlock ID Westport Drain Flow station	Core sites
29	Turlock ID Harding Drain	Core sites
30	Turlock ID Lateral 6 & 7 at Levee	Core sites
33	Hospital Creek	Intermittent, BMP
34	Ingram Creek	Core sites, BMP
36	Del Puerto Creek Flow Station	Core sites, BMP
44	San Luis Drain End	Core sites
53	Salt Slough at Wolfsen Road	Wetland
57	Ramona Lake Drain	Core sites, BMP
59	SJR Laird Park	Core sites
60	Moffit 1 South	Wetland
61	Deadman's Slough	Wetland
62	Mallard Slough	Wetland
63	Inlet C Canal	Wetland
80	South Marsh-1-Inlet	Wetland
81	South Marsh-1-Outlet	Wetland
82	South Marsh-3-Inlet	Wetland
83	South Marsh-3-Outlet	Wetland
95	Ramona drain at Ramona Lake	BMP, Intermittent

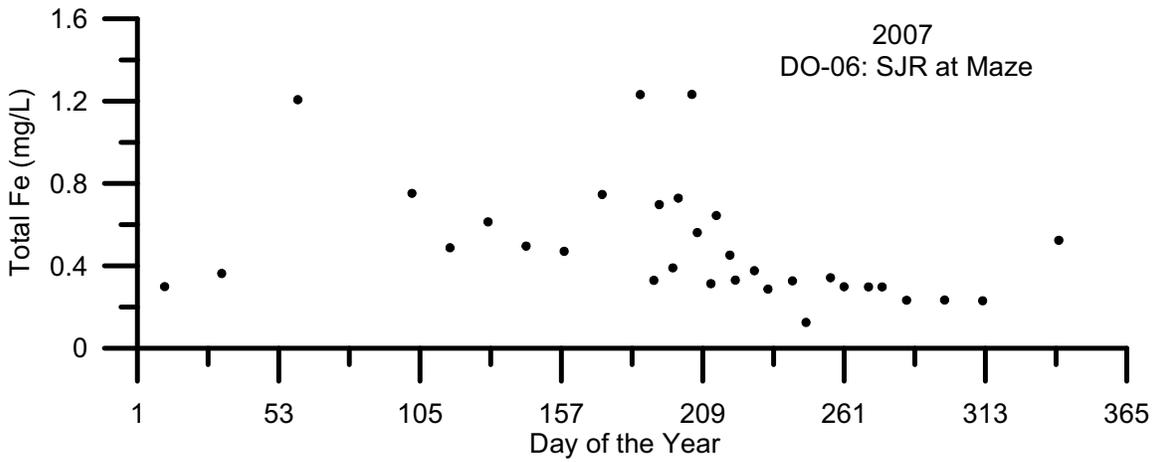
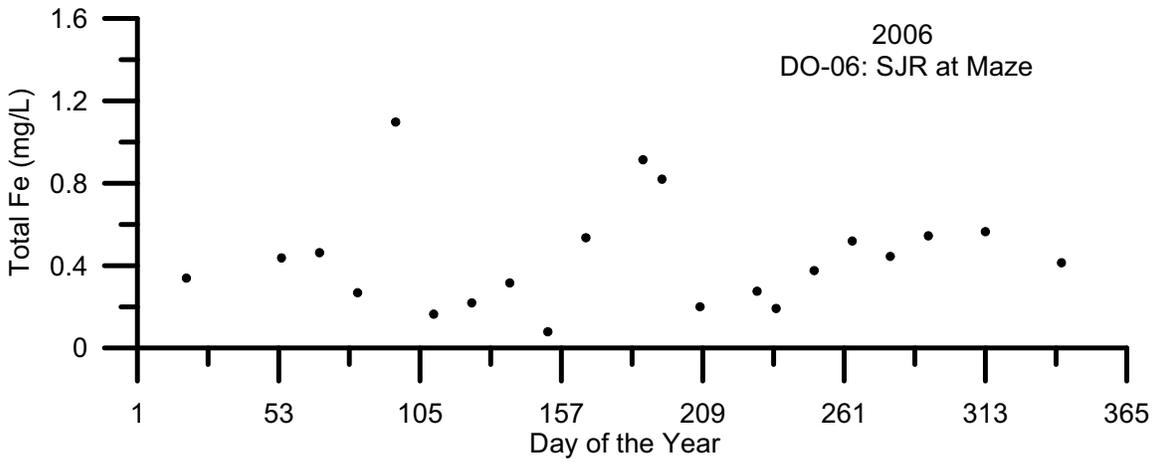
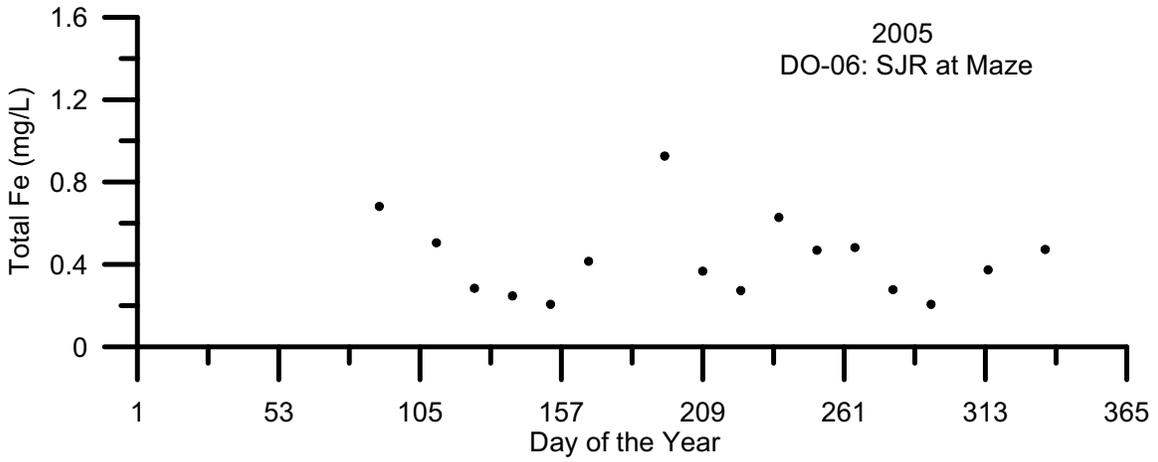
Figure 1: EERP Sampling Site Map of SJR Watershed and Tributaries

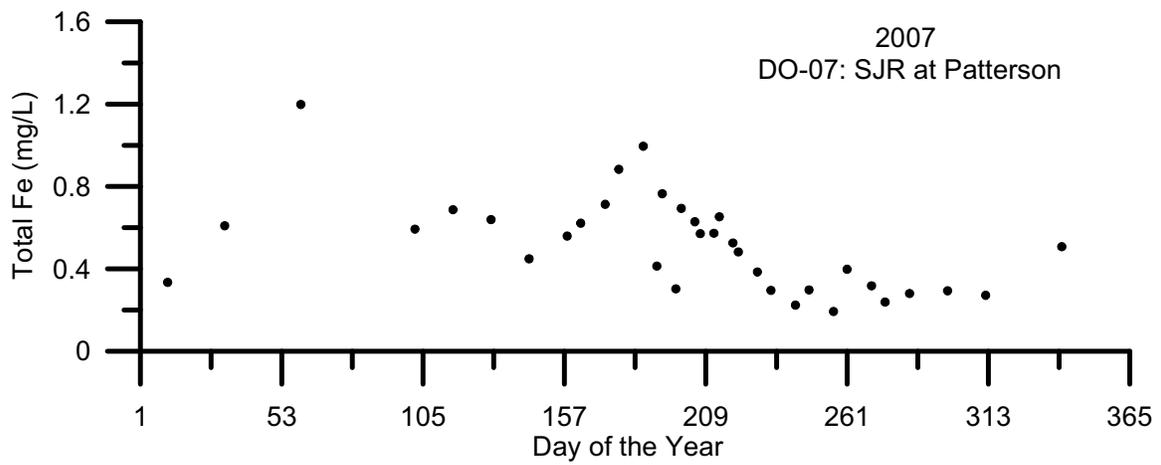
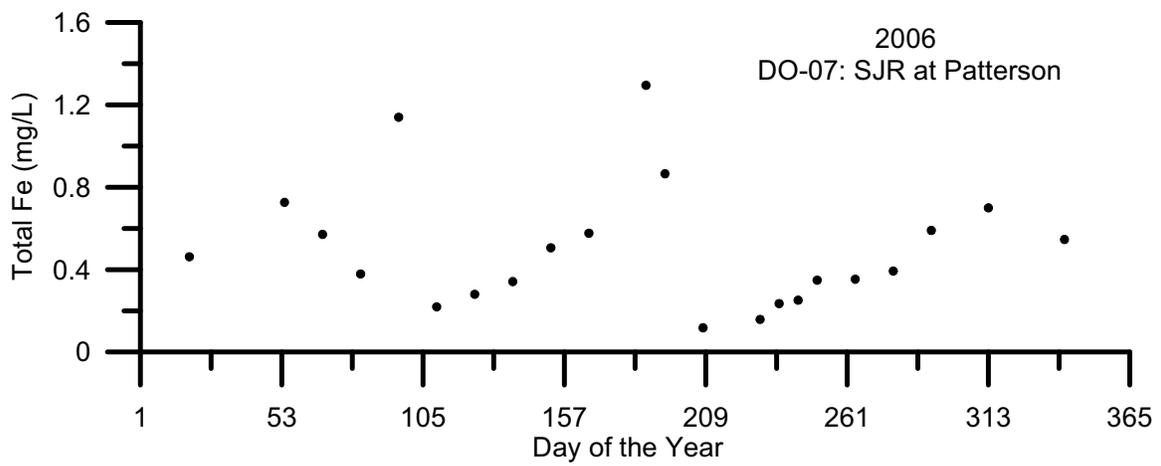
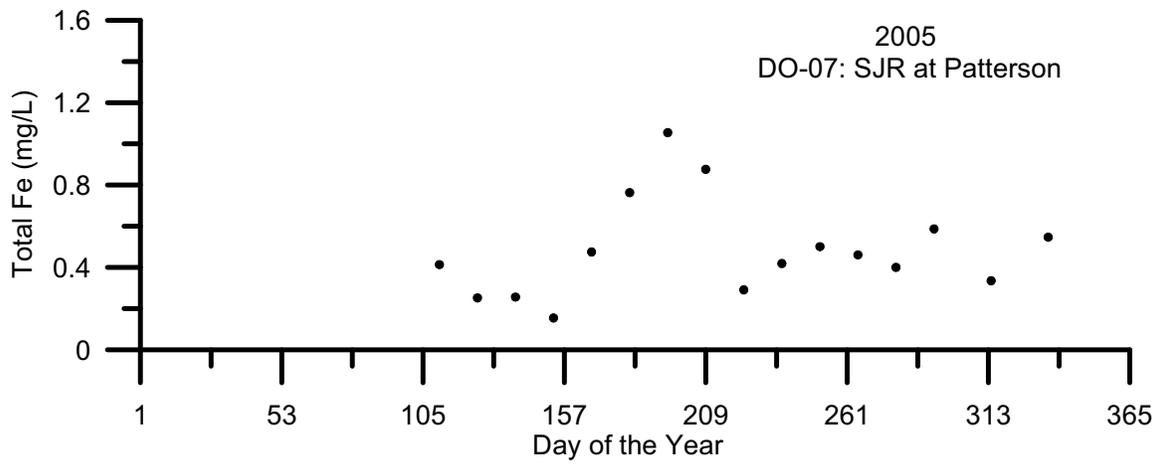


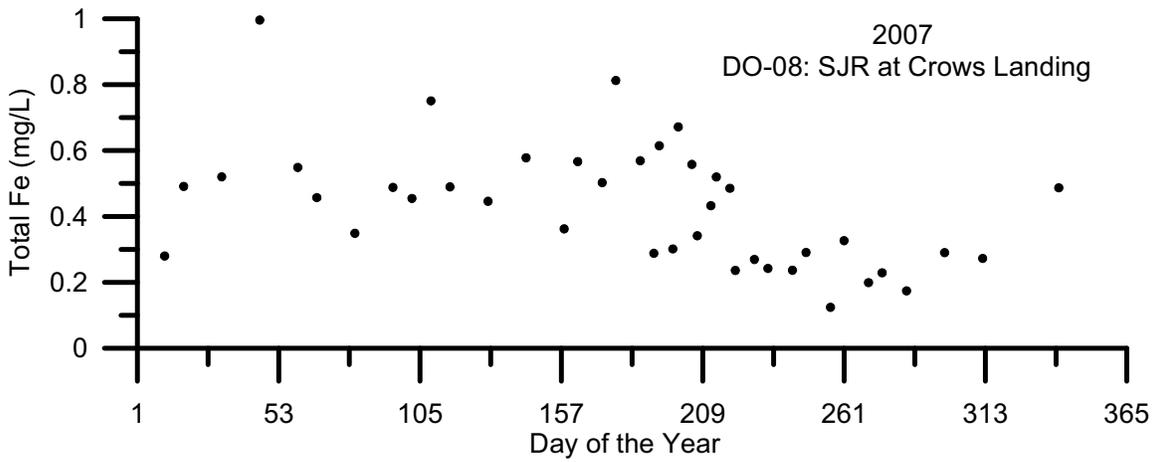
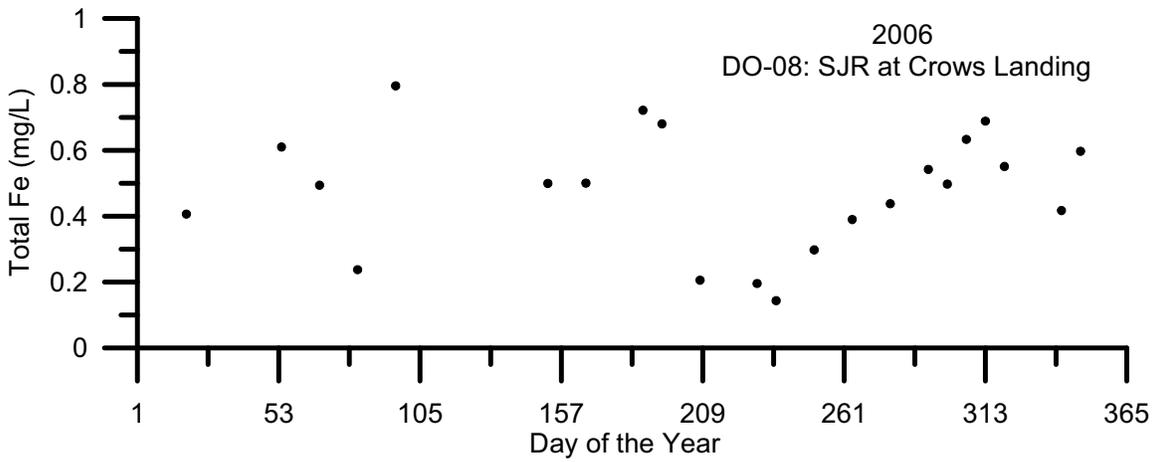
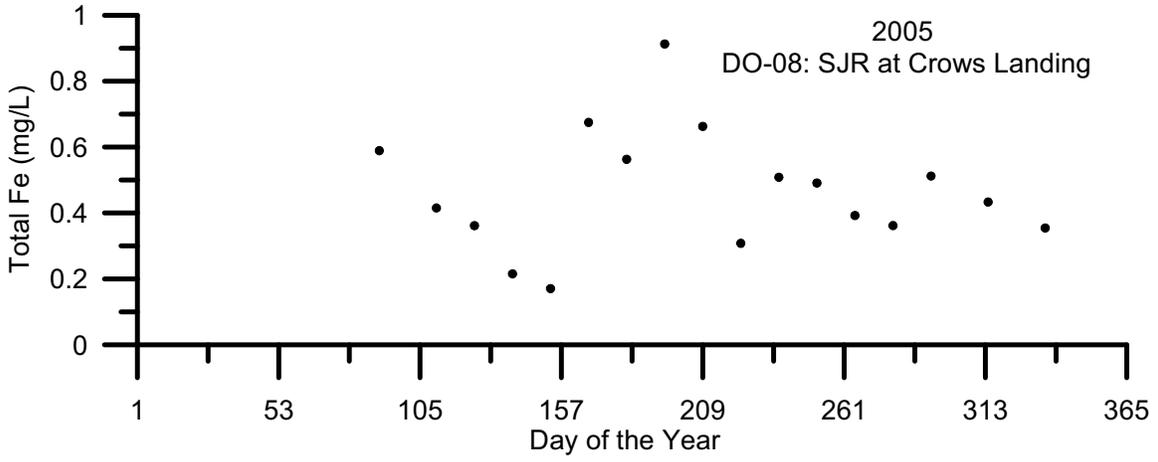
Figures 2 -103: Temporal Plots of Total Fe By Site ID

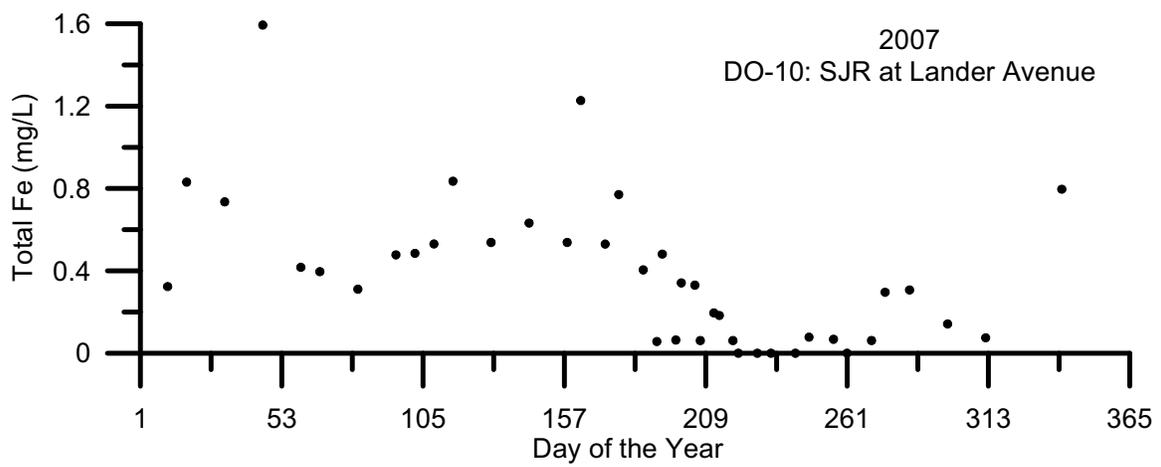
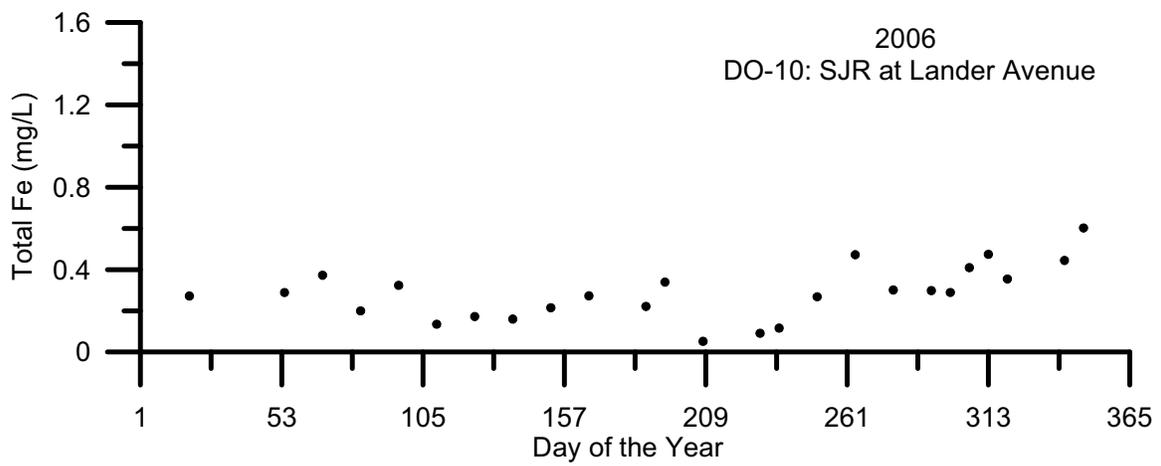
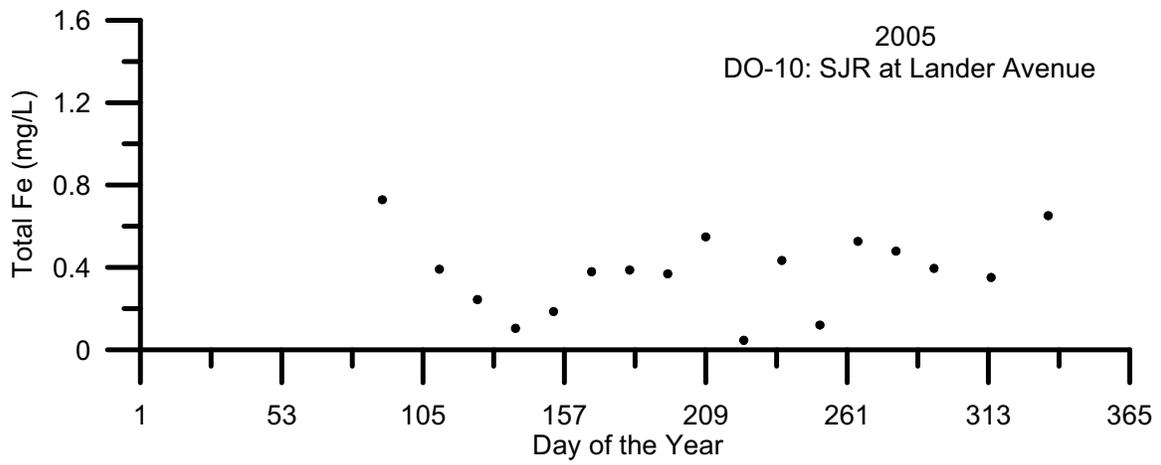


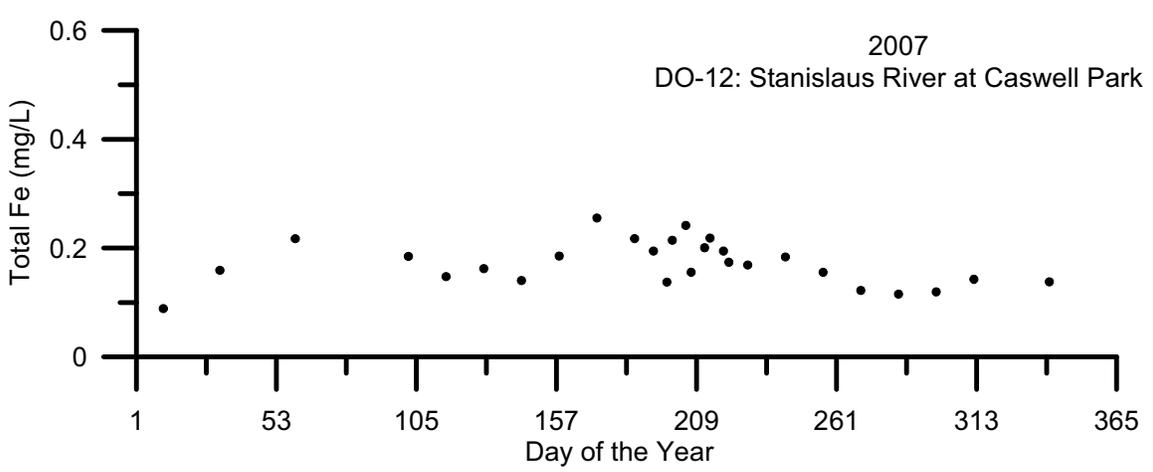
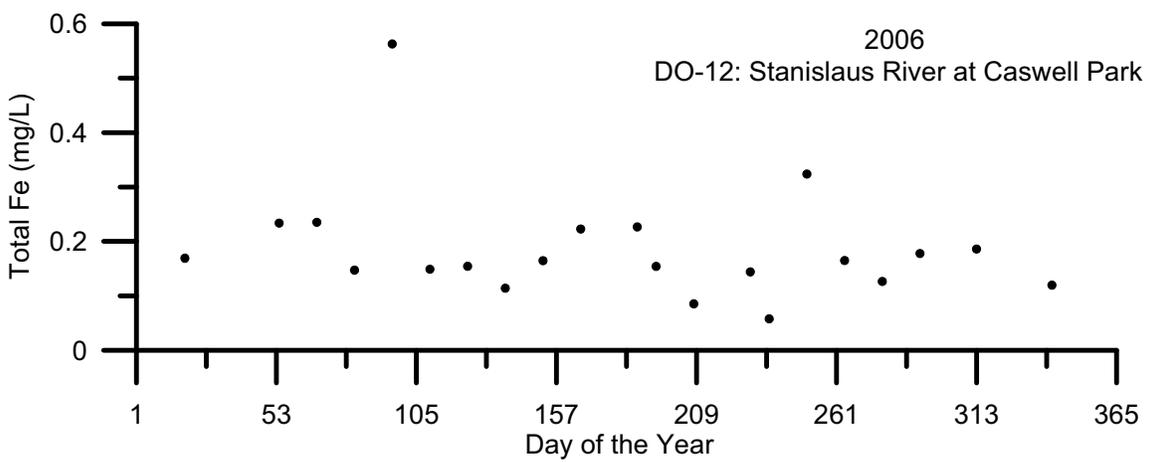
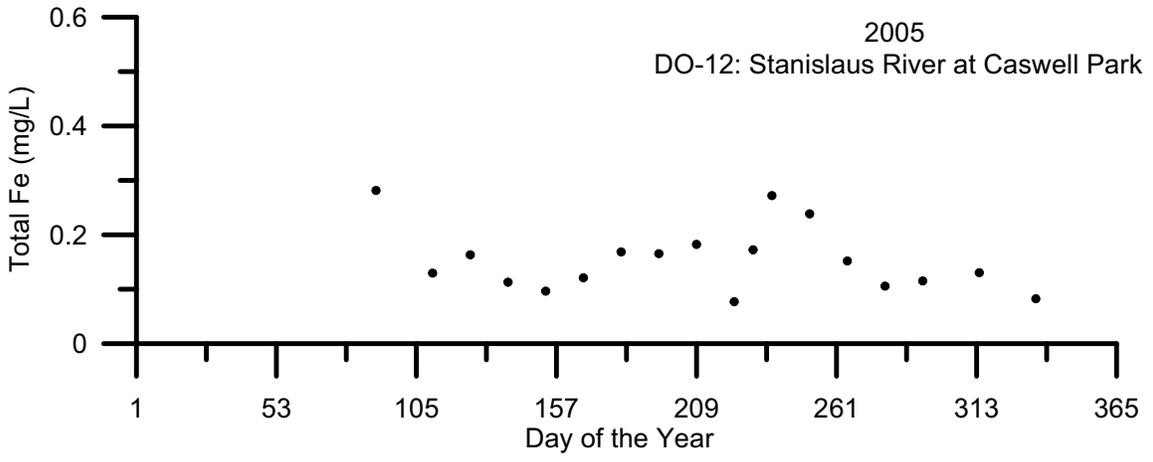


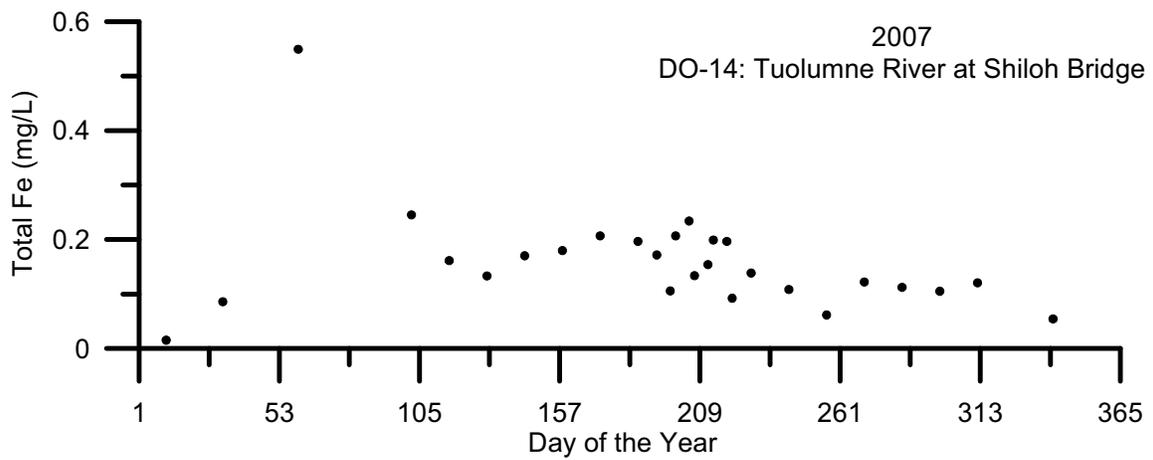
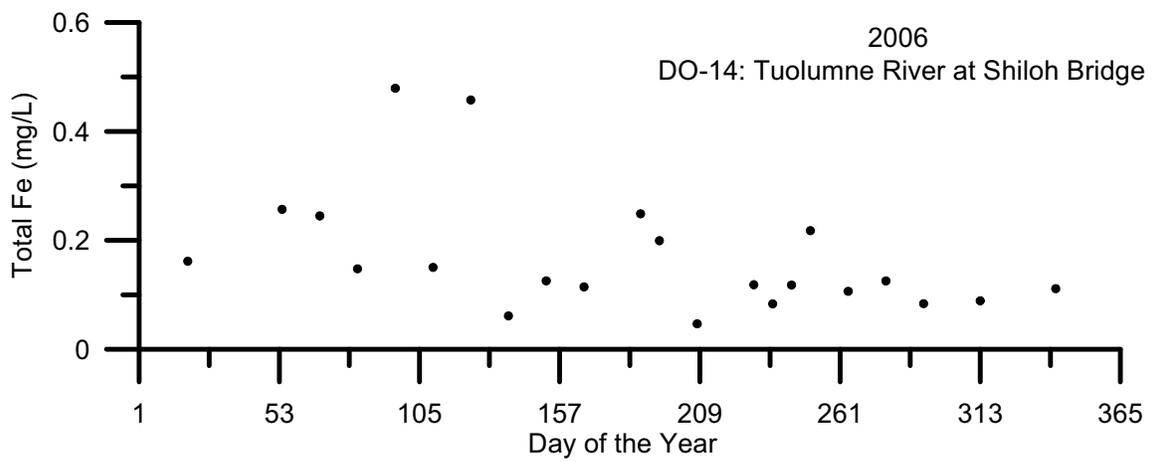
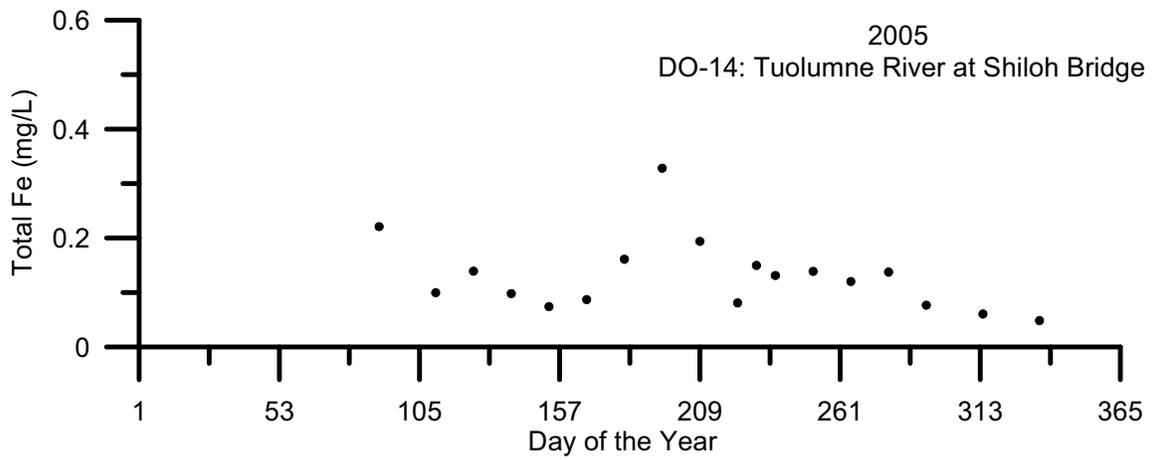


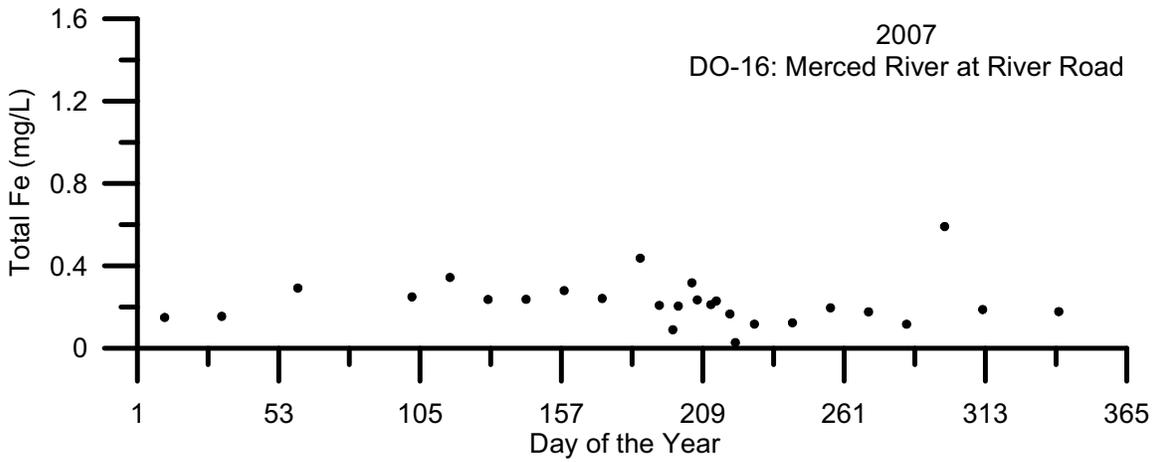
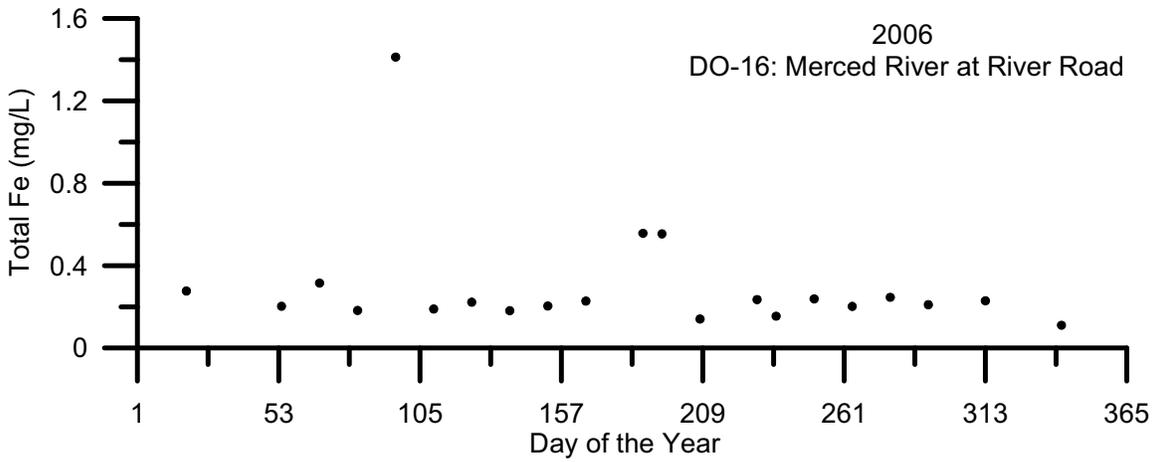
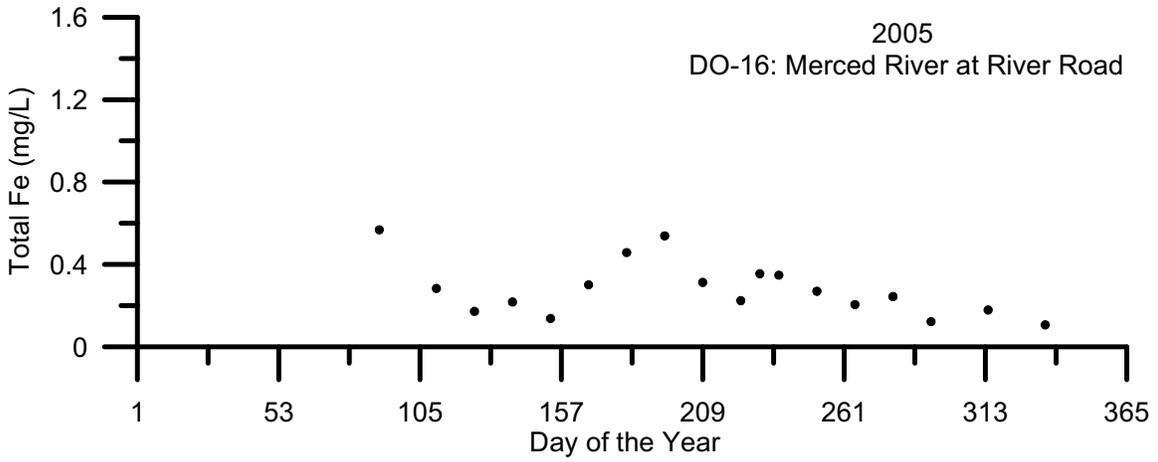


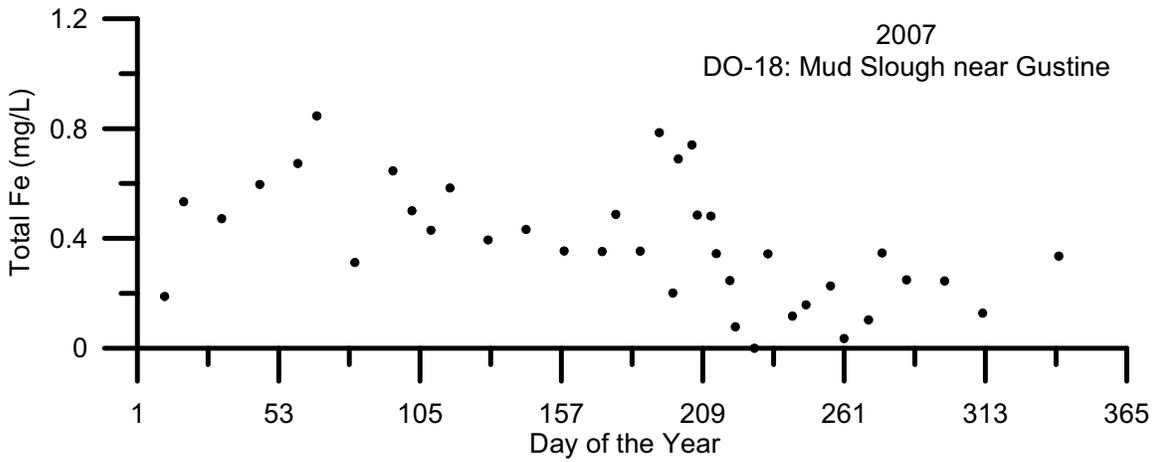
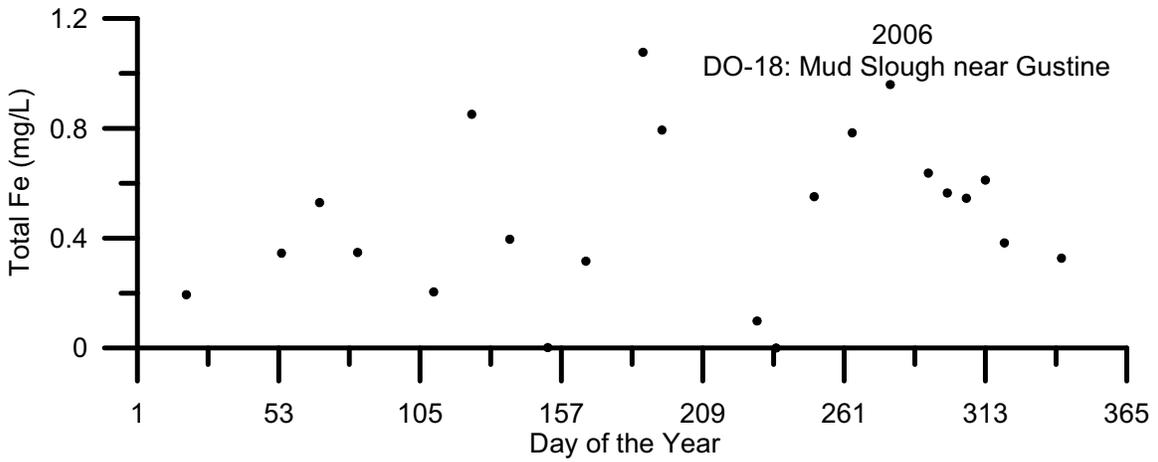
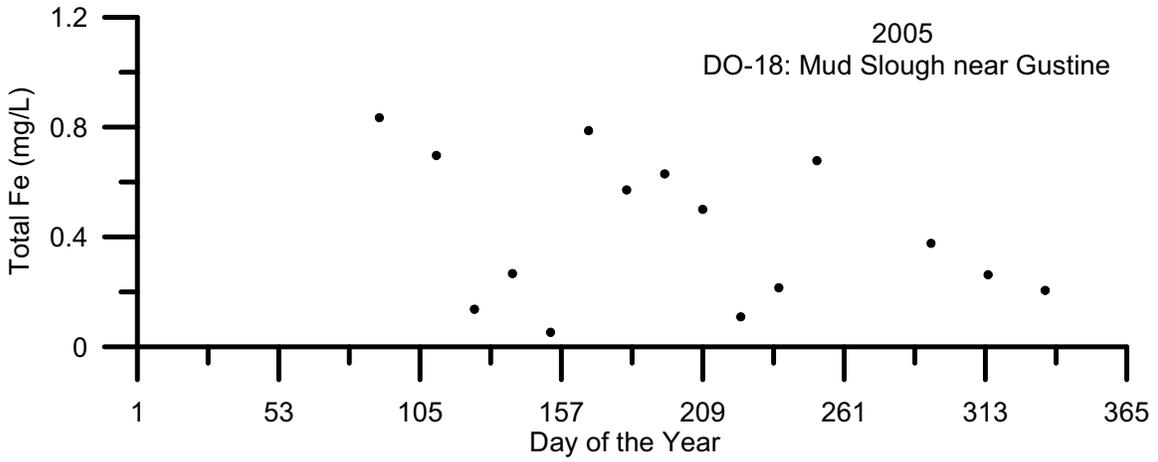


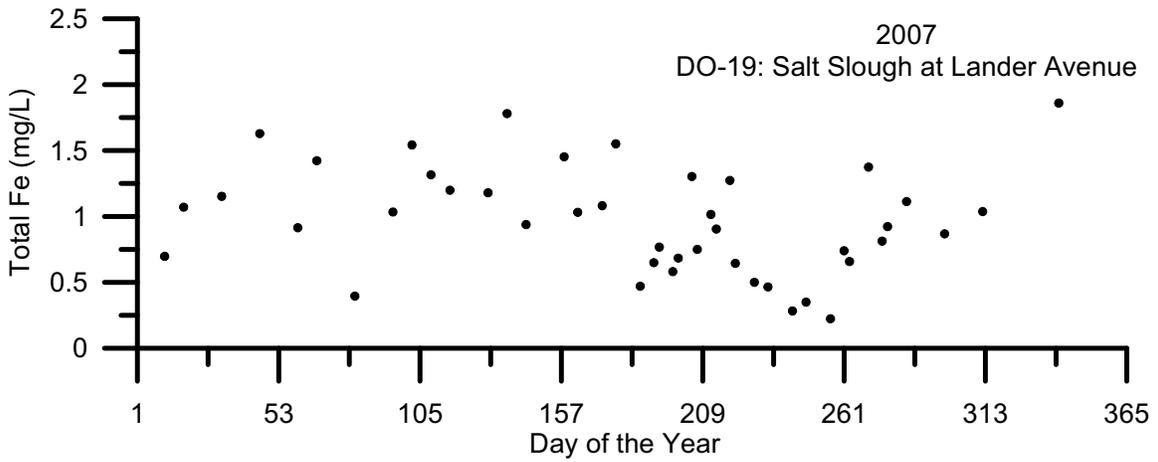
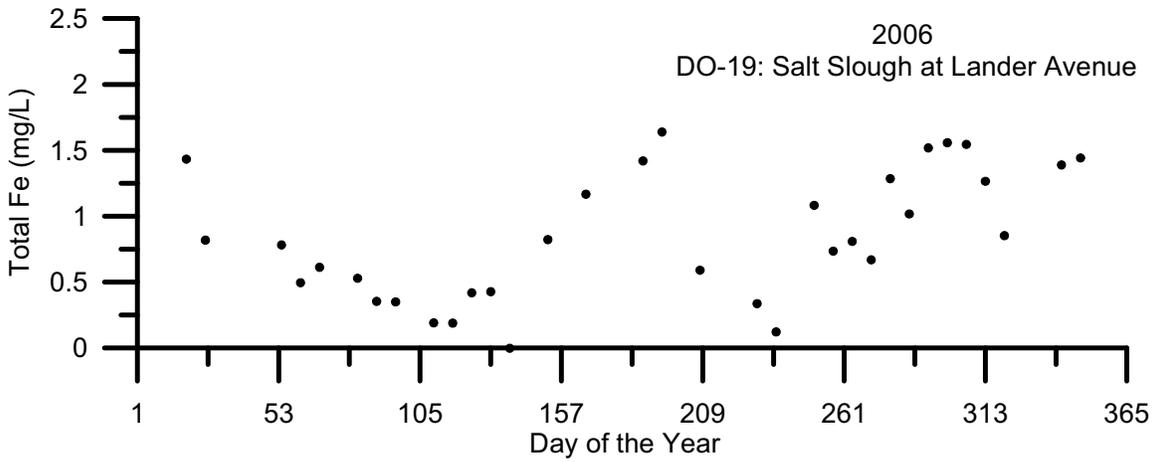
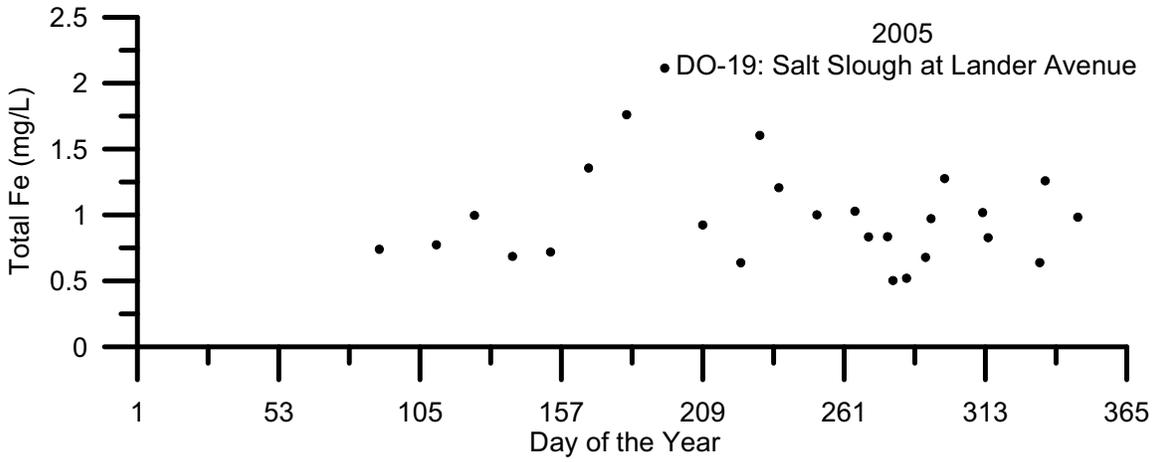


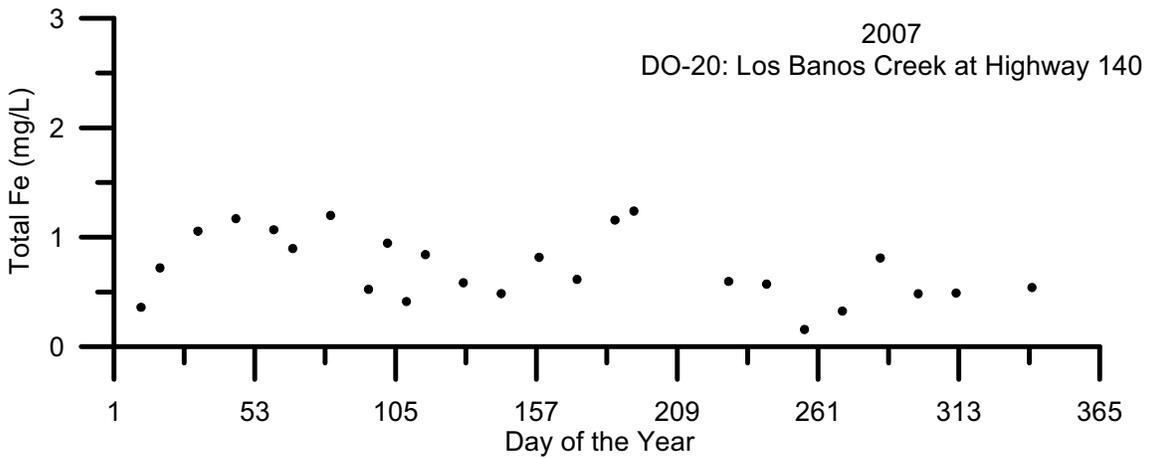
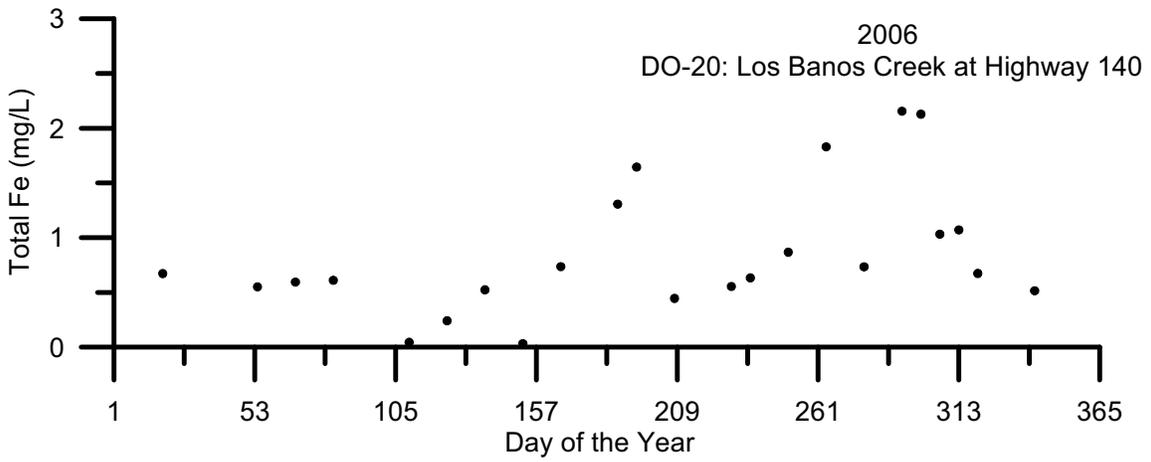
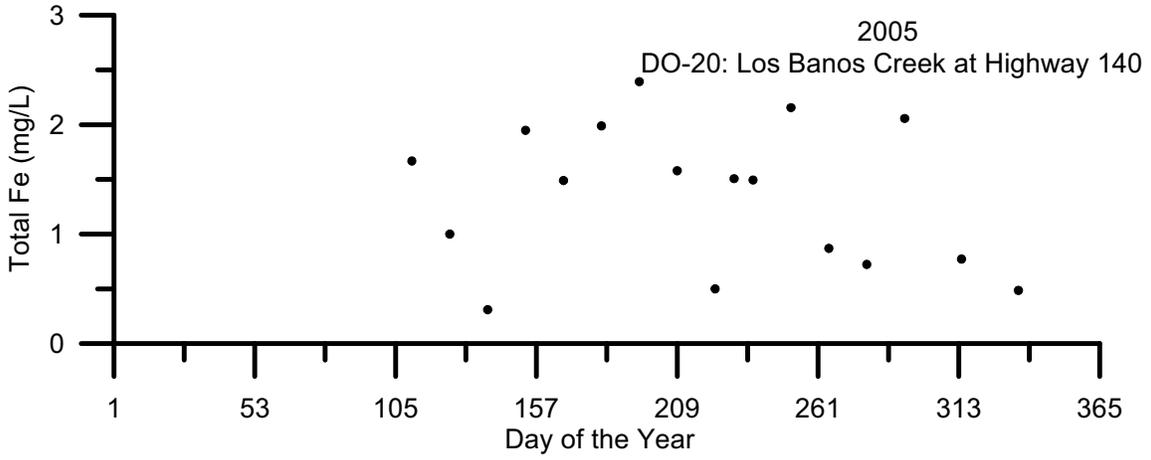


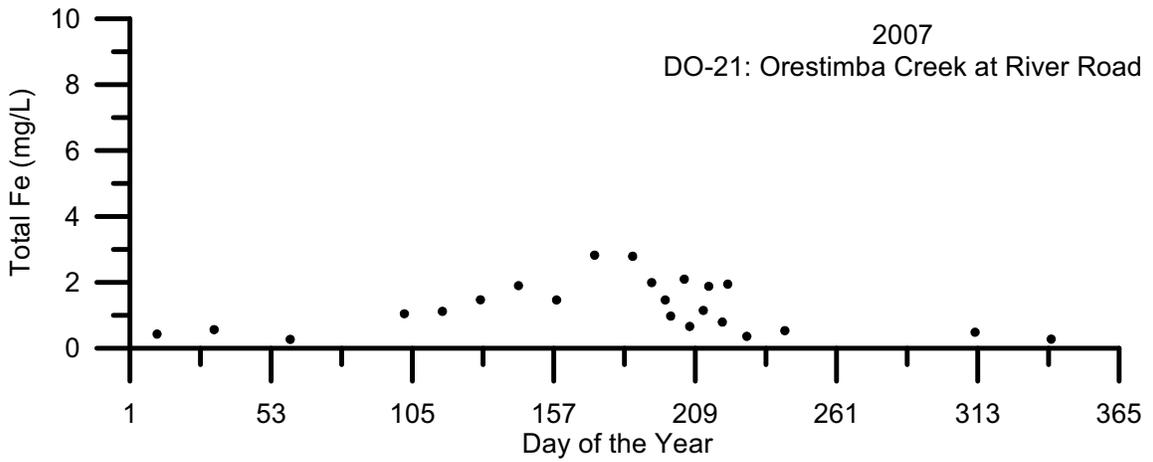
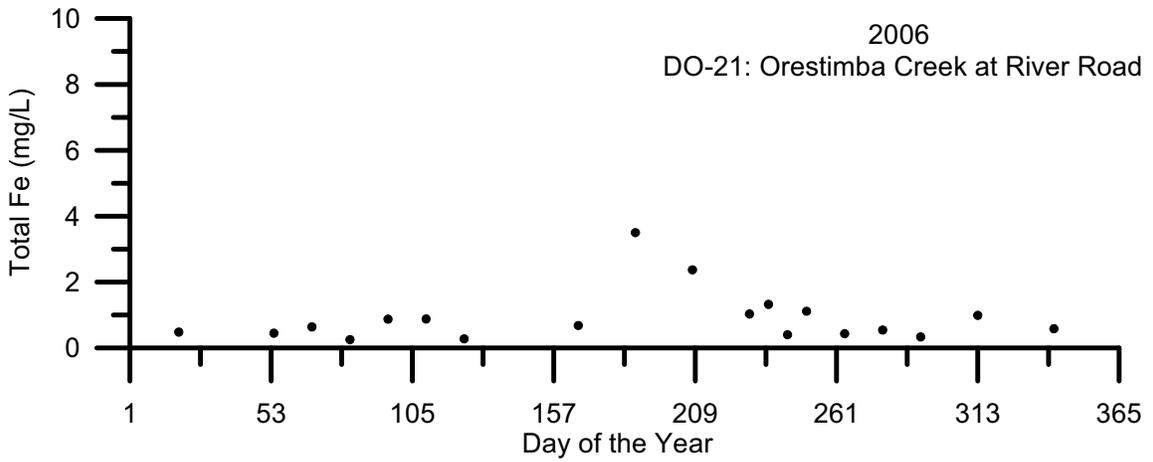
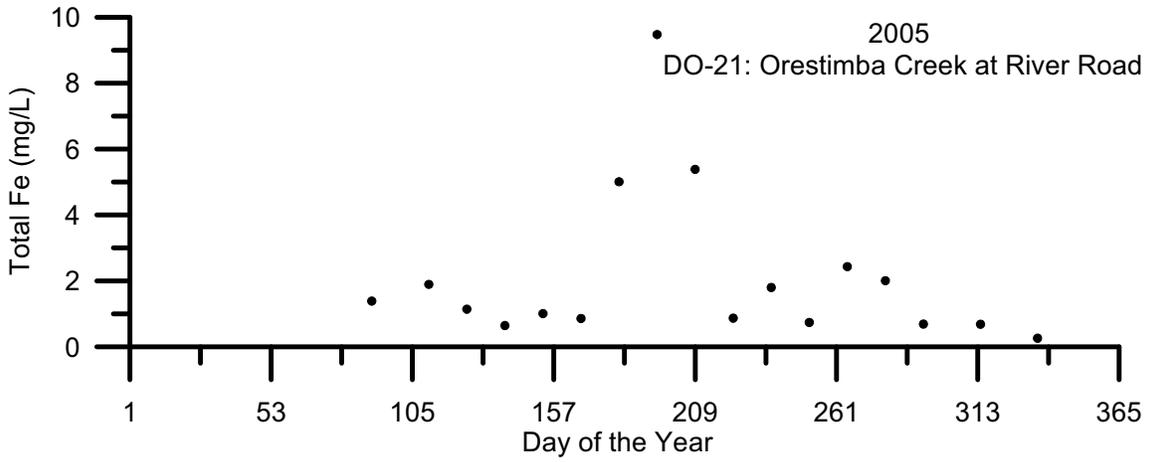


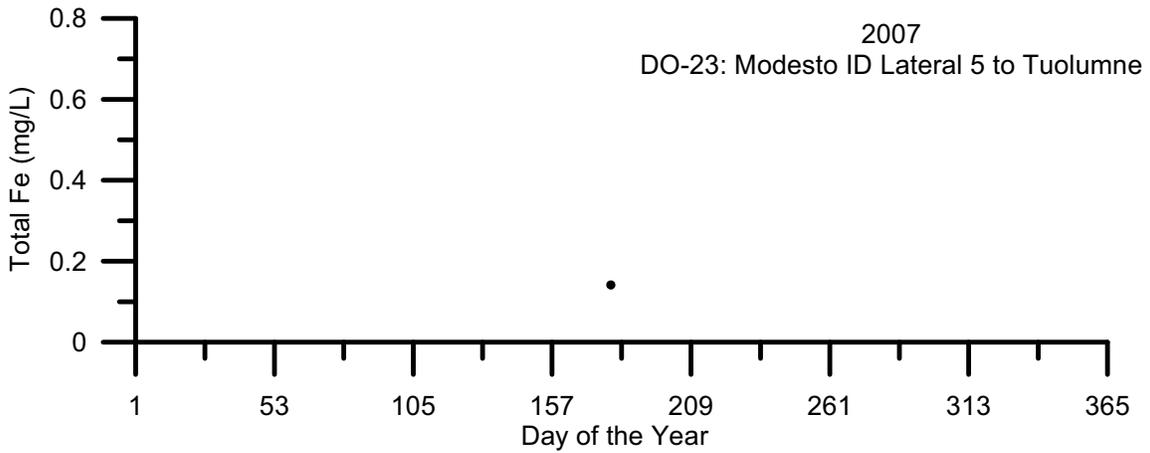
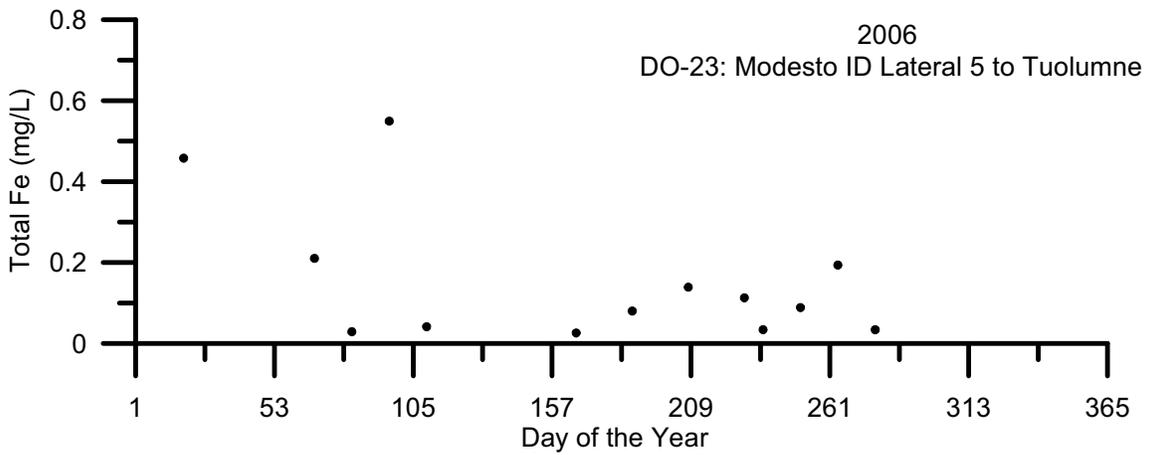
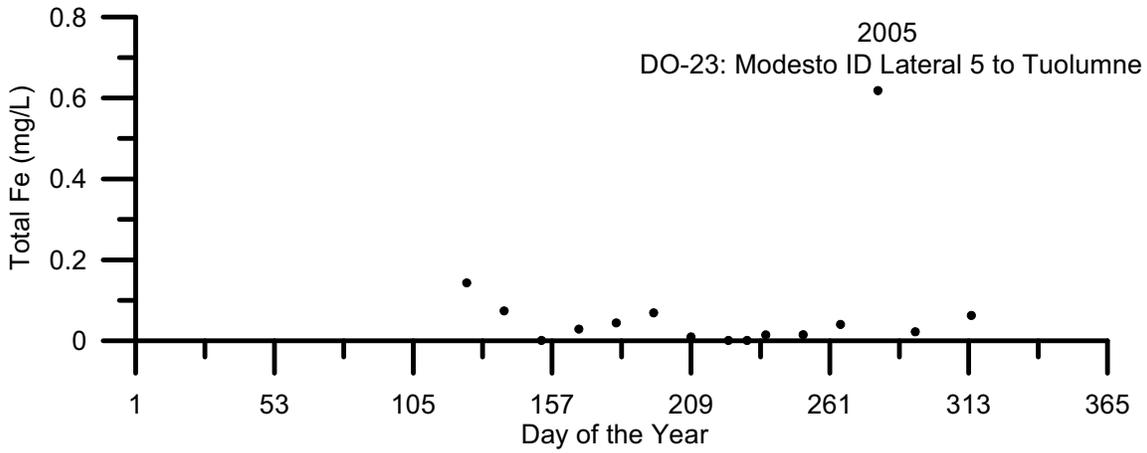


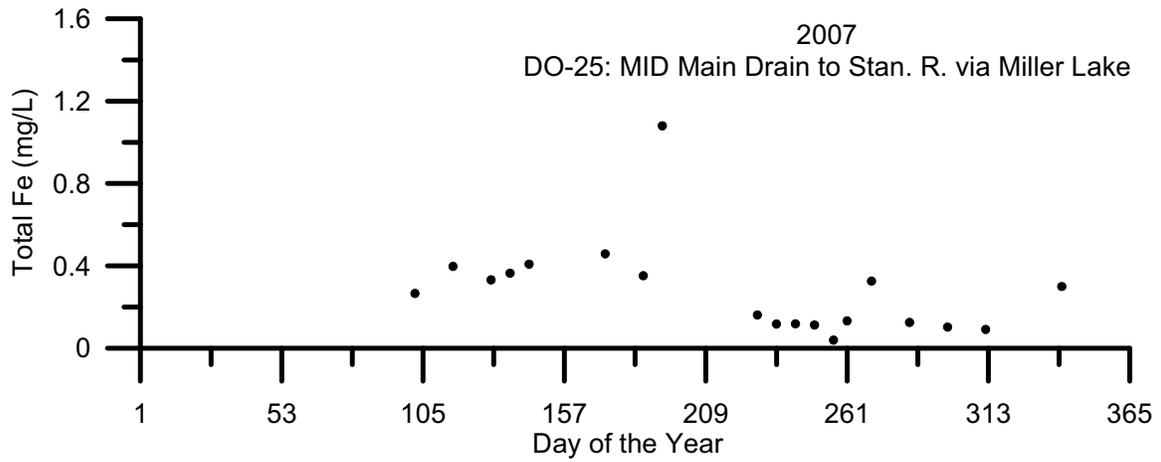
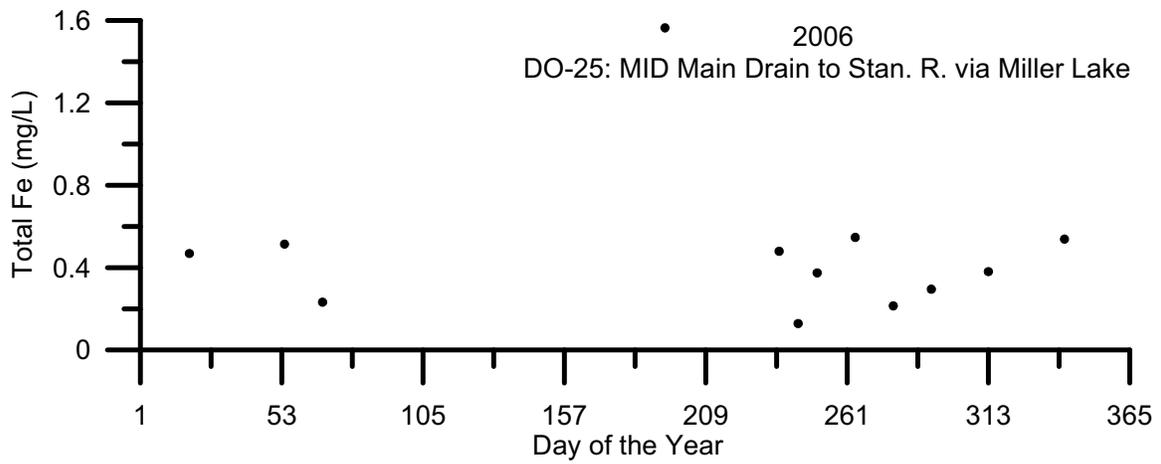
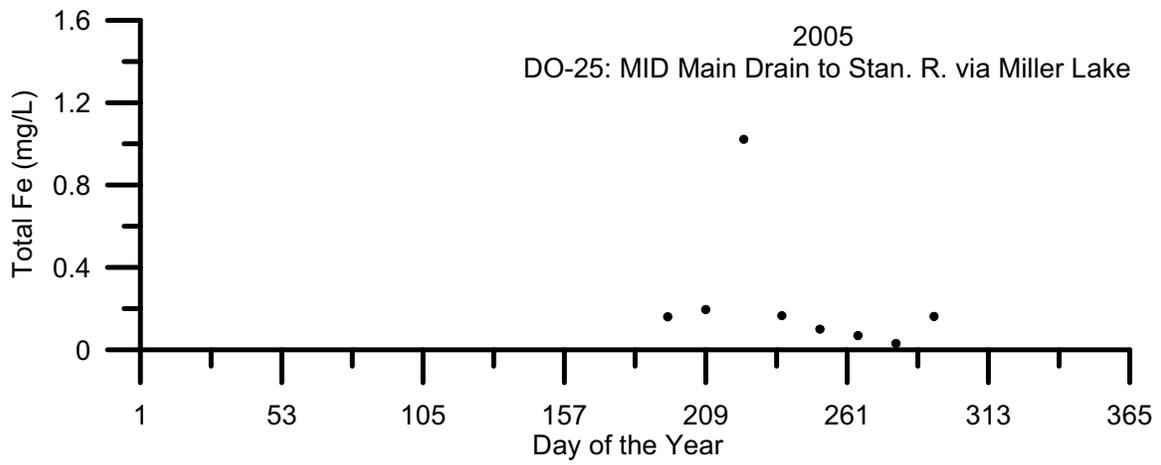


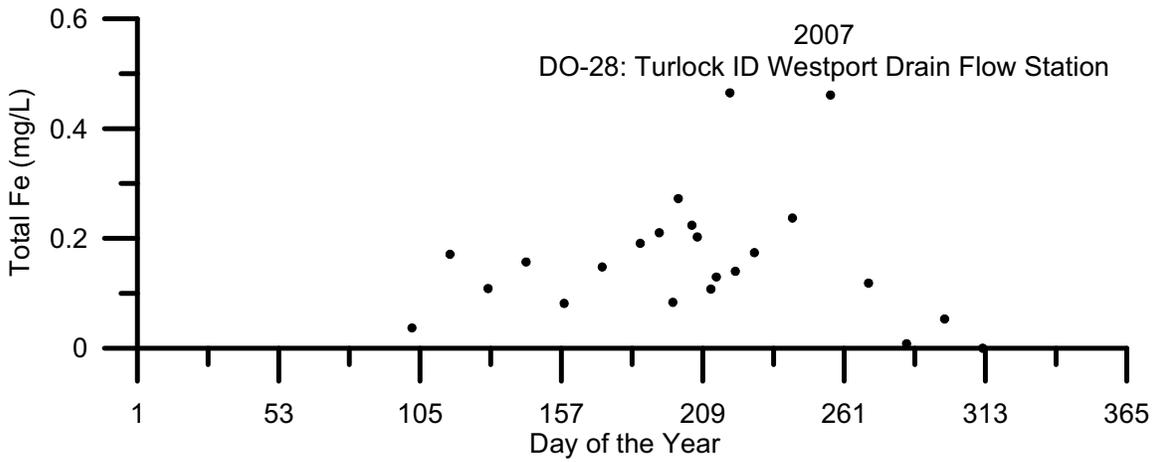
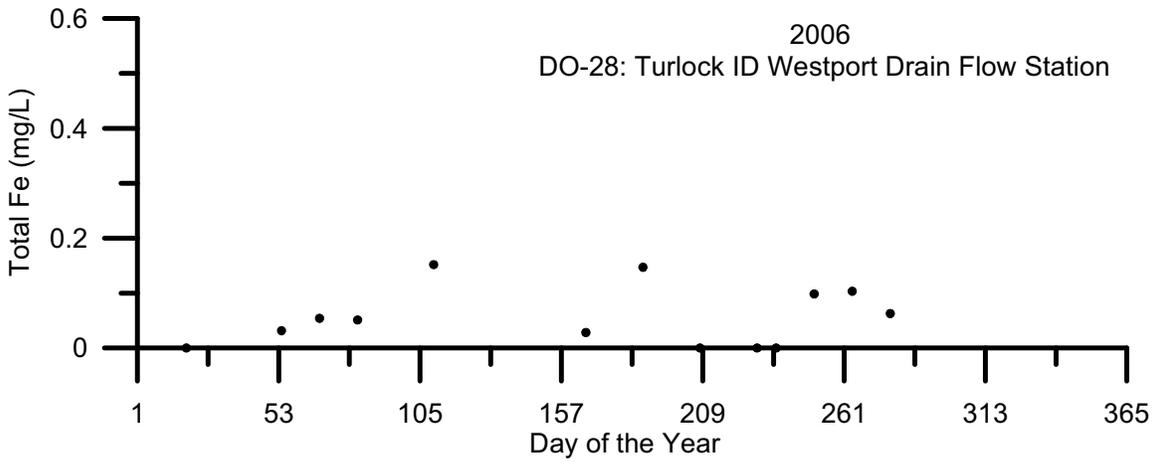
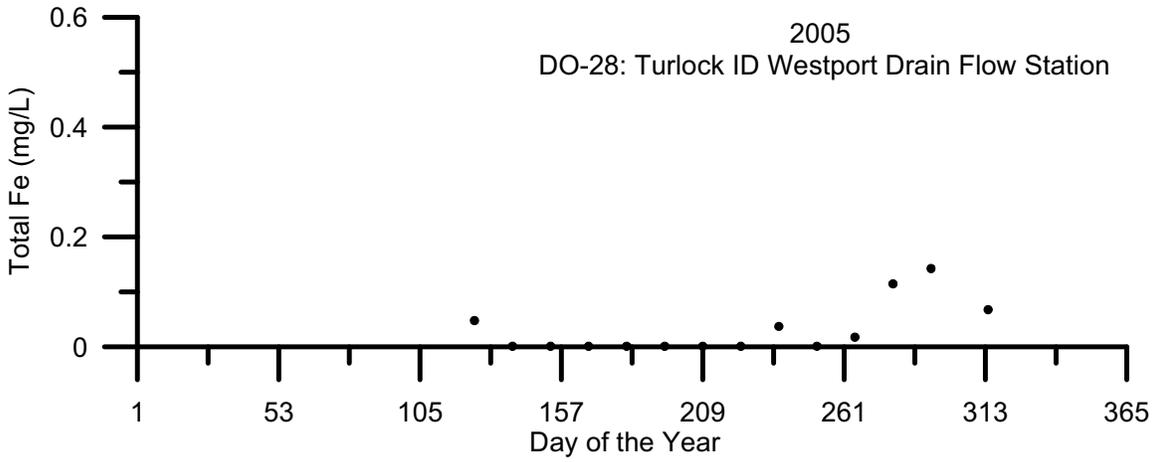


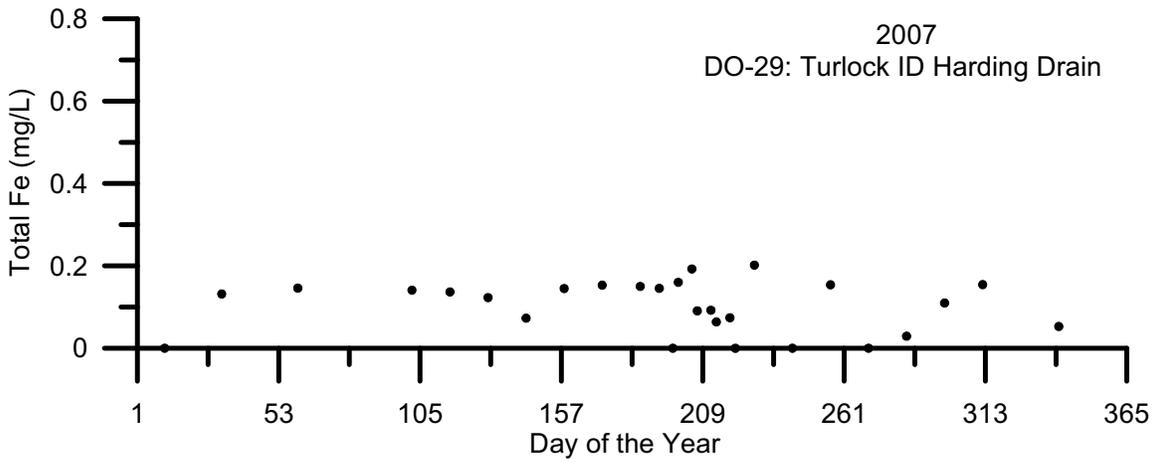
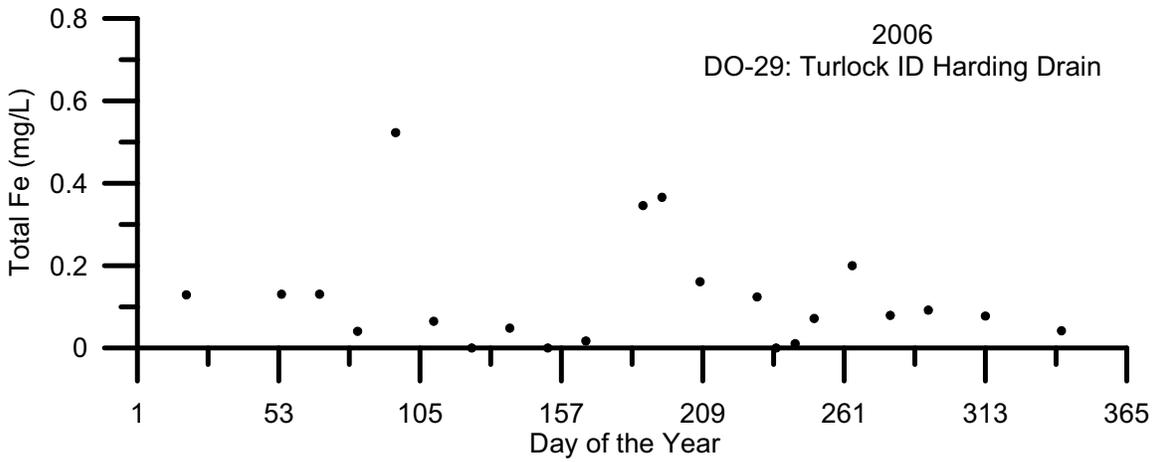
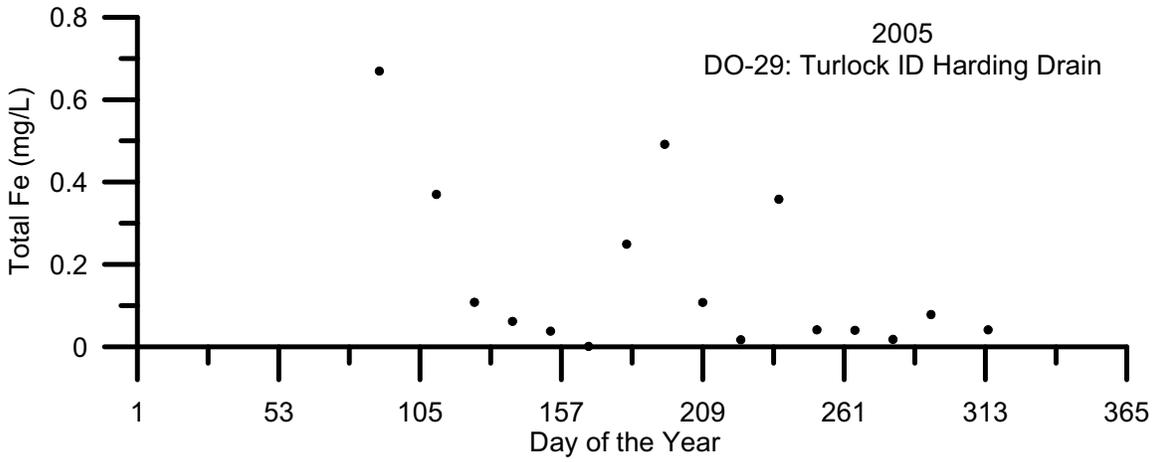


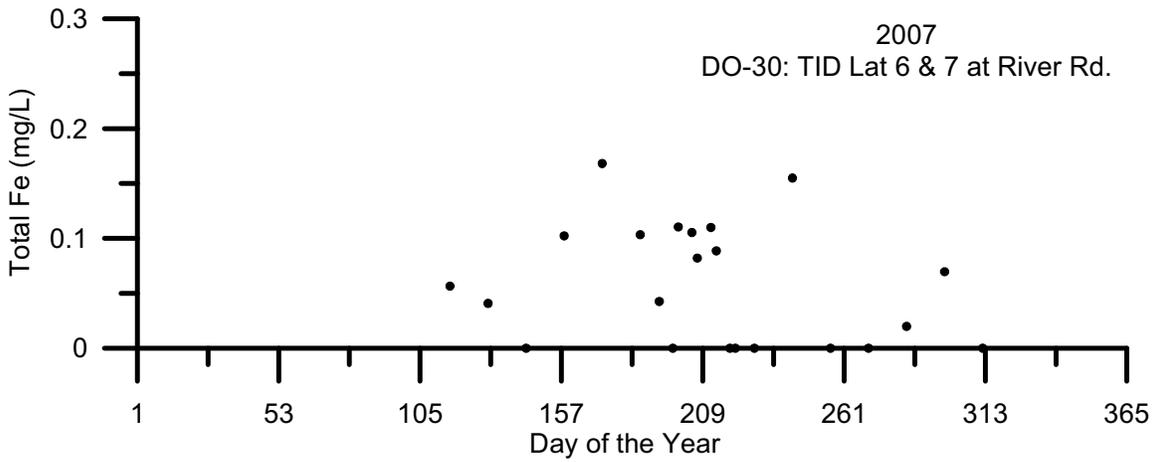
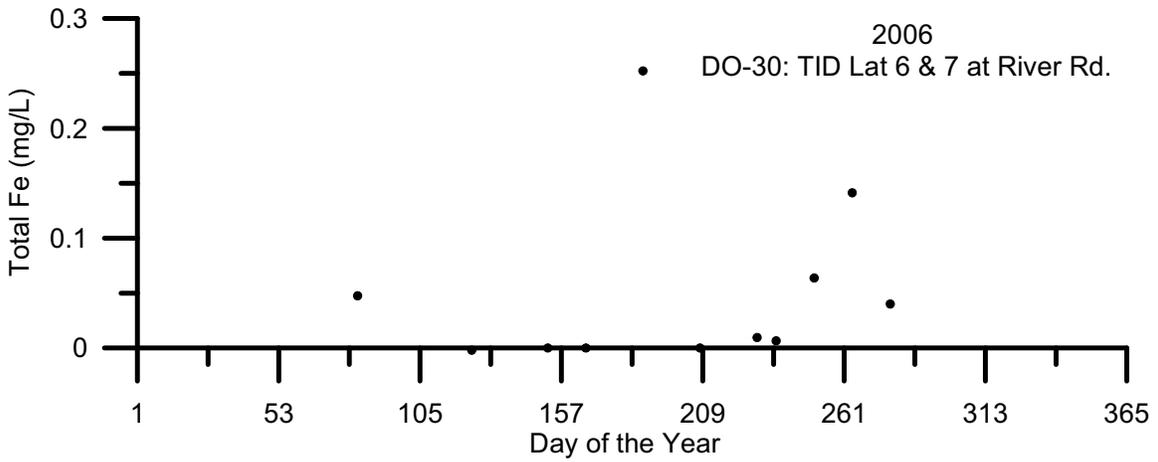
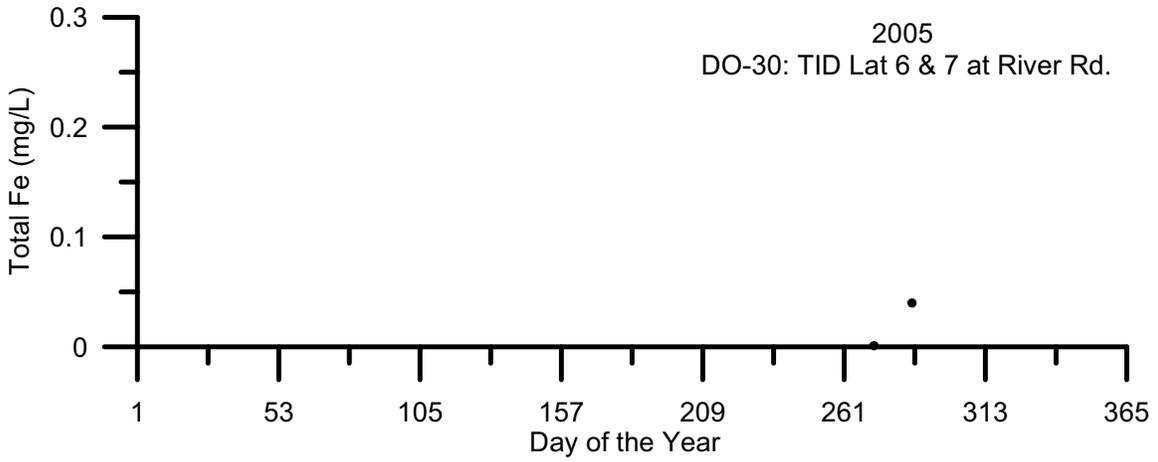


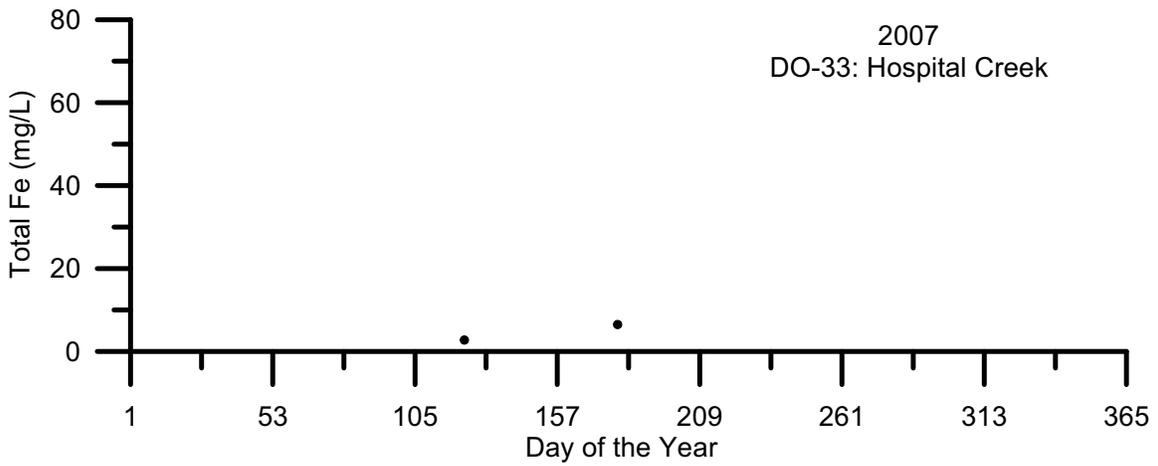
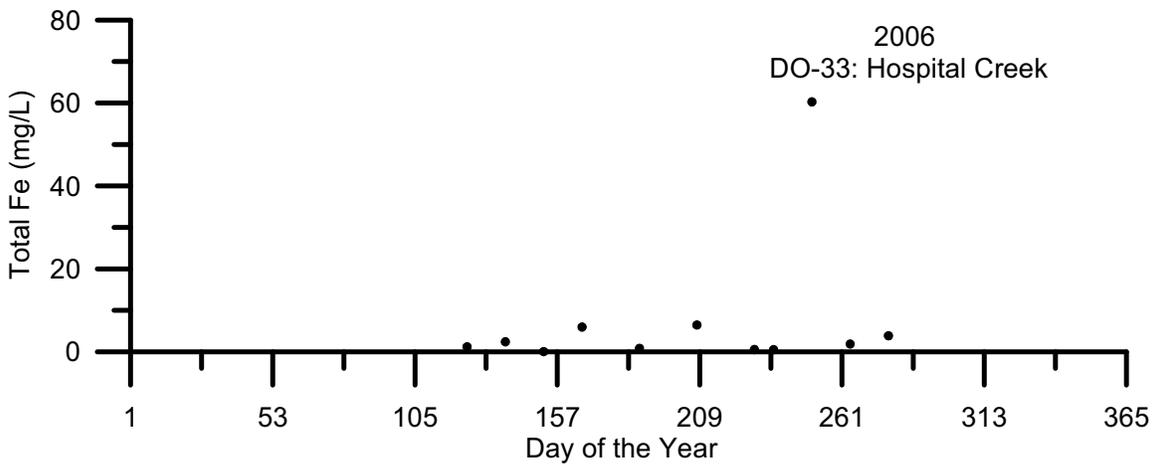
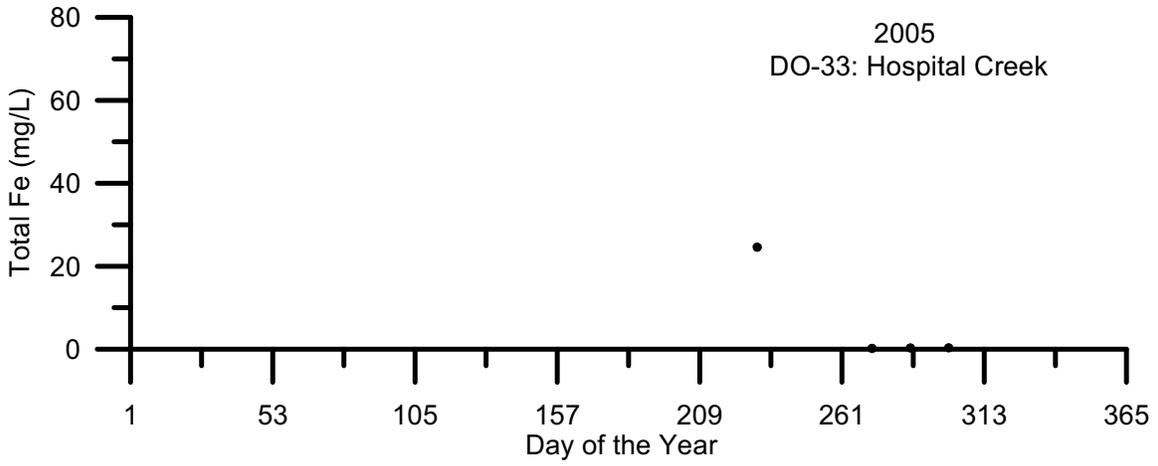


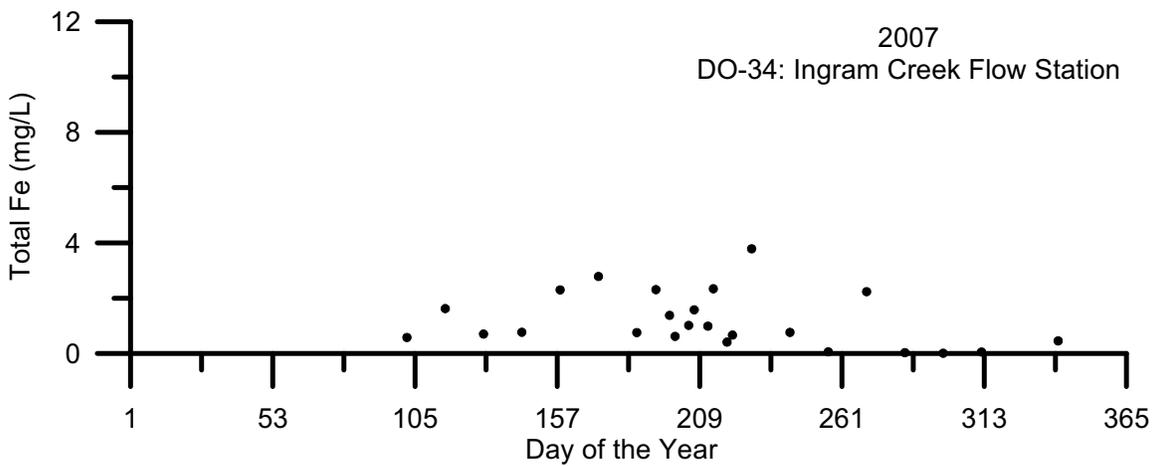
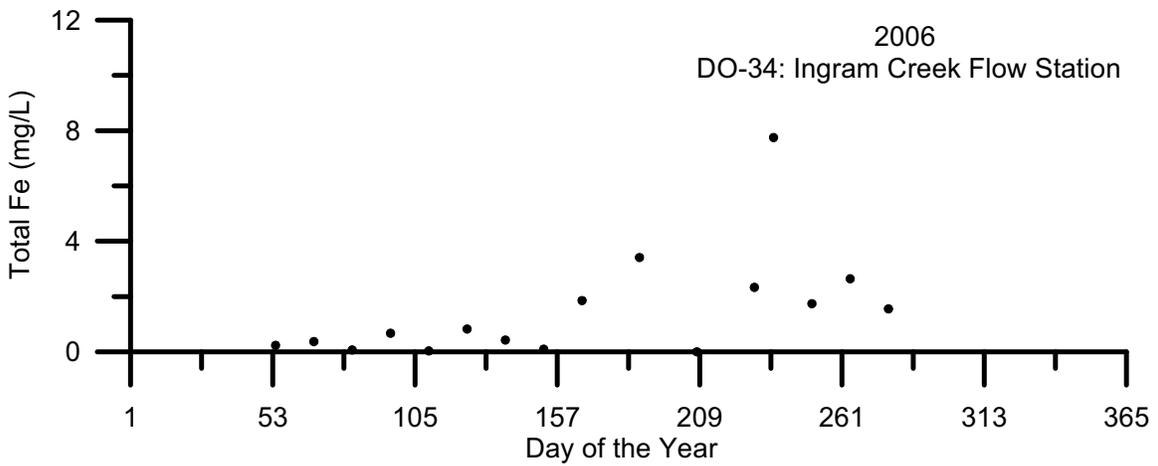
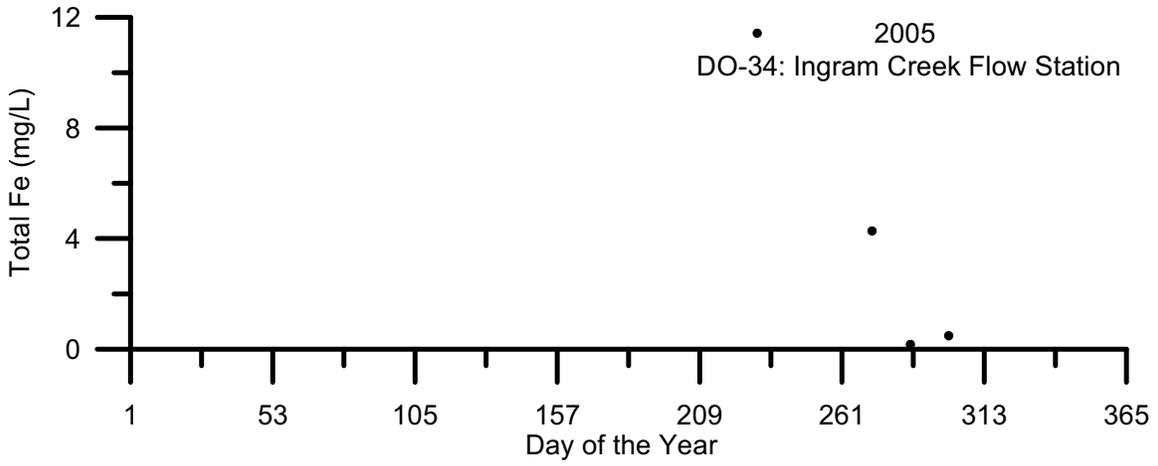


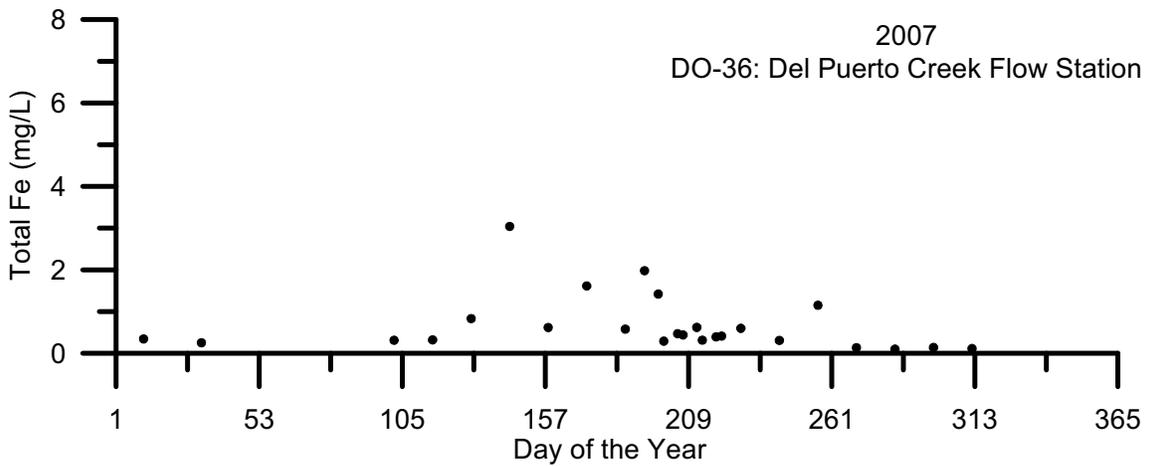
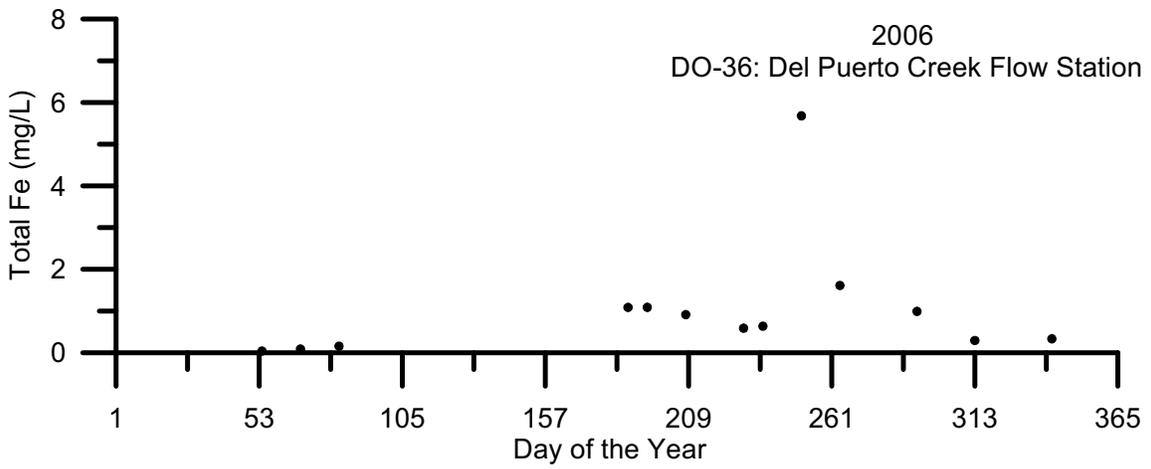
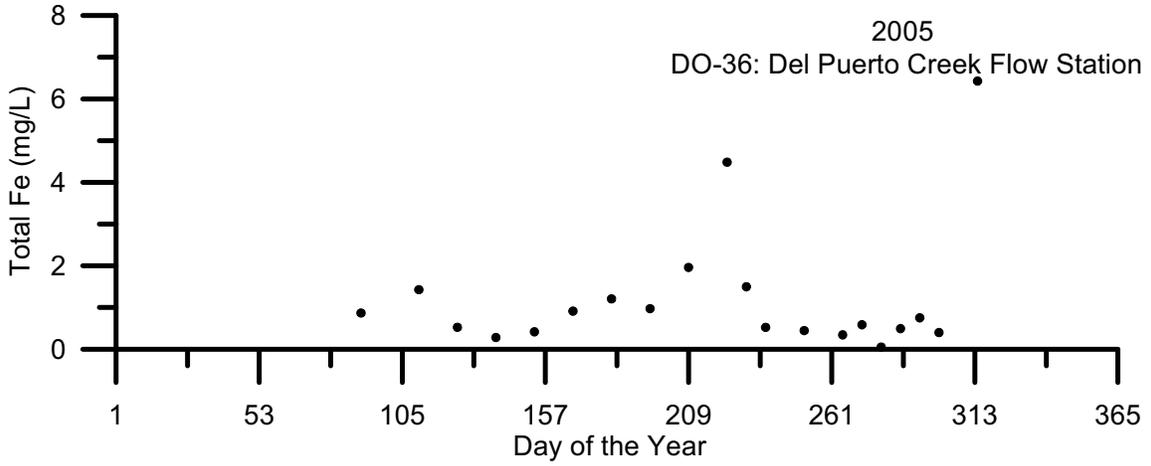


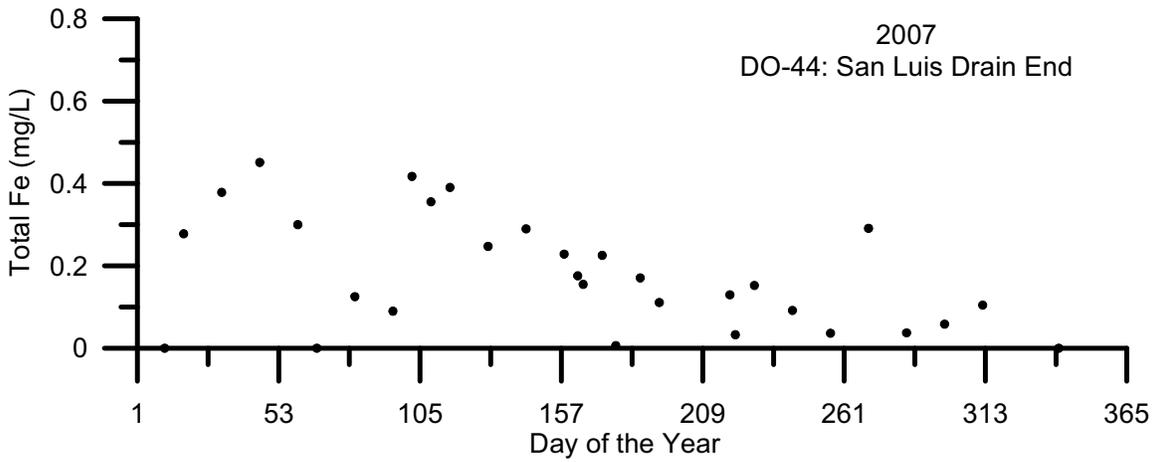
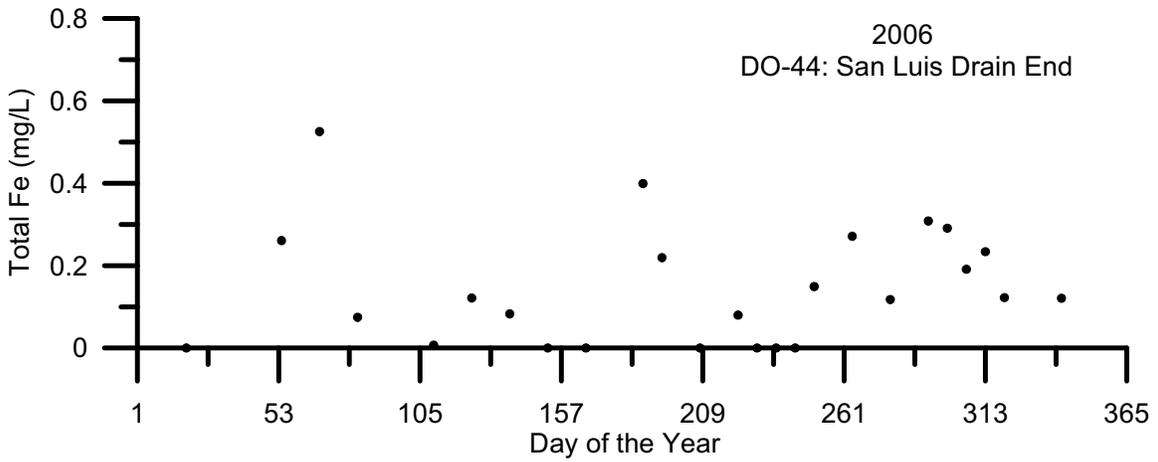
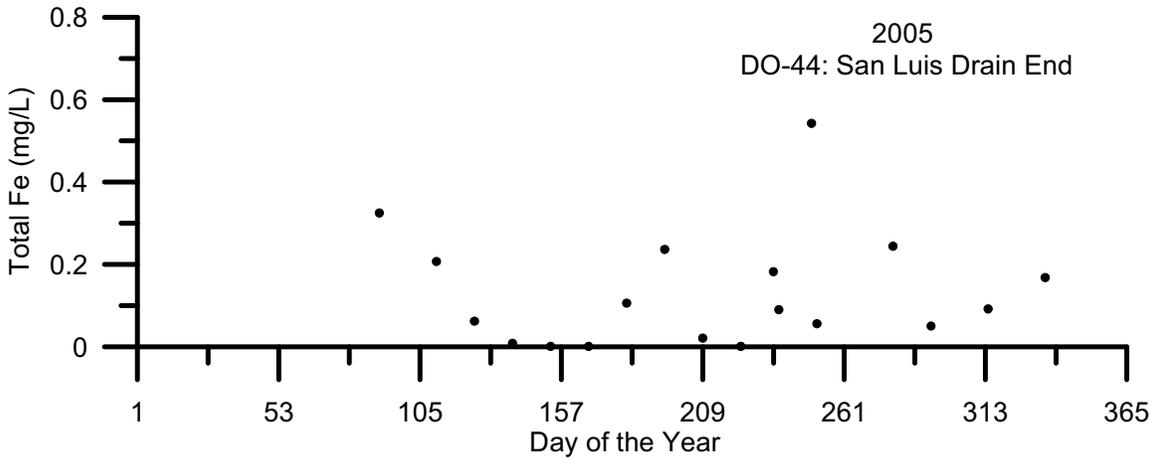


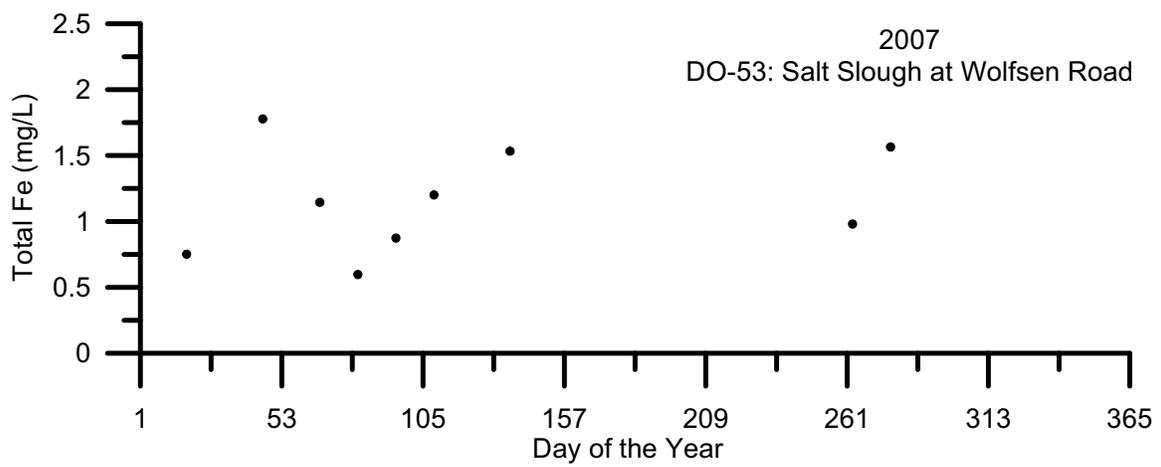
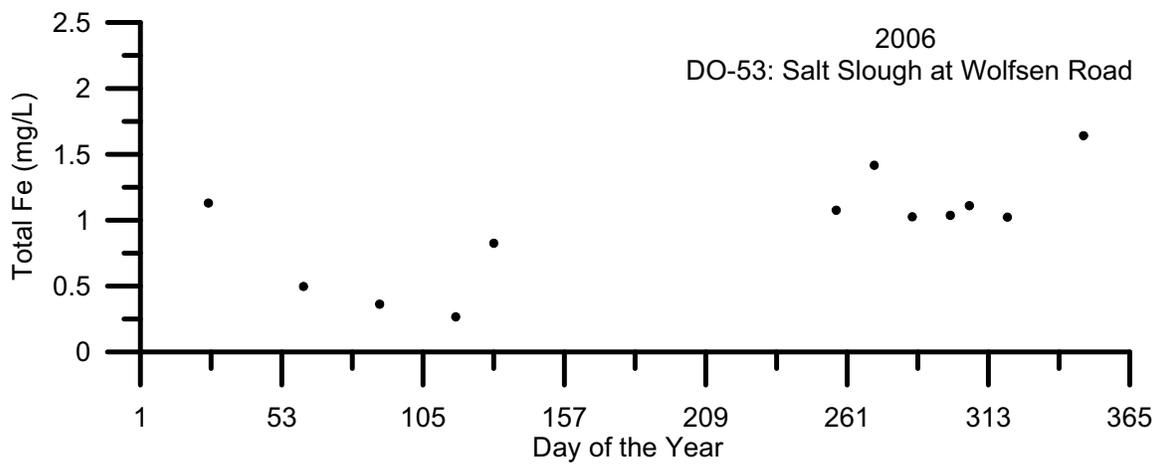
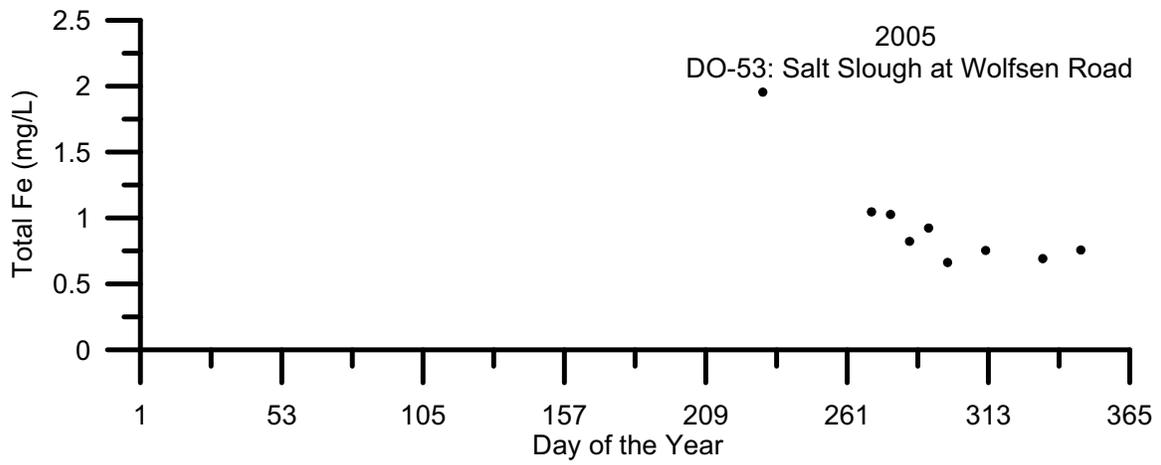


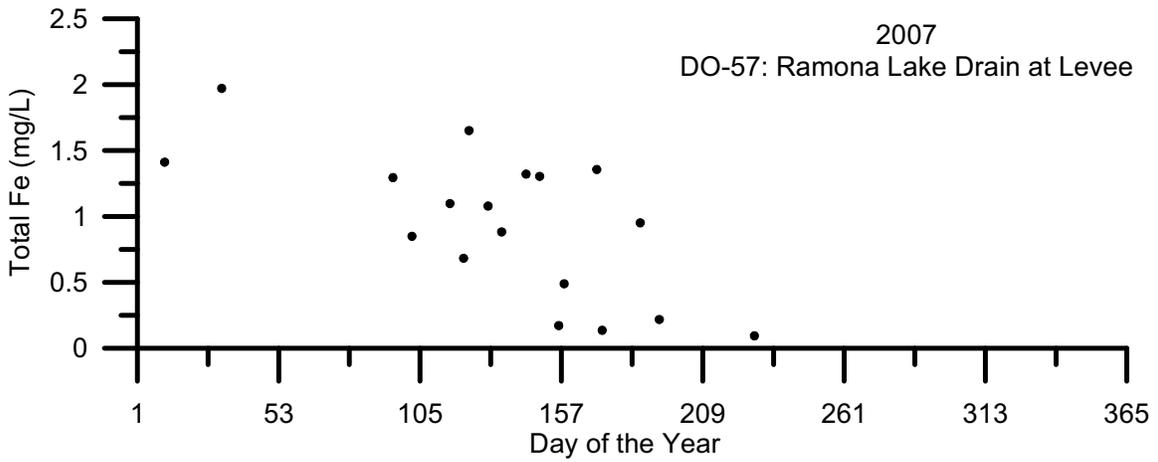
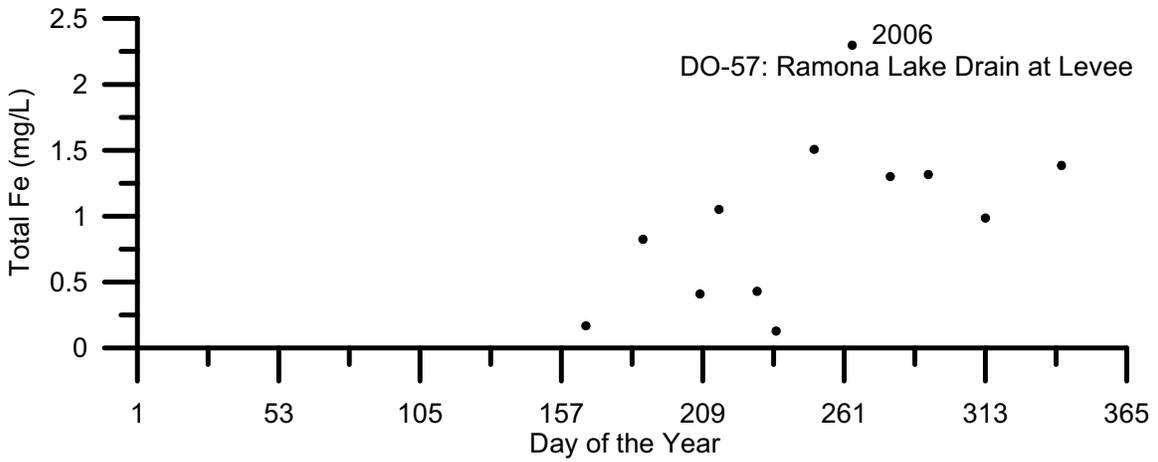
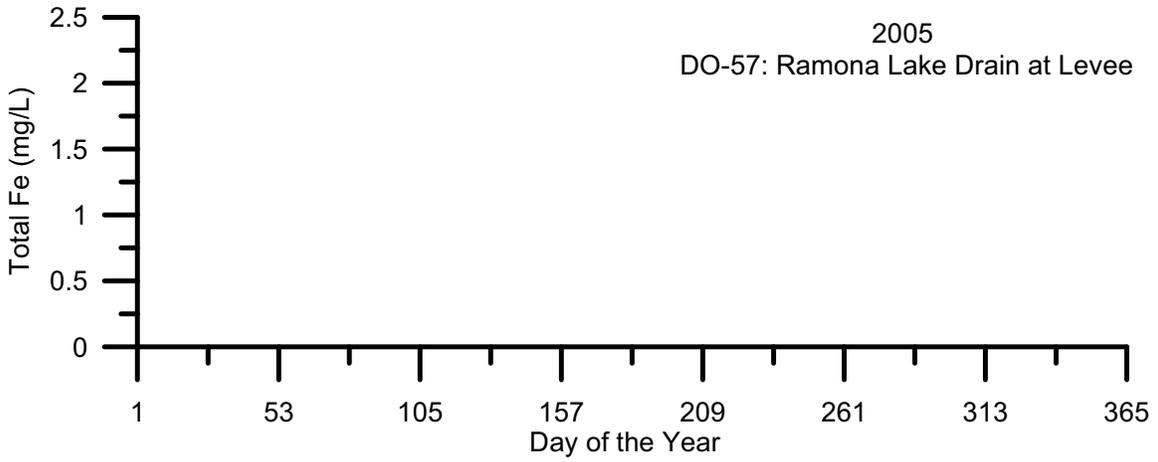


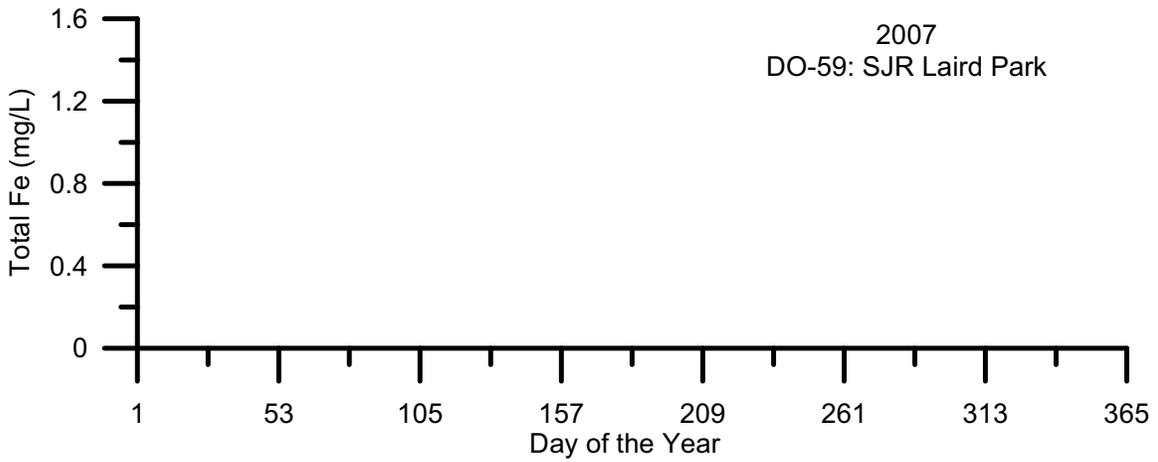
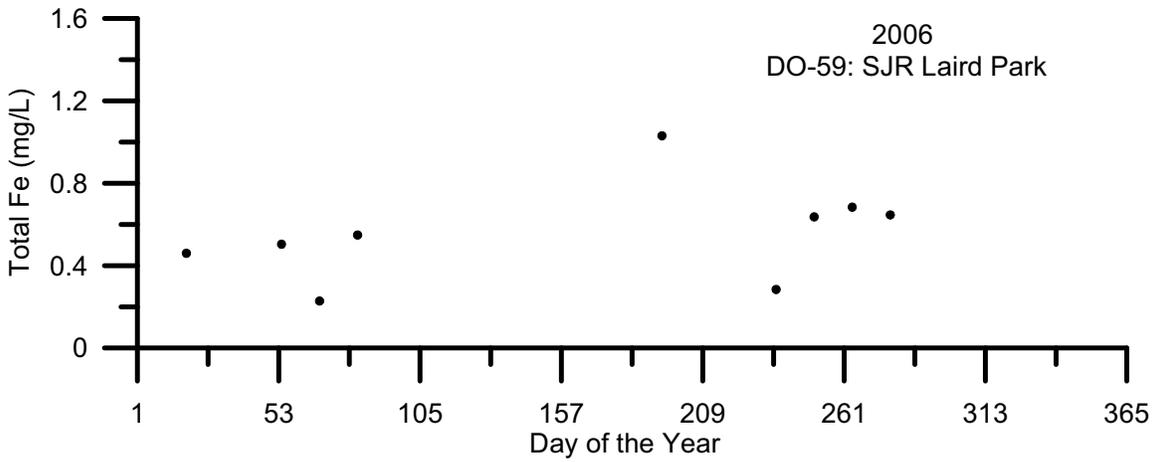
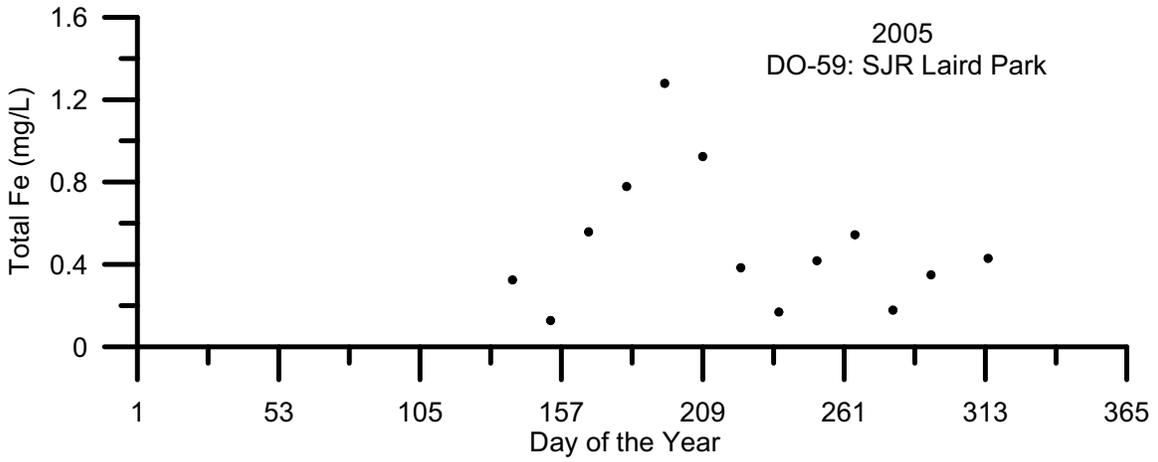


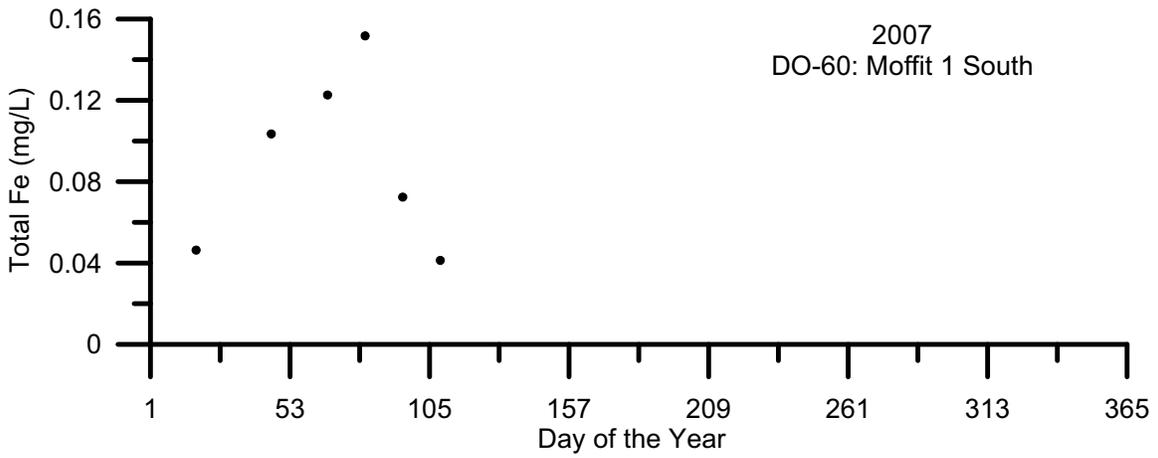
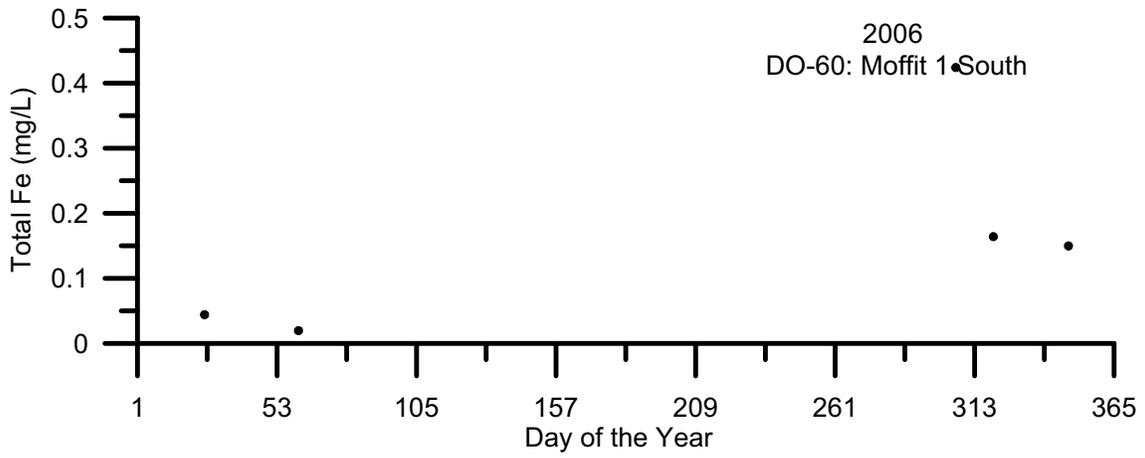
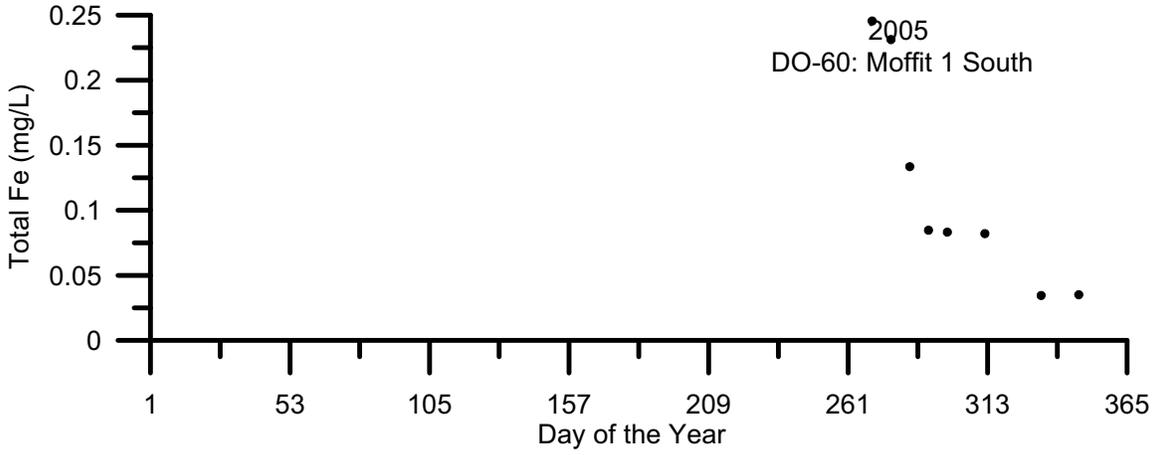


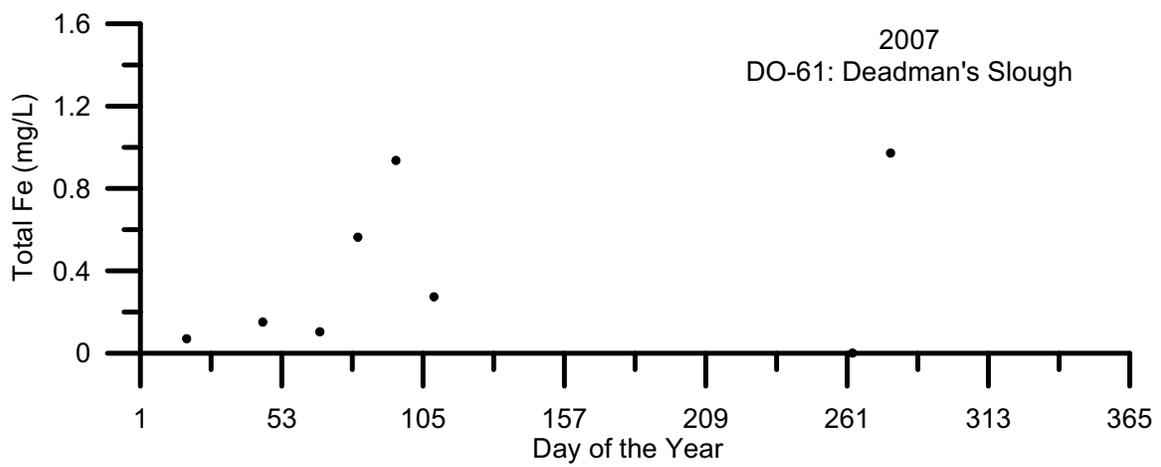
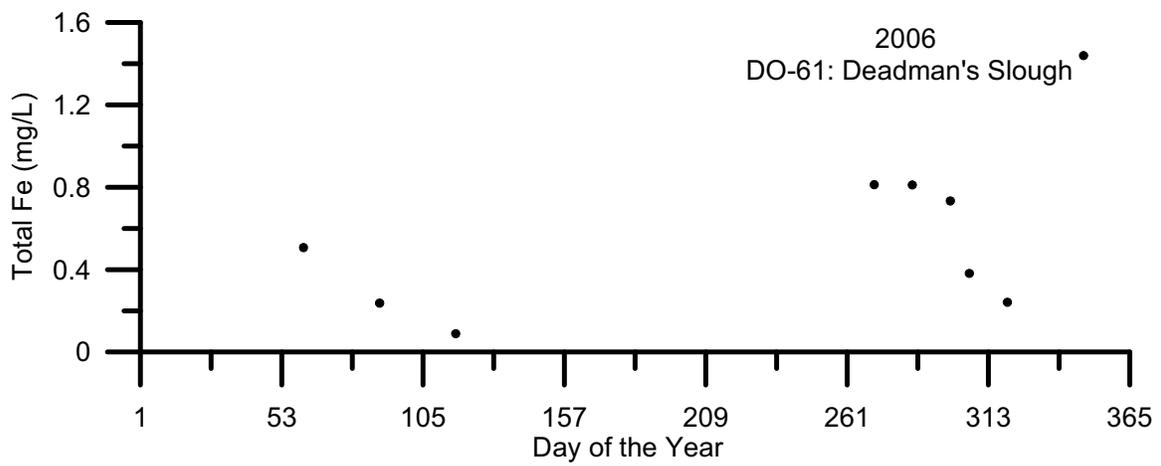
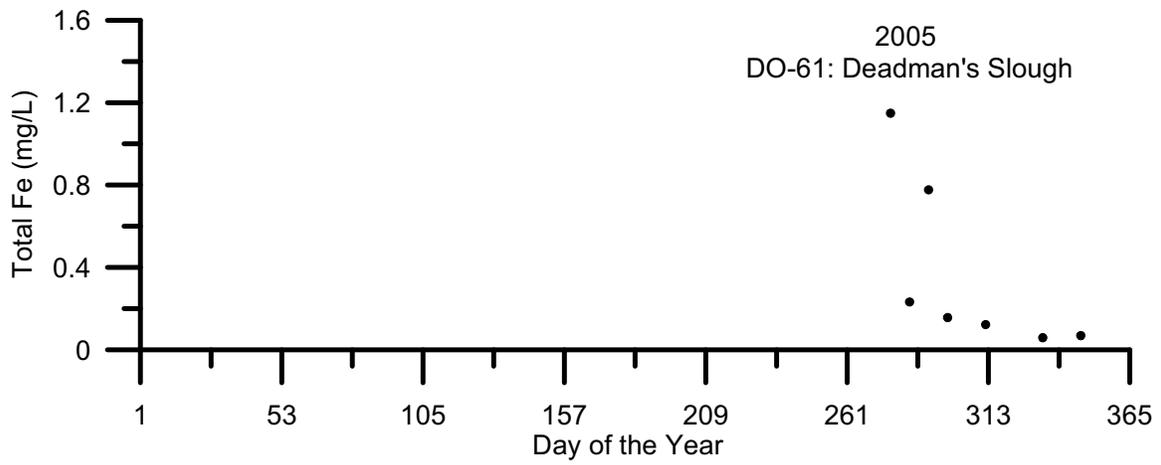


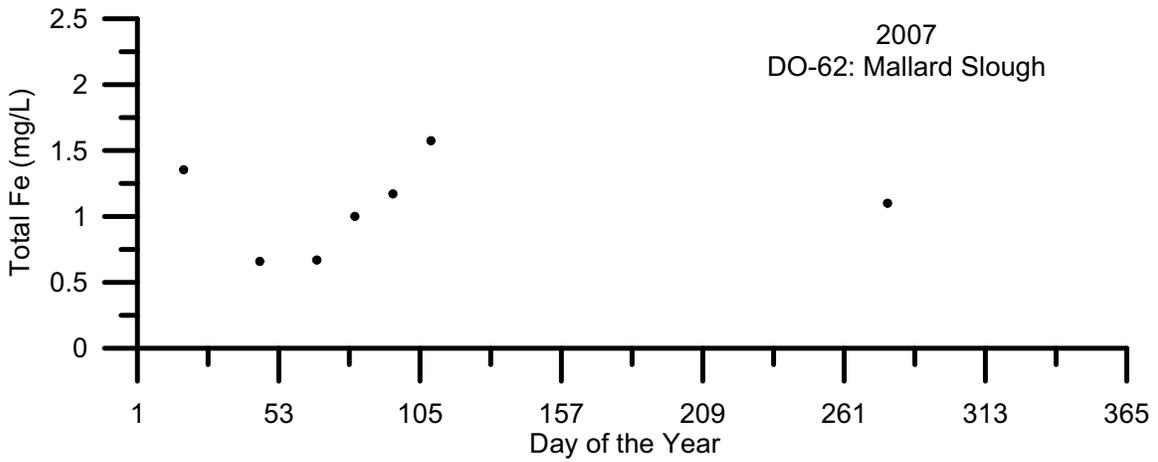
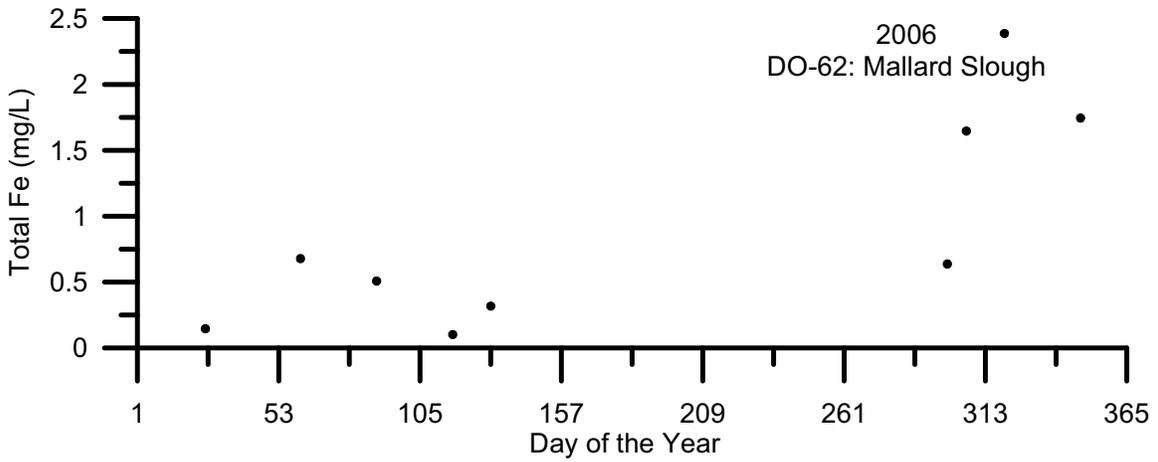
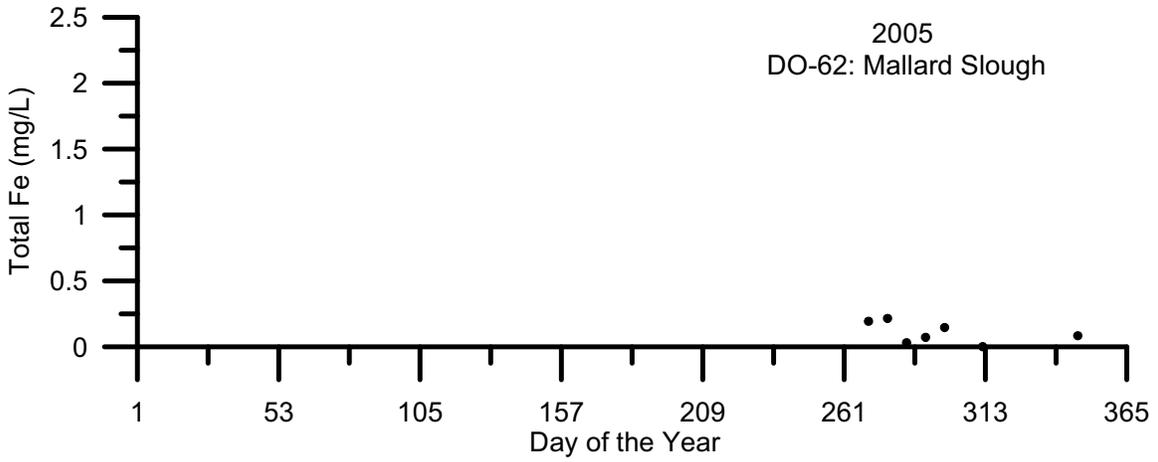


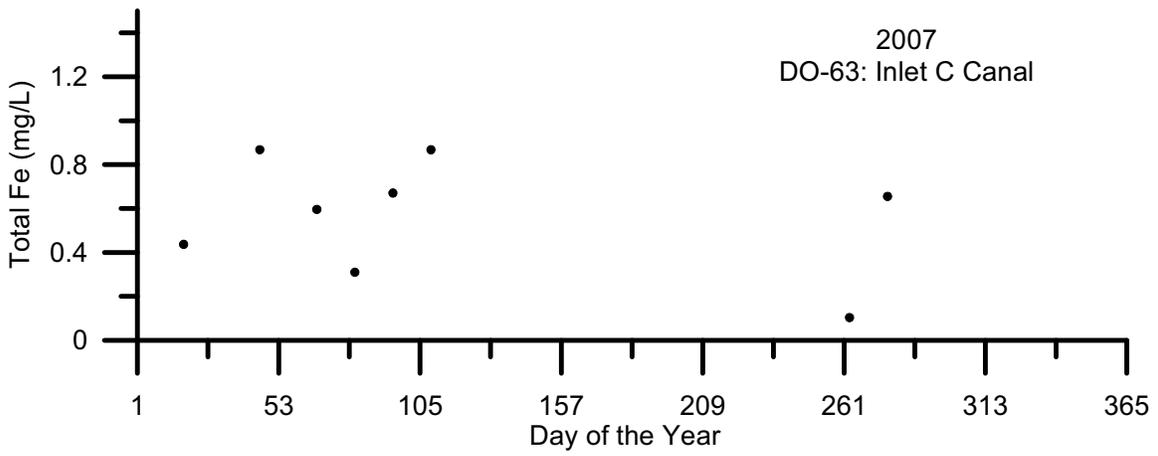
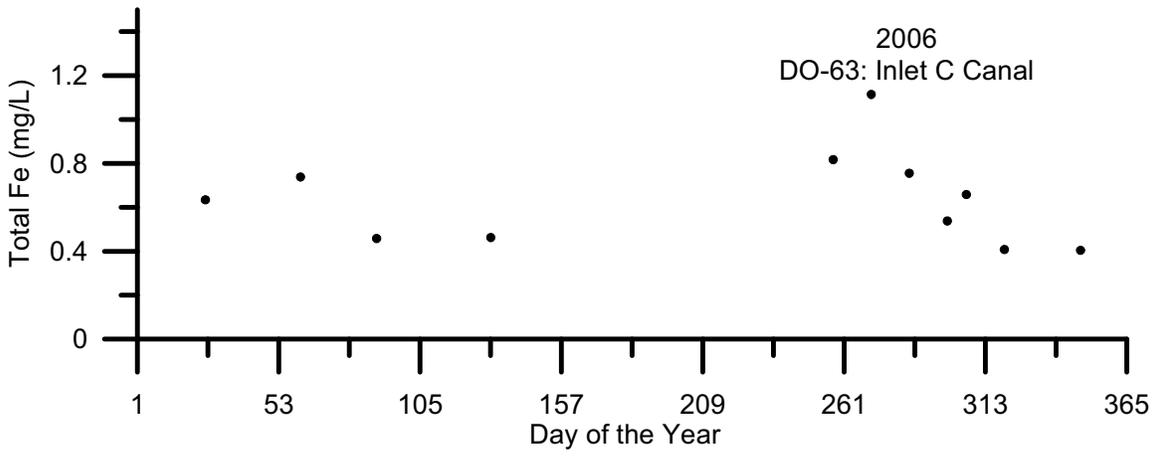
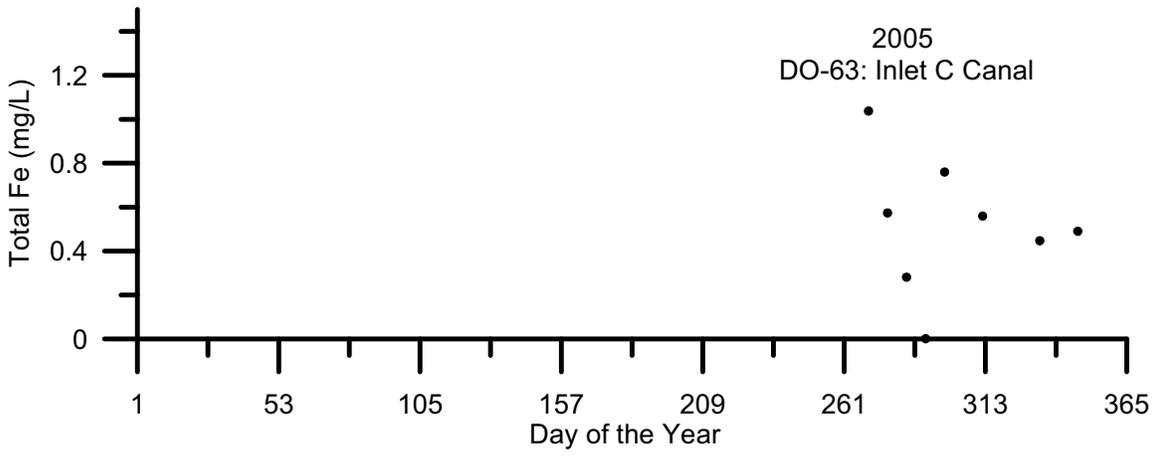


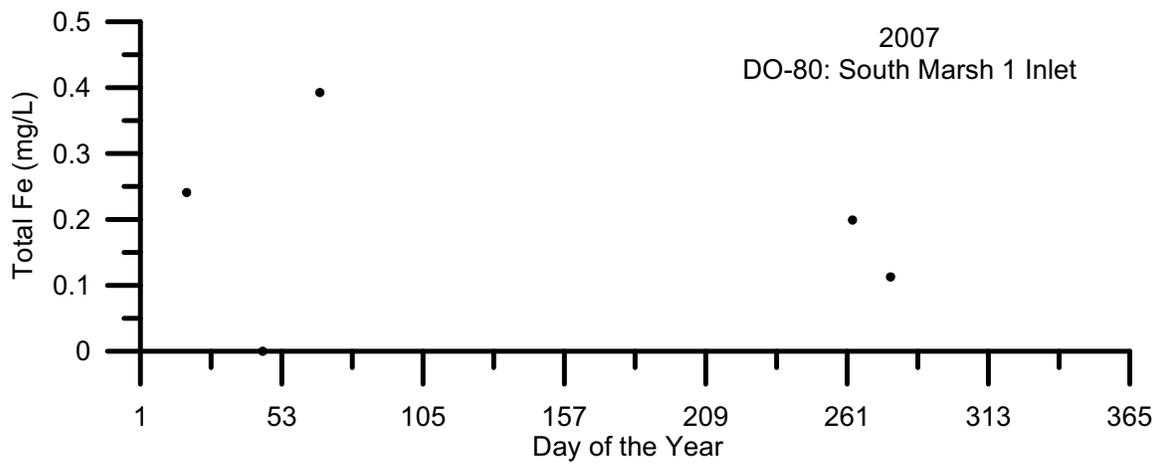
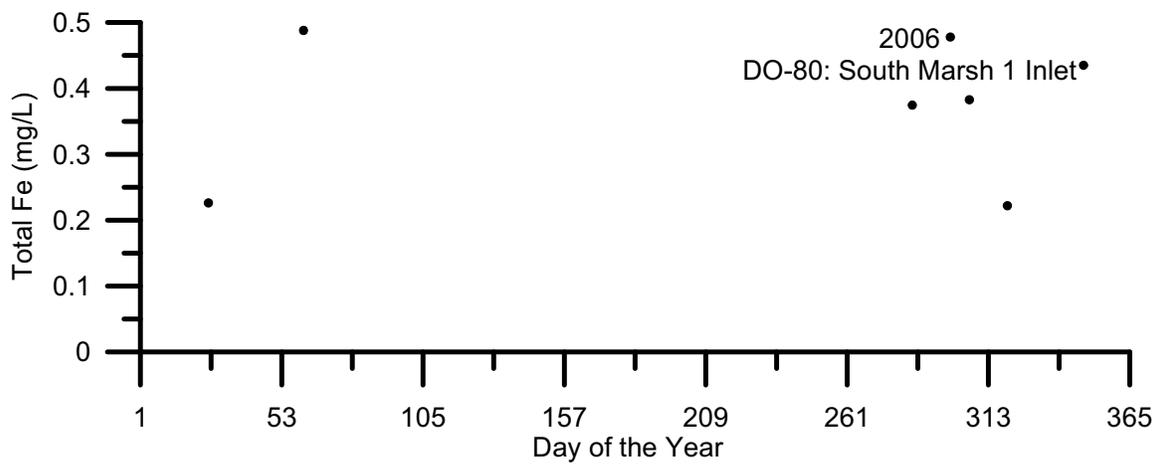
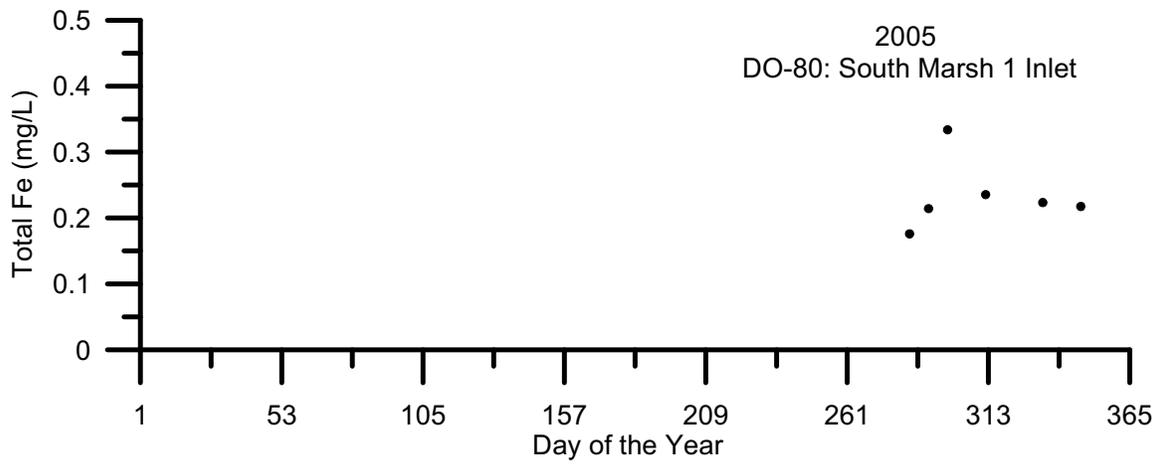


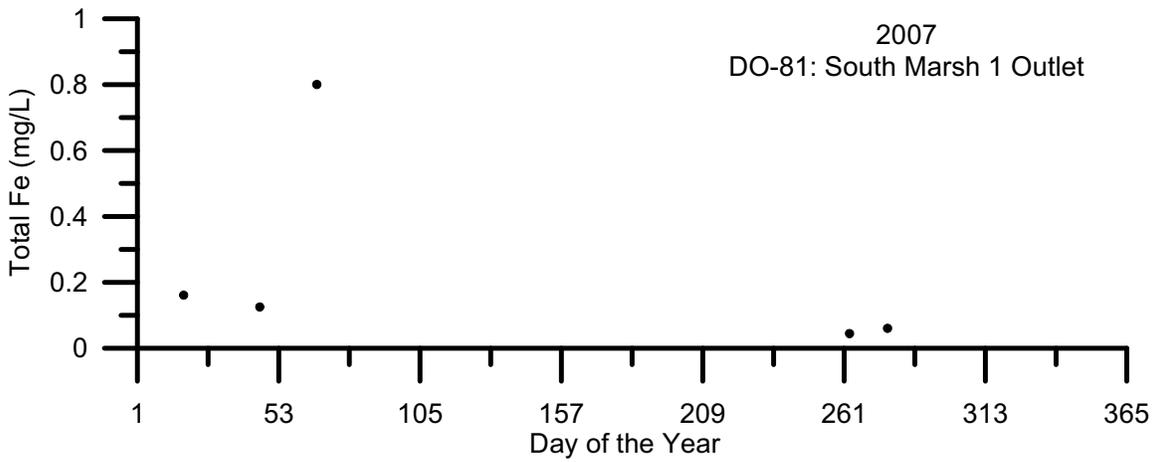
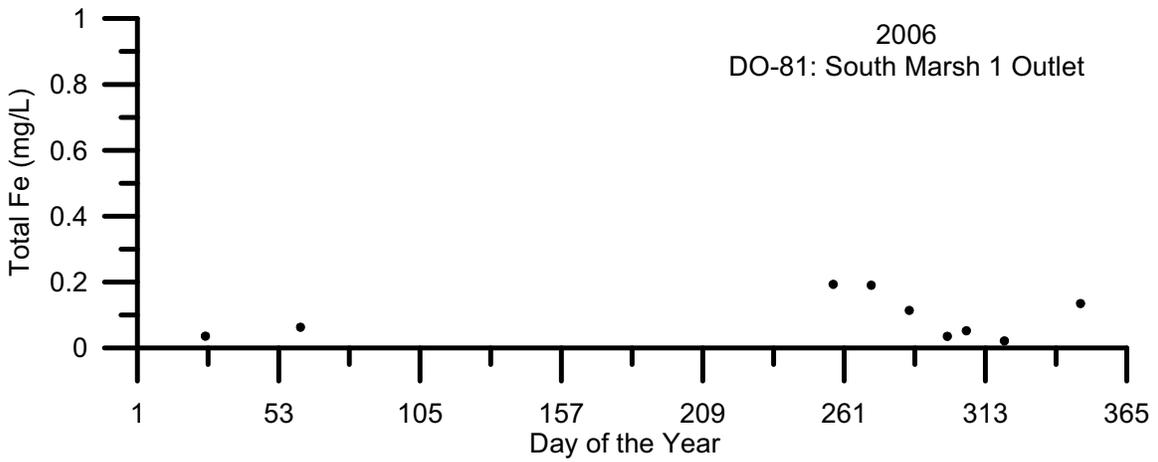
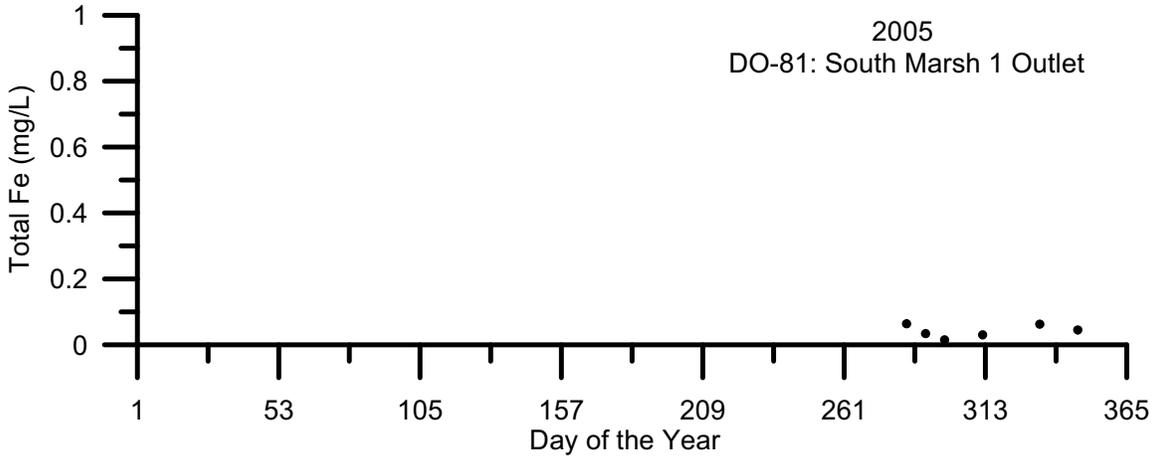


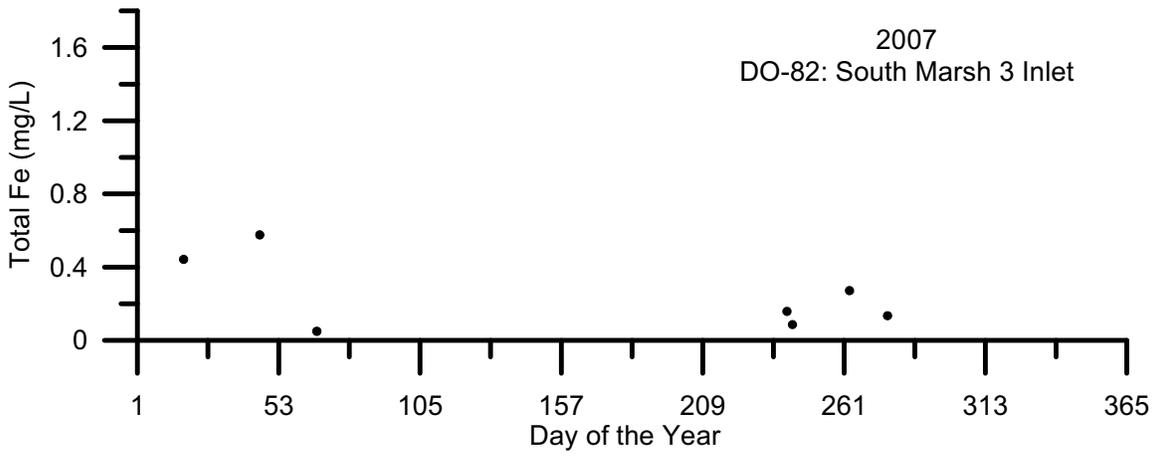
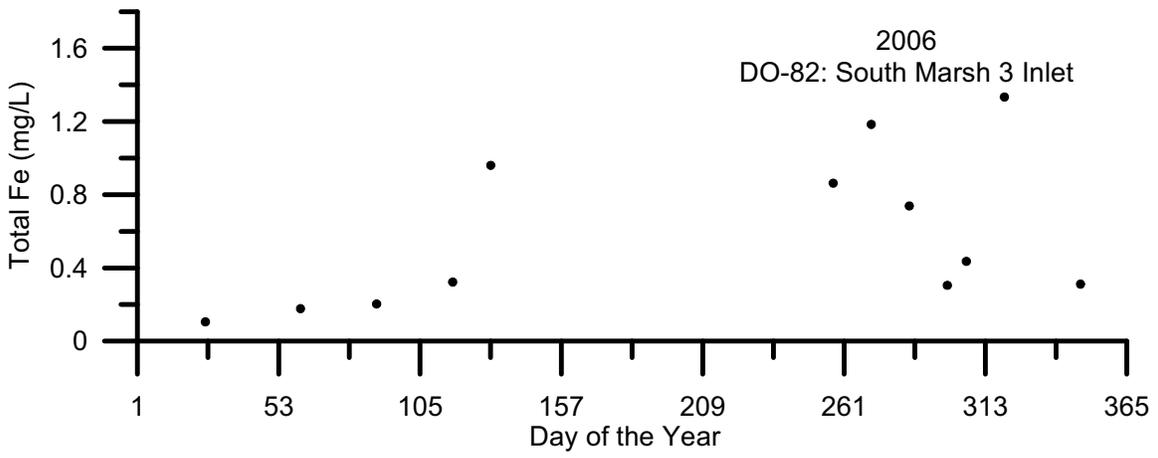
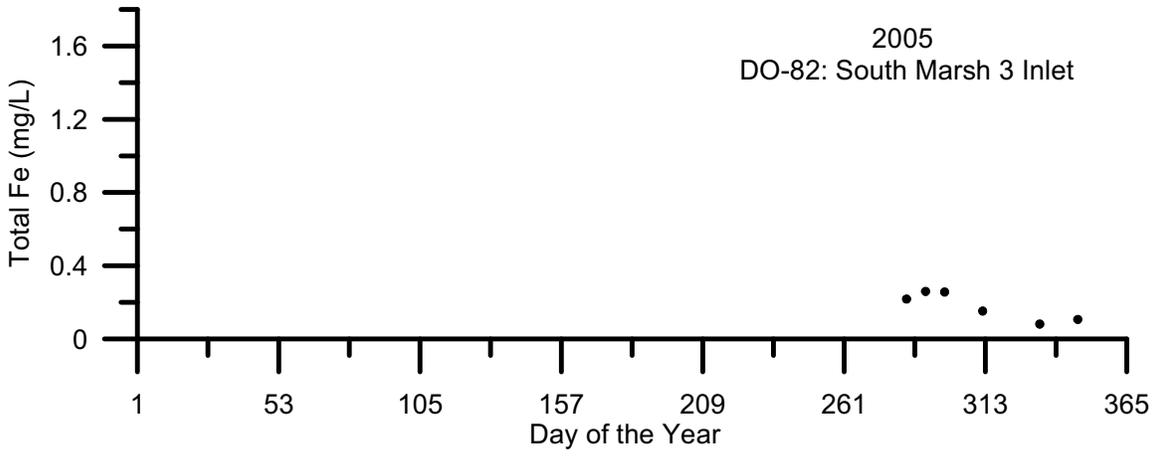


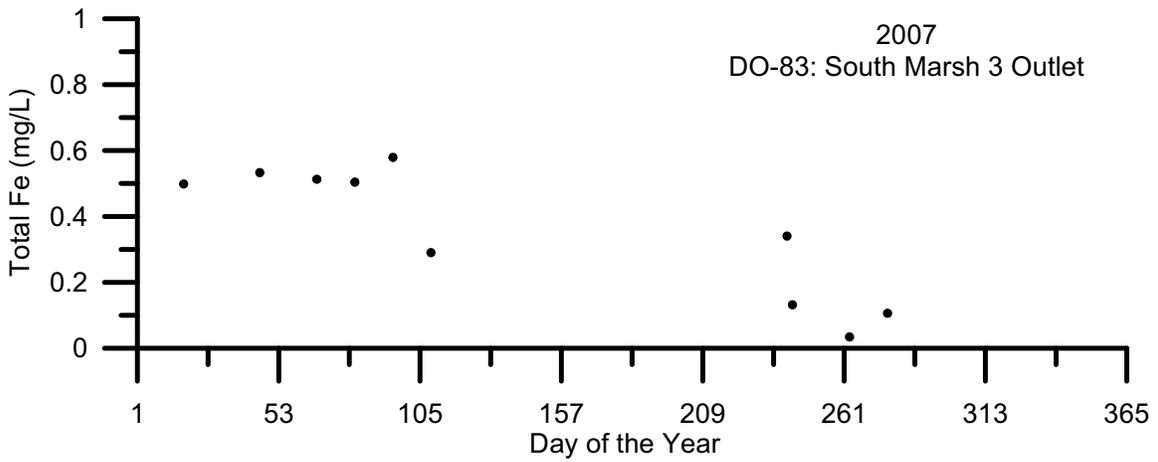
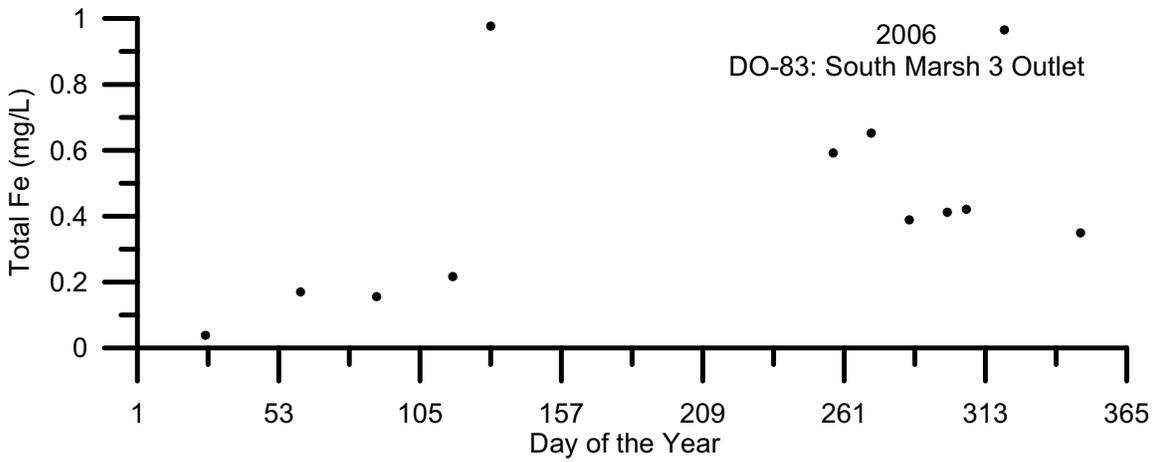
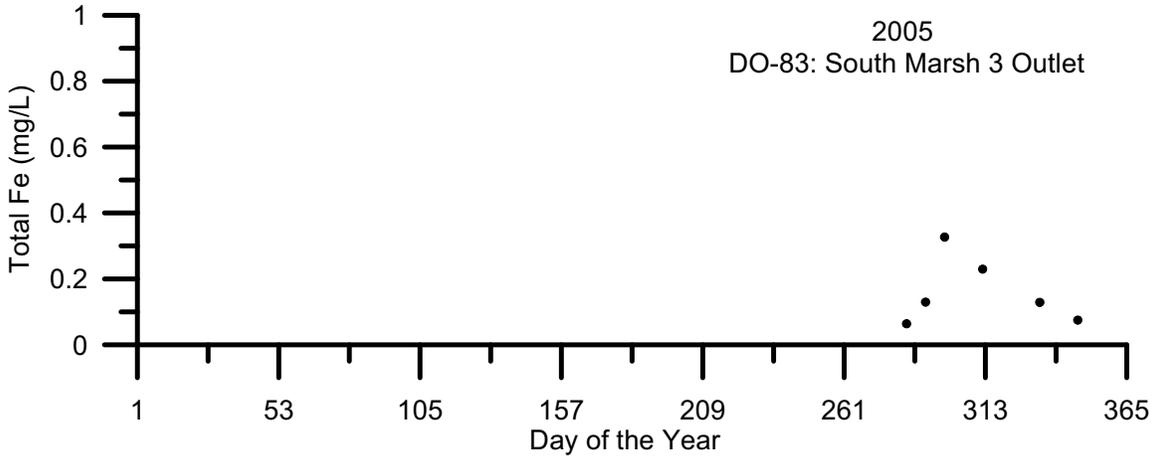


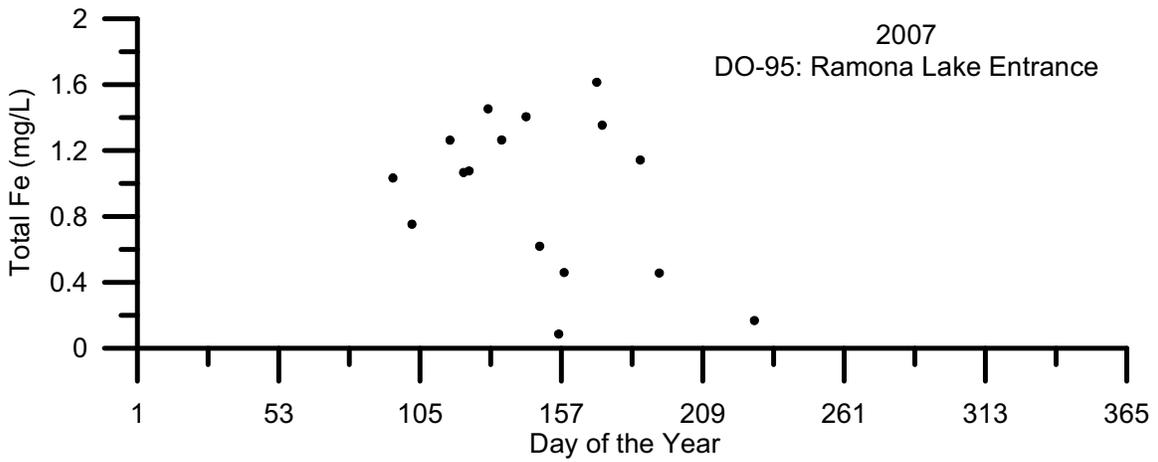
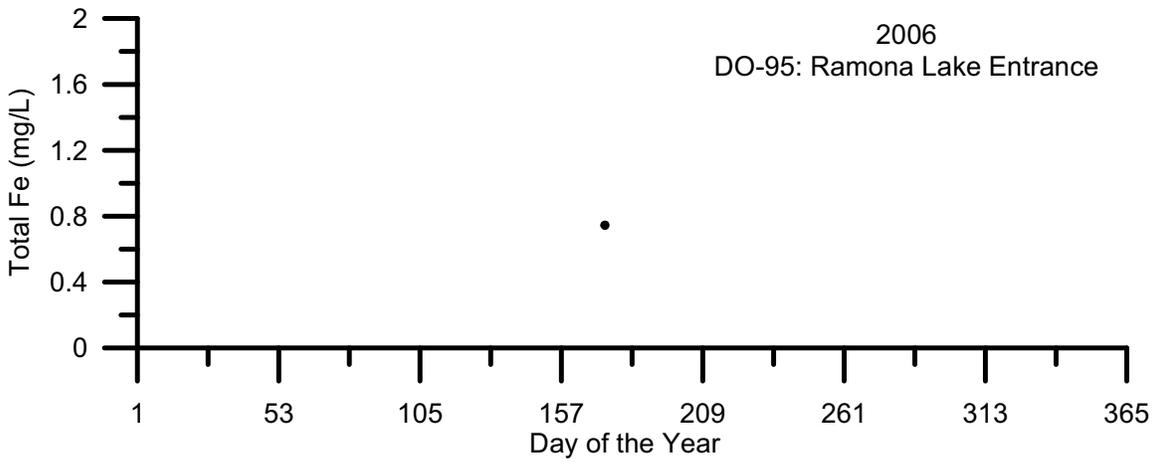
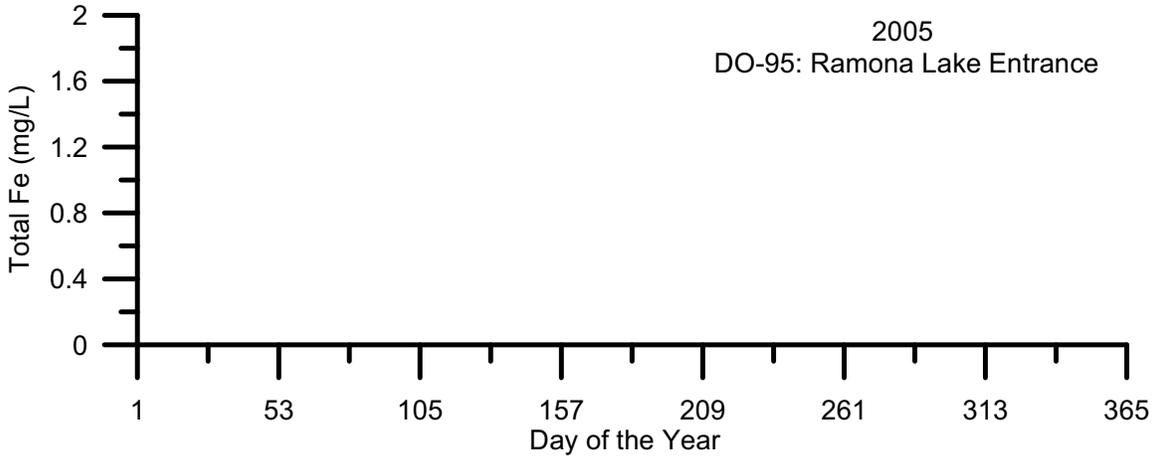














Temporal Plots of 2005-2007 Total Ammonia Data from the Upstream San Joaquin River

Chelsea Spier¹
Remie Burks¹
Sharon Borglin¹
Randy Dahlgren²
Jeremy Hanlon¹
Justin Graham¹
William Stringfellow¹

February 2008

¹Environmental Engineering Research Program
School of Engineering & Computer Sciences
University of the Pacific
3601 Pacific Avenue, Sears Hall
Stockton, CA 95211

²UC Davis, Davis, CA

Introduction

The San Joaquin River (SJR) supports one of the most productive agricultural regions in the world and its productivity is heavily dependant on irrigated agriculture. A consequence of irrigated agriculture is the production of return flows conveyed down gradient drains that eventually discharge to surface waters. Agricultural drainage may have significant nutrient load and can impact algae growth and general water quality in the SJR. Individual farmers and agricultural organizations, such as drainage authorities, are in need of tools to manage the environmental impacts of agricultural activities (Stringfellow, 2008).

For the years 2005 through 2007, sites throughout the San Joaquin Valley watershed were sampled to assess the overall water quality in the region. One thousand nine hundred and ninety-six (1996) individual surface water samples were collected and analyzed and WQ was assessed at 113 locations in the SJR basin (Borglin et al., 2008). Samples were processed and analyzed by the Environmental Engineering Research Program (EERP) laboratory at the University of the Pacific as well as at the University of California, Davis, Dahlgren Lab. This report includes temporal plots of total ammonia (NH₃-N) data analyzed by the EERP laboratory starting in the summer of 2007.

Methods

Depth integrated field samples were collected during 2005-2007 in the upper San Joaquin River in accordance with EERP Field Standard Operating Procedures Protocol Book (Graham 2008). Water samples were collected in glass 1000 mL bottles (Wheaton Science Products, Millville, NJ), 1000 mL HDPE Trace-Clean narrow mouth plastic bottles (VWR International), 250 mL HDPE Trace-Clean wide mouth plastic bottles (VWR International) as well as 40 mL trace clean vials with PTFE septa (ICChem, Rockwood, TN). Bottles were labeled with the appropriate sample number, site name and sampling date. All bottles were rinsed with sample water prior to sample collection. Some sites required a bucket to collect sample water because of accessibility from a high bridge or platform. For these sites, the bucket was pre-rinsed with sample water and sample bottles were filled using a rinsed funnel. Care was taken to distribute water simultaneously to all sample bottles (rather than sequentially). Samples were immediately stored at 4 °C after sampling (cooler temperature was recorded in the lab upon delivery) and transported to the EERP lab on the day of sampling.

Within 24 hours of collection samples were aliquotted and stored at -20°C until total NH₃-N analysis could be completed.

Starting in 2007 total NH₃-N was quantified using the TL-2800 Ammonia analyzer made by Timberline Instruments (Boulder, CO). Total NH₃-N sample was mixed with caustic solution (to pH 11-13). NH₃ (g) diffused across as gas-permeable membrane and is dissolved in a buffer solution. The dissolution of NH₃ (g) in the buffer causes a change in conductivity which can be correlated to NH₃ (g). The Timberline instrument automates the mixing of the caustic solution with the sample and pumping of the buffer solution through the gas permeable membrane (Borglin, et al, 2008) and (Carlson, 1990). The resulting peak is proportional to the NH₃-N concentration. The reportable limit for this

method is 0.03 mg/L NH₃-N.

Results/Discussion

With each set of NH₃-N field samples analyzed in the EERP laboratory, quality assurance samples including a lab duplicate, field duplicate, matrix spike, matrix spike duplicate, calibration check standards, laboratory control standard, trip blank, and lab blanks were also analyzed. 98.25% of all quality assurance samples were within a passing range (Spier et al, 2008). Proficiency check samples, standards with unknown concentration to the laboratory analyst, were run approximately twice a year. Three proficiency check samples were analyzed for NH₃-N in the EERP laboratory during 2007, and all of these samples were found to be within the acceptable range. Samples were measured ranging from 0.0-4.26 mg/L NH₃-N. The average concentration of NH₃-N in samples collected was 0.15 mg/L NH₃-N. NH₃-N was also analyzed at UC Davis on all of the same water samples and has a high correlation to values measured by EERP. When all data points measured by the two labs are compared they have $r^2=0.956$ (Spier et al, 2008). NH₃-N samples measured by EERP have about 91.1% as much NH₃-N as the same samples measured by UCD (Figure 2).

All of the quality assurance samples that failed were lab banks that had higher concentrations than the detection limit. The baseline can drift for various reasons on the Timberline instrument. Normally lab blanks are run at the beginning and end of each set of samples run on the Timberline instrument. To help catch baseline drift sooner, and prevent this problem lab blanks are now also run after every 20 samples in addition to being run at the beginning and end of each set. EERP used to measure NH₃-N using the Nessler method SM 4500-NH₃ C (APHA, 1992). The nessler method had a much higher detection limit, a lower correlation to UCD's NH₃-N values, and also created mercury waste so it is no longer used by EERP to measure NH₃-N.

These temporal plots (Figures 3-103) as well as the plot comparing EERP's laboratory data to UCD's data (Figure 2) created an easy visual way to find outliers and double check data entry for possible mistakes. For the purpose of these plots any data points that were slightly negative were changed to 0.

References

- American Public Health Association (APHA). 1992. Standard Methods for the Examination of Water and Wastewater, 18th Edition. American Public Health Association, Washington, DC.
DC.
- Borglin, S., W. Stringfellow, J. Hanlon. 2005. Standard Operating Procedures for the Up-Stream Dissolved Oxygen TMDL Project. LBNL/Pub-937.
- Borglin, S., Burks, R., Hanlon, J., Graham, J., Spier, C., Stringfellow, W., and Dahlgren, R., (2008) Methods overview, quality assurance, and quality control, University of the Pacific, Stockton, CA
- Borglin, S.E., Burks, R.D., Hanlon, J.S., Stringfellow, W.T. (2008) EERP Lab Protocol

- Book, University of the Pacific, Stockton, CA.
- Carlson, R.M., R. Cabrera, J. Paul, J. Quick and R.Y. Evans. 1990. Rapid direct determination of ammonium and nitrate in soil and plant tissue extracts. *Comm. Soil Sci. Plant Anal.* 21:1519-1530.
- Graham, J., Hanlon, J.S., Stringfellow, W.T., (2008) EERP Field Protocol Book, University of the Pacific, Stockton, CA.
- Stringfellow, W.T., et al., (2008) Evaluation of Vegetated Ditches, Ponds, and Wetlands as BMPs for Mitigating the Water Quality Impact of Irrigated Agriculture in the San Joaquin Valley, University of the Pacific, Stockton, CA
- Spier, C., Burks, R., Borglin, S.E. (2008) Comparison of Methods for Analyzing Ammonia, Nitrate and Total Nitrogen, University of the Pacific, Stockton, CA.
- YSI Environmental Operations Manual (2005) 6-Series Environmental Monitoring Systems, Yellow Springs, OH.

Table 1: EERP Sampling Site List

DO Number	Site Name	Type
4	SJR at Mossdale	Core sites
5	SJR at Vernalis-McCune Station (River Club)	Core sites, BMP
6	SJR at Maze	Core sites, BMP
7	SJR at Patterson	Core sites, BMP
8	SJR at Crows Landing	Core sites, BMP
10	SJR at Lander Avenue	Core sites
12	Stanislaus River at Caswell Park	Core sites
14	Tuolumne River at Shiloh Bridge	Core sites
16	Merced River at River Road	Core sites
18	Mud Slough near Gustine	Core sites, Wetland
19	Salt Slough at Lander Avenue	Core sites, Wetland
20	Los Banos Creek Flow Station	Core sites, Wetland
21	Orestimba Creek at River Road	Core sites, BMP
23	Modesto ID Lateral 5 to Tuolumne	Core sites
25	Modesto ID Main Drain to Stan. R. via Miller Lake	Core sites
28	Turlock ID Westport Drain Flow station	Core sites
29	Turlock ID Harding Drain	Core sites
30	Turlock ID Lateral 6 & 7 at Levee	Core sites
33	Hospital Creek	Intermittent, BMP
34	Ingram Creek	Core sites, BMP
36	Del Puerto Creek Flow Station	Core sites, BMP
44	San Luis Drain End	Core sites
53	Salt Slough at Wolfsen Road	Wetland
57	Ramona Lake Drain	Core sites, BMP
59	SJR Laird Park	Core sites
60	Moffit 1 South	Wetland
61	Deadman's Slough	Wetland
62	Mallard Slough	Wetland
63	Inlet C Canal	Wetland
80	South Marsh-1-Inlet	Wetland
81	South Marsh-1-Outlet	Wetland
82	South Marsh-3-Inlet	Wetland
83	South Marsh-3-Outlet	Wetland
95	Ramona drain at Ramona Lake	BMP, Intermittent

Figure 1: EERP Sampling Site Map of SJR Watershed and Tributaries

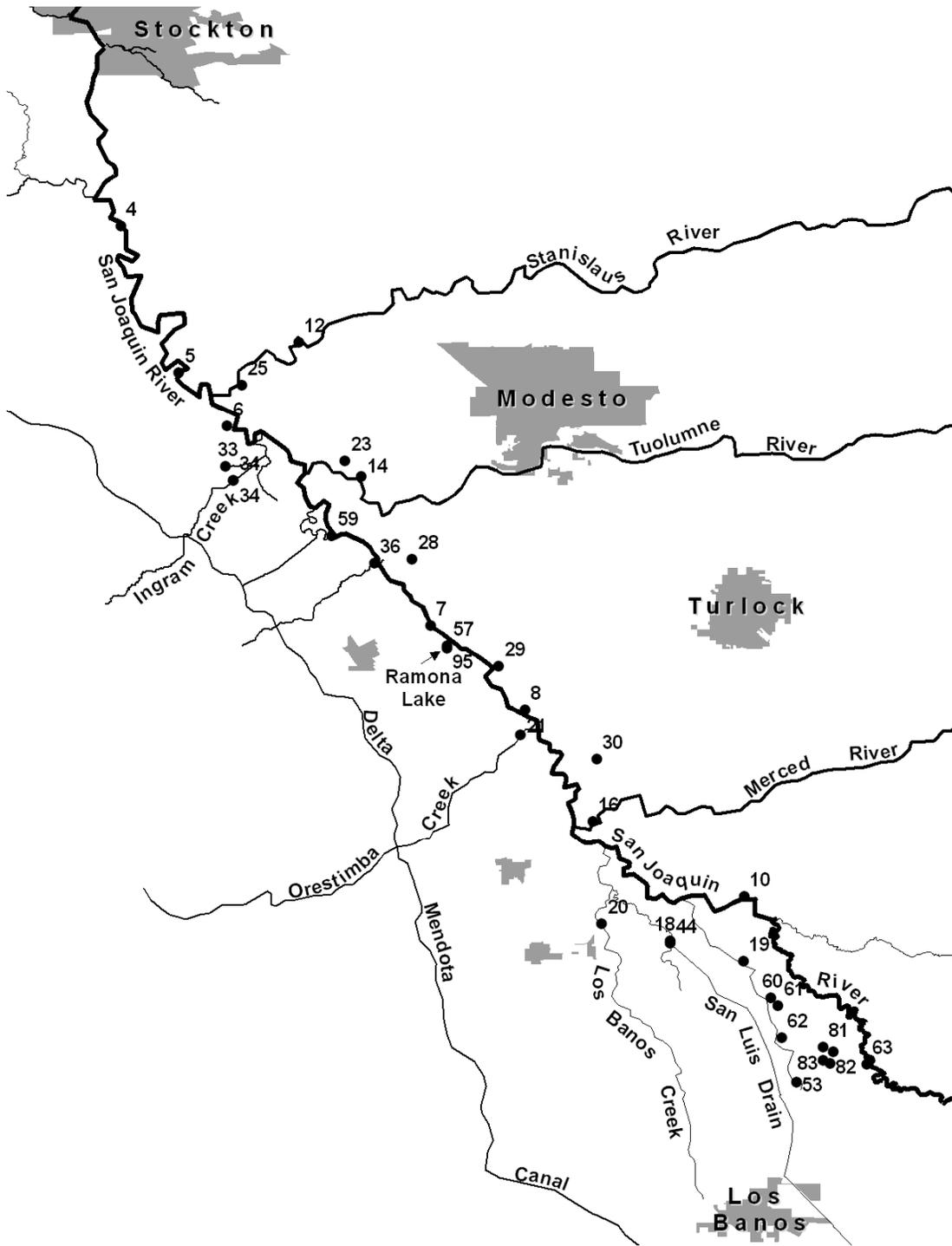
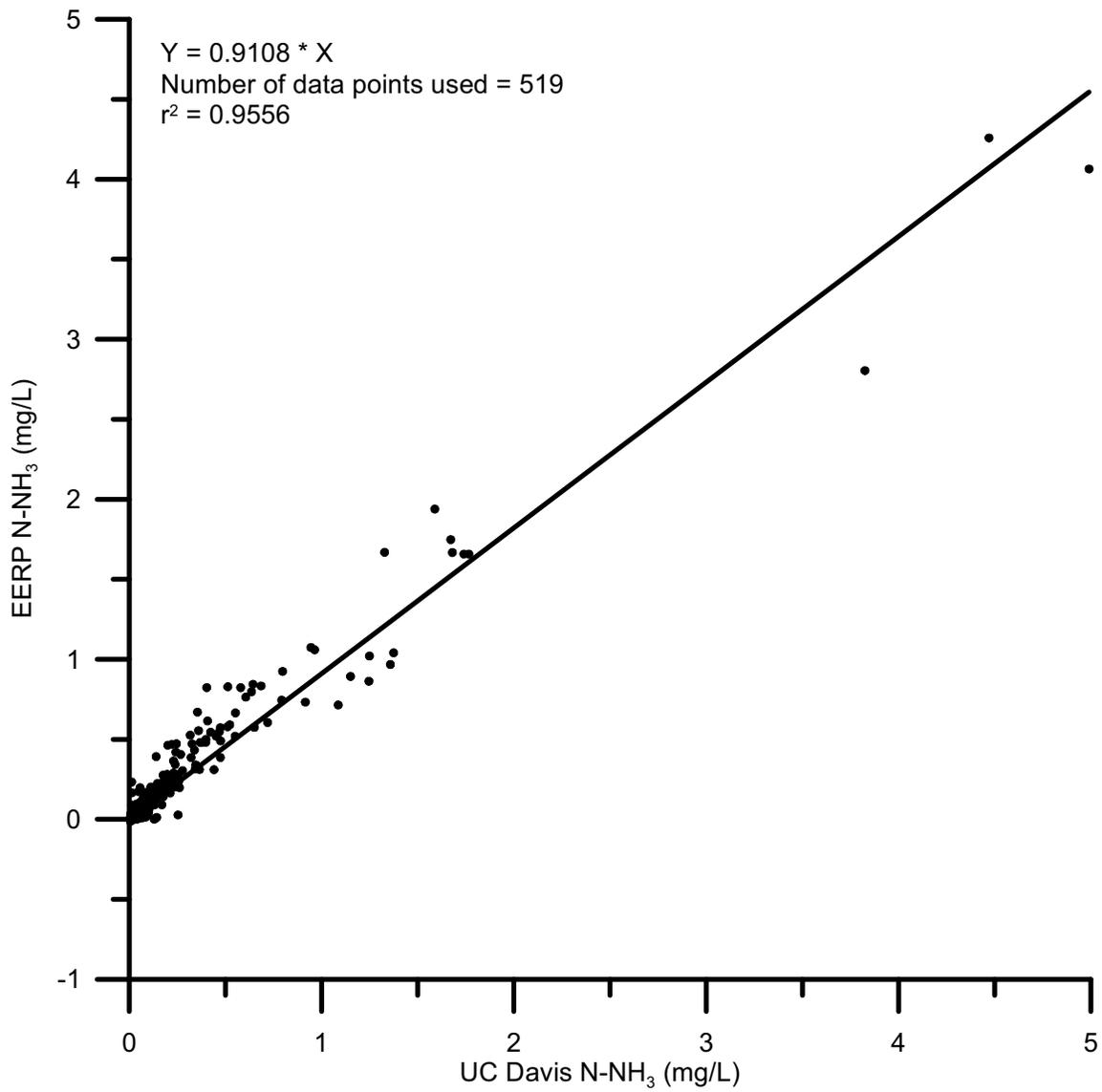
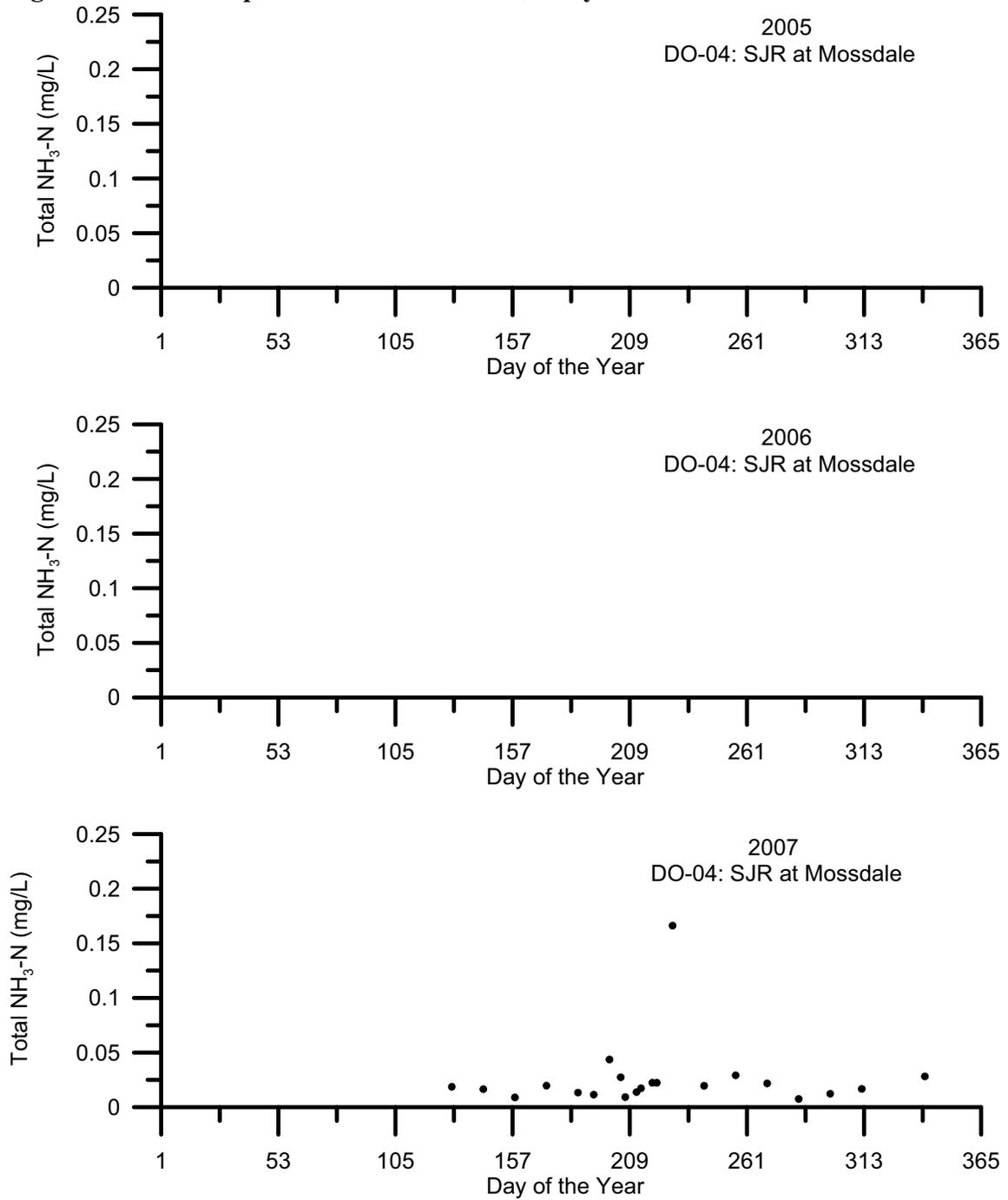
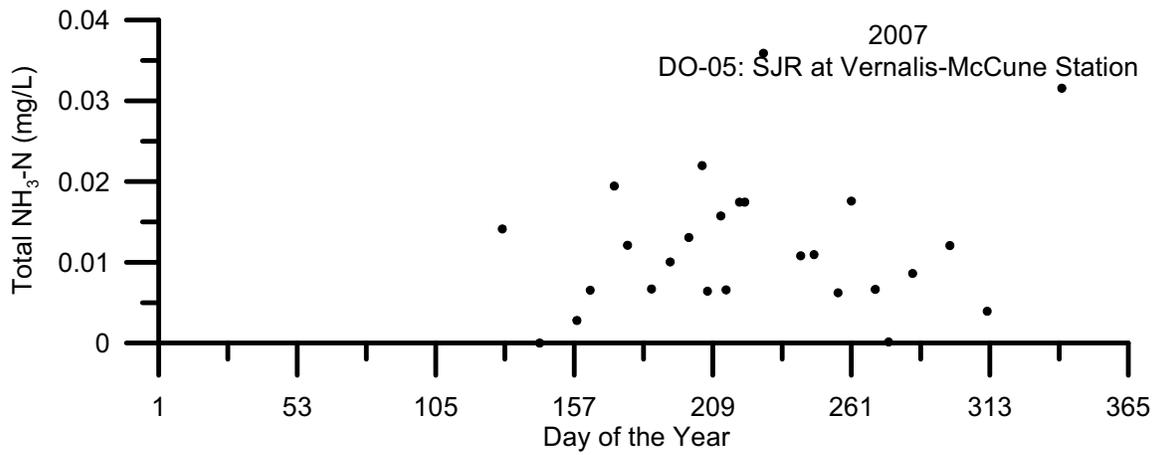
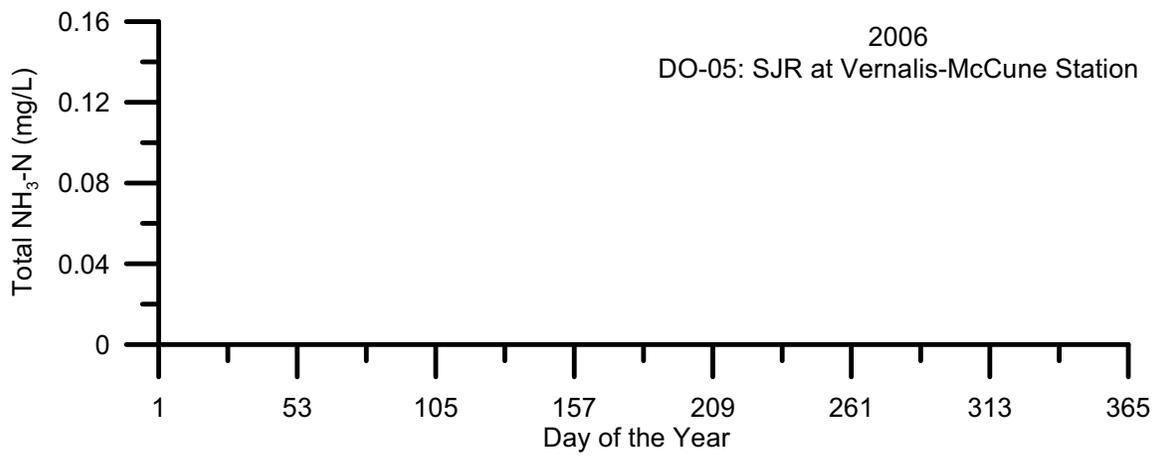
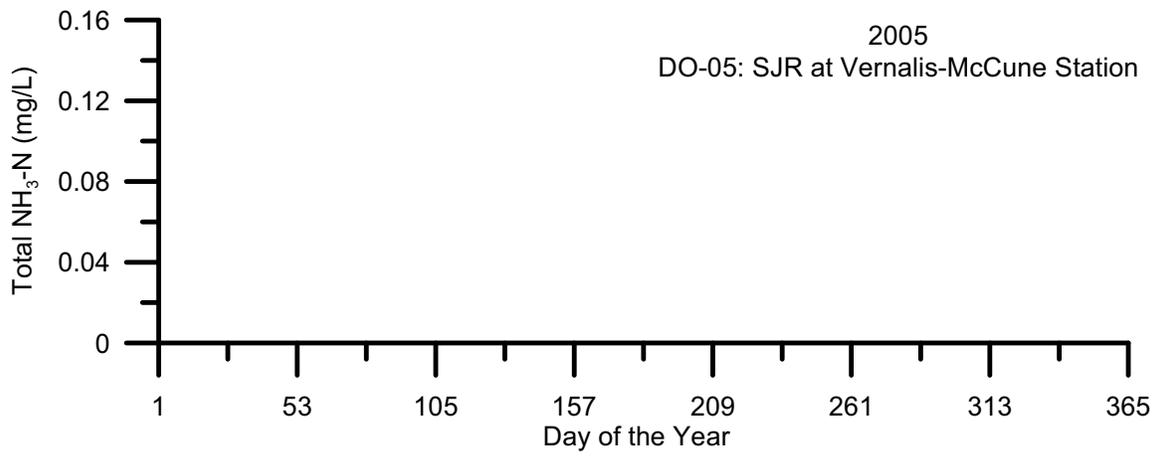


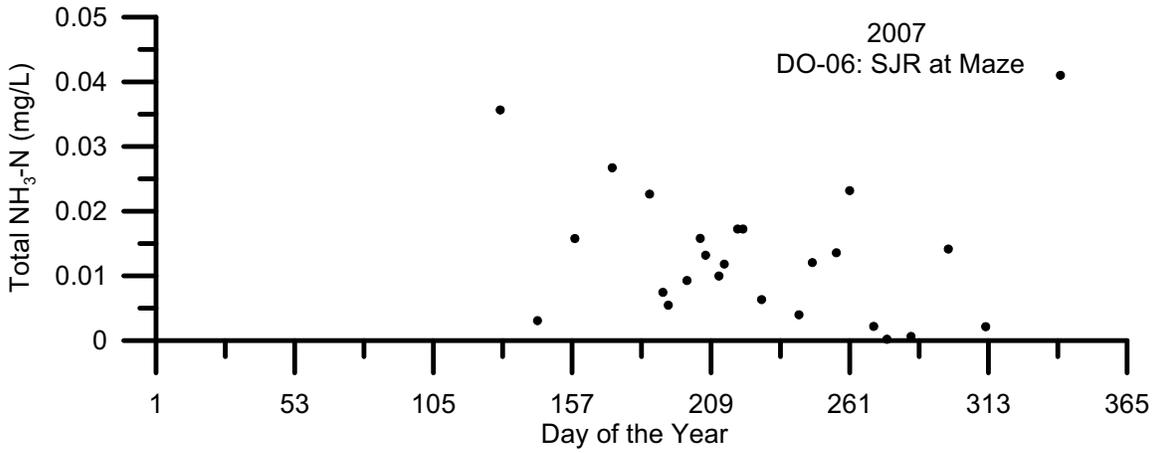
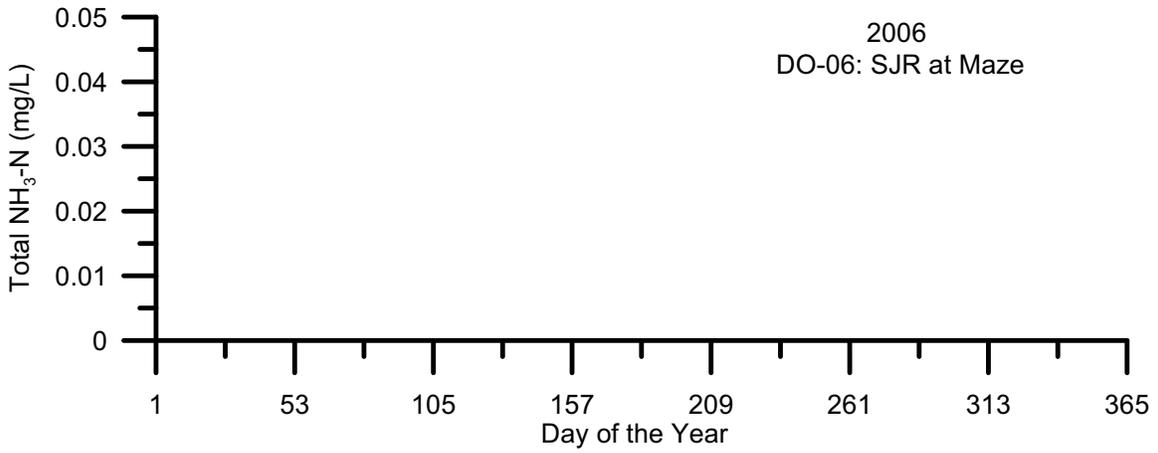
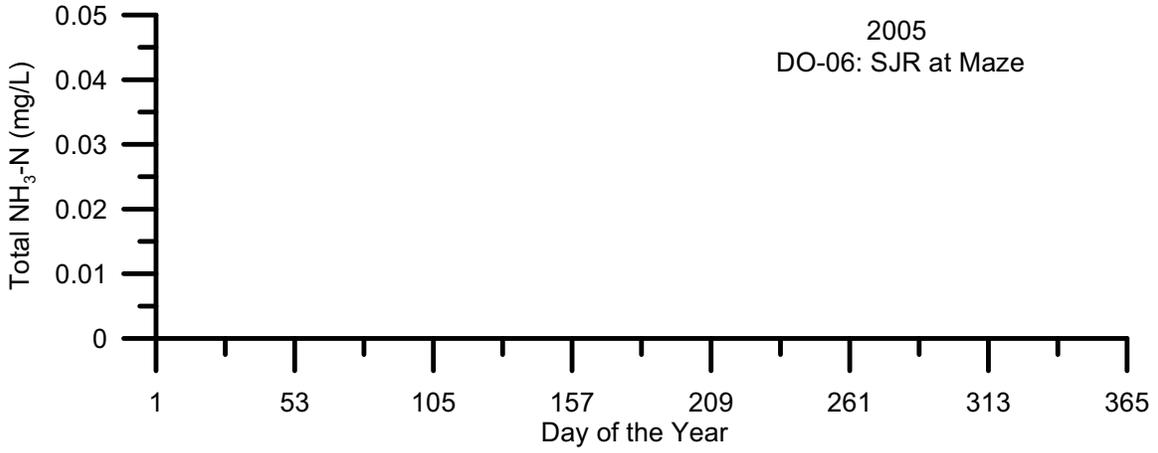
Figure 2: UCD vs EERP Timberline Plot of NH₃-N .

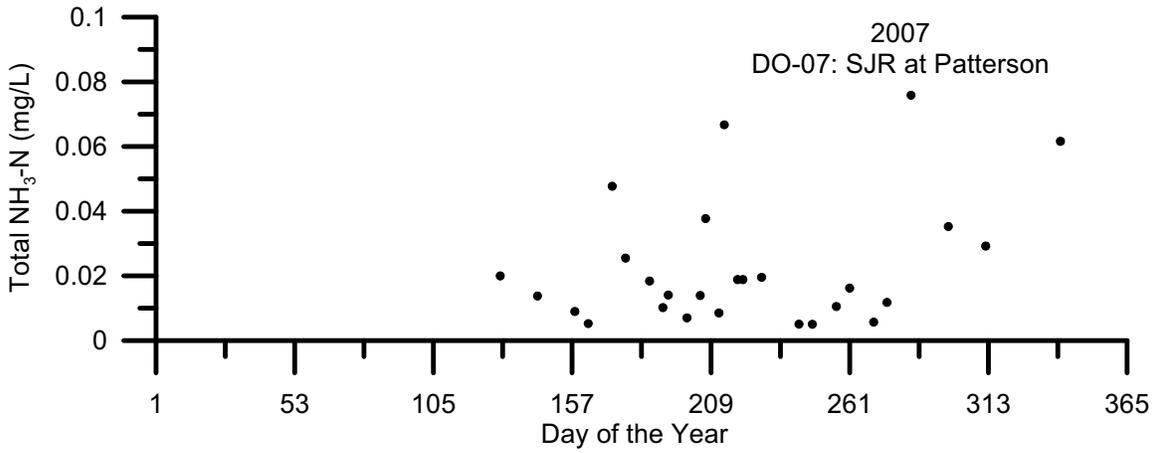
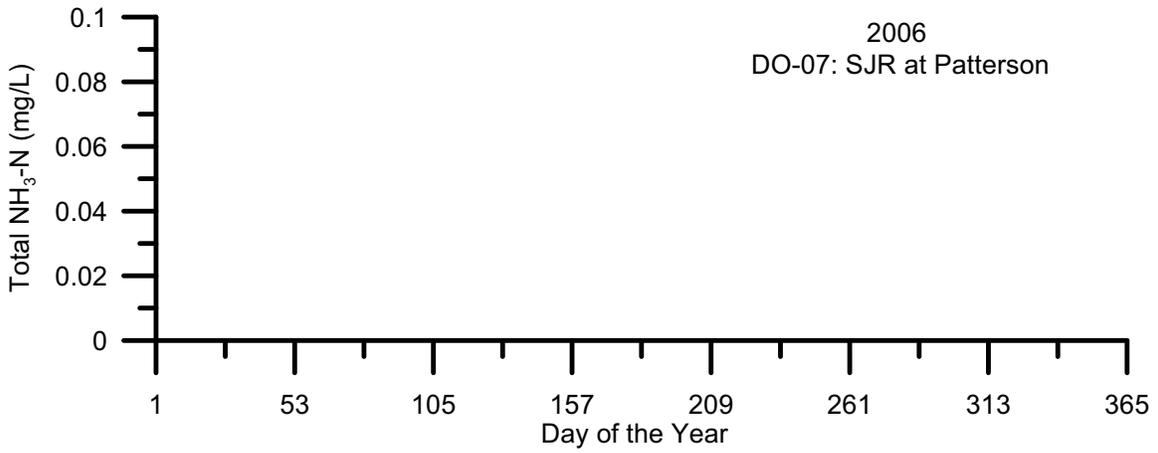
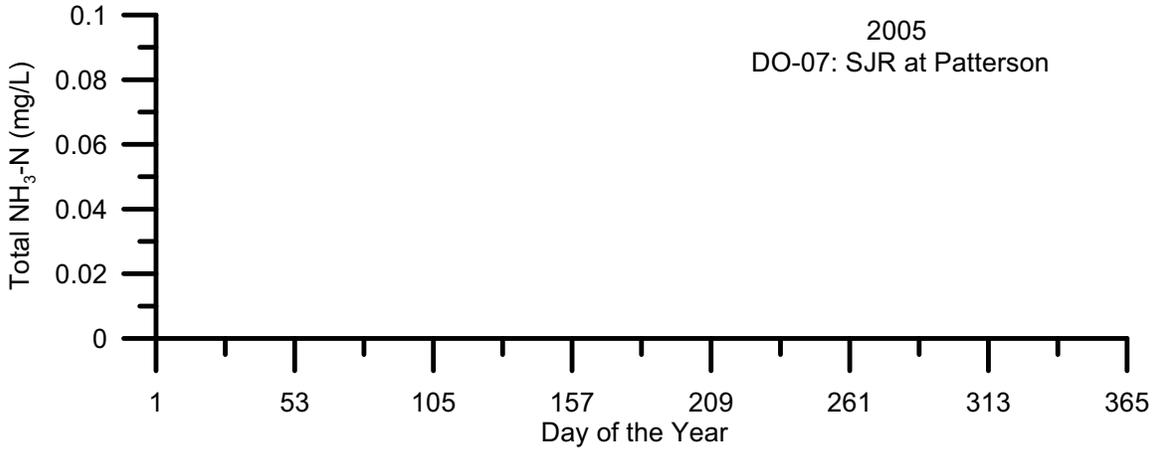


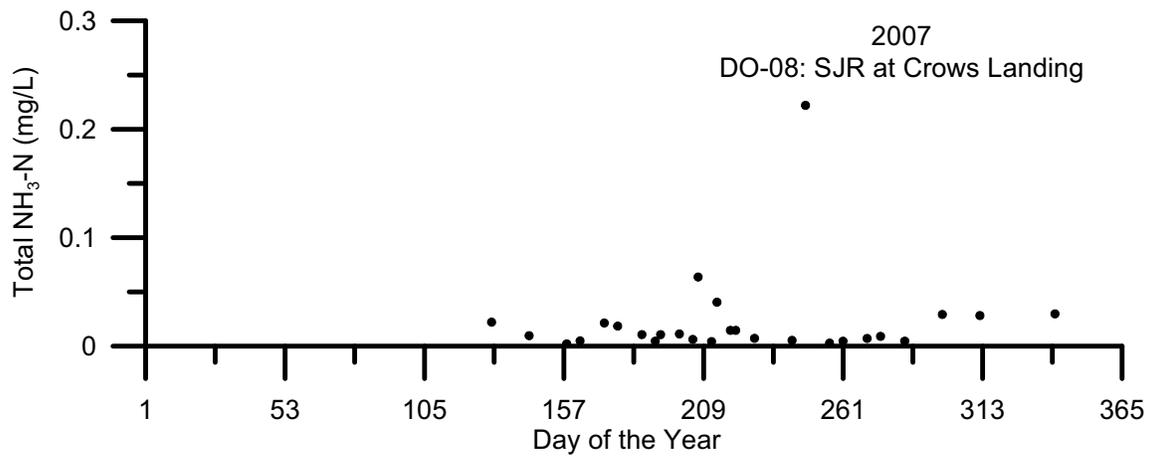
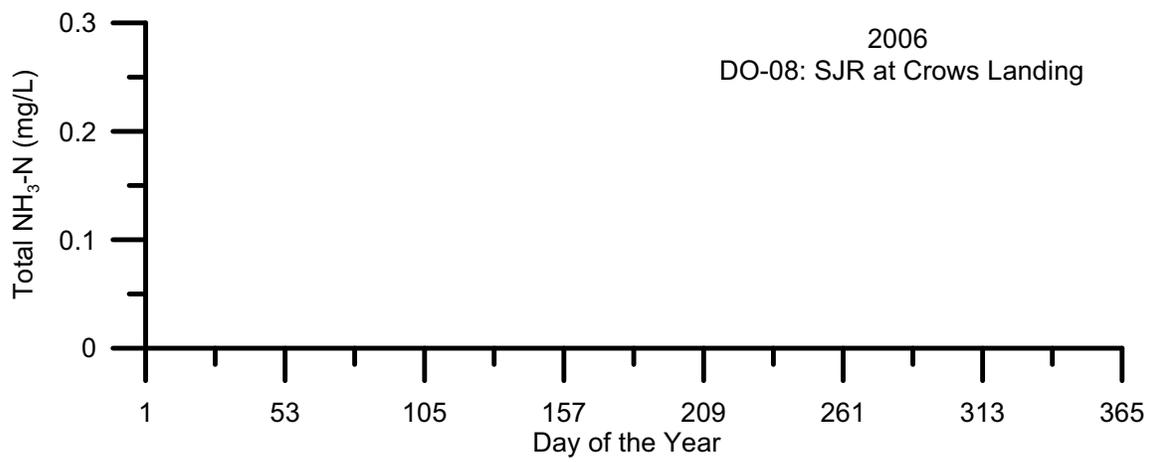
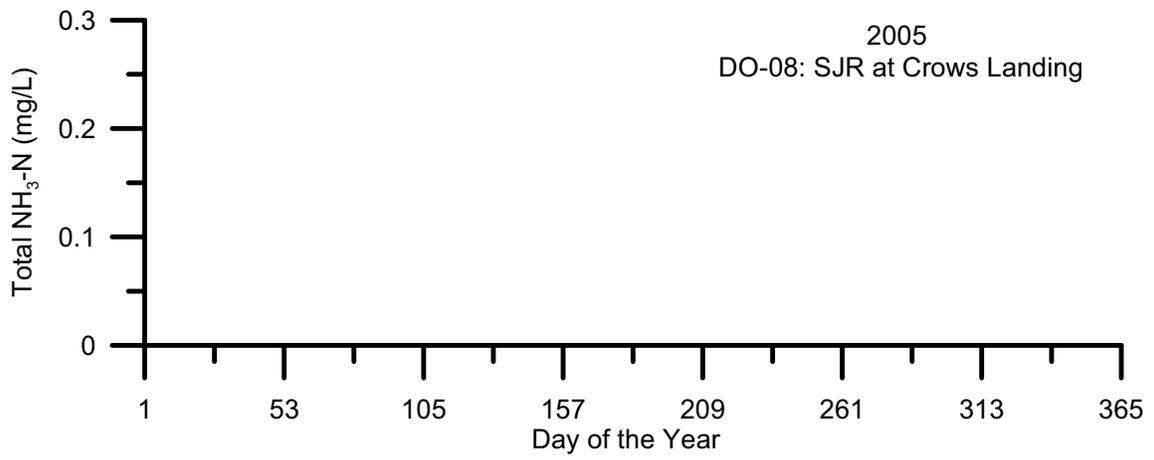
Figures 3 -104: Temporal Plots of Total NH₃-N By Site ID

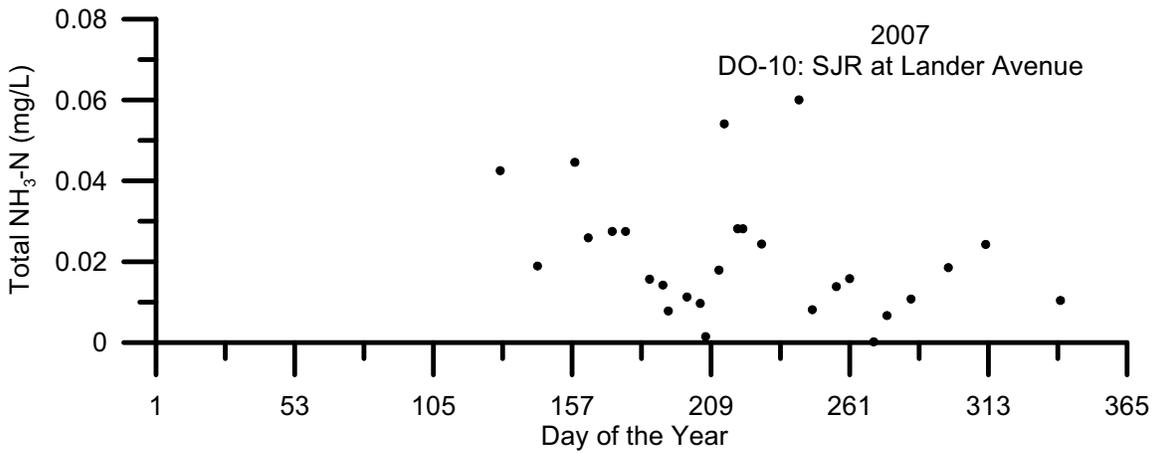
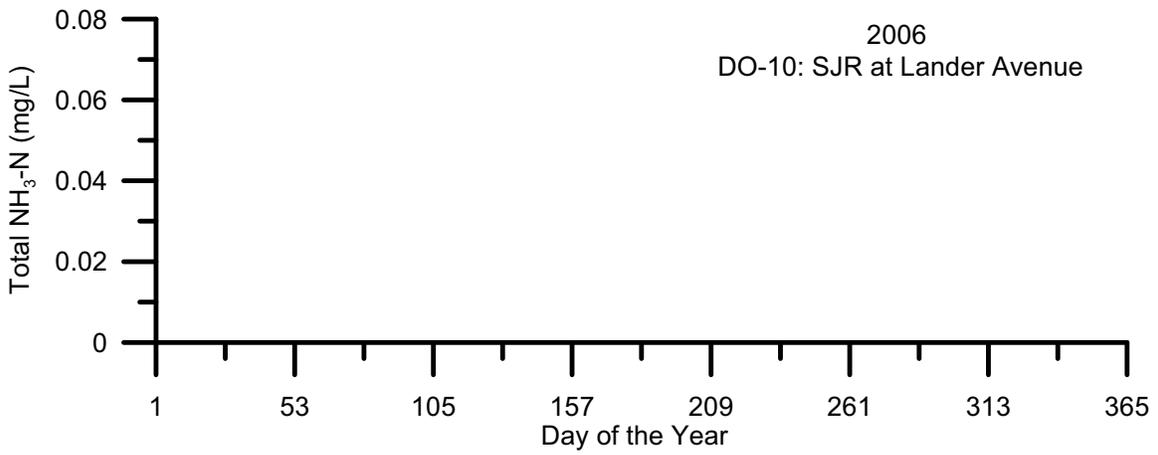
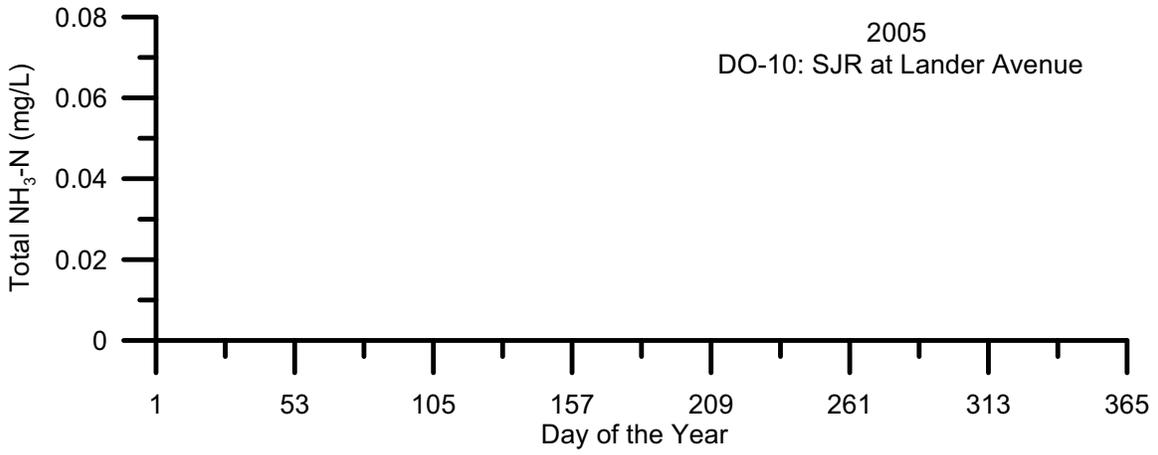


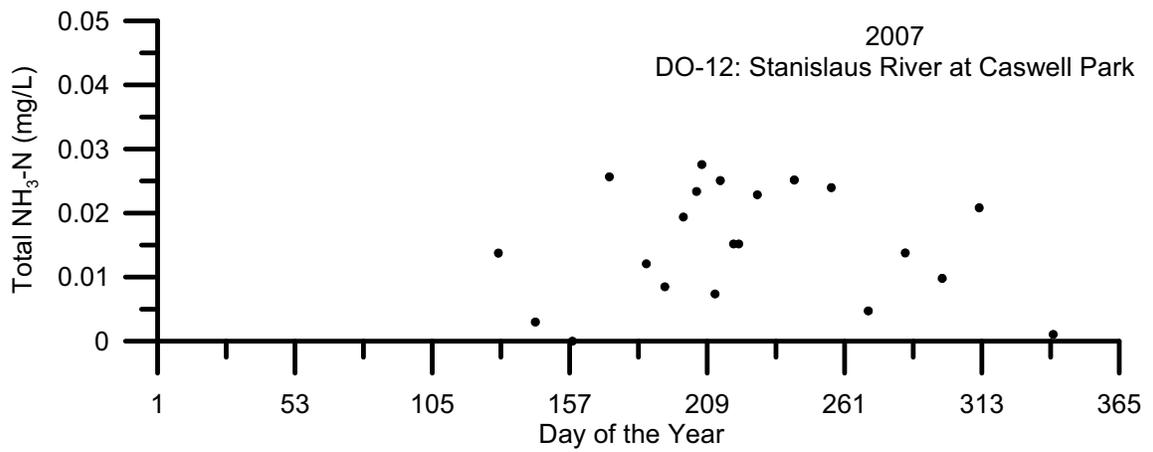
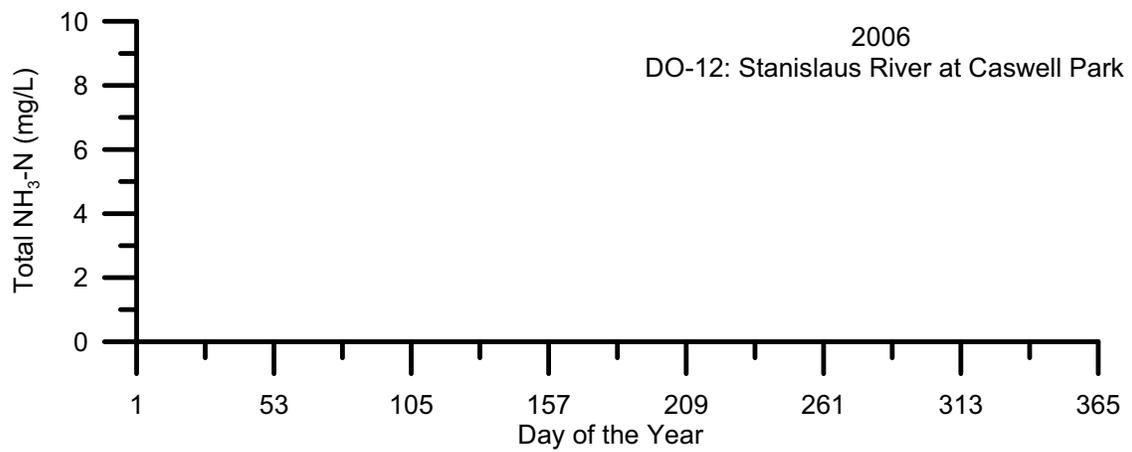
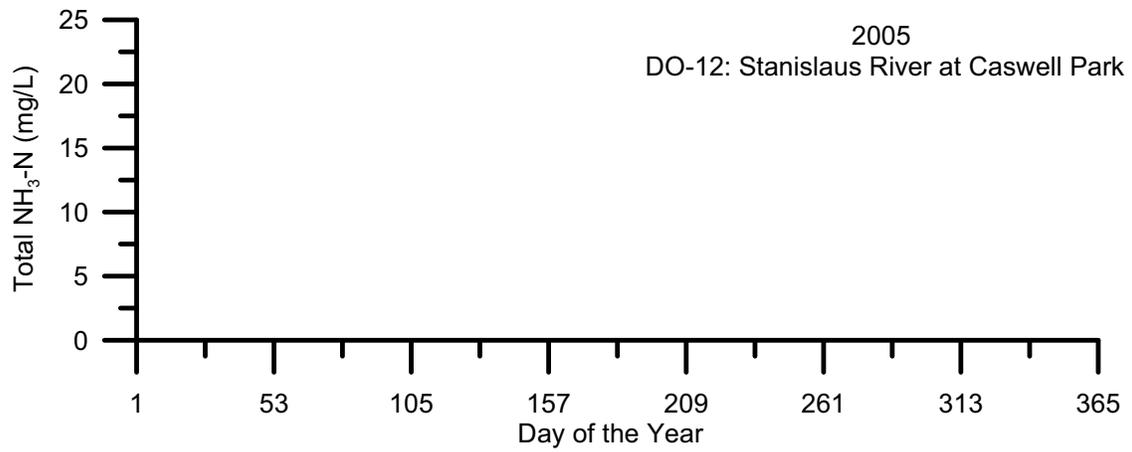


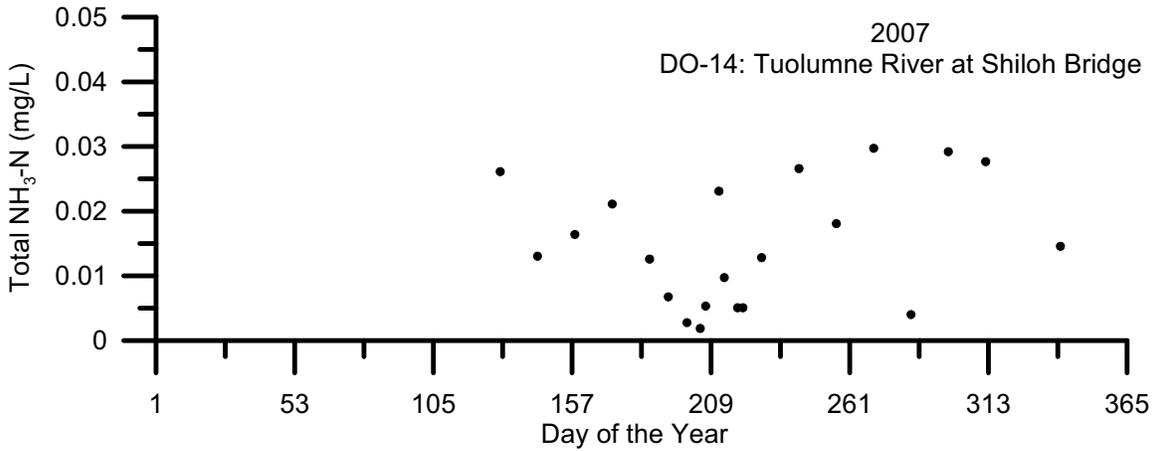
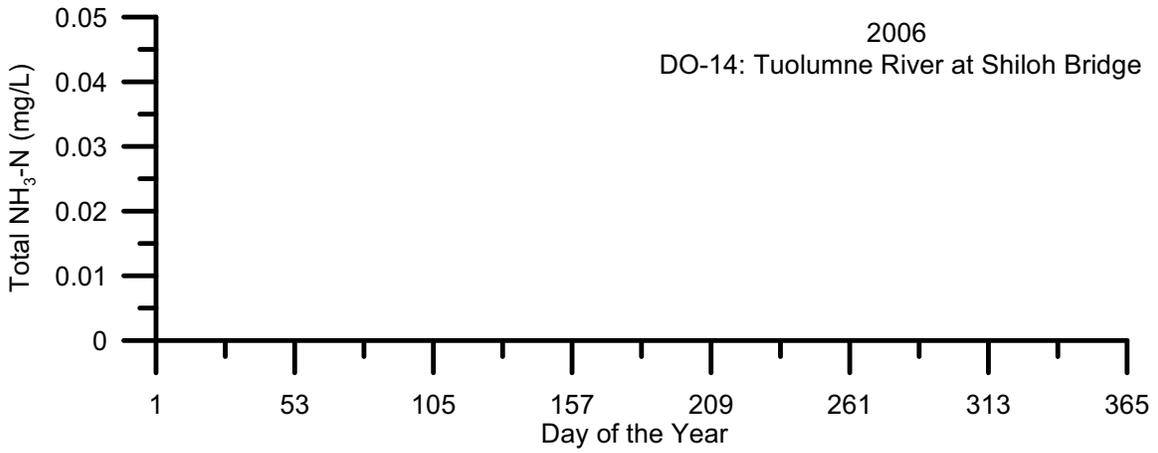
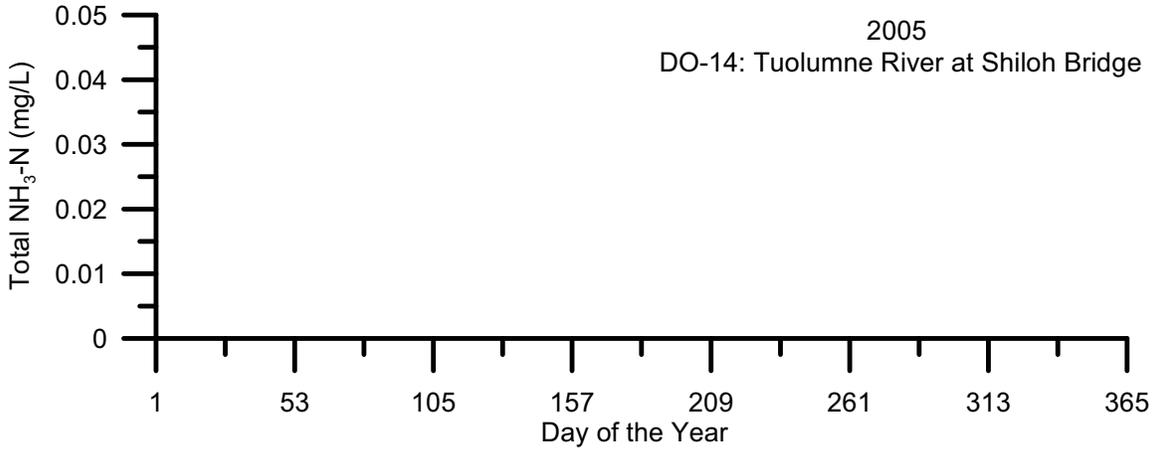


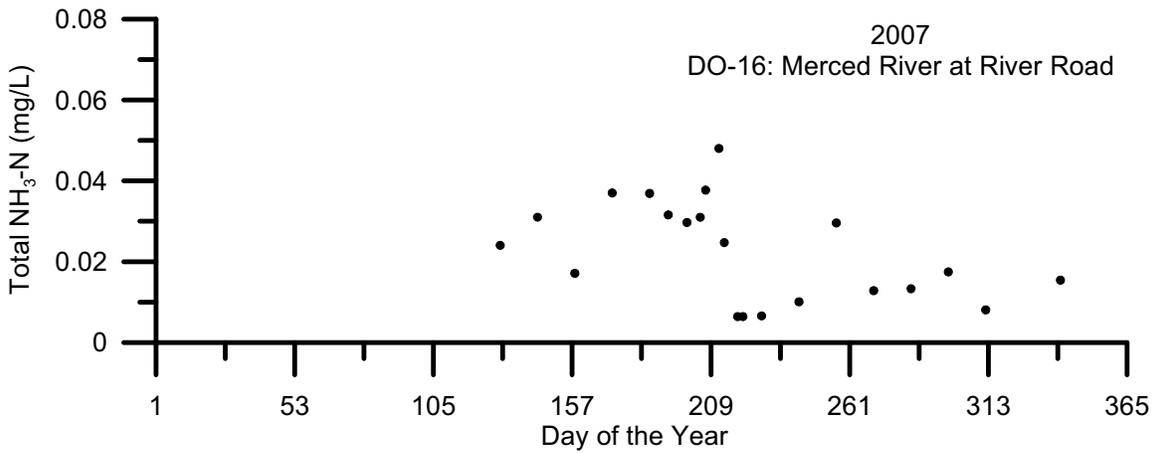
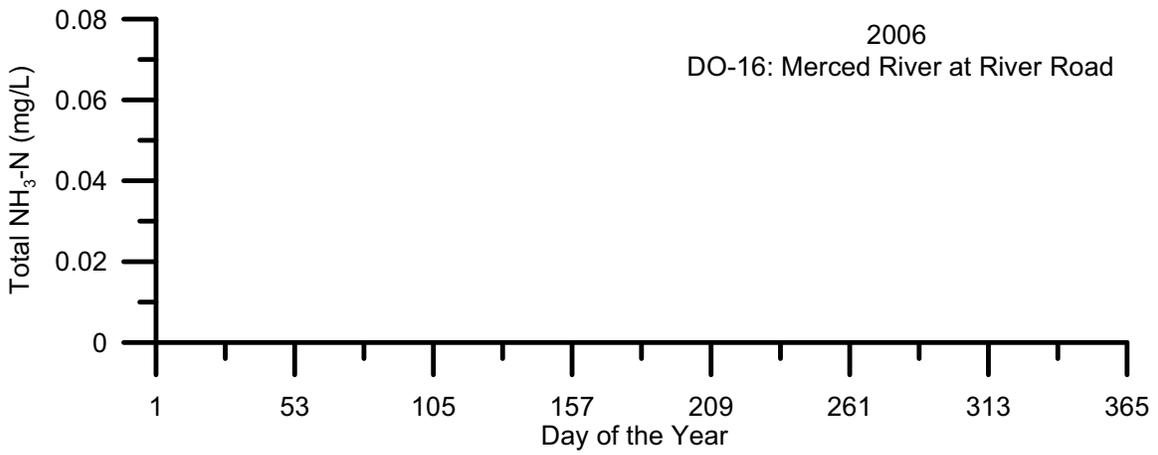
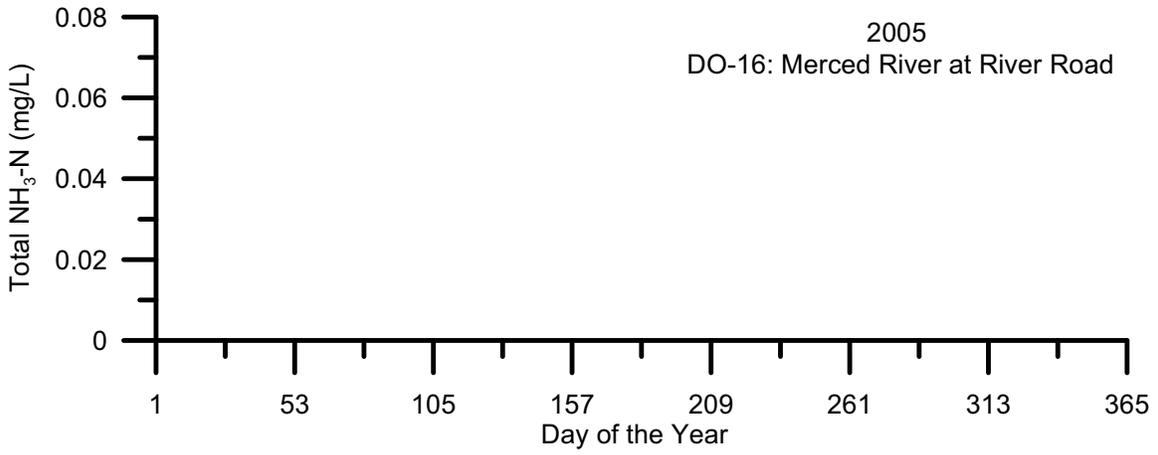


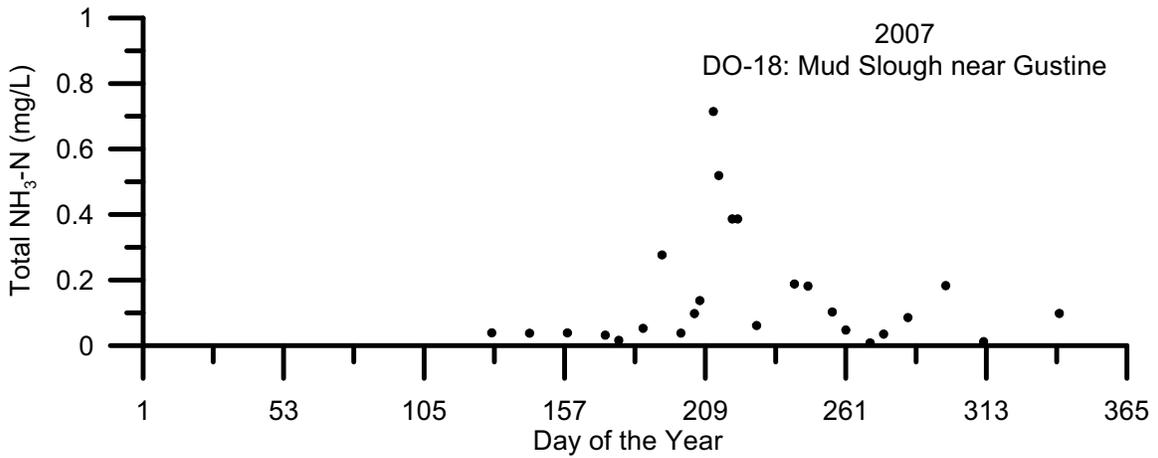
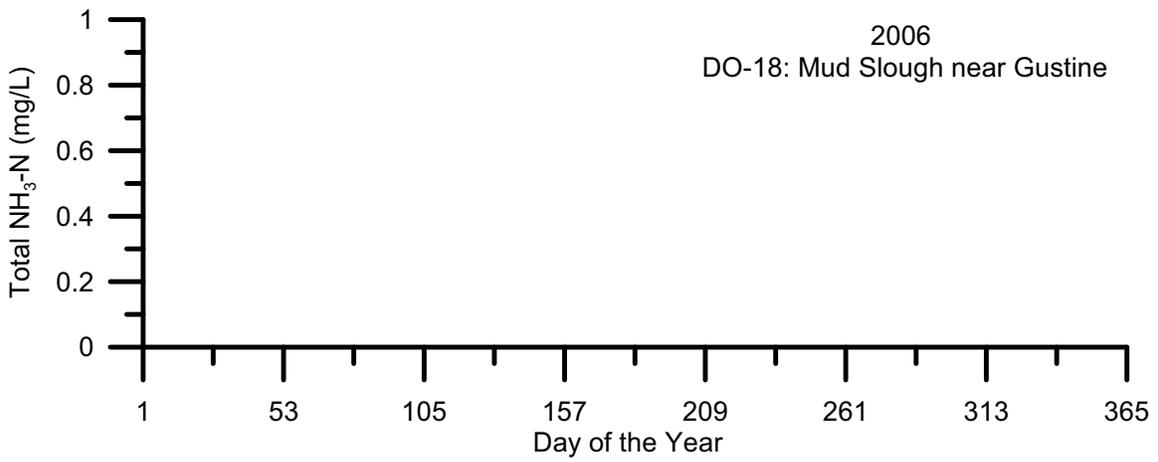
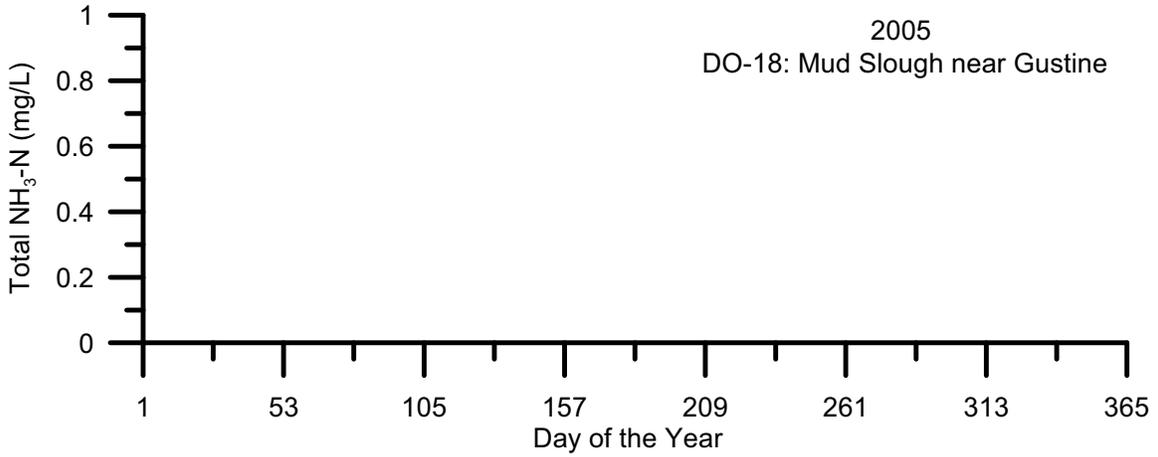


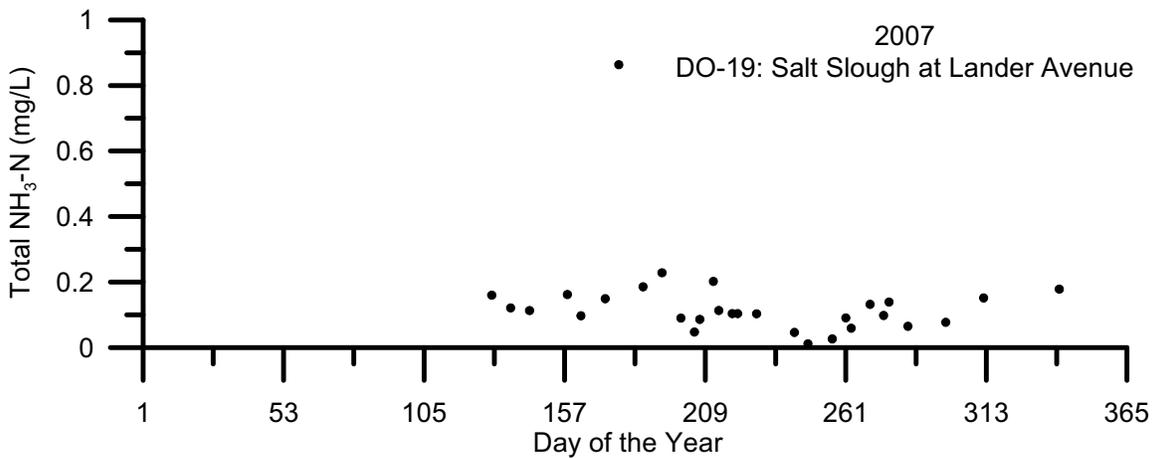
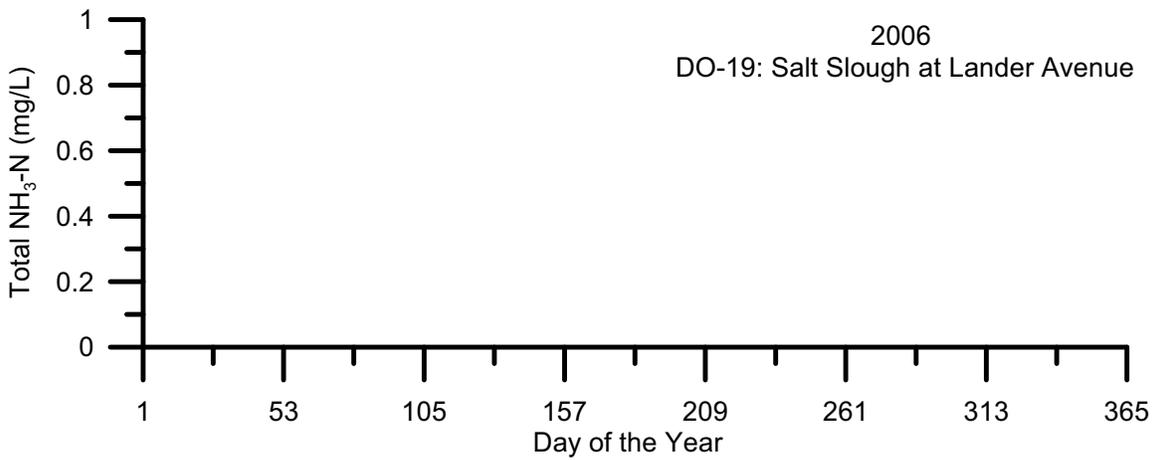
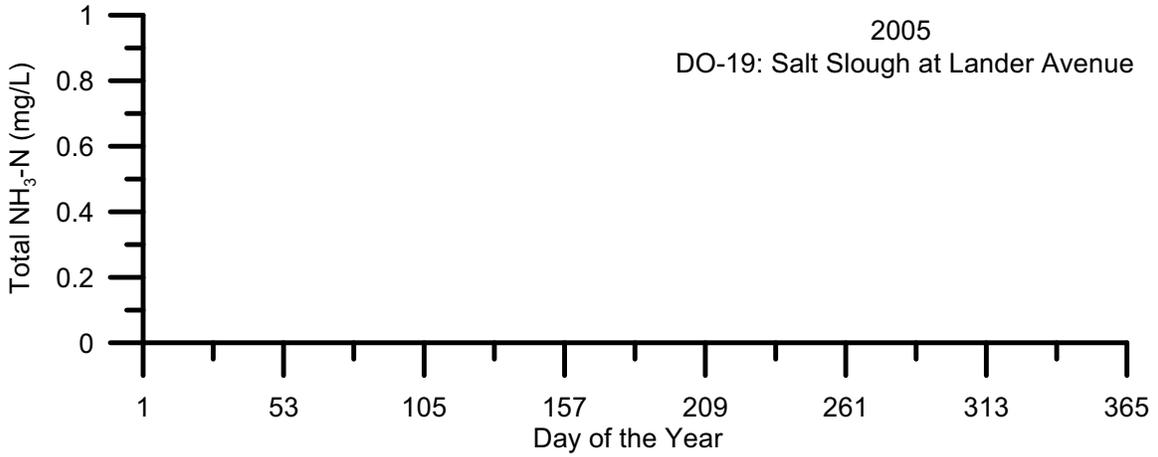


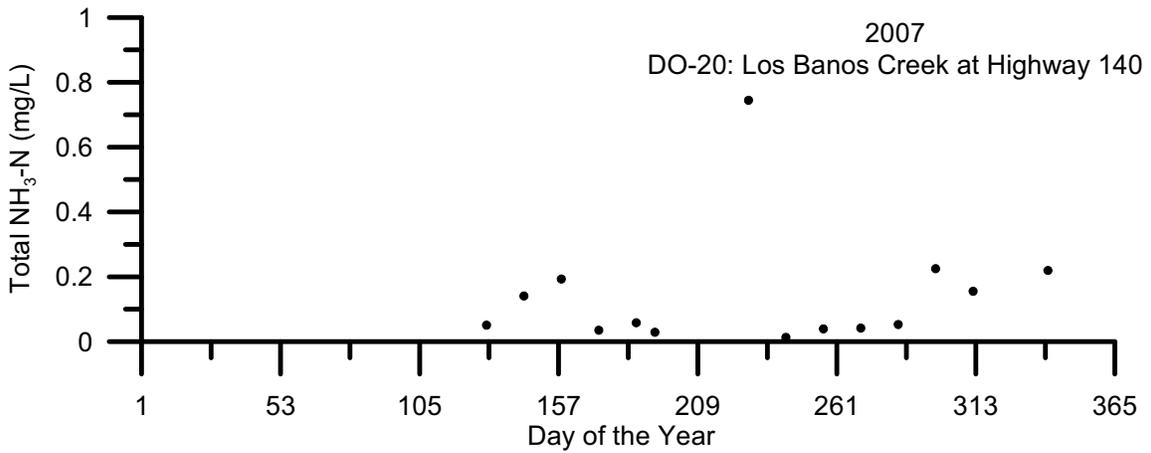
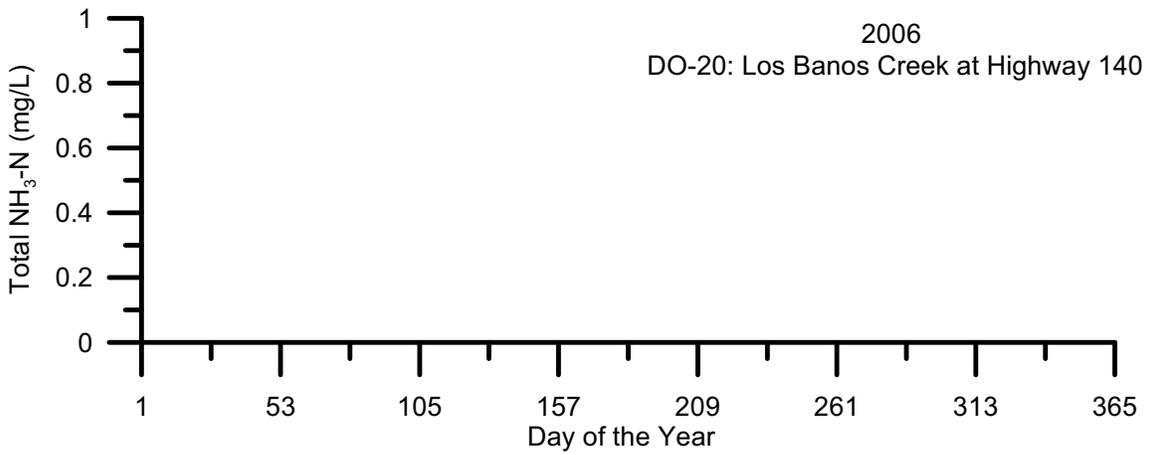
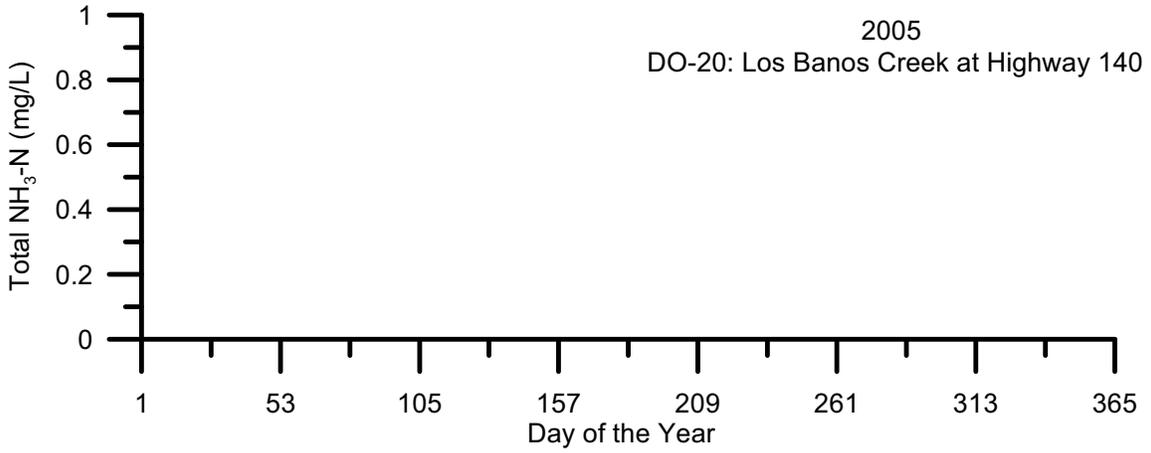


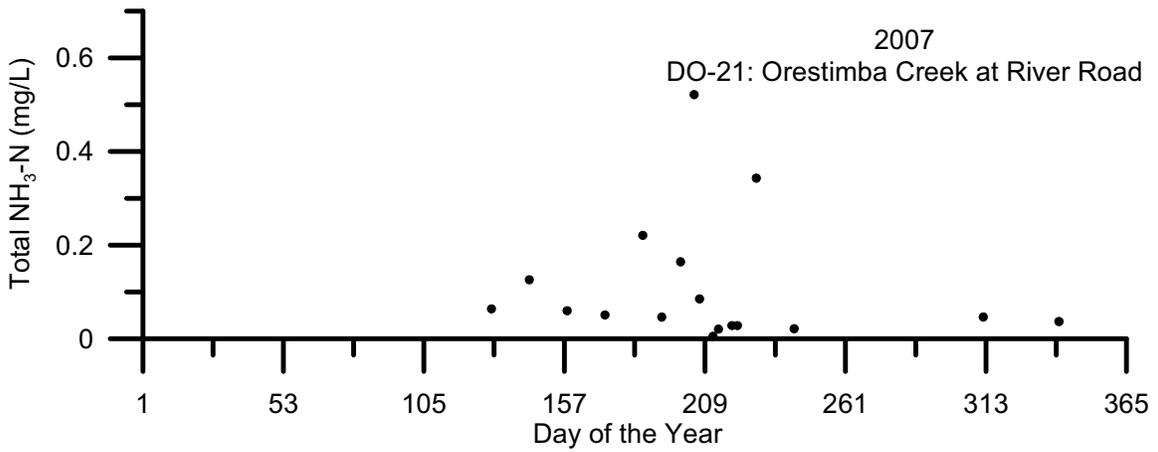
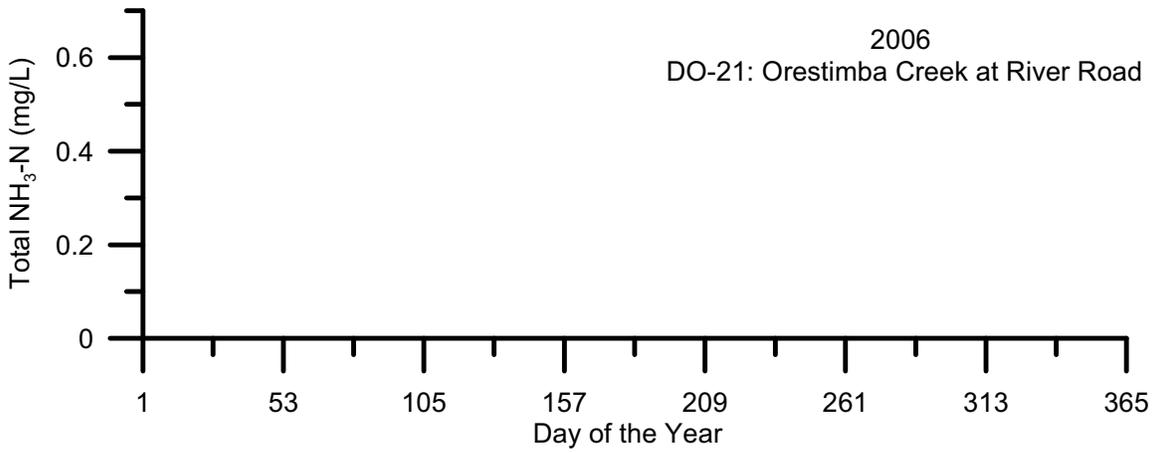
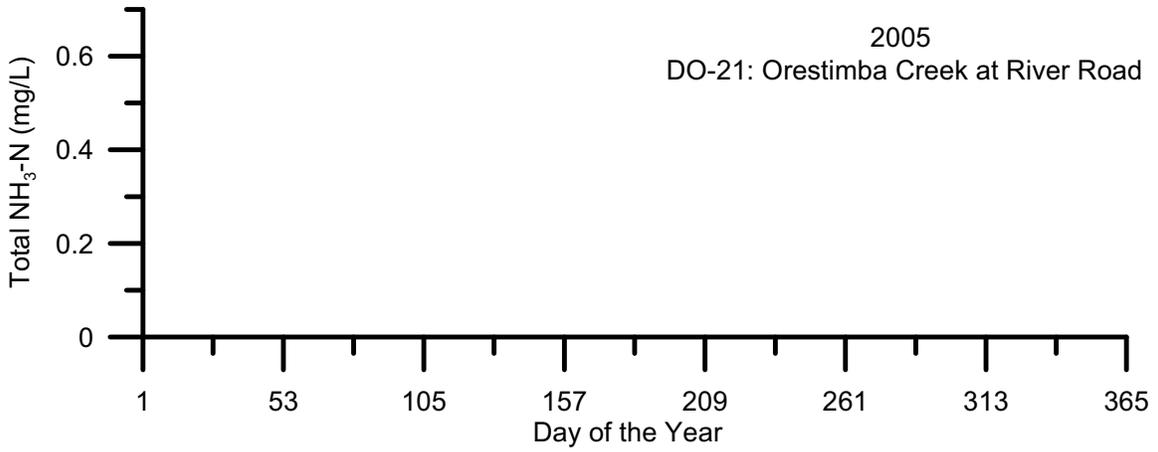


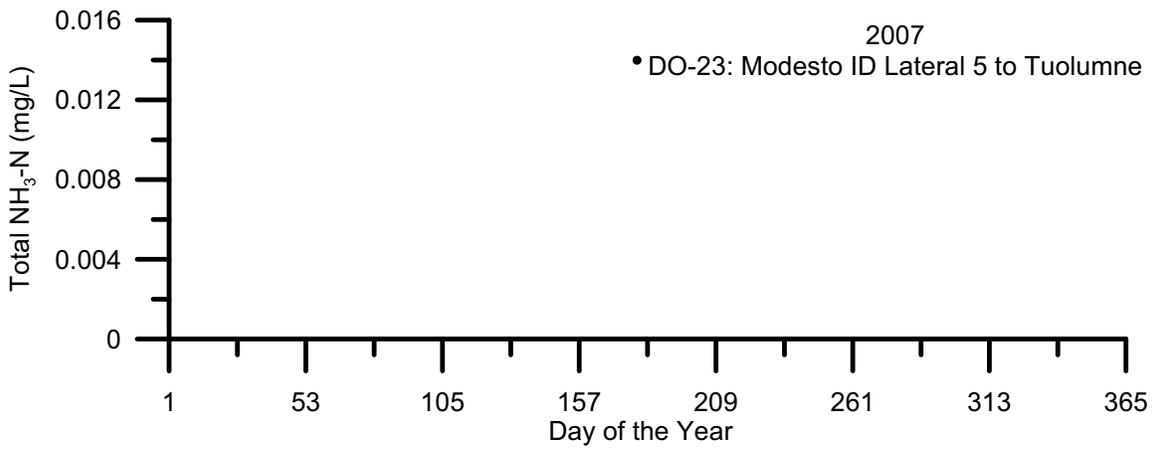
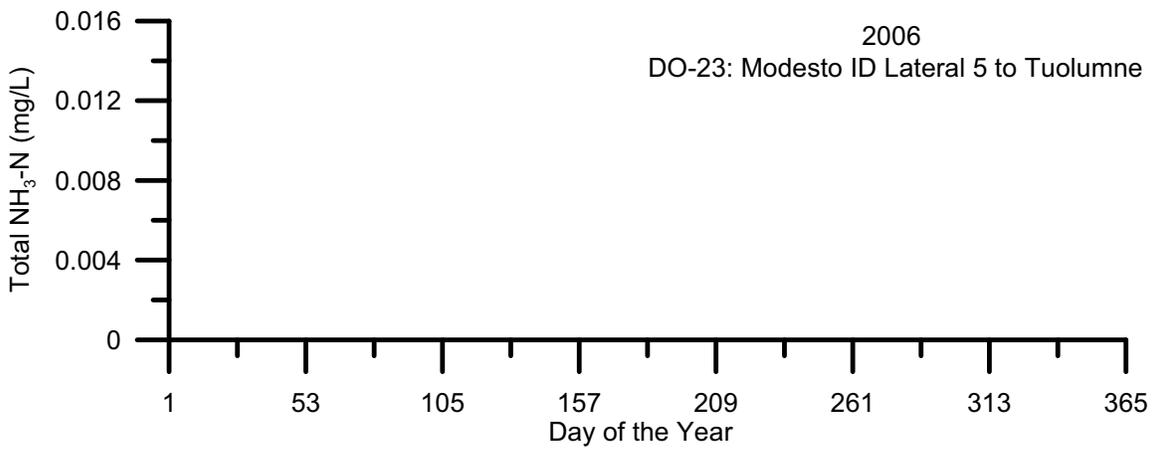
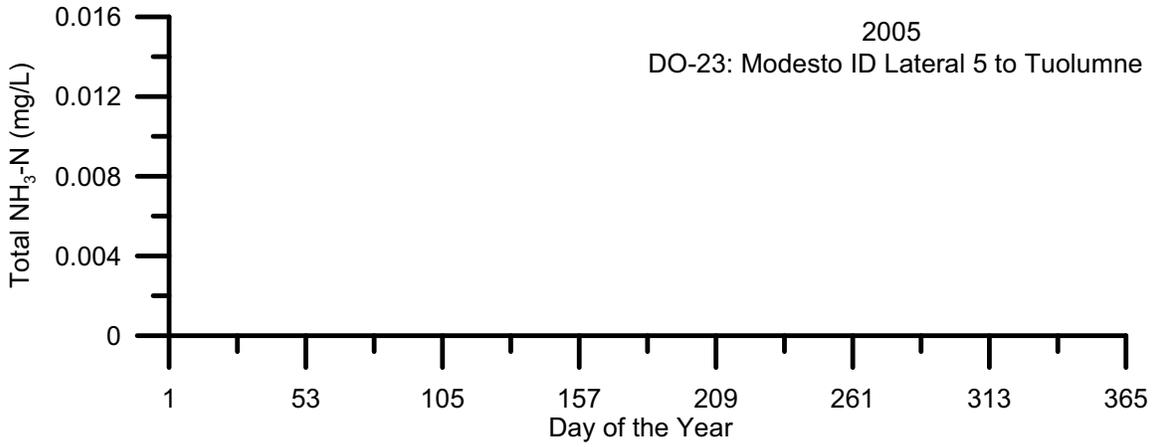


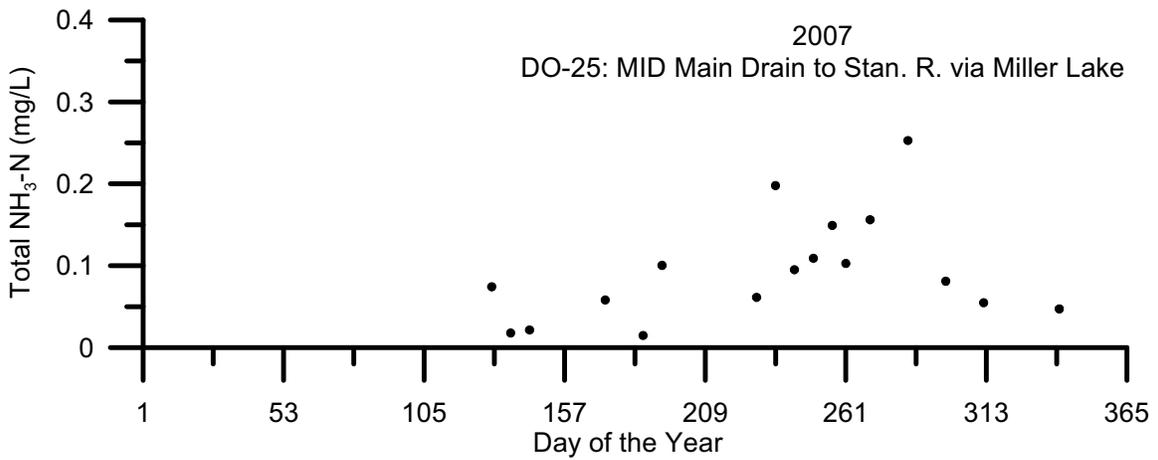
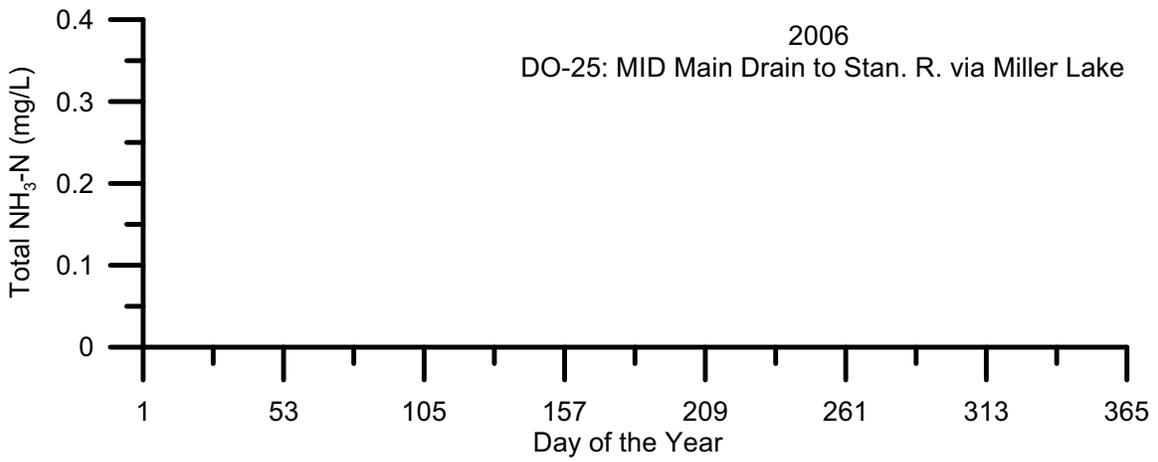
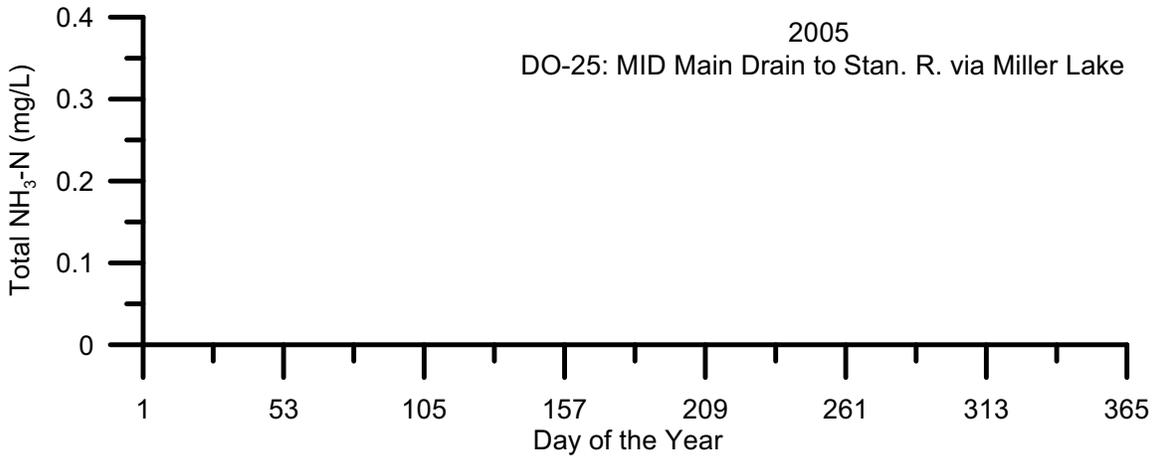


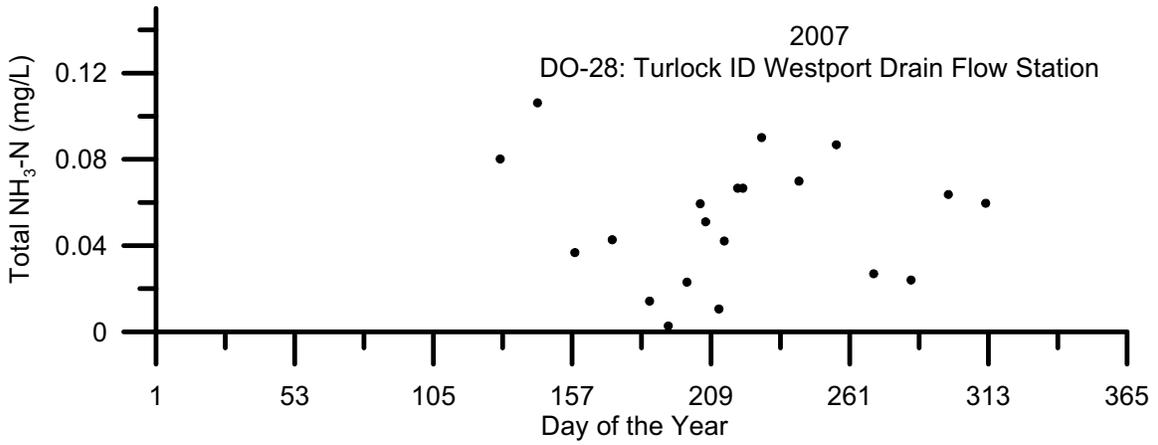
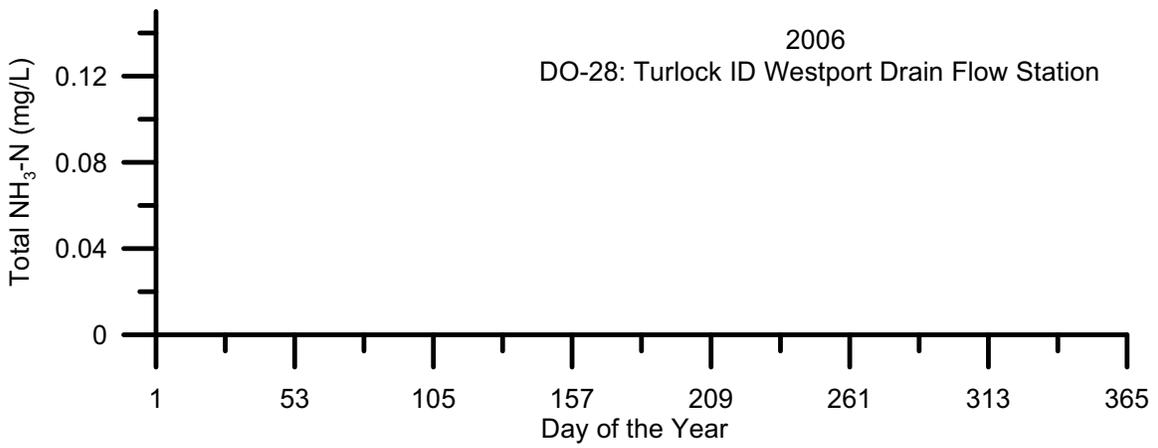
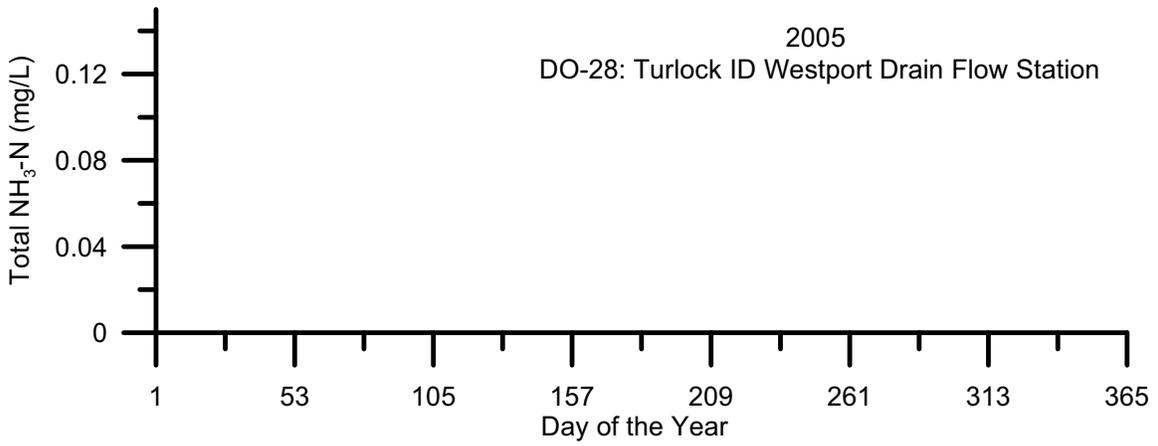


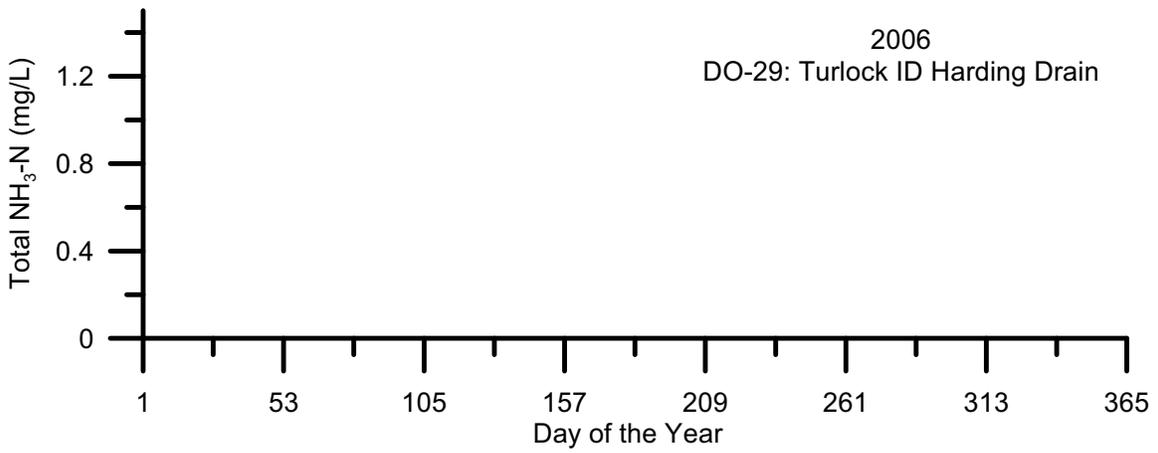
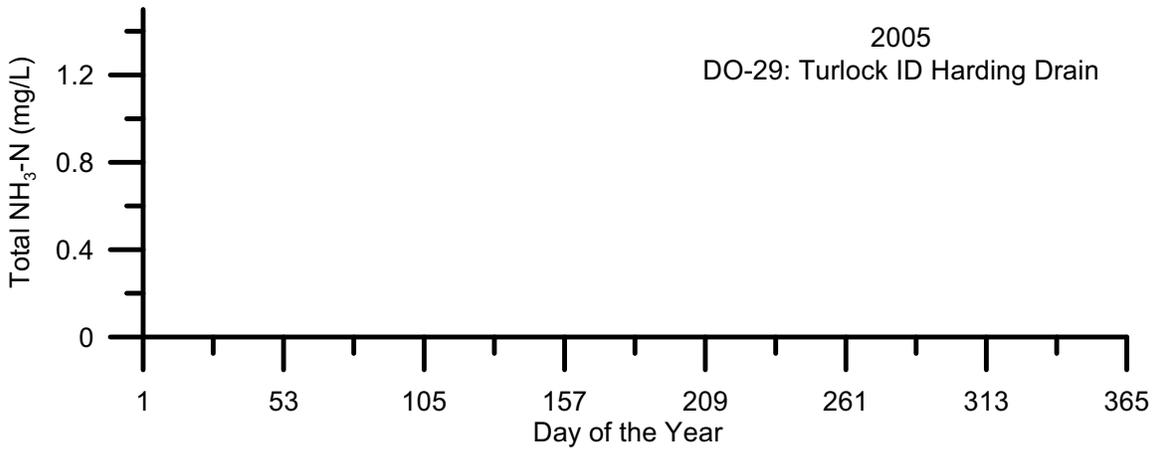


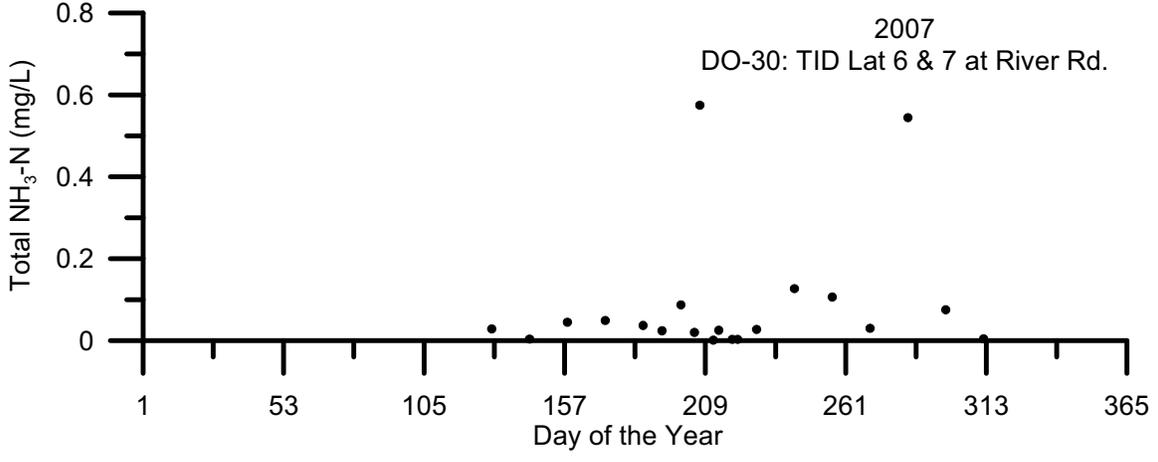
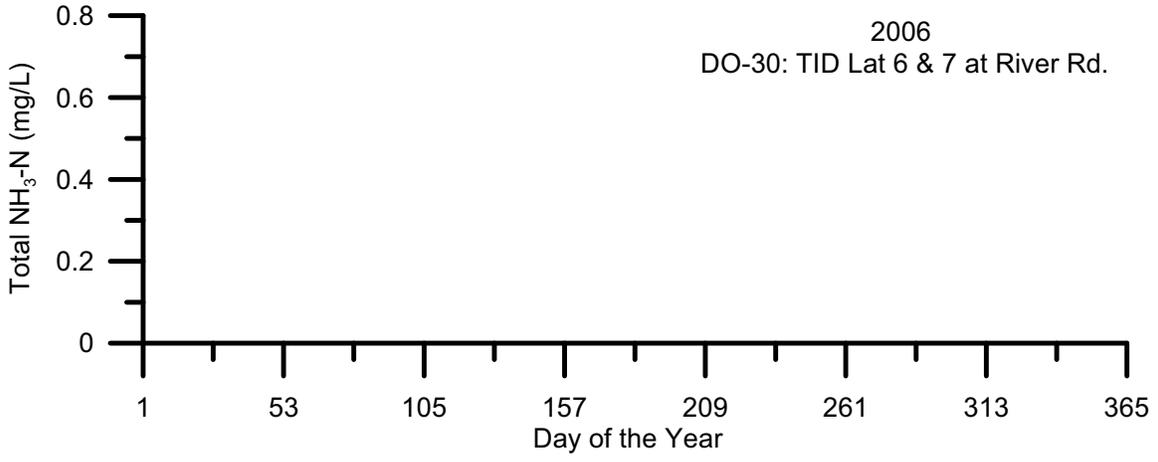
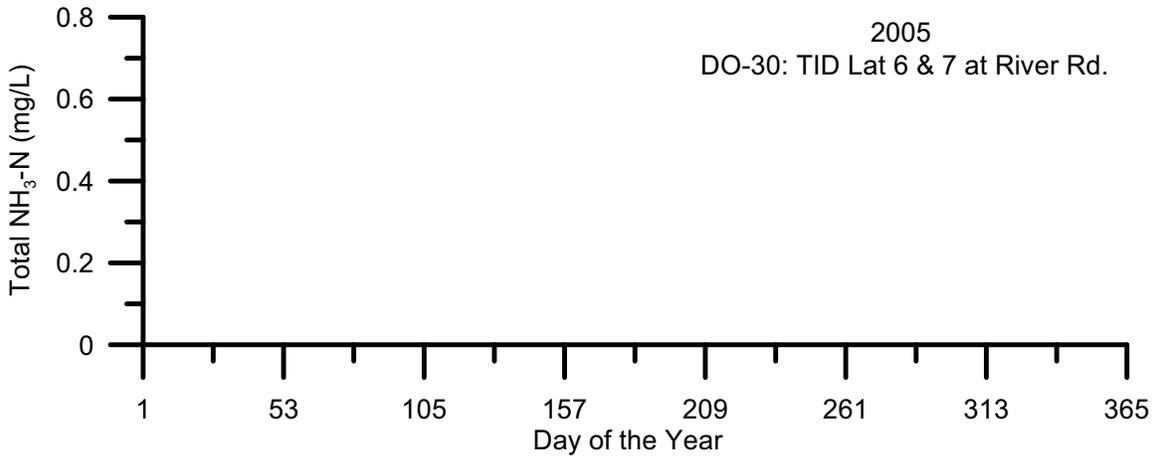


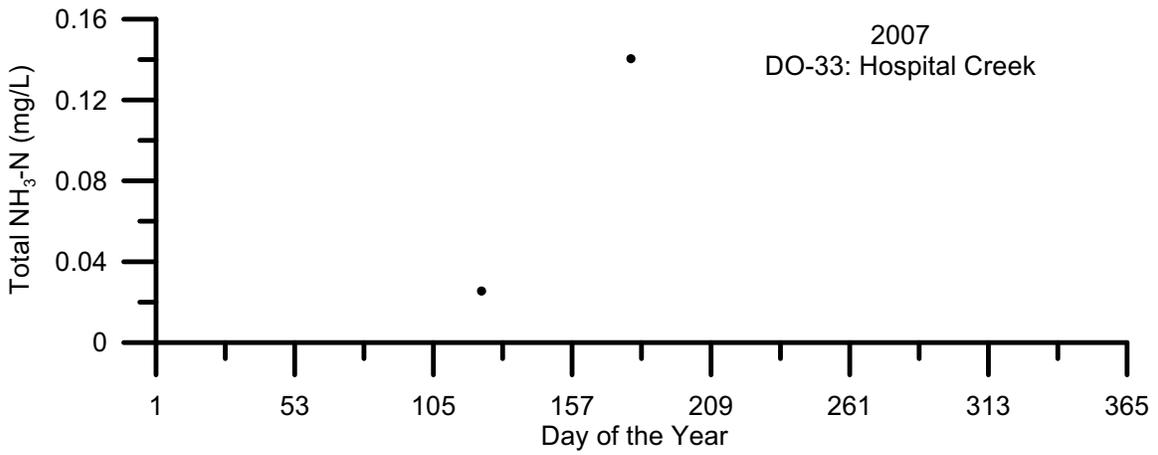
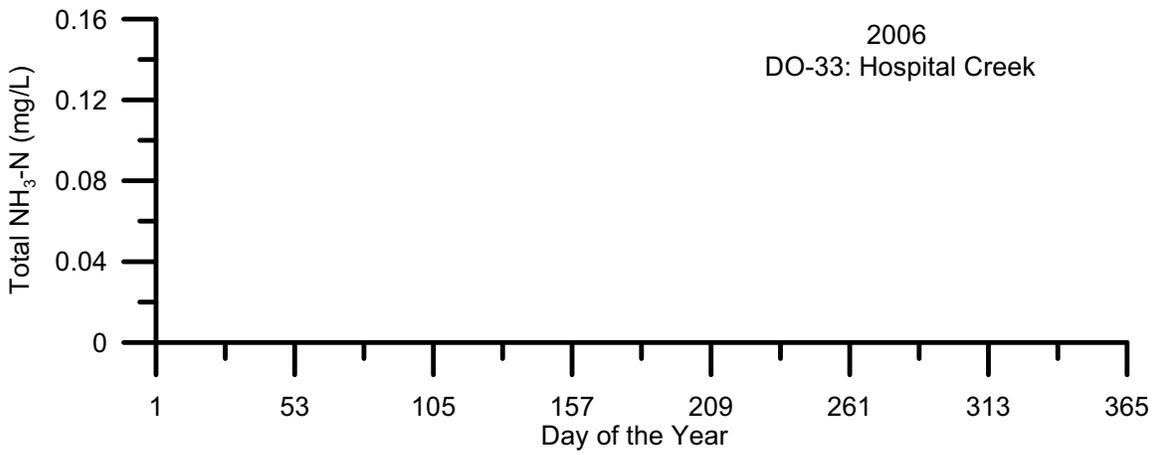
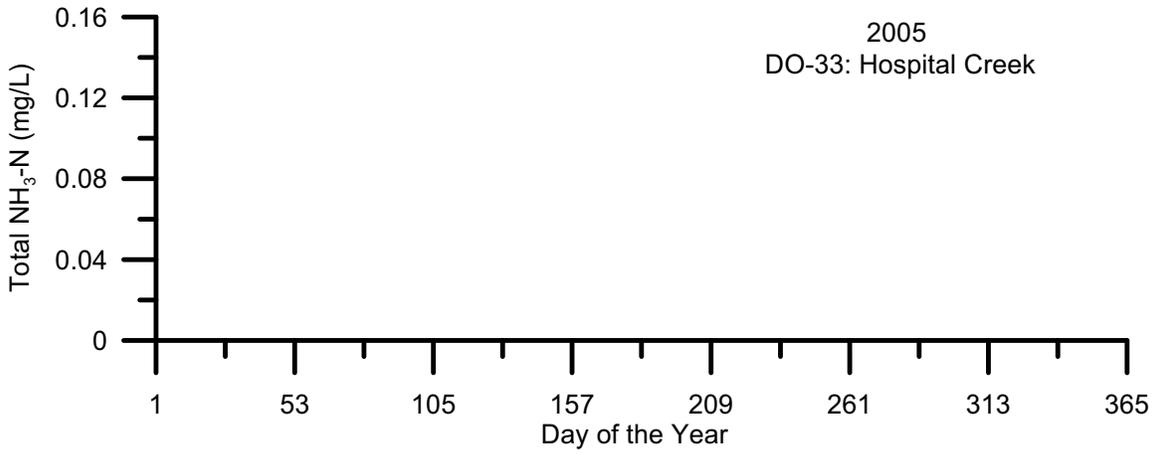


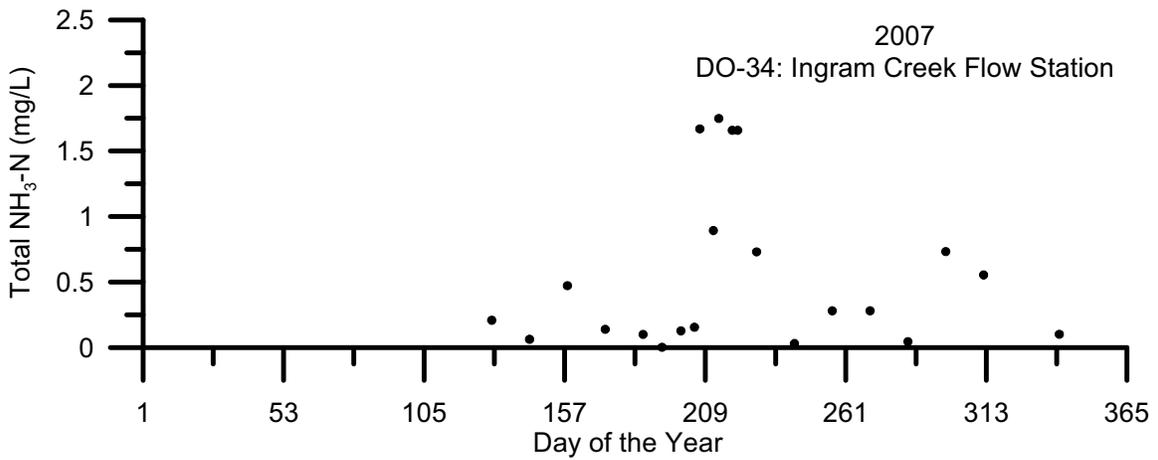
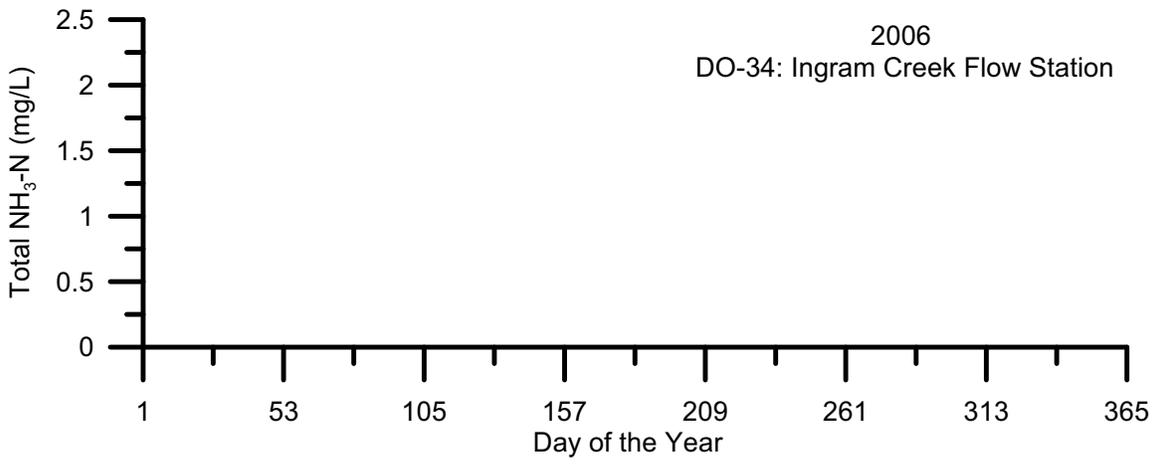
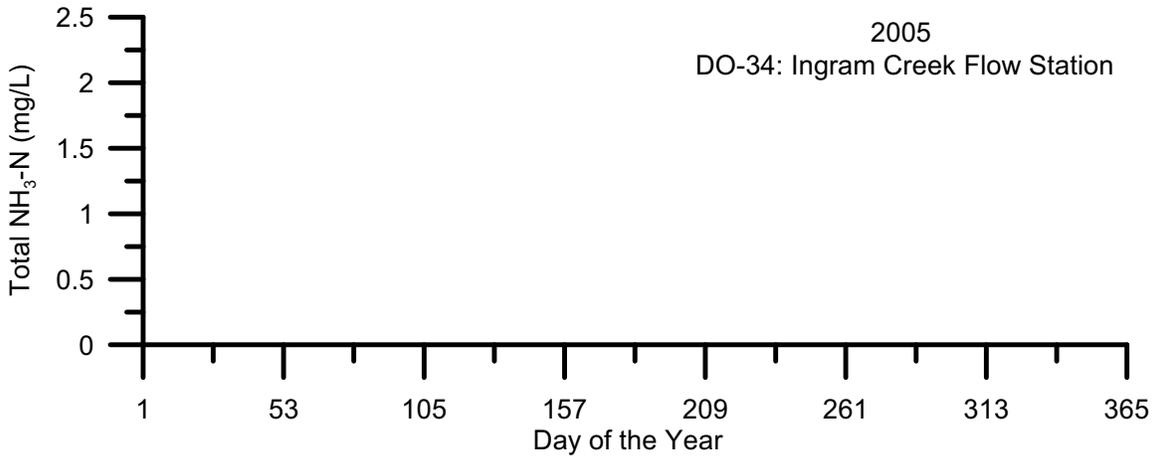


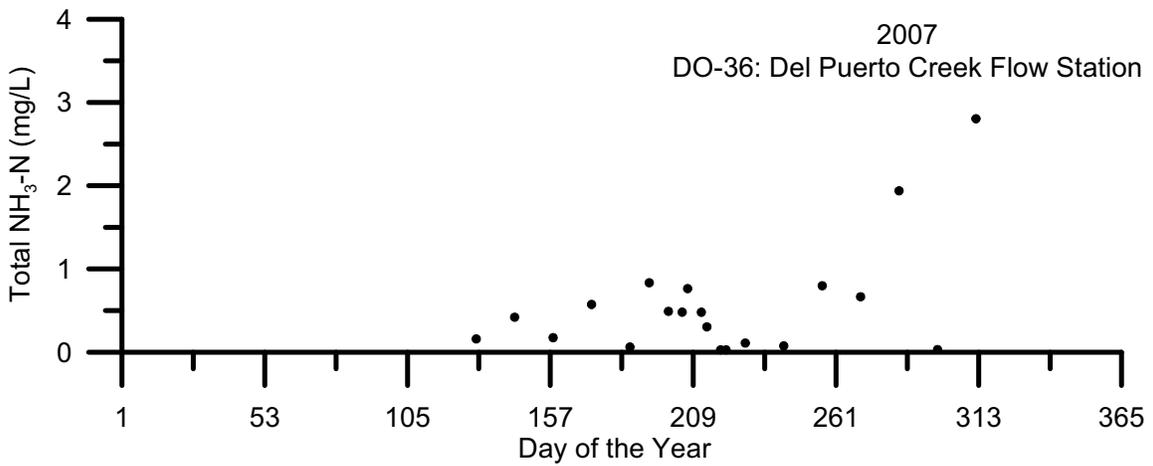
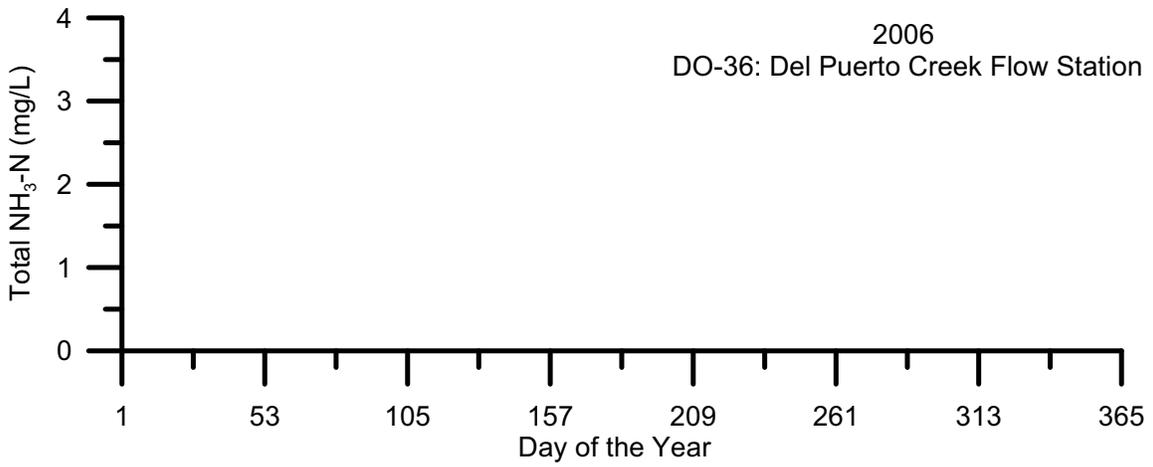
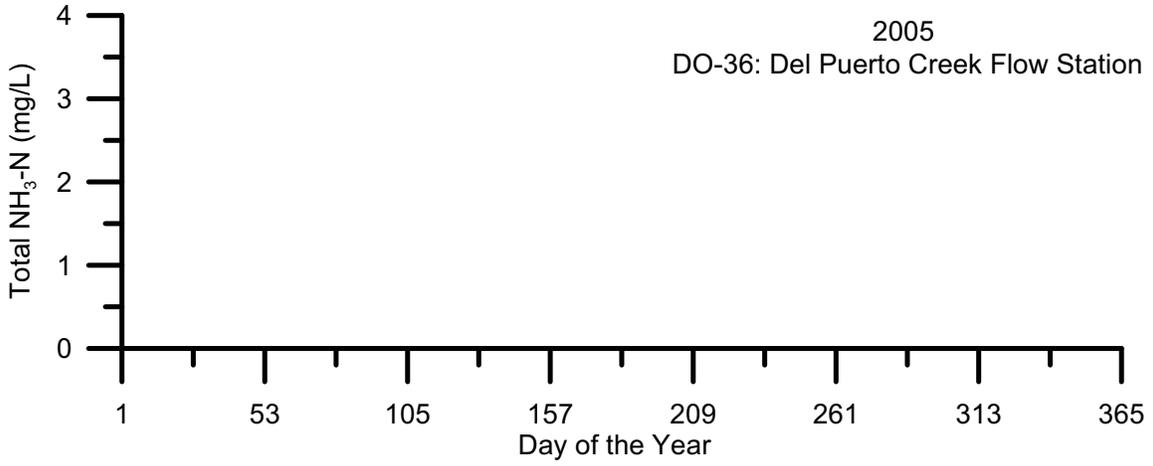


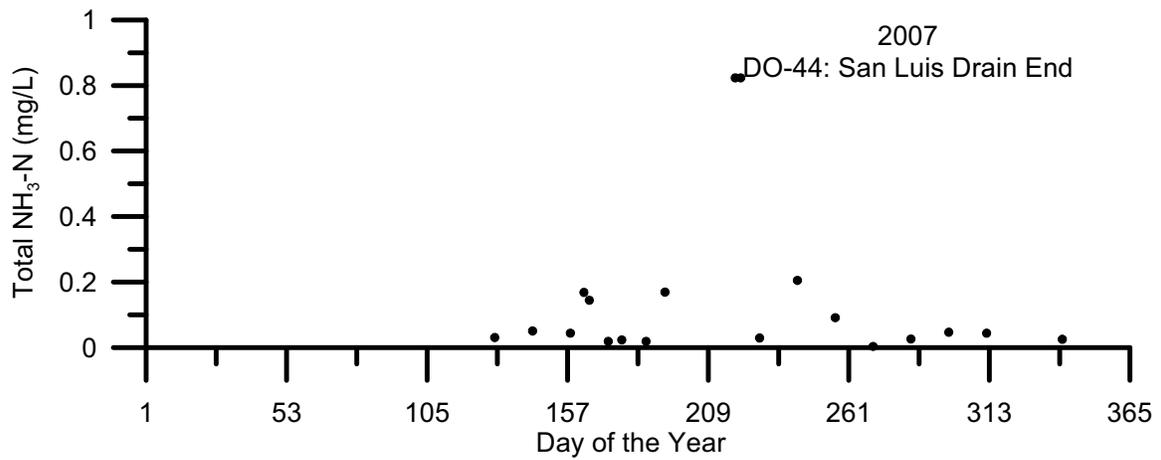
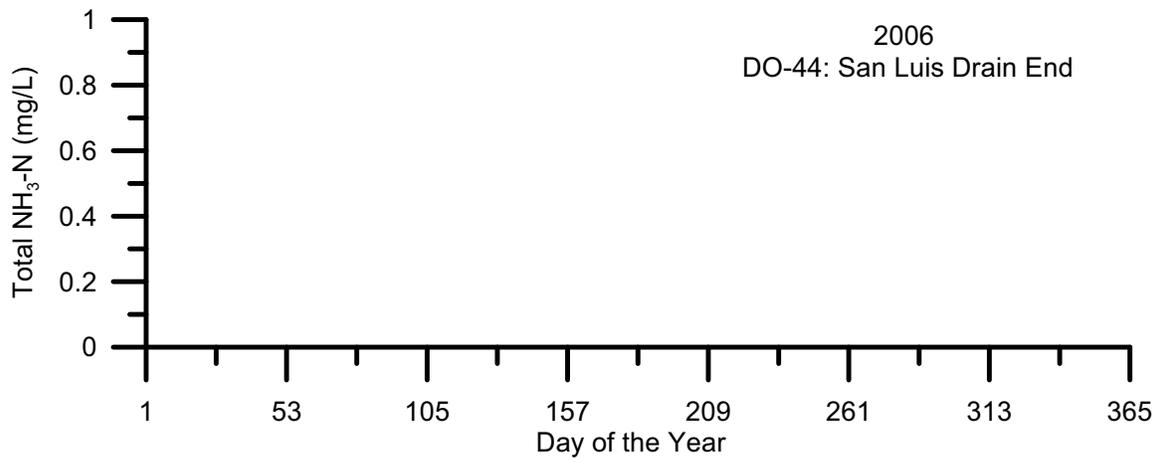
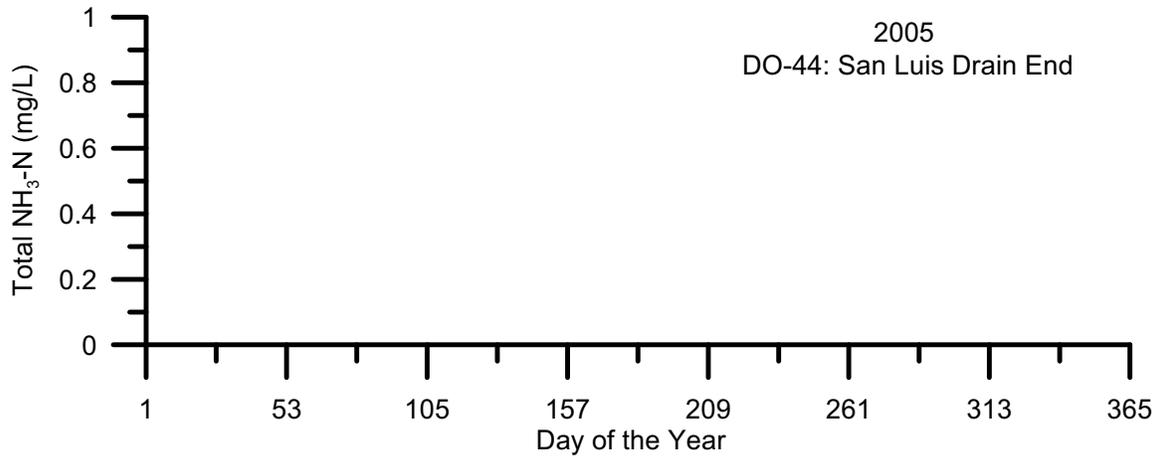


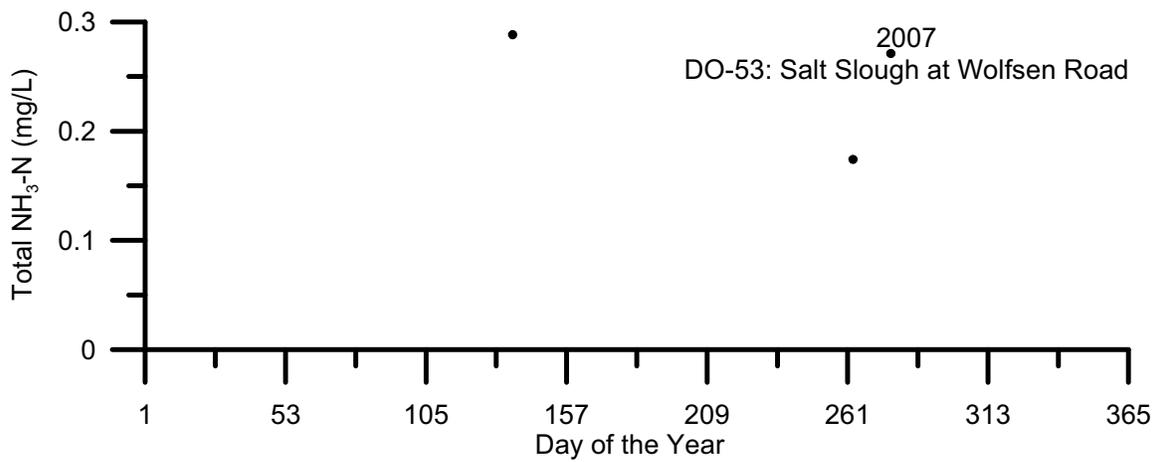
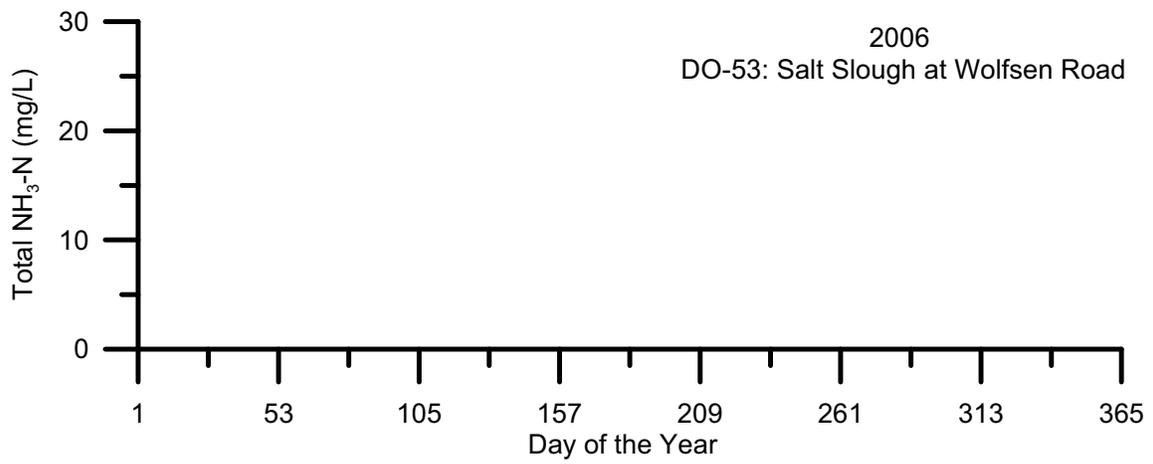
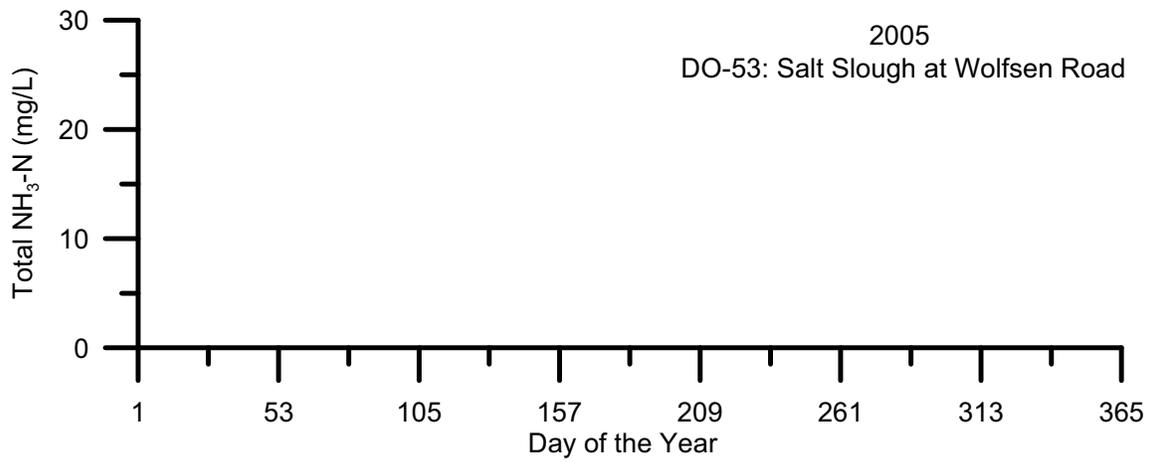


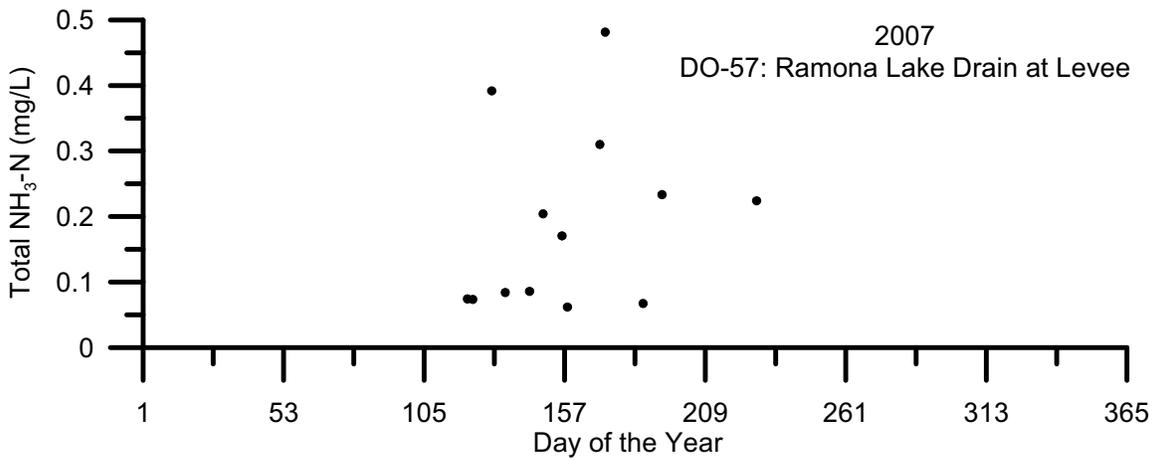
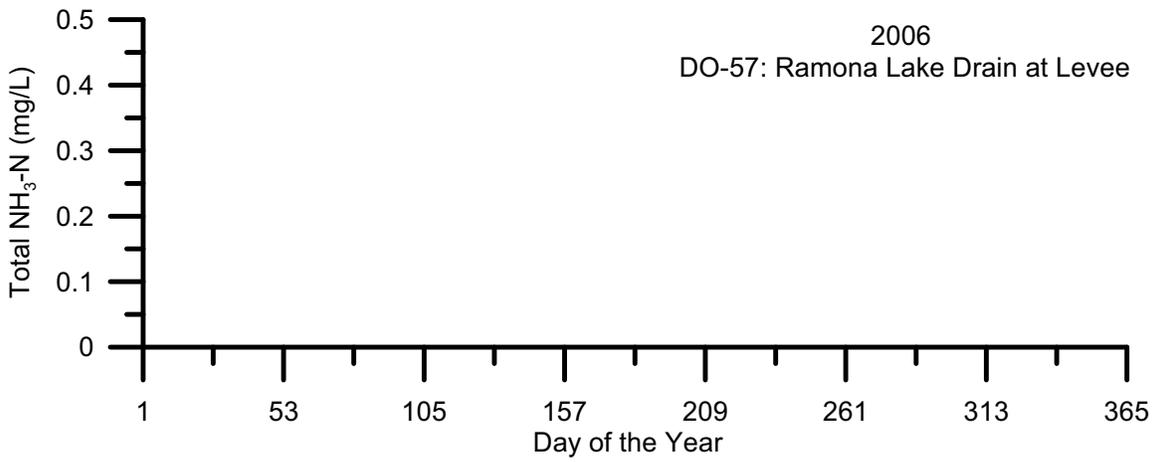
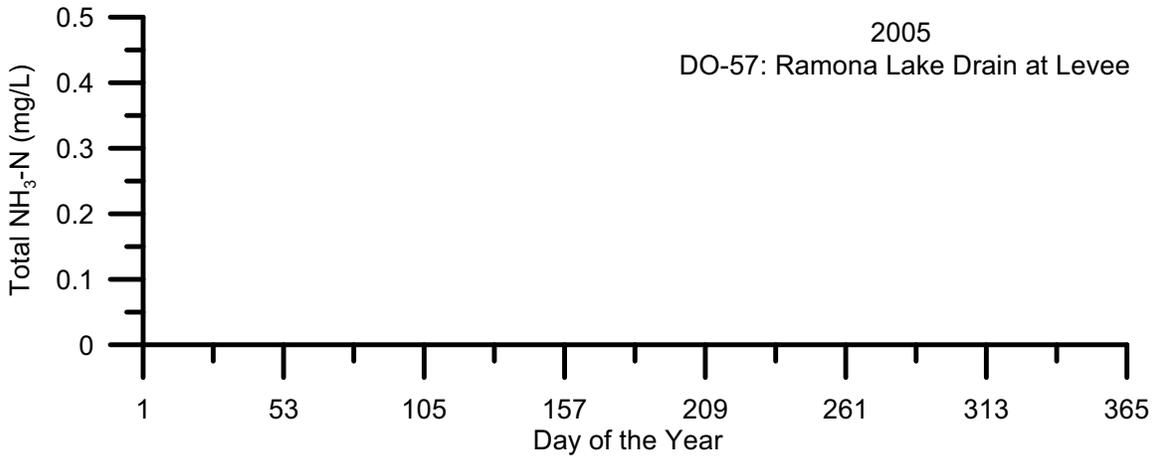


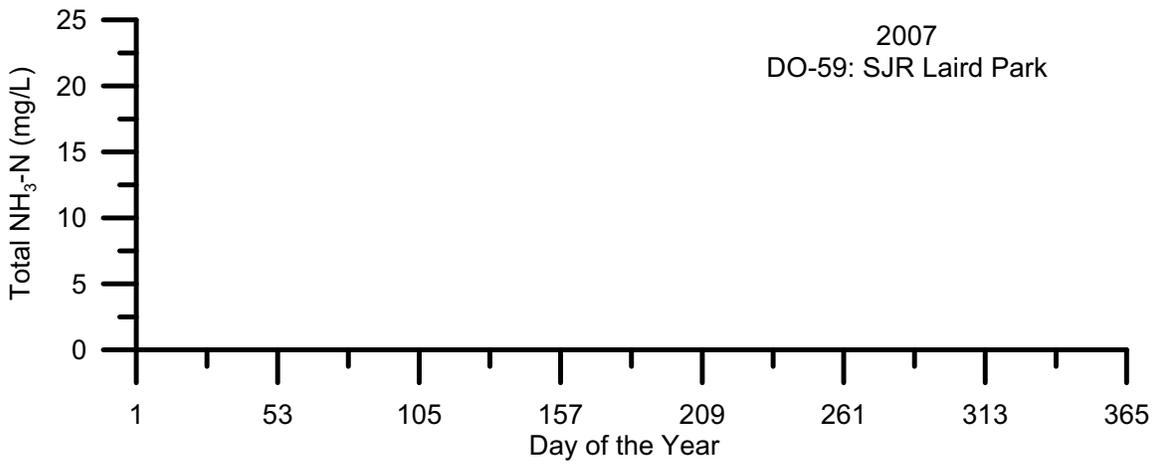
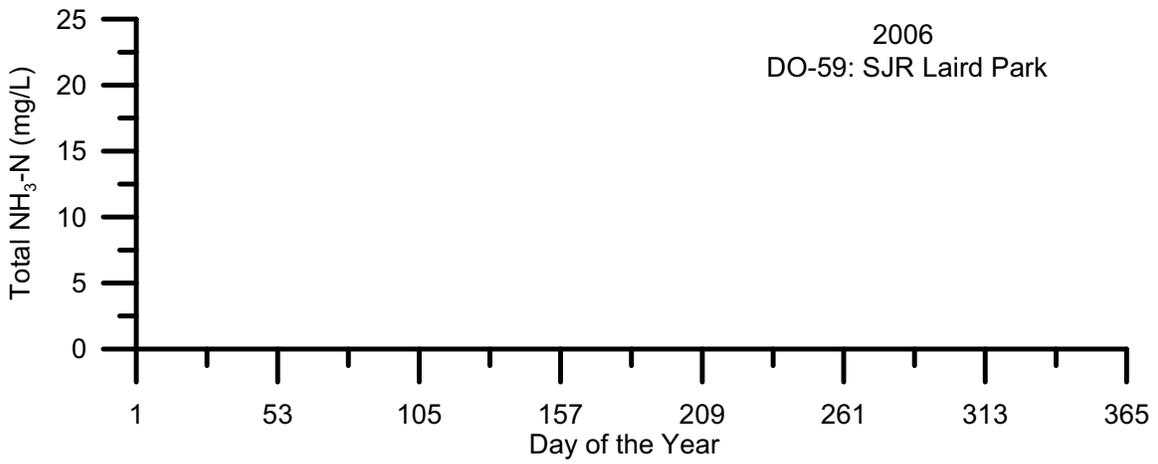
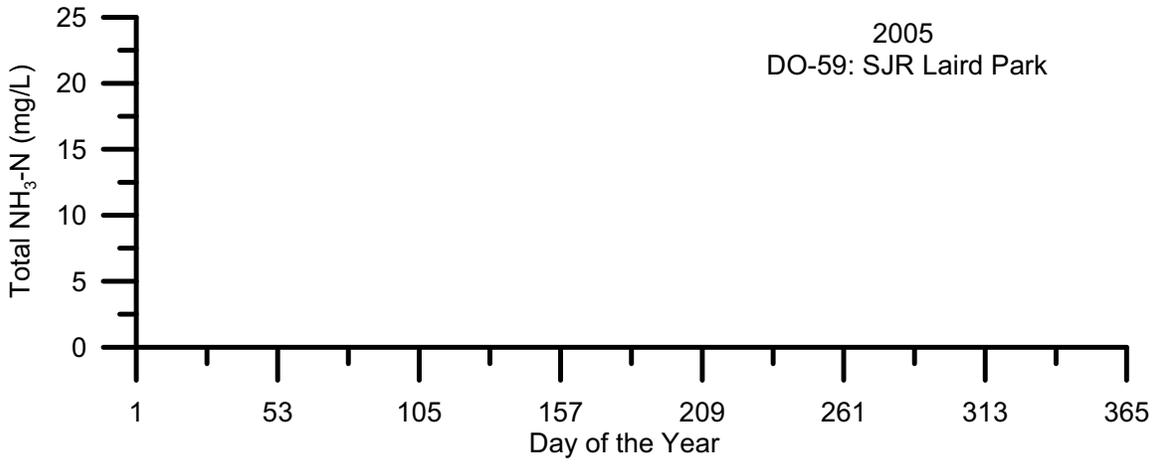


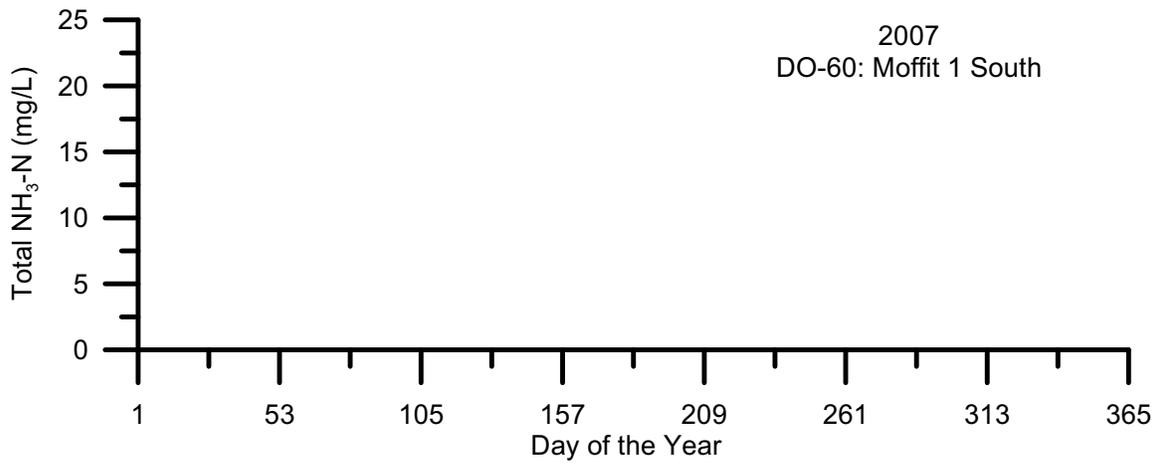
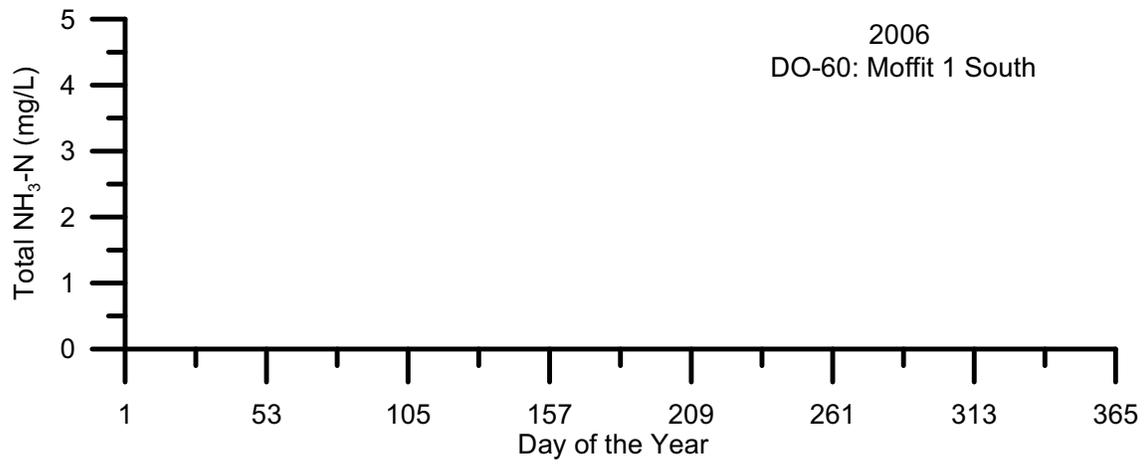
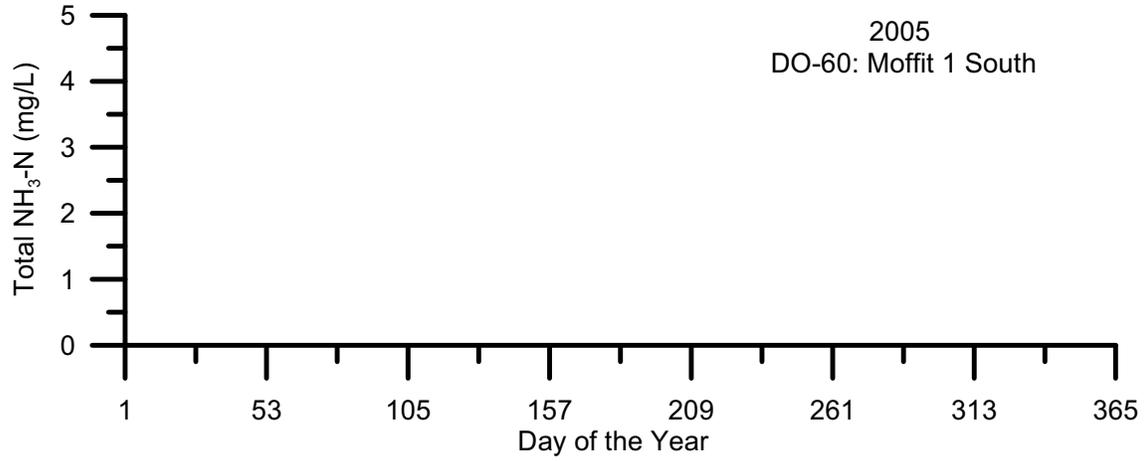


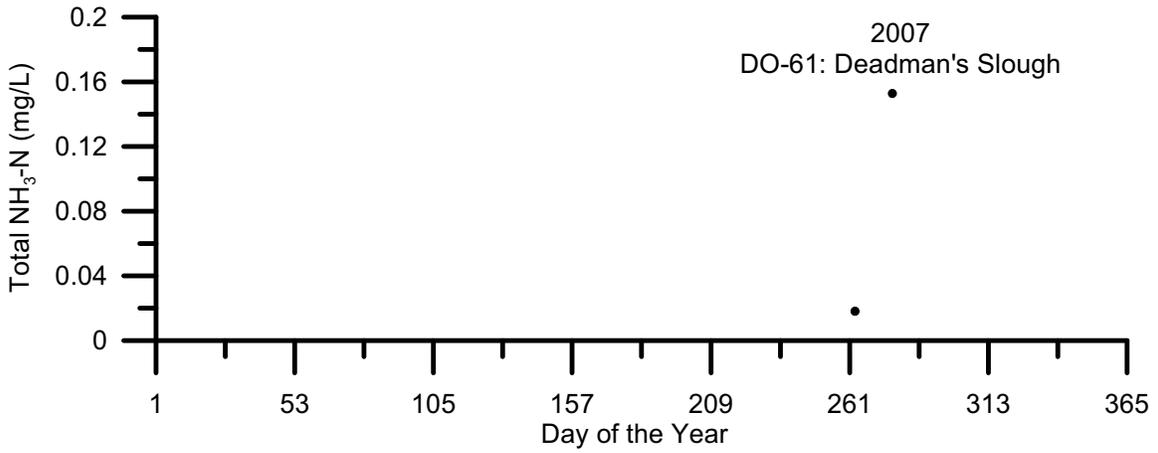
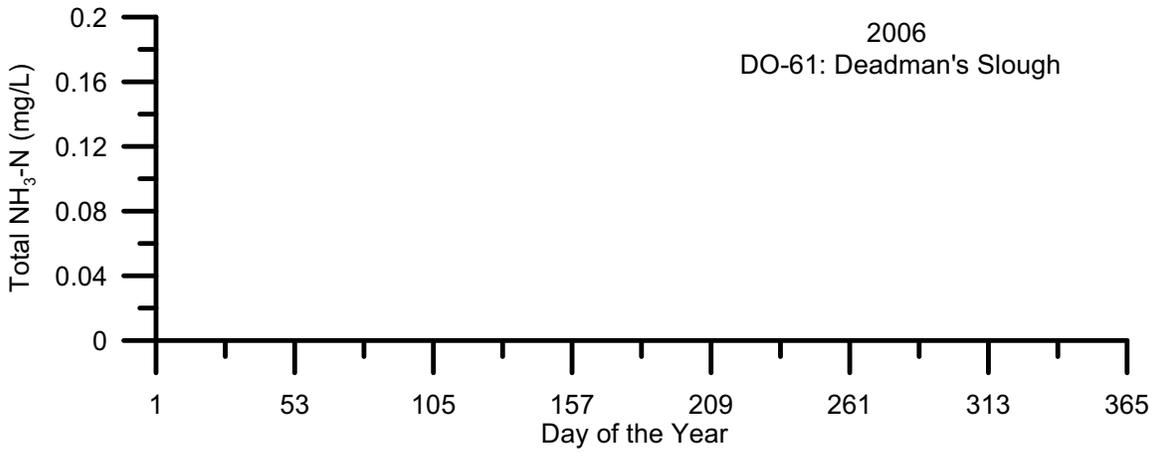
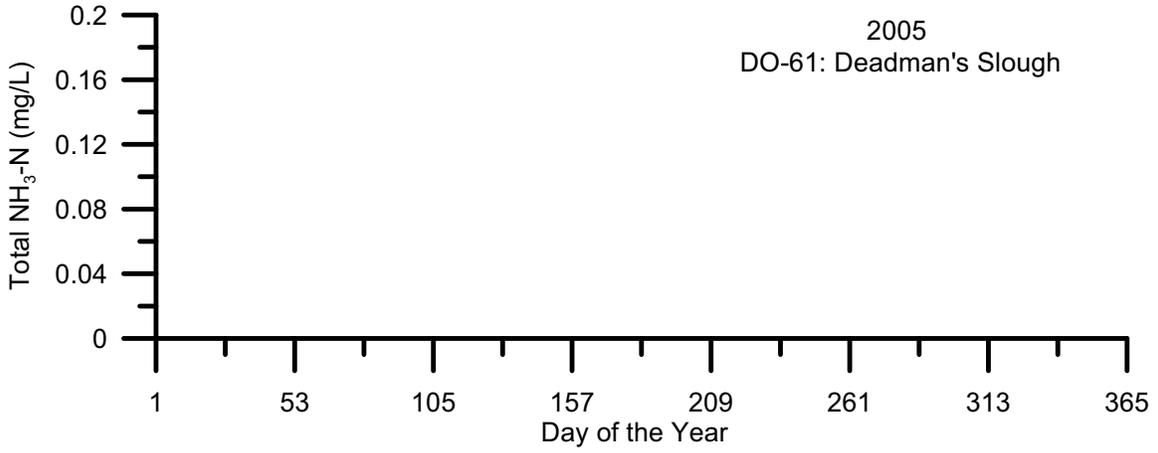


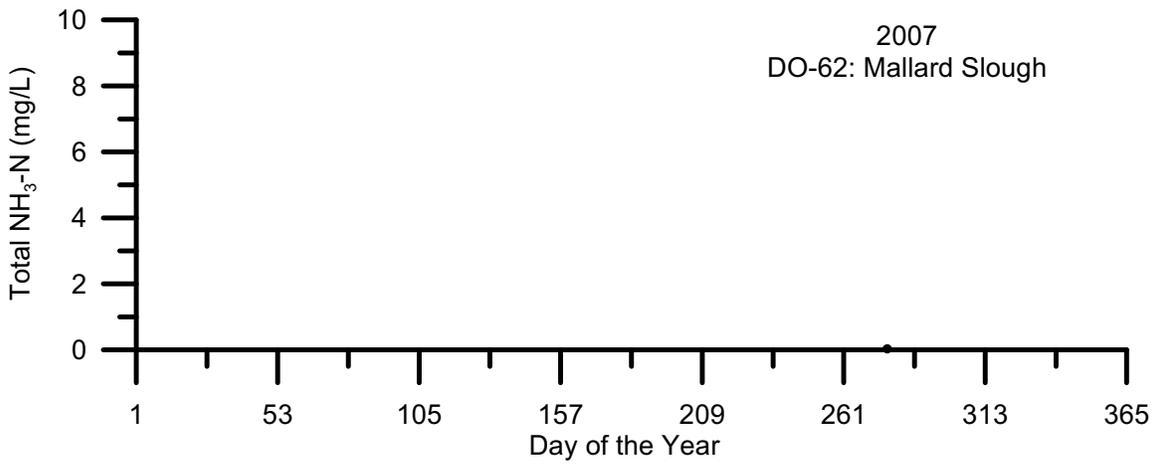
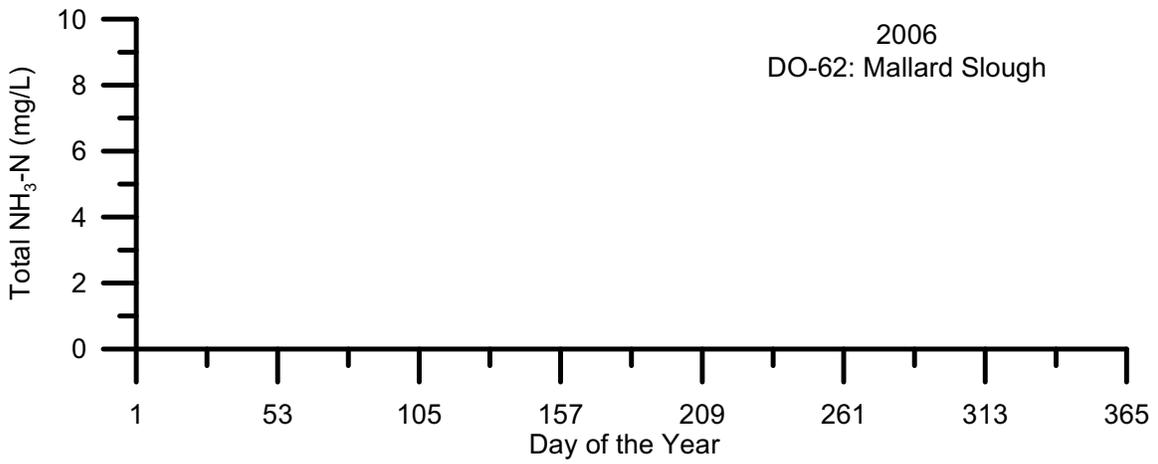
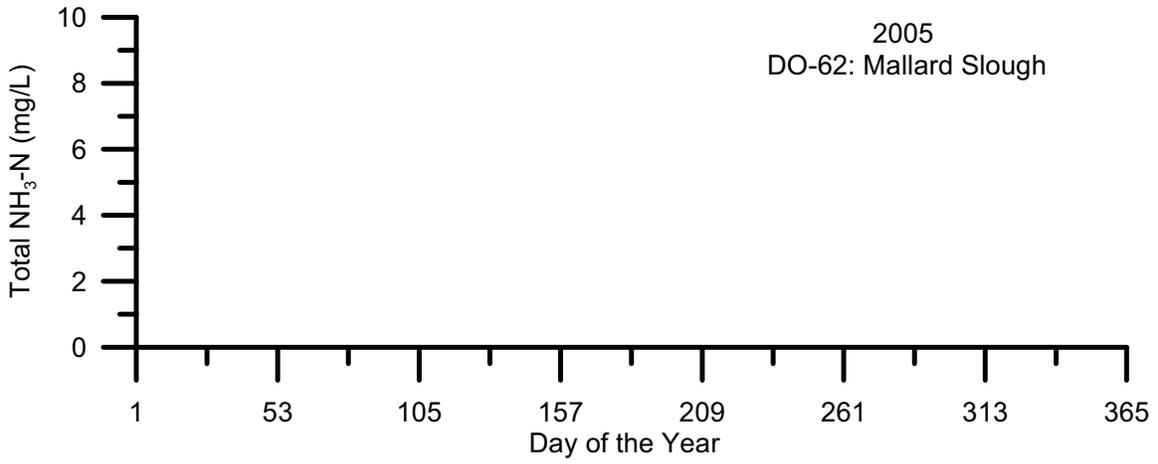


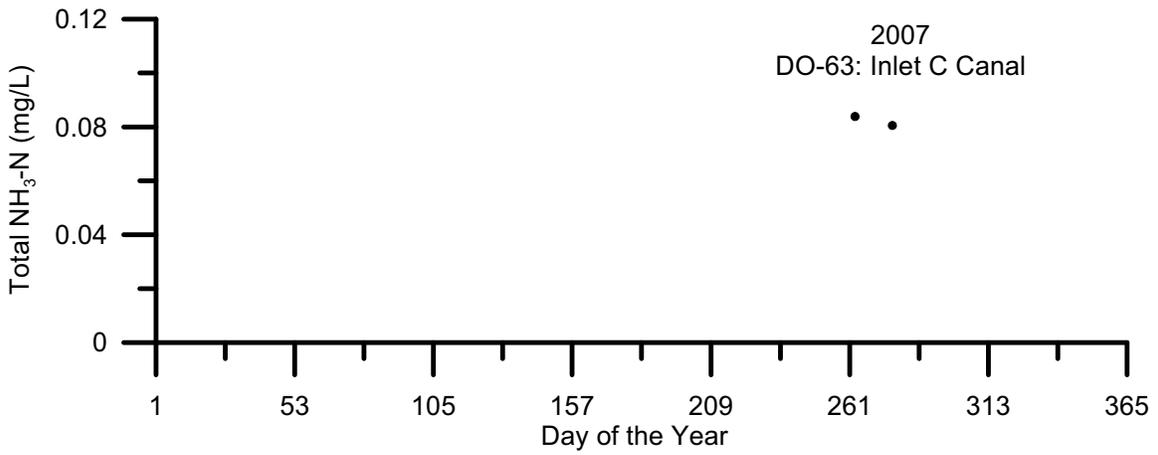
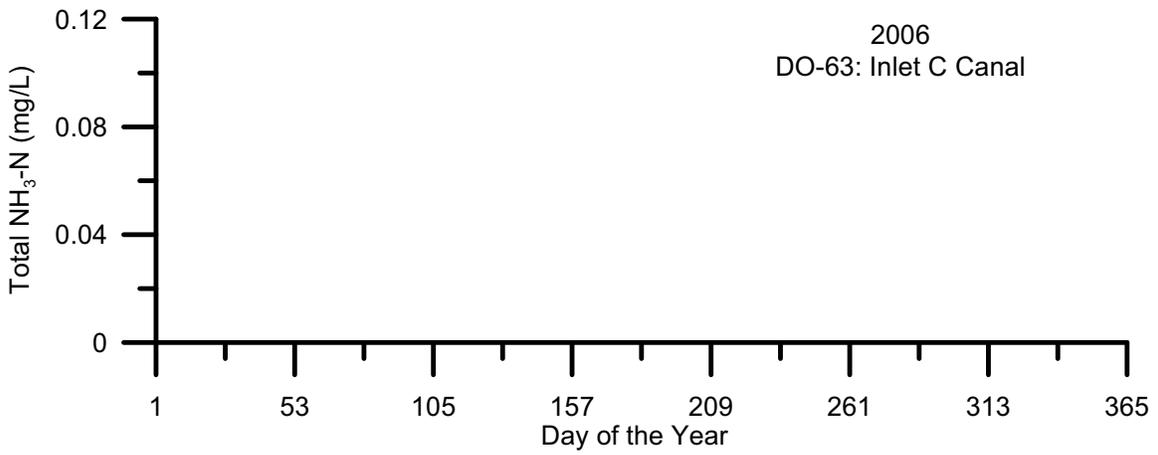
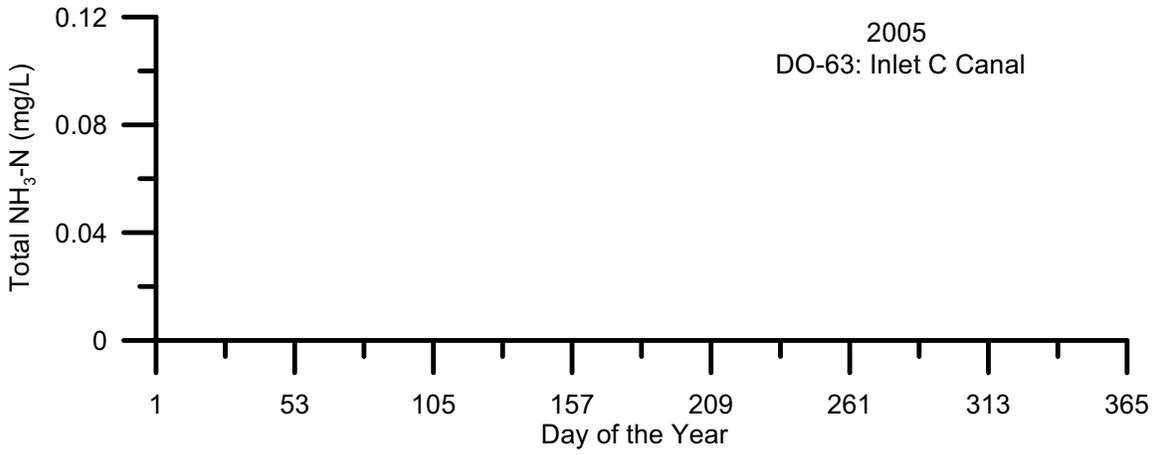


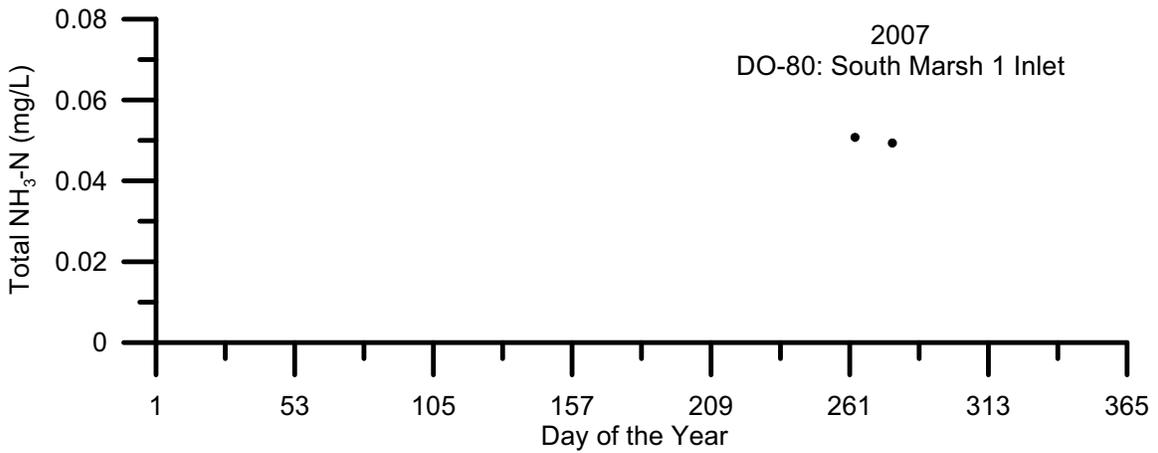
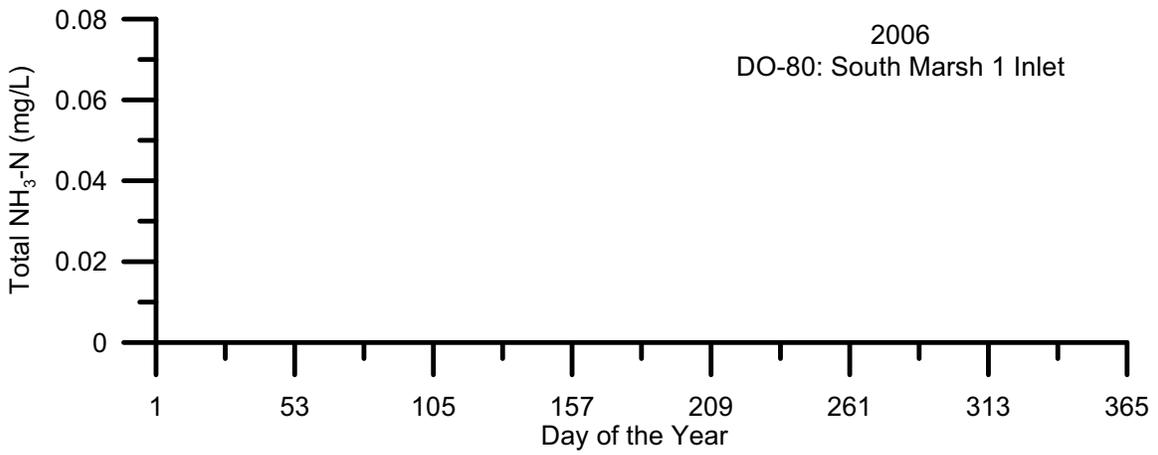
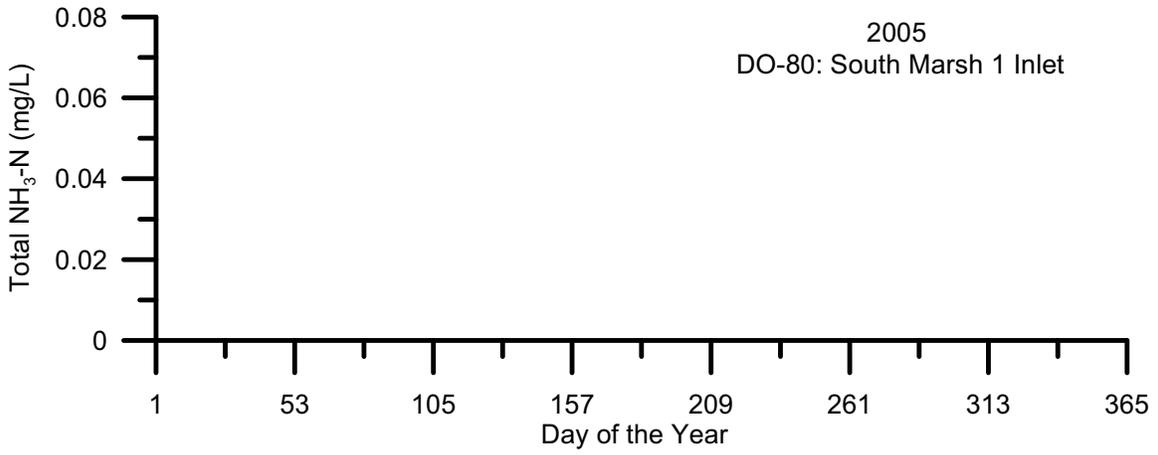


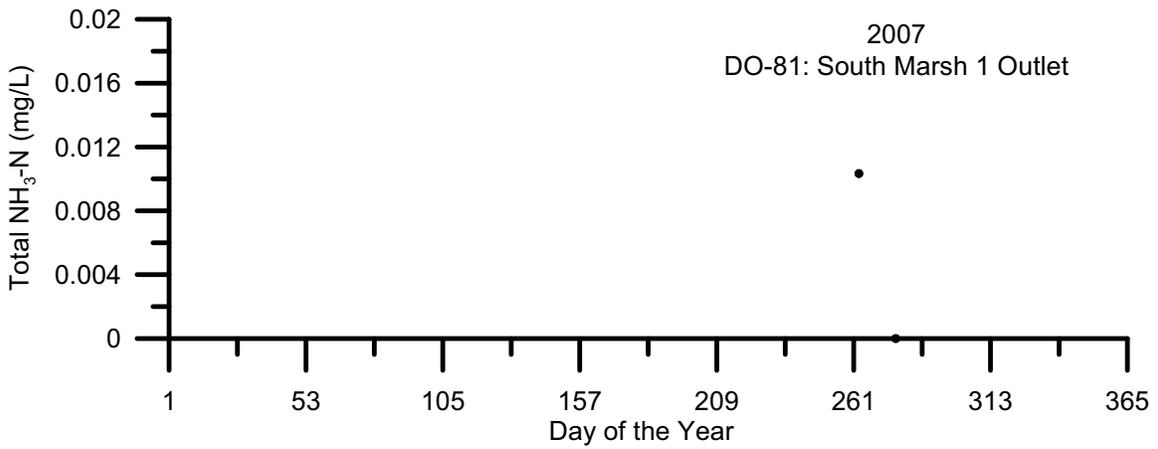
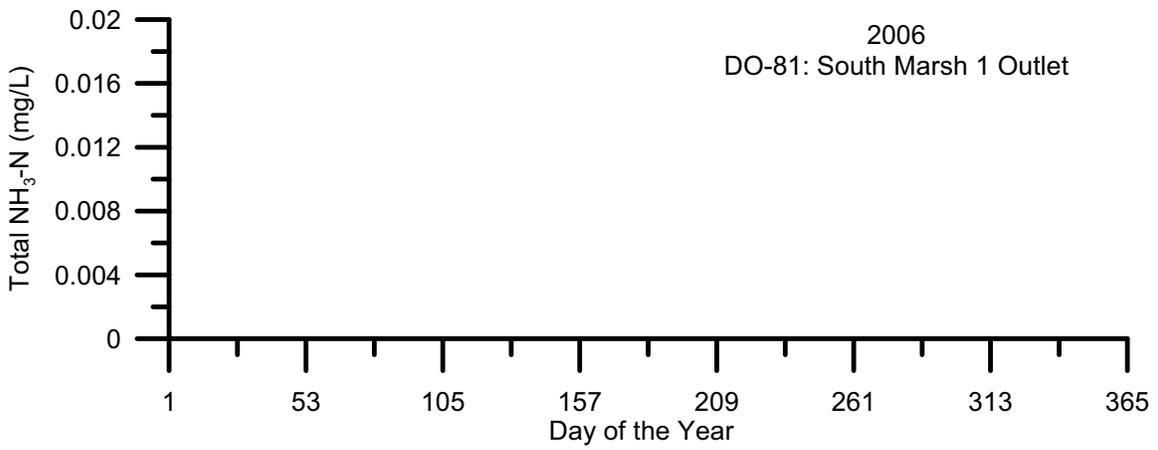
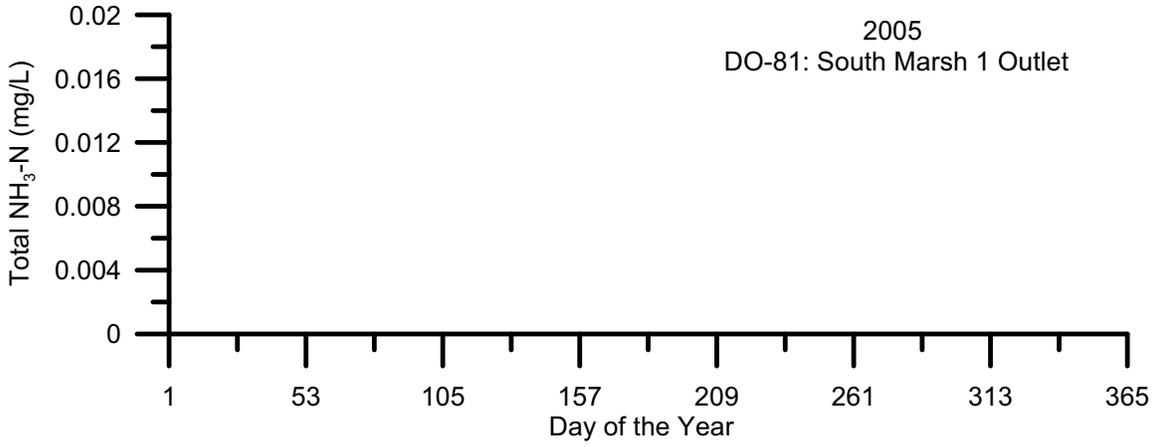


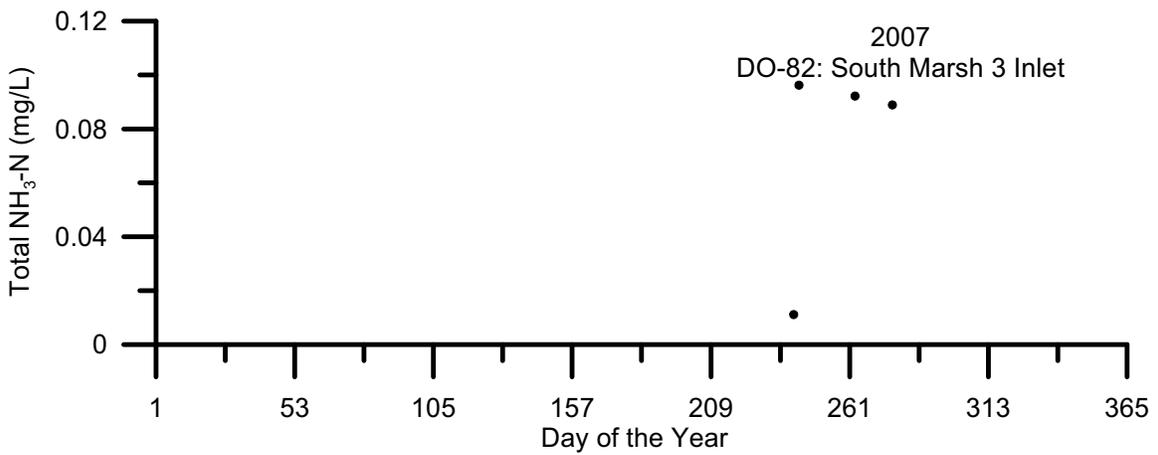
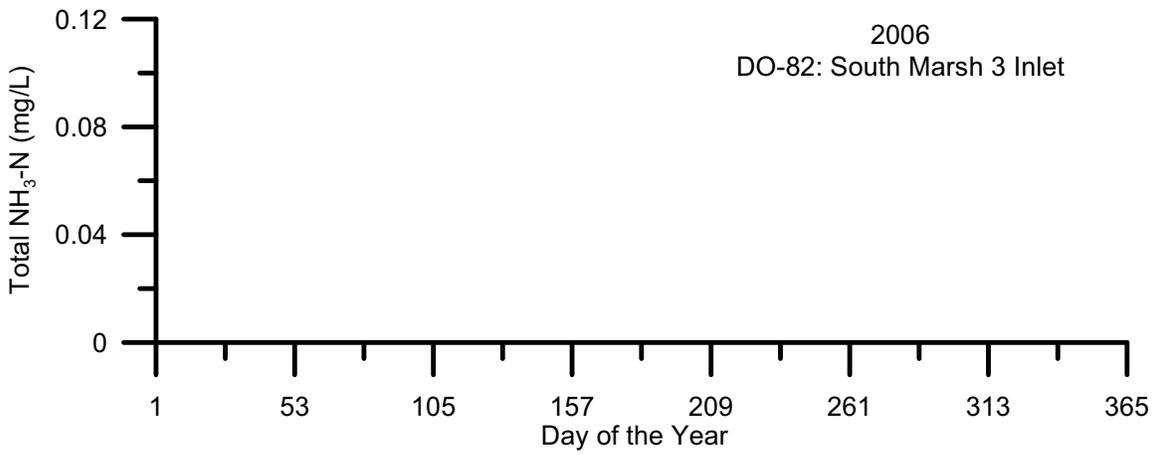
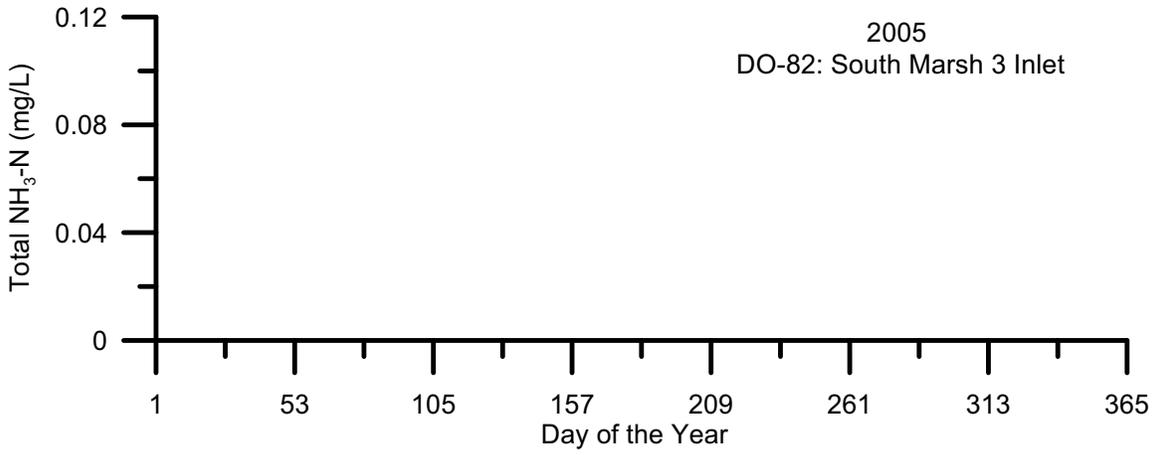


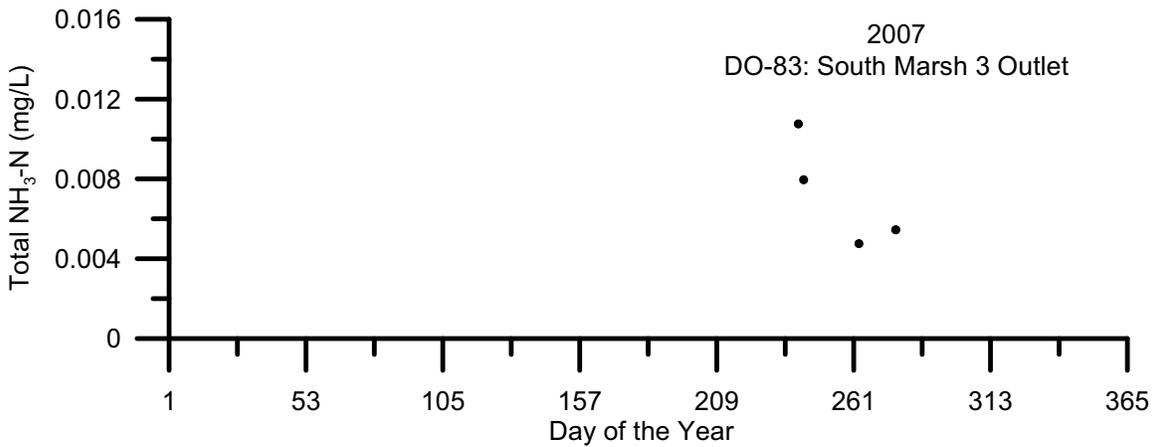
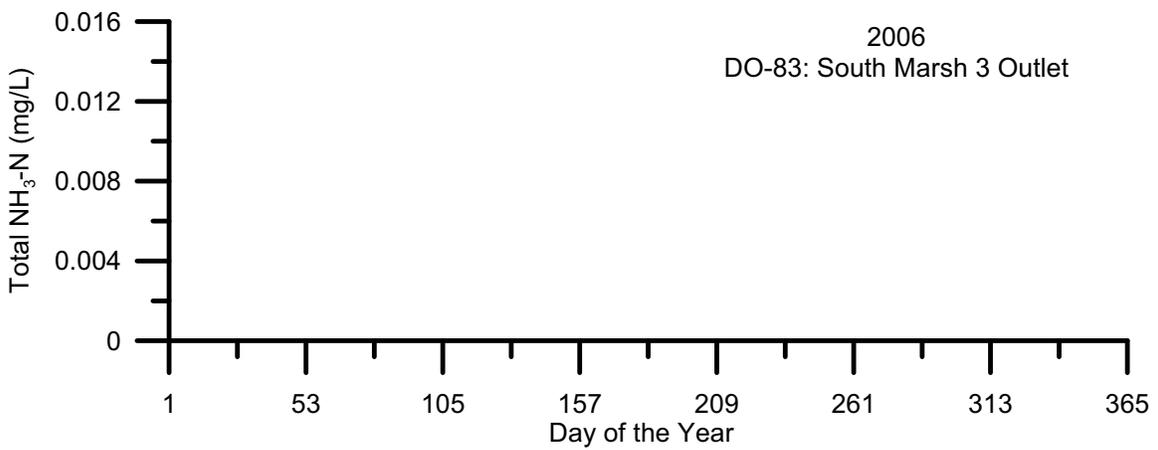
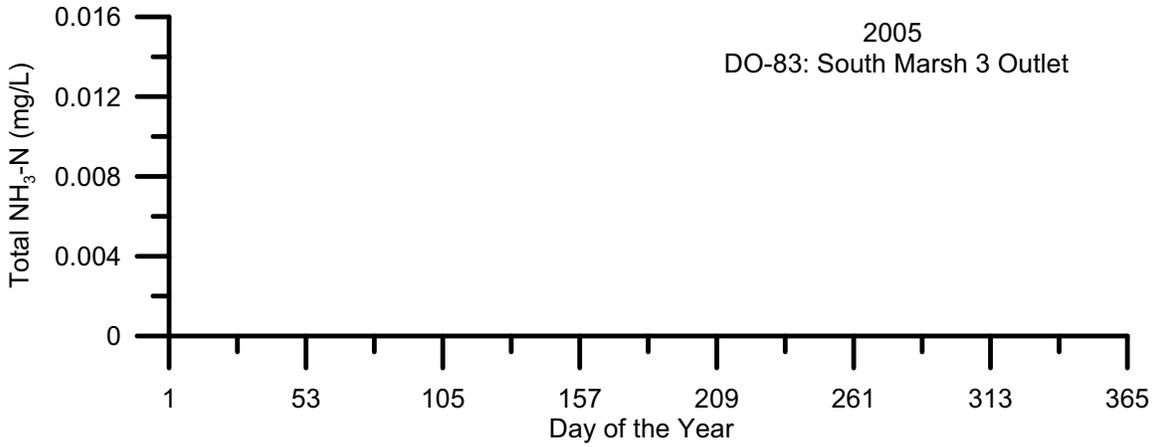


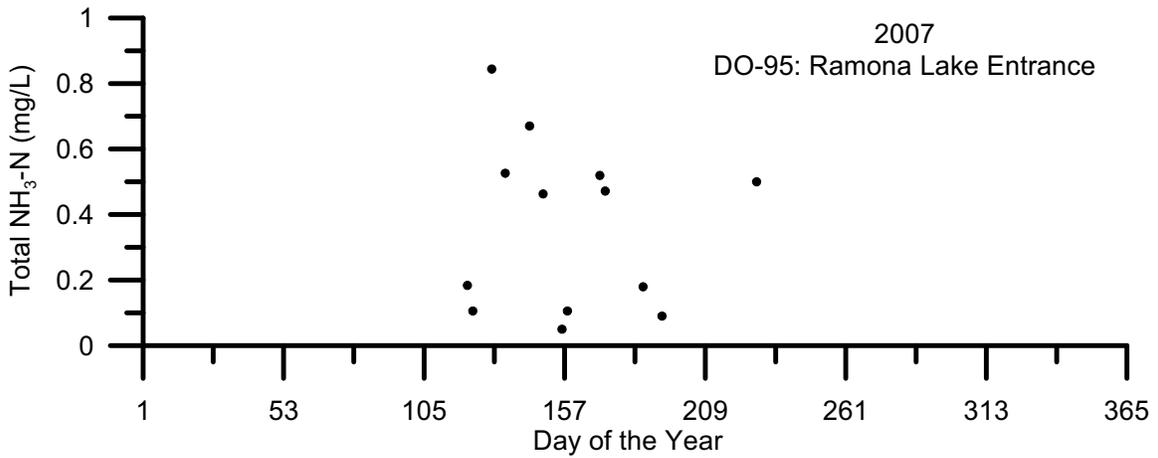
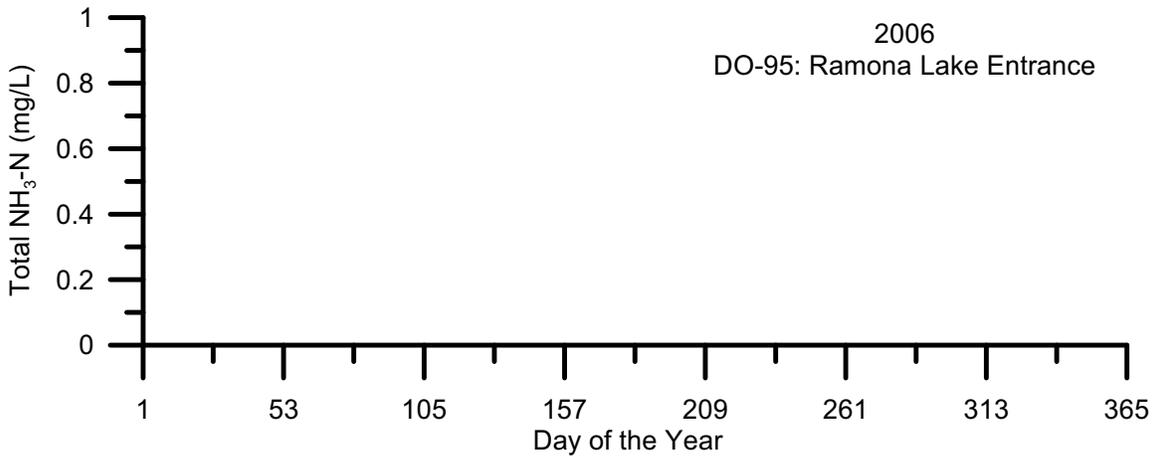
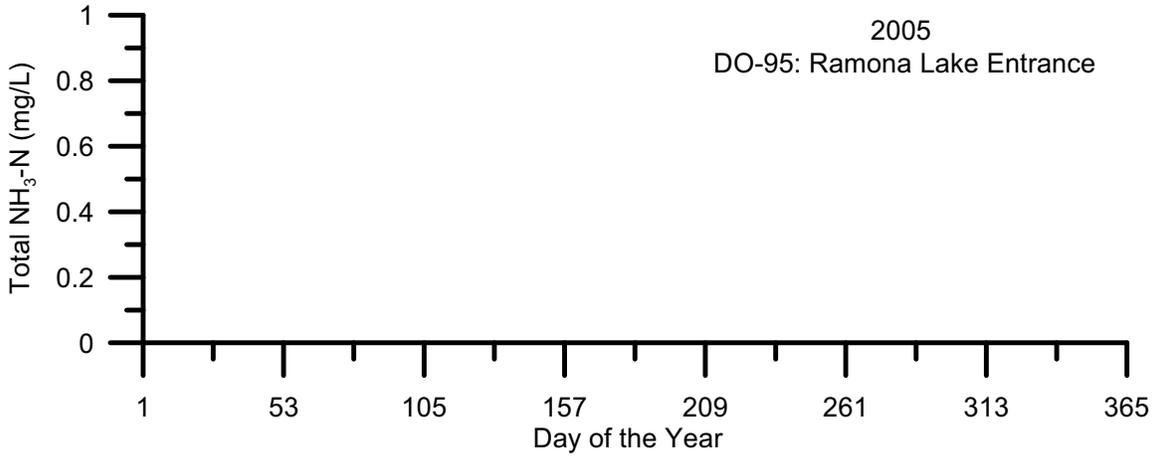














**Analysis of Volatile Suspended Solid
Concentrations in the San Joaquin River
Watershed
2005-2007**

*Remie Burks
Chelsea Spier
Sharon Borglin
Jeremy Hanlon
Justin Graham
William Stringfellow*

February 2008

*Environmental Engineering Research Program
University of the Pacific
3601 Pacific Avenue, Sears Hall
Stockton CA 95211*

Introduction

The San Joaquin River (SJR) supports one of the most productive agricultural regions in the world and its productivity is heavily dependant on irrigated agriculture. A consequence of irrigated agriculture is the production of return flows conveyed down gradient drains that eventually discharge to surface waters. Agricultural drainage may have significant nutrient load and can impact algae growth and general water quality in the SJR. Individual farmers and agricultural organizations, such as drainage authorities, are in need of tools to manage the environmental impacts of agricultural activities (Stringfellow, 2008).

For the years 2005 through 2007, sites throughout the San Joaquin Valley watershed were sampled to assess the overall water quality in the region. One thousand nine hundred and ninety-six (1996) individual surface water samples were collected and analyzed and WQ was assessed at 113 sampling locations in the SJR basin (Borglin et al., 2008). Samples were processed and analyzed by the Environmental Engineering Research Program (EERP) laboratory at the University of the Pacific as well as at the University of California, Davis, Dahlgren Lab. This report presents temporal plots of volatile suspended solids (VSS) for all sites sampled in the SJR from 2005-2007.

Methods

Field sampling consisted of collecting water samples, measuring water quality with a YSI Sonde 6600 with MDS650 hand-held display, and recording of field conditions at sites within the study area per *EERP Field Protocol Book* (Graham, 2008).

Water samples were collected in glass 1000 mL bottles (Wheaton Science Products, Millville, NJ), 1000 mL HDPE Trace-Clean narrow mouth plastic bottles (VWR International), 250 mL HDPE Trace-Clean wide mouth plastic bottles (VWR International) as well as 40 mL trace clean vials with PTFE septa (ICChem, Rockwood, TN) in accordance with requirements for different lab analysis and volume requirements.

Samples were immediately stored at 4°C after sampling (cooler temperature was recorded in the lab upon delivery) and transported to the lab on the day of sampling.

Samples were received by the laboratory the same day they were sampled and stored at 4°C until filtering and analysis. Samples were collected, preserved, stored, and analyzed by methods outlined in *Standard Methods for the Analysis of Water and Wastewater*, (APHA, 2005, 1998). Total suspended solids (TSS) and volatile suspended solids (VSS) were analyzed by SM 2540 D and E (APHA, 2005). Typically 1000 mL of sample was filtered on pre-weighed, pre-combusted, Whatman GF/F filters. The filters were placed in an aluminum dish and dried at 105°C under vacuum to constant weight. After drying the filter and dish were allowed to cool in a dessicator and were weighed for TSS determination. The dried and weighted filters were subsequently combusted at 550°C for 6 hours and reweighed for VSS determination. Mineral suspended solids (MSS) concentration was calculated by subtracting VSS from TSS.

Results and Discussion

Volatile suspended solids (VSS) analysis was performed routinely over the years 2005-2007 with no modification to the standard method (APHA, 2005). A 94.44% passage rate for all QA parameters over the three years was observed (Borglin et. al., 2008).

References

Stringfellow, W.T., et al., (2008), *Evaluation of Vegetated Ditches, Ponds, and Wetlands as BMPs for Mitigating the Water Quality Impact of Irrigated Agriculture in the San Joaquin Valley*, University of the Pacific, Stockton, CA

Graham, J., Hanlon, J.S., Stringfellow, W.T., (2008), *EERP Field Protocol Book*, University of the Pacific, Stockton, CA.

Borglin, S., W. Stringfellow, J. Hanlon. (2005), *Standard Operating Procedures for the Up-Stream Dissolved Oxygen TMDL Project*, LBNL/Pub-937.

Borglin, S., Burks, R., Hanlon, J., Graham, J., Spier, C., Stringfellow, W., and Dahlgren, R., (2008) *Methods overview, quality assurance, and quality control*, University of the Pacific, Stockton, CA

American Public Health Association, (2005), *Standard Methods for the Examination of Water and Wastewater*, American Public Health Association, Washington, D.C.

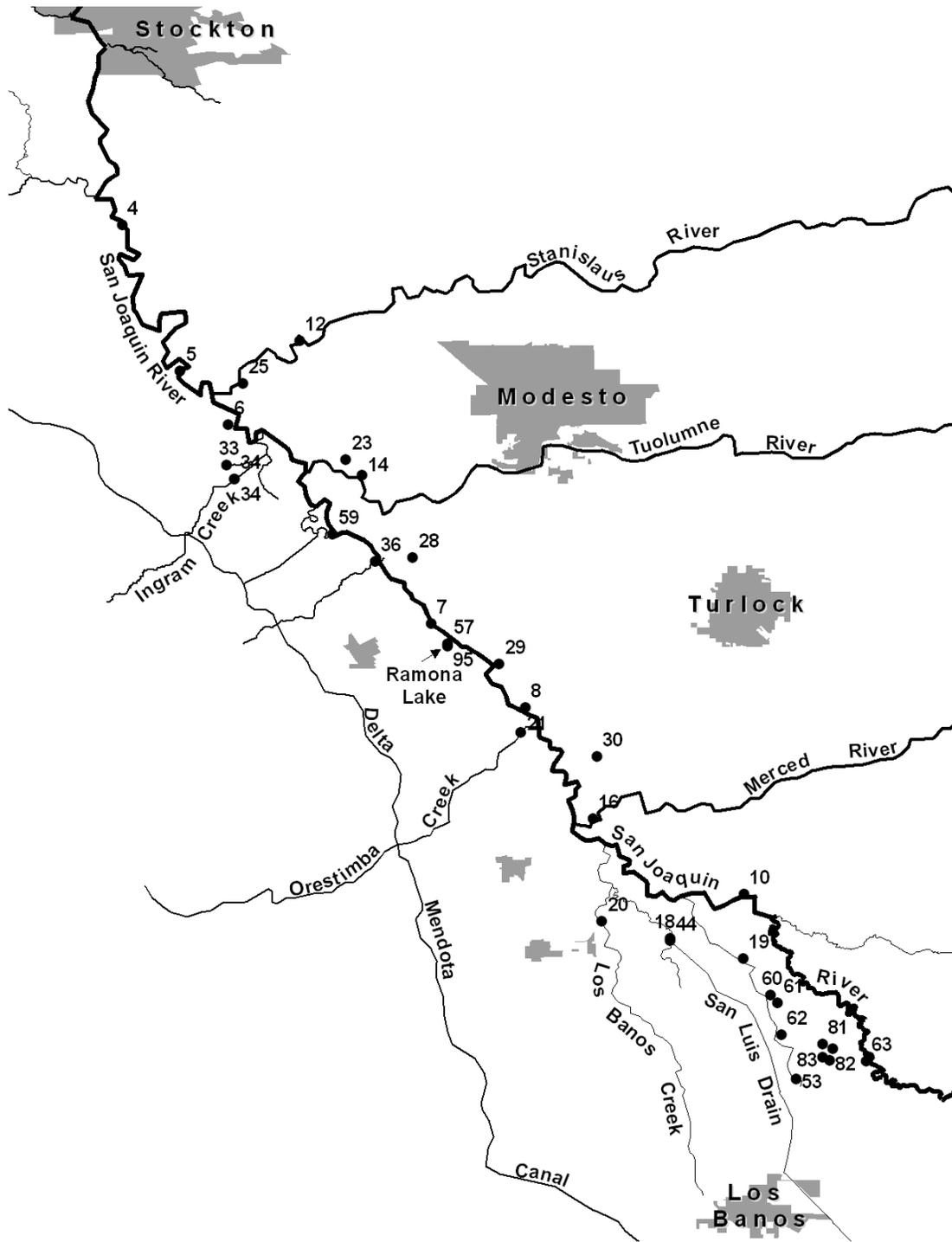
Borglin, S.E., Burks, R.D., Hanlon, J.S., Stringfellow, W.T. (2008) *EERP Lab Protocol Book*, University of the Pacific, Stockton, CA.

YSI Environmental Operations Manual, (2005), 6-Series Environmental Monitoring Systems, Yellow Springs, OH.

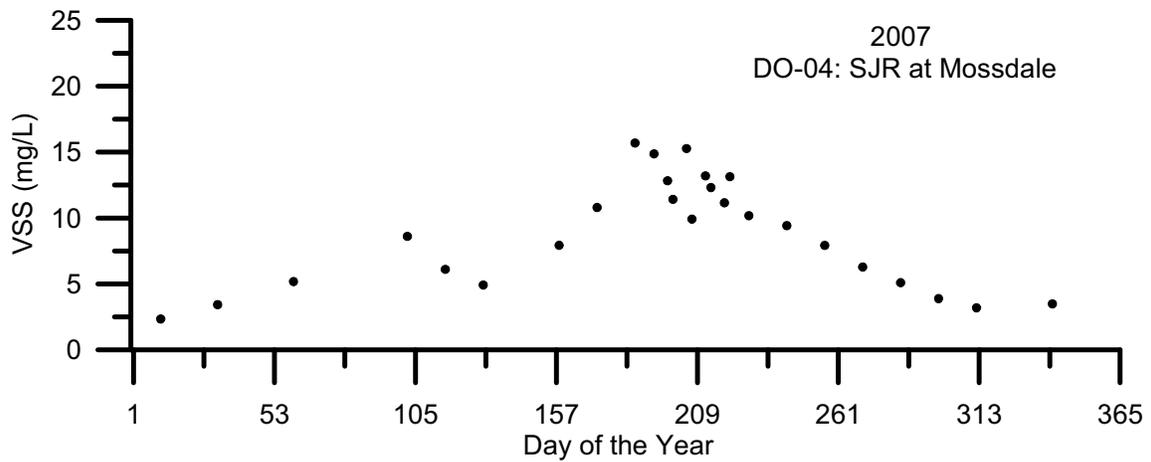
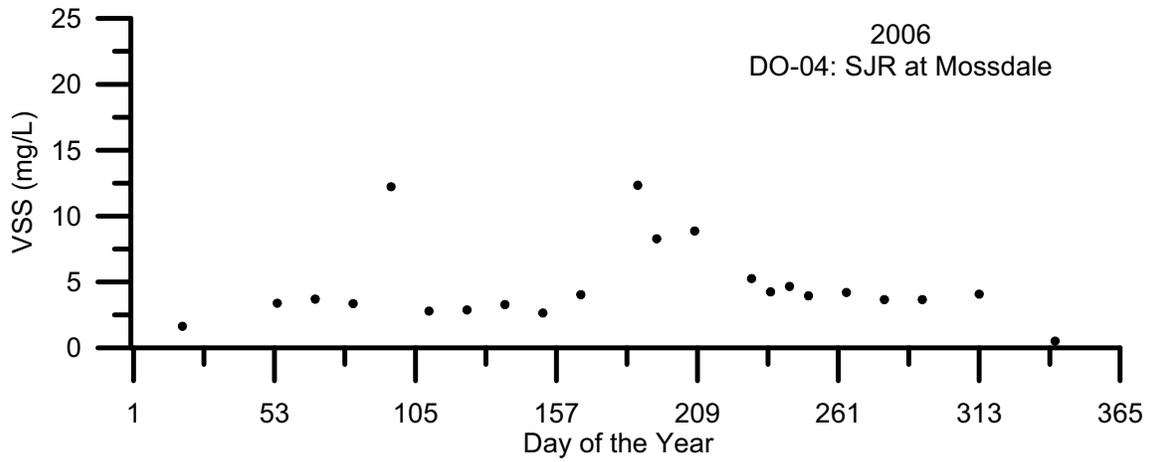
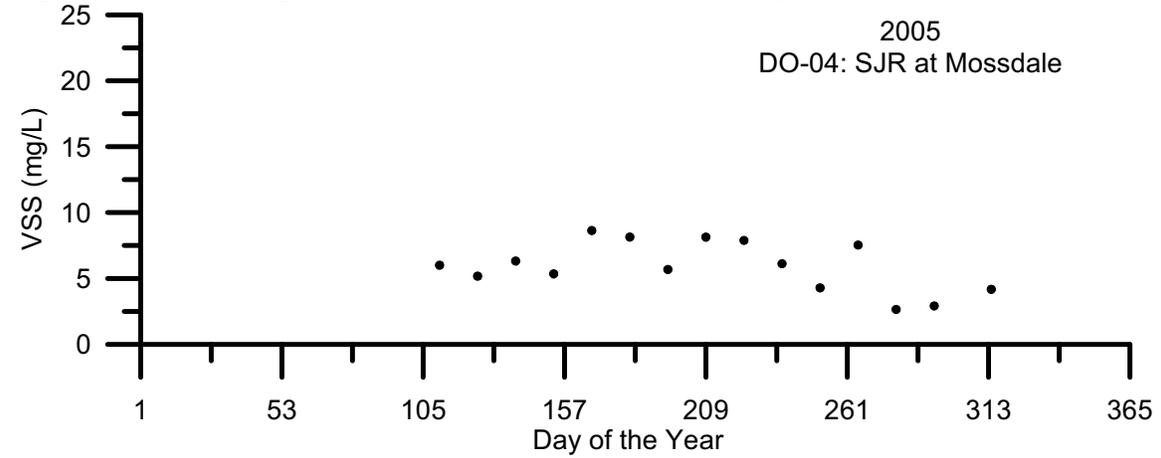
Table 1: EERP Sampling Site List

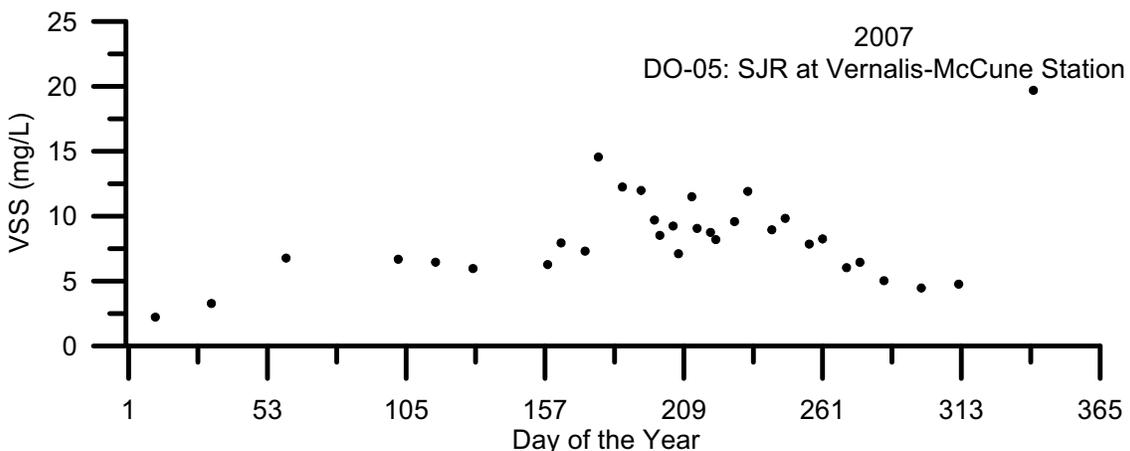
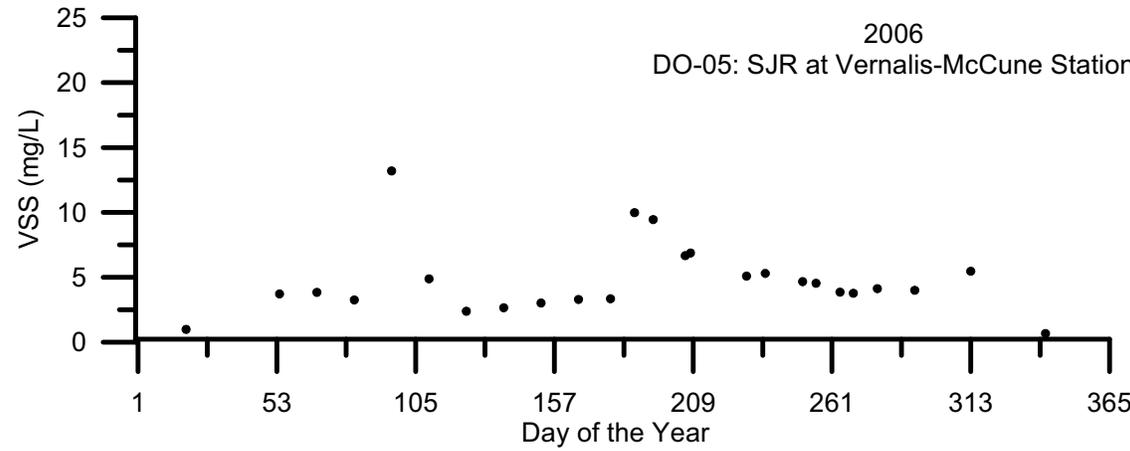
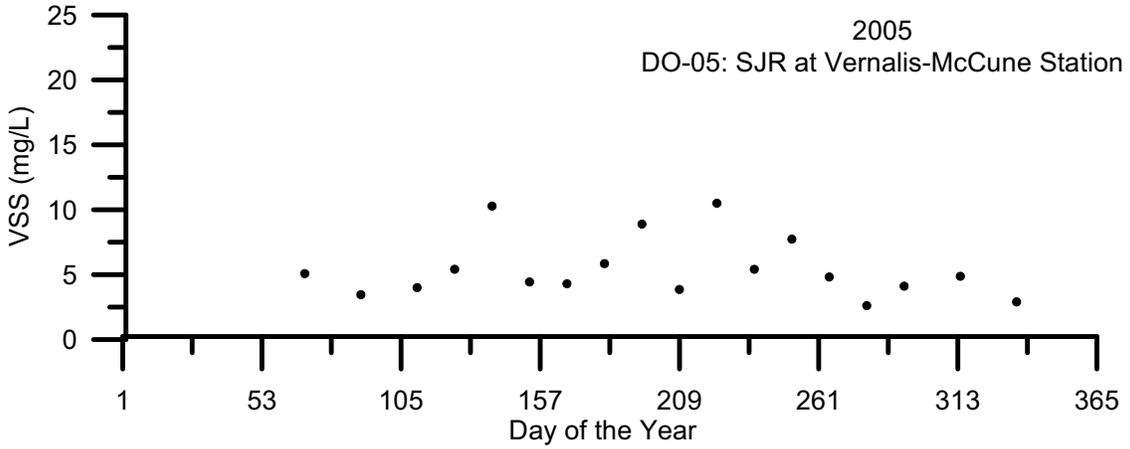
DO Number	Site Name	Type
4	SJR at Mossdale	Core sites
5	SJR at Vernalis-McCune Station (River Club)	Core sites, BMP
6	SJR at Maze	Core sites, BMP
7	SJR at Patterson	Core sites, BMP
8	SJR at Crows Landing	Core sites, BMP
10	SJR at Lander Avenue	Core sites
12	Stanislaus River at Caswell Park	Core sites
14	Tuolumne River at Shiloh Bridge	Core sites
16	Merced River at River Road	Core sites
18	Mud Slough near Gustine	Core sites, Wetland
19	Salt Slough at Lander Avenue	Core sites, Wetland
20	Los Banos Creek Flow Station	Core sites, Wetland
21	Orestimba Creek at River Road	Core sites, BMP
23	Modesto ID Lateral 5 to Tuolumne	Core sites
25	Modesto ID Main Drain to Stan. R. via Miller Lake	Core sites
28	Turlock ID Westport Drain Flow station	Core sites
29	Turlock ID Harding Drain	Core sites
30	Turlock ID Lateral 6 & 7 at Levee	Core sites
33	Hospital Creek	Intermittent, BMP
34	Ingram Creek	Core sites, BMP
36	Del Puerto Creek Flow Station	Core sites, BMP
44	San Luis Drain End	Core sites
53	Salt Slough at Wolfsen Road	Wetland
57	Ramona Lake Drain	Core sites, BMP
59	SJR Laird Park	Core sites
60	Moffit 1 South	Wetland
61	Deadman's Slough	Wetland
62	Mallard Slough	Wetland
63	Inlet C Canal	Wetland
80	South Marsh-1-Inlet	Wetland
81	South Marsh-1-Outlet	Wetland
82	South Marsh-3-Inlet	Wetland
83	South Marsh-3-Outlet	Wetland
95	Ramona drain at Ramona Lake	BMP, Intermittent

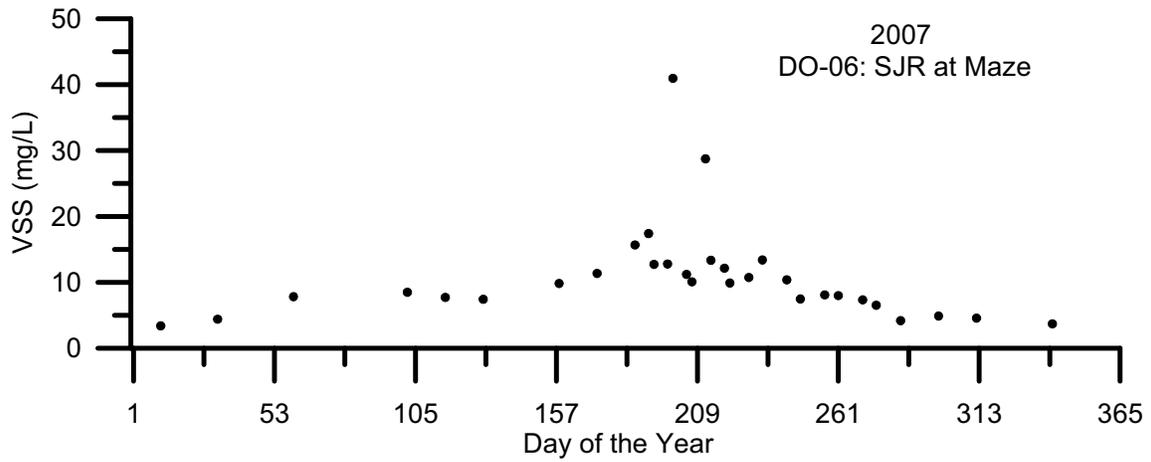
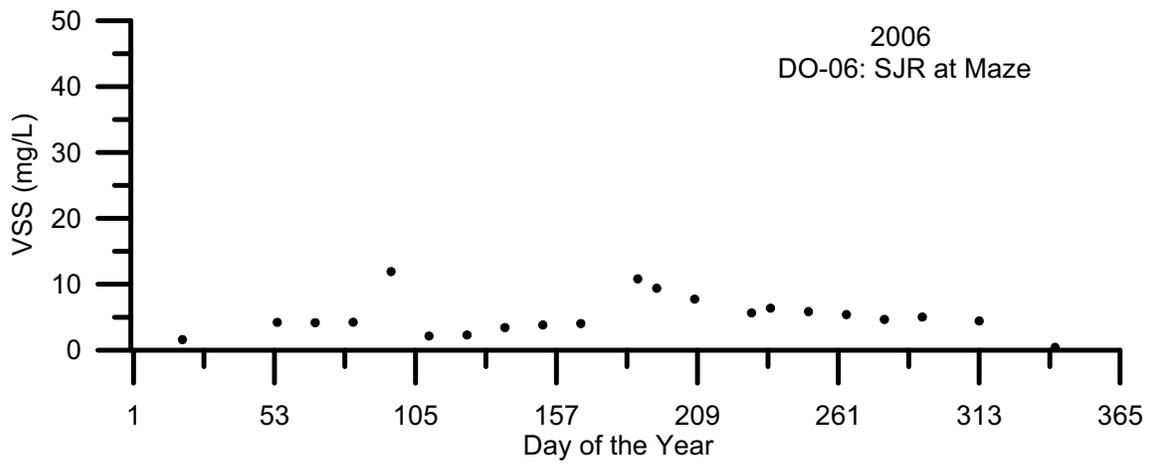
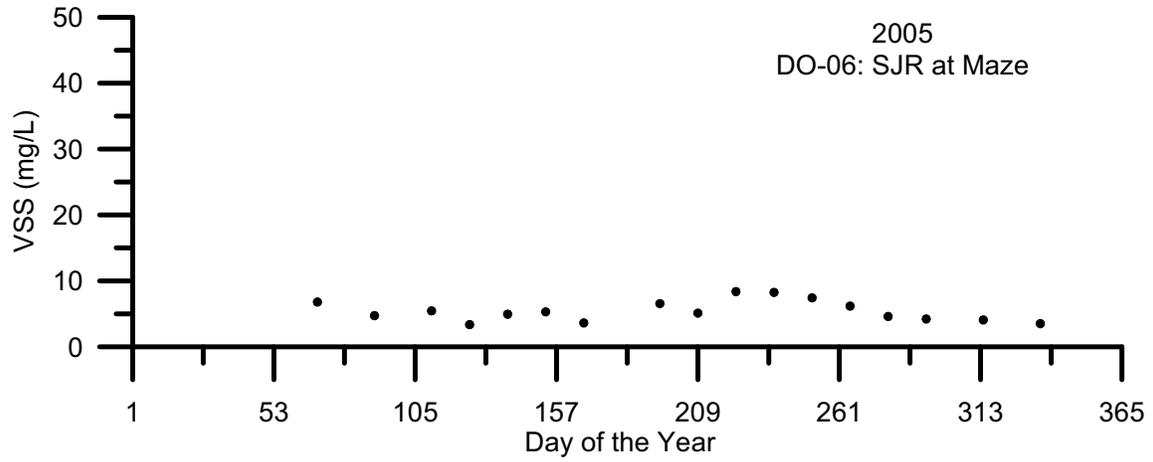
Figure 1: EERP Sampling Site Map of SJR Watershed and Tributaries

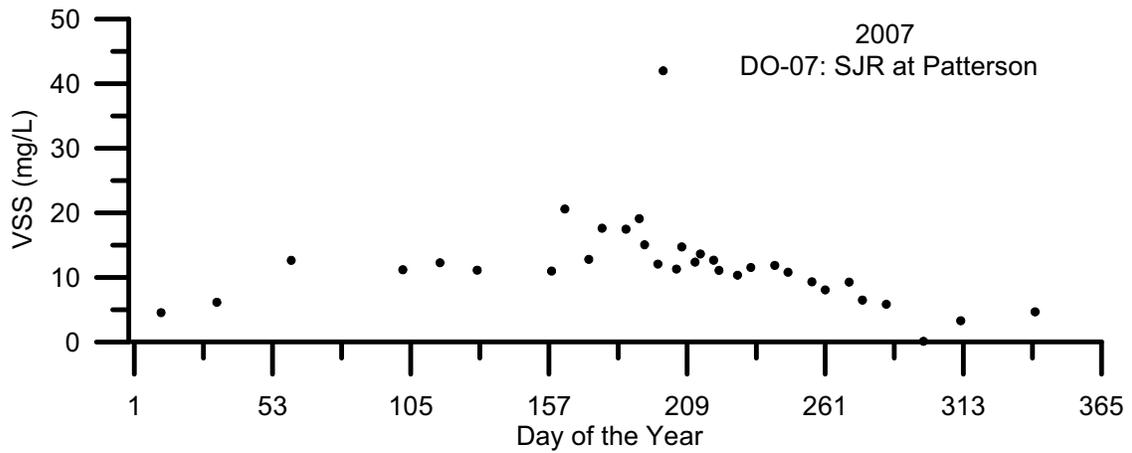
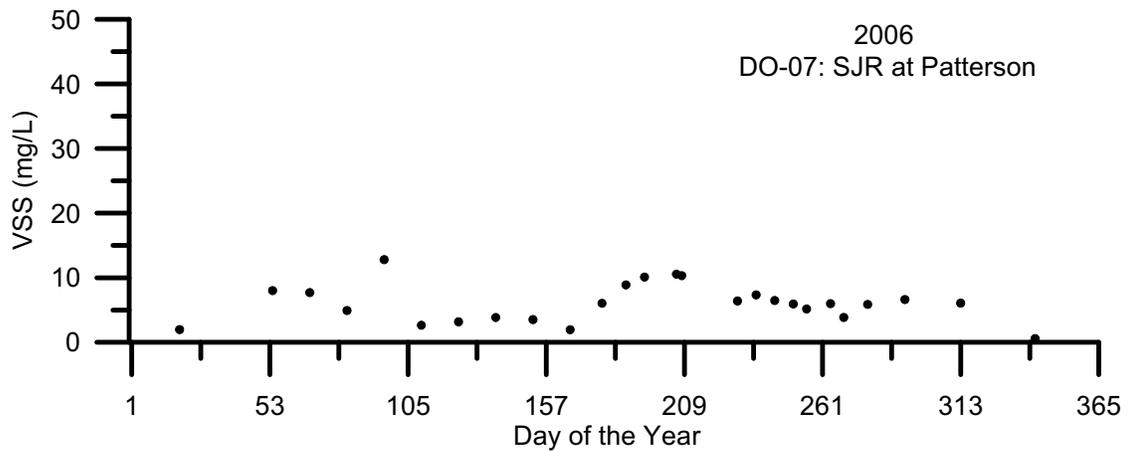
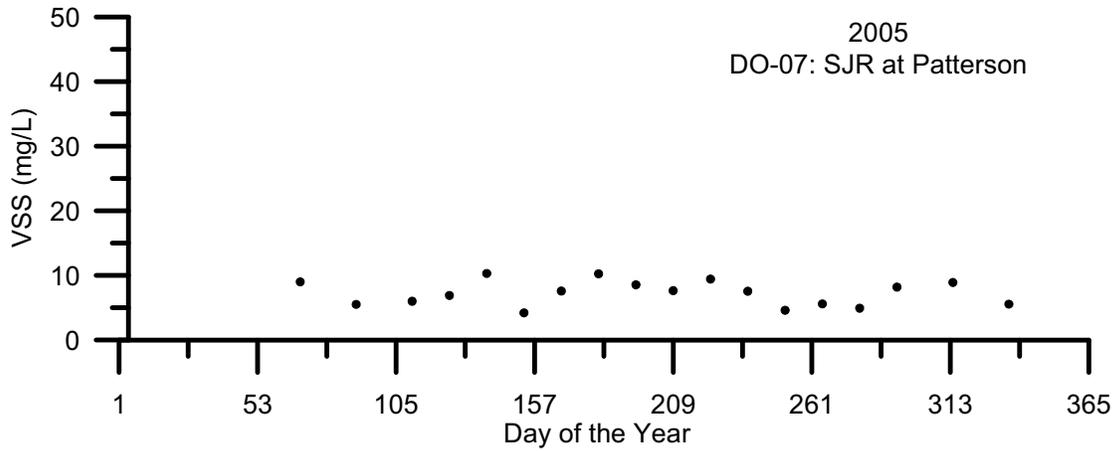


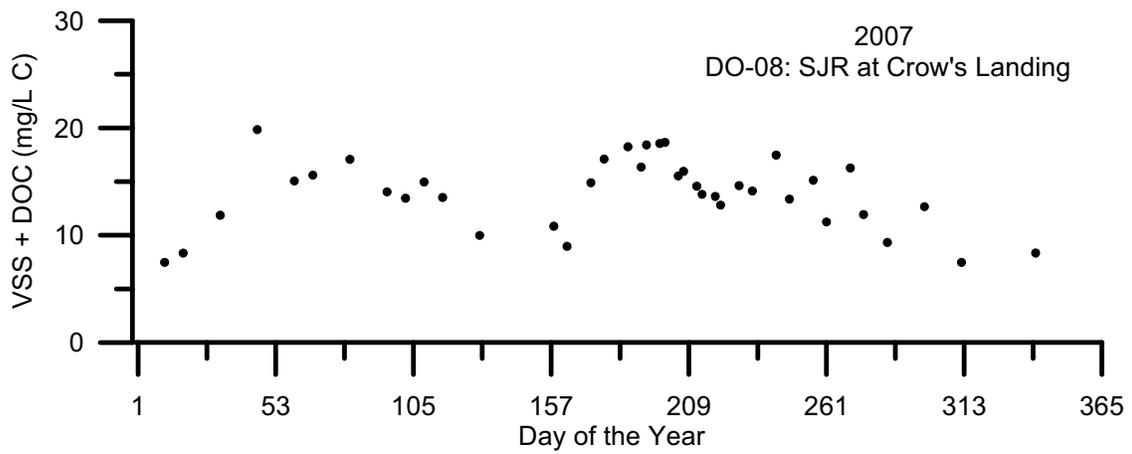
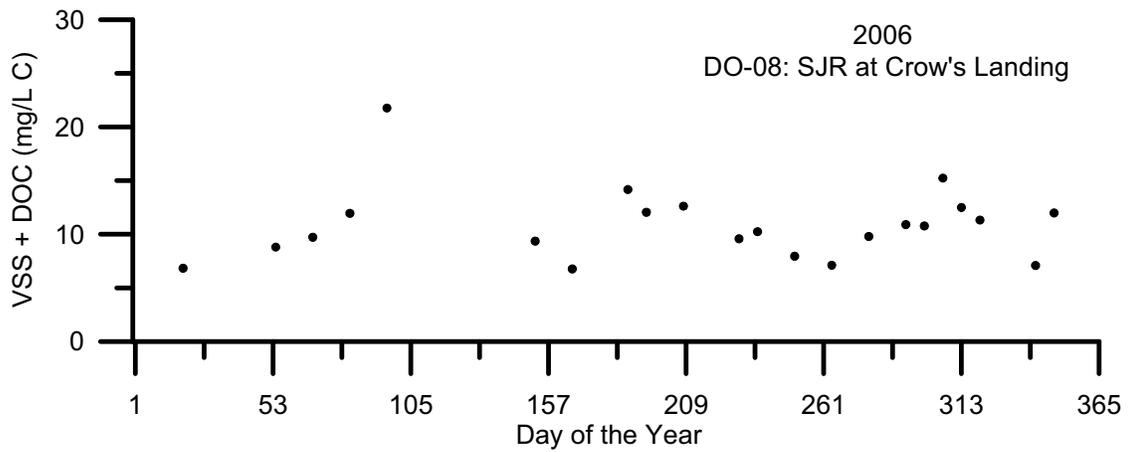
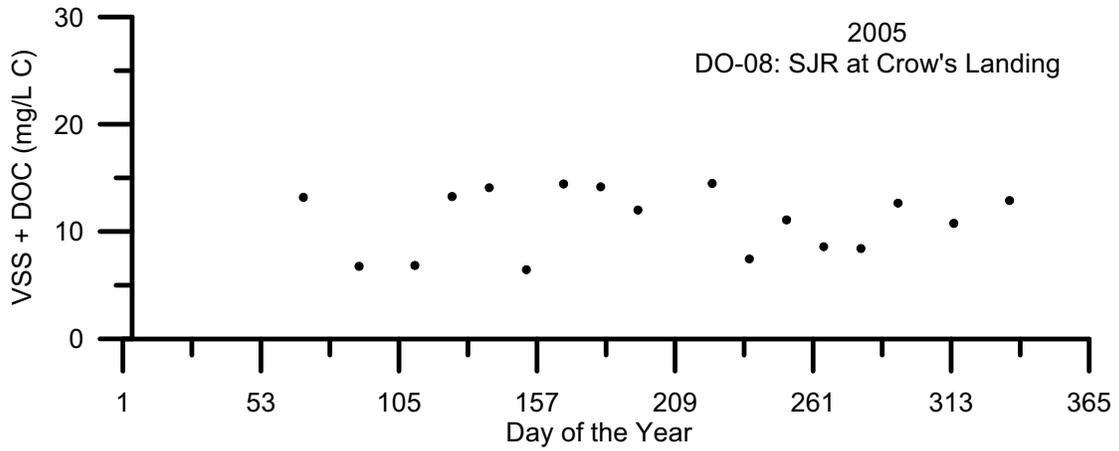
Figures 2 -101: Temporal Plots of VSS Concentrations By Site ID

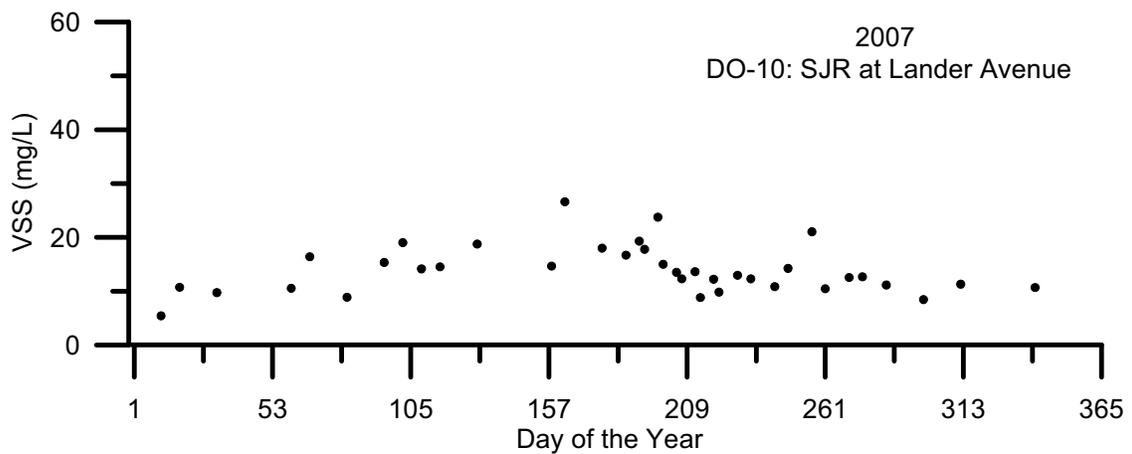
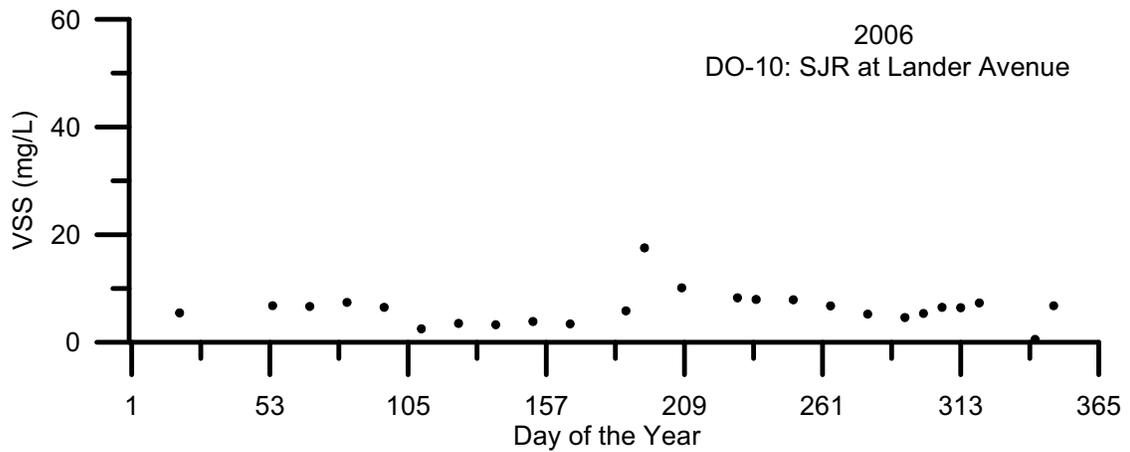
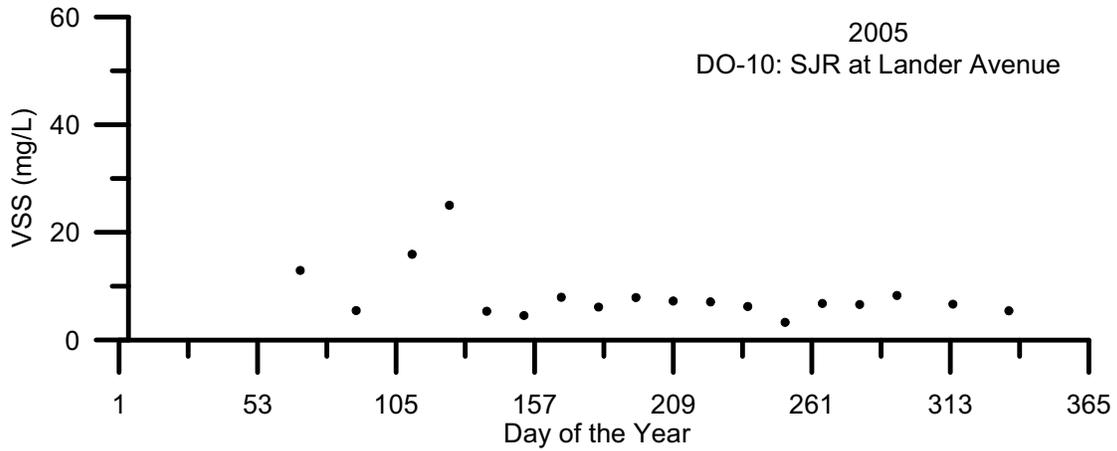


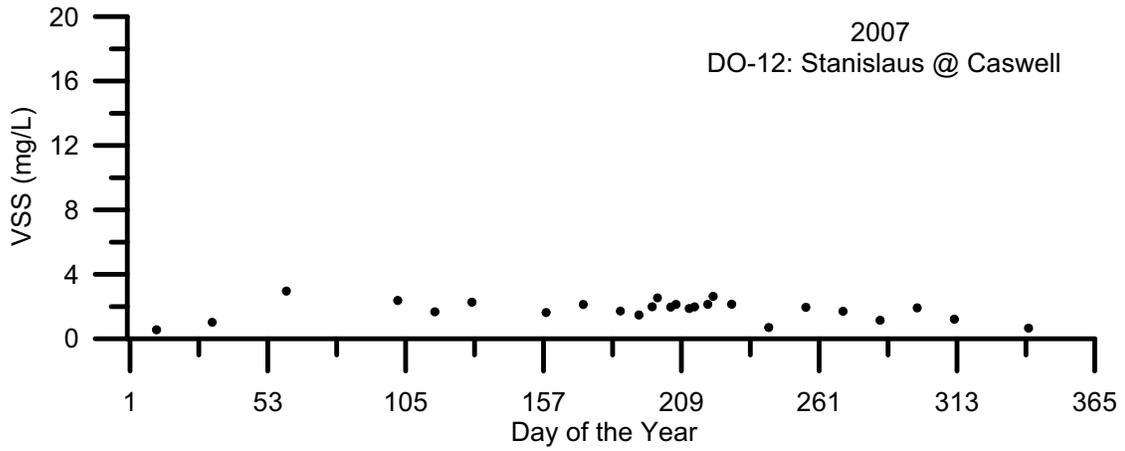
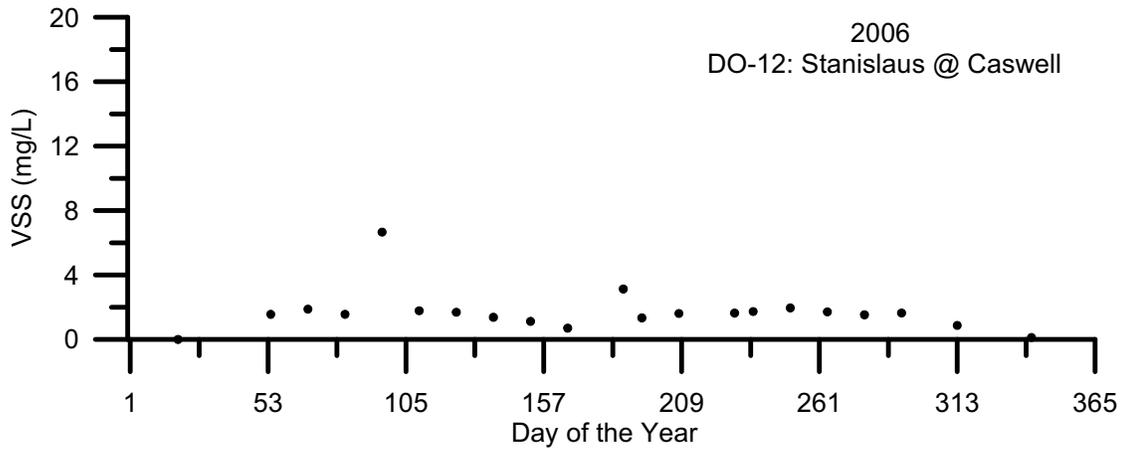
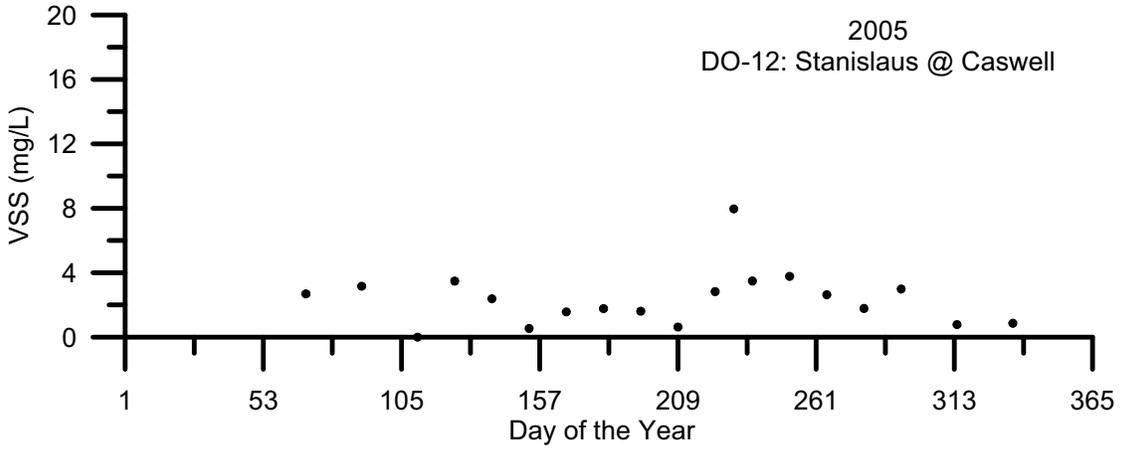


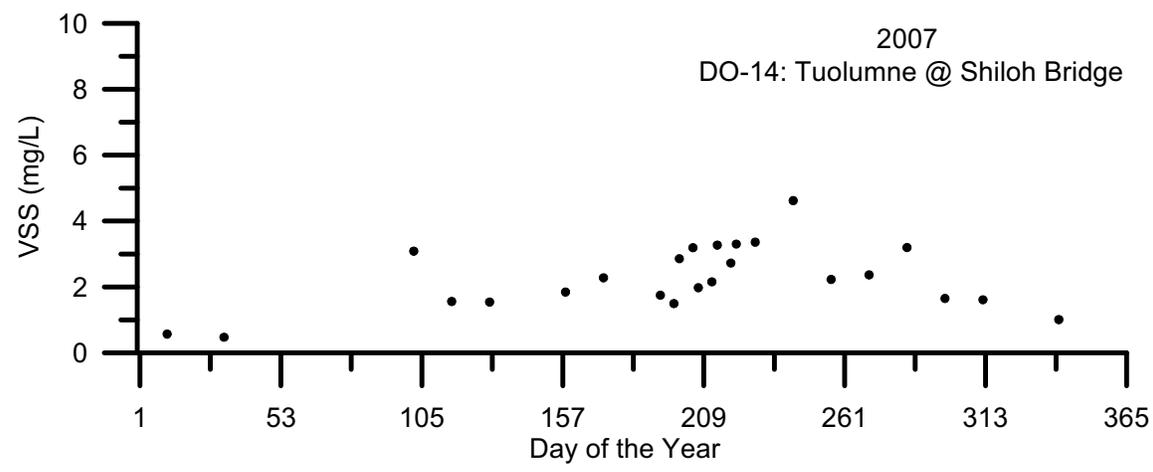
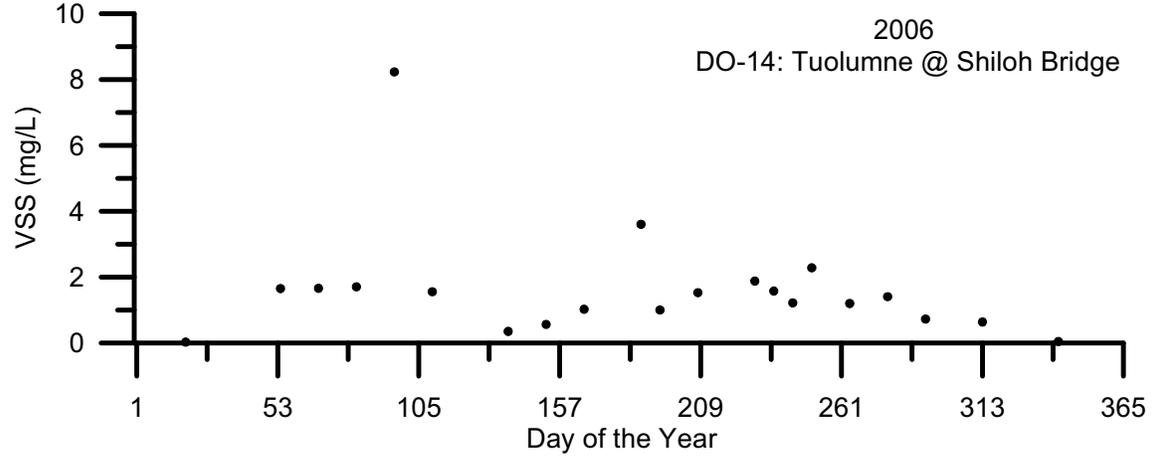
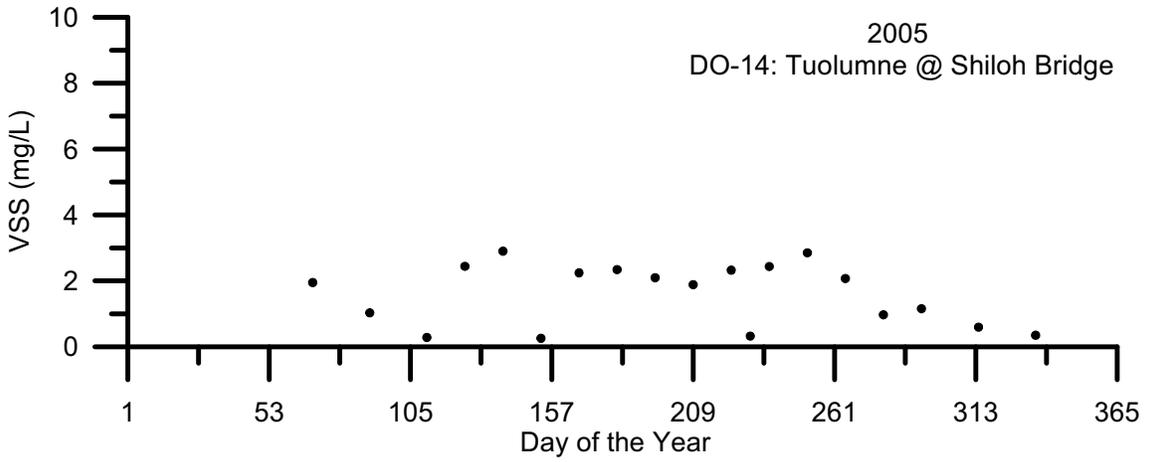


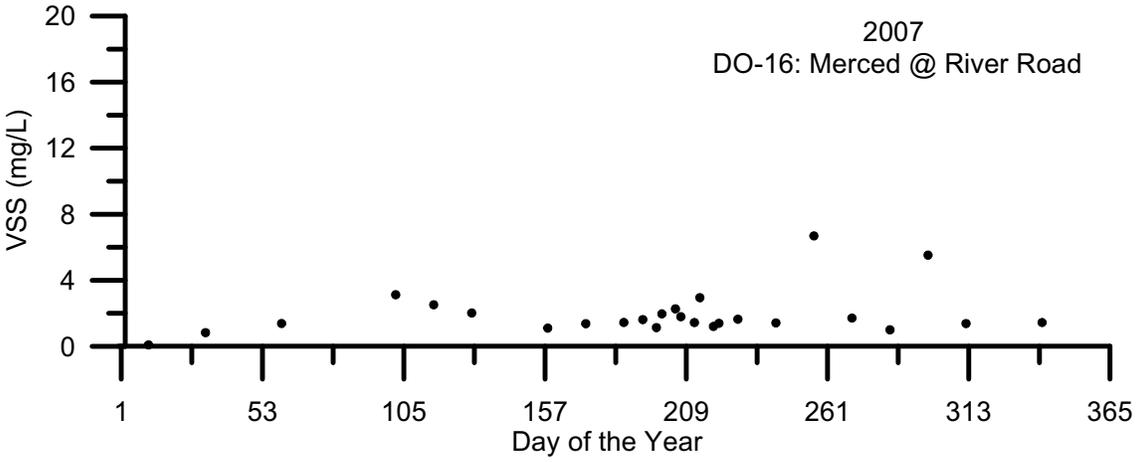
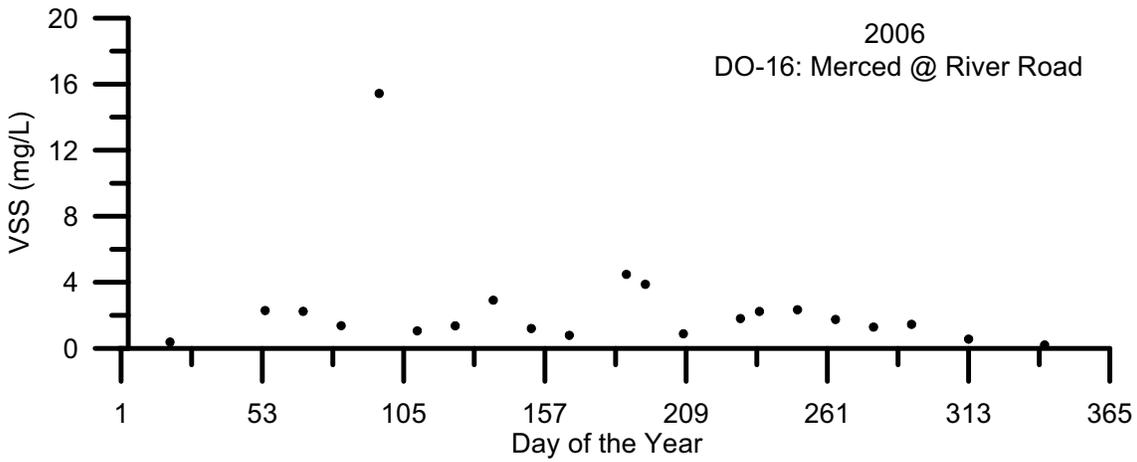
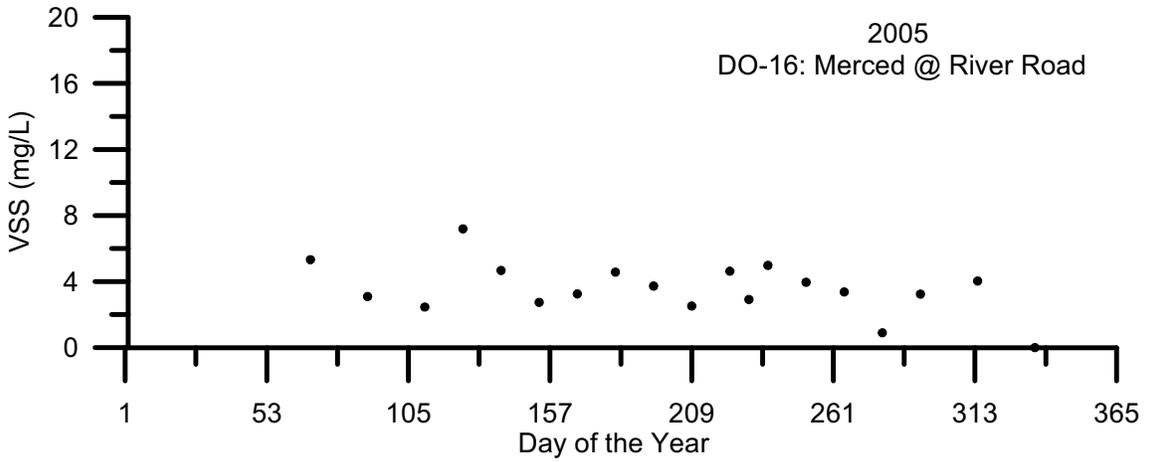


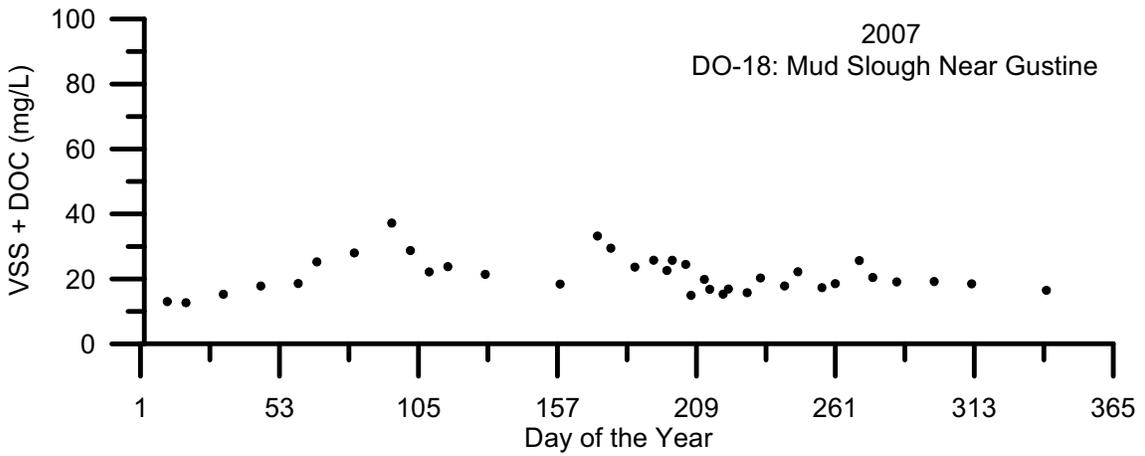
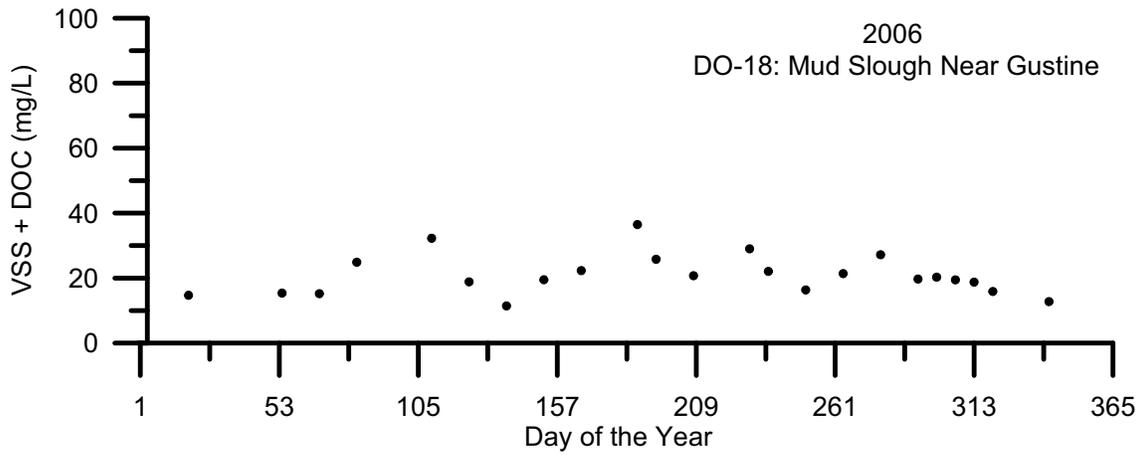
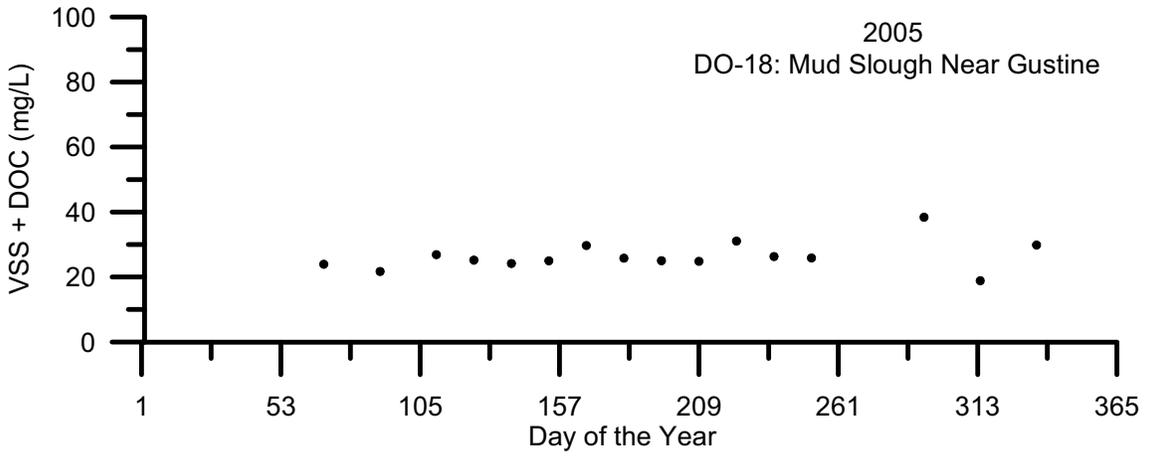


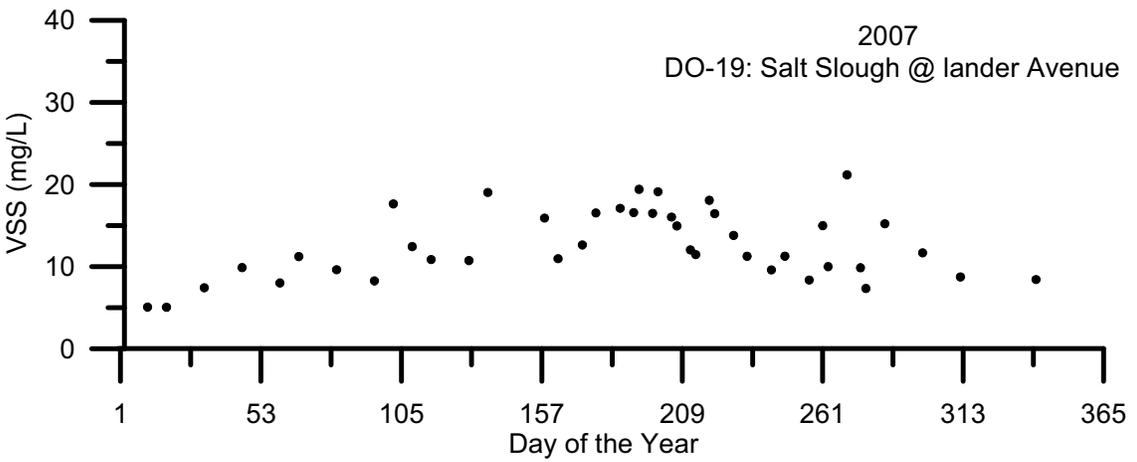
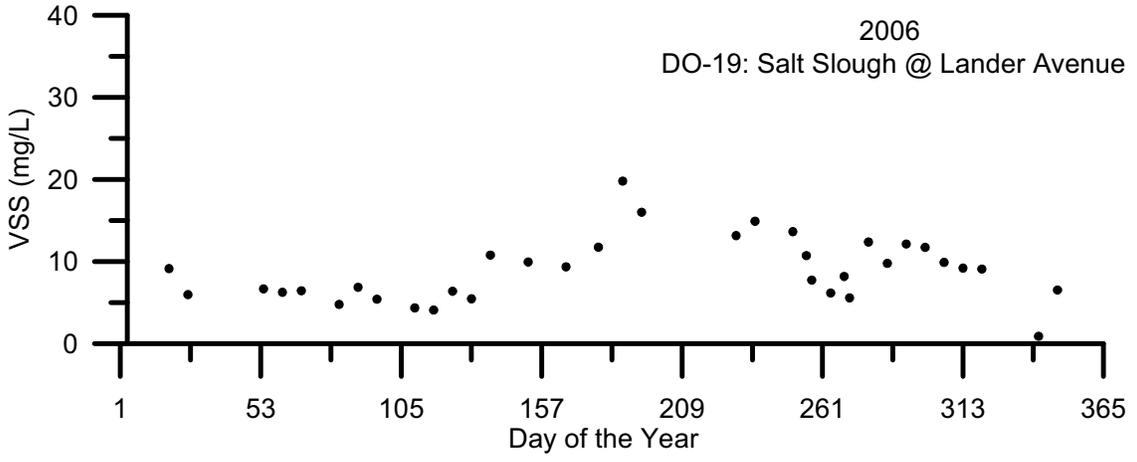
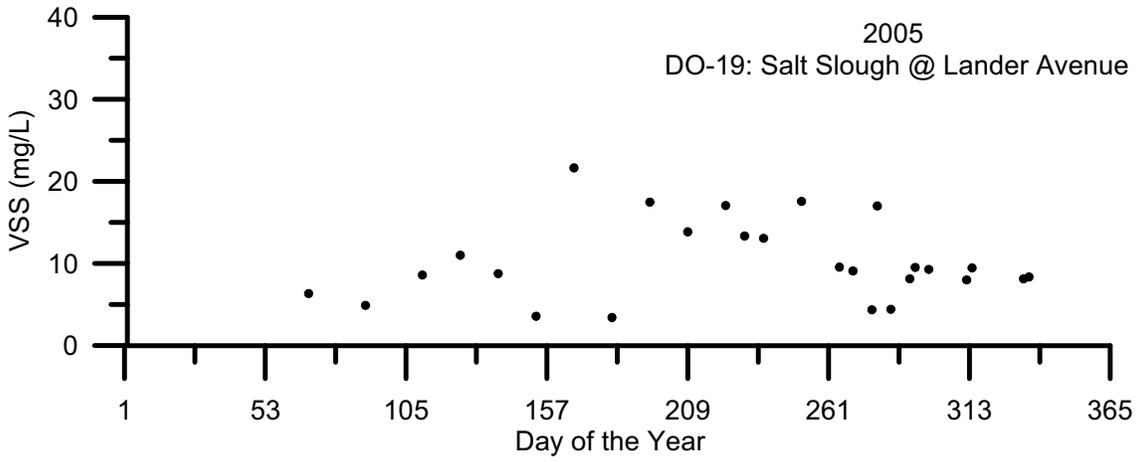


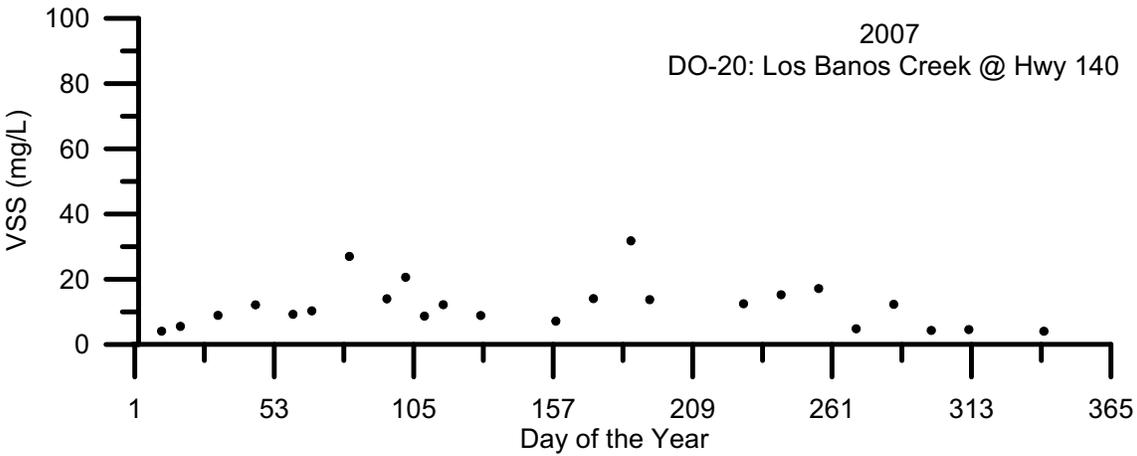
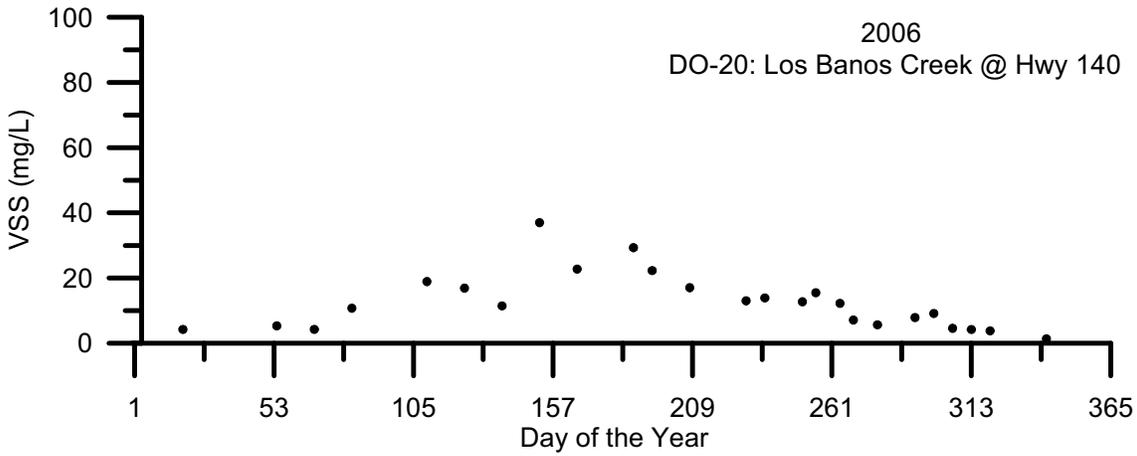
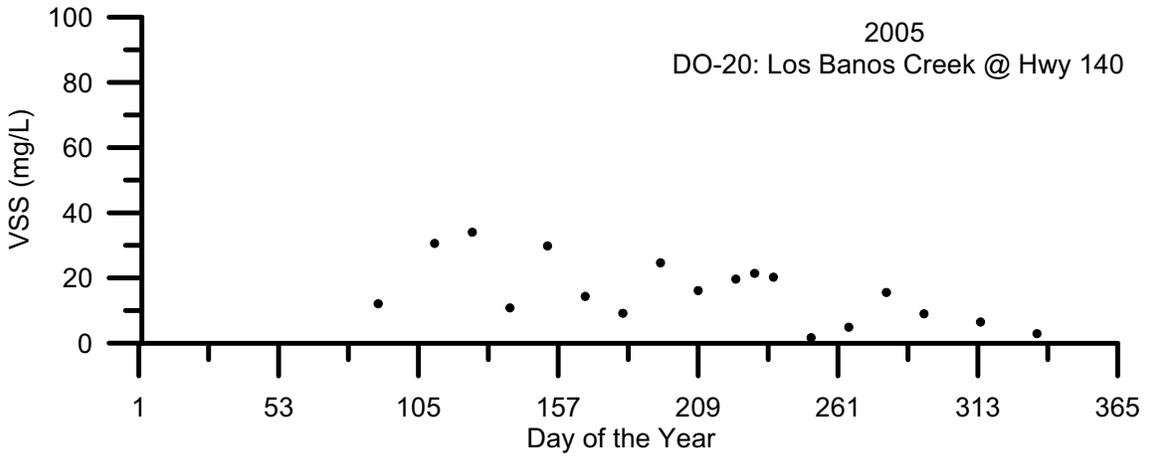


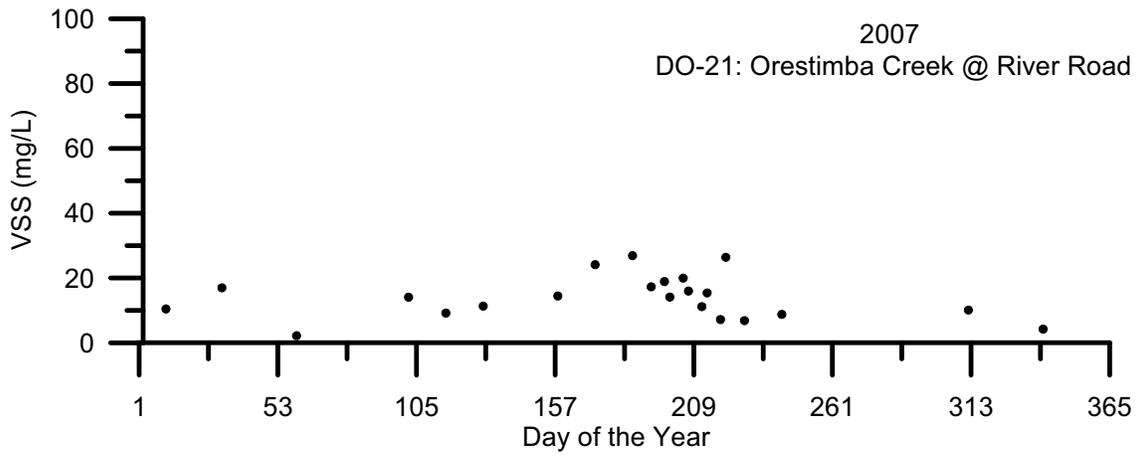
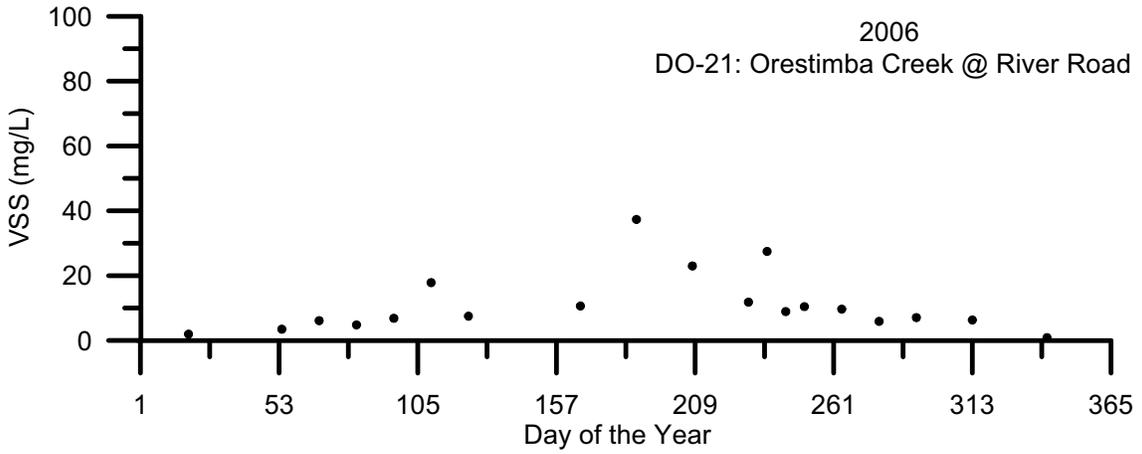
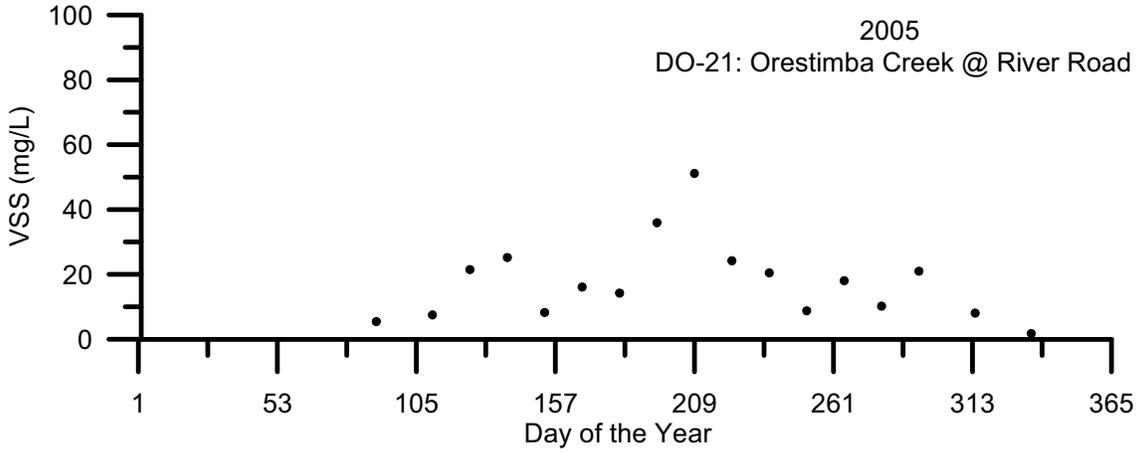


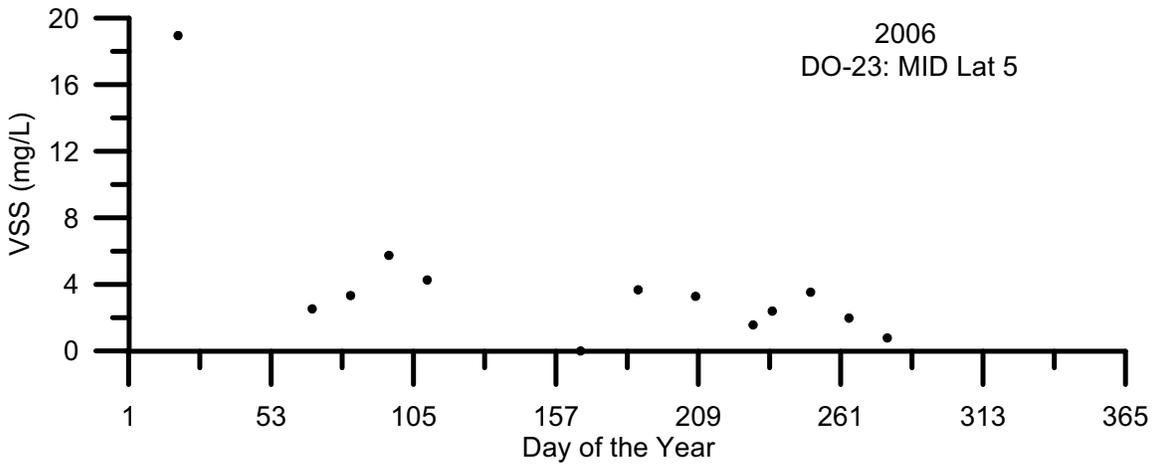
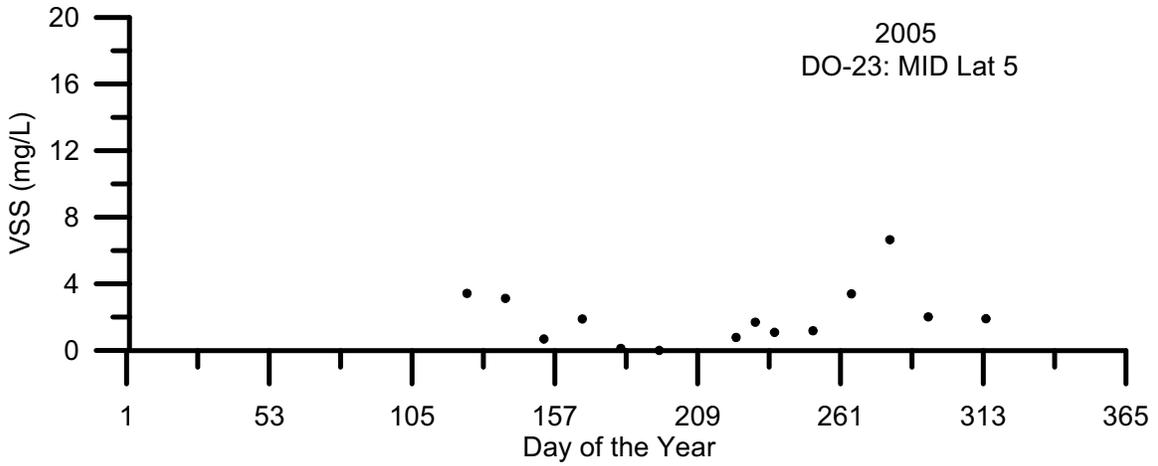


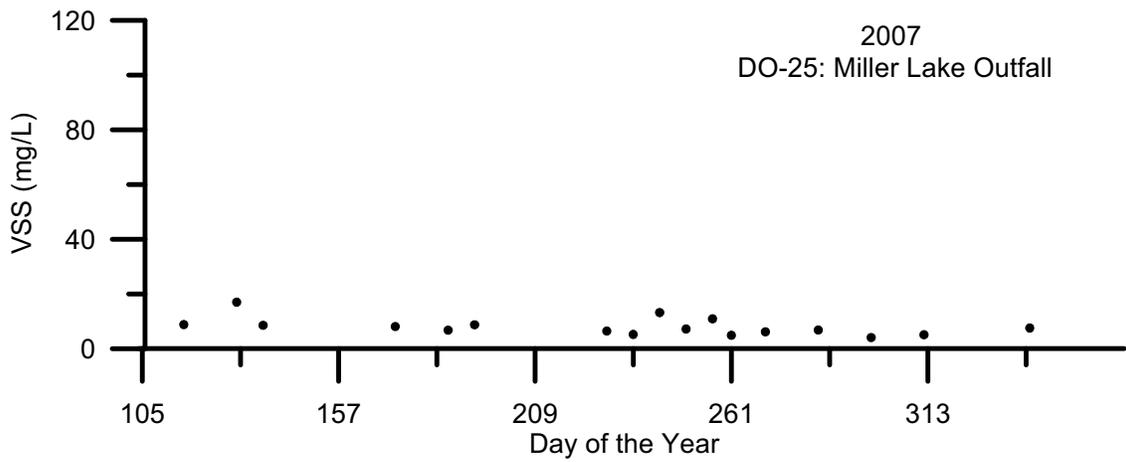
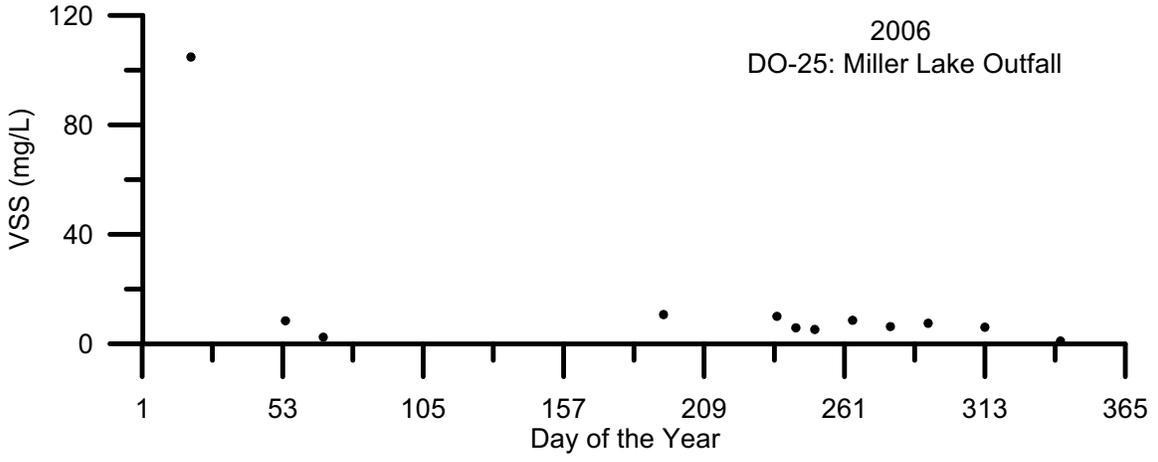
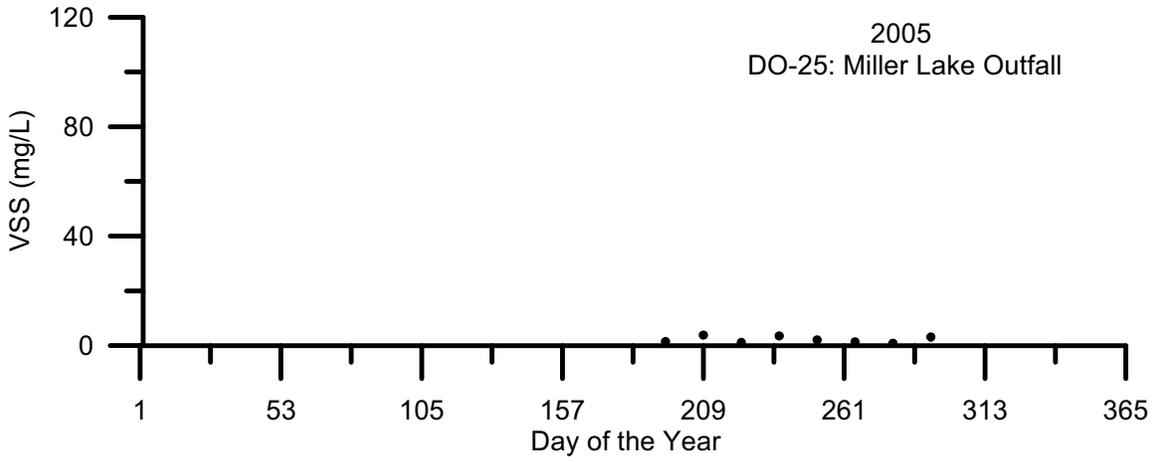


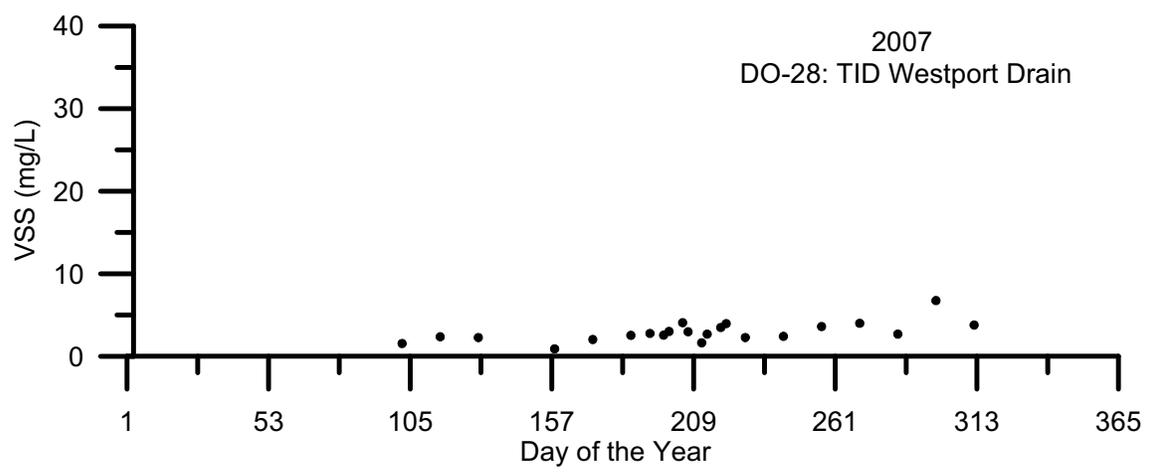
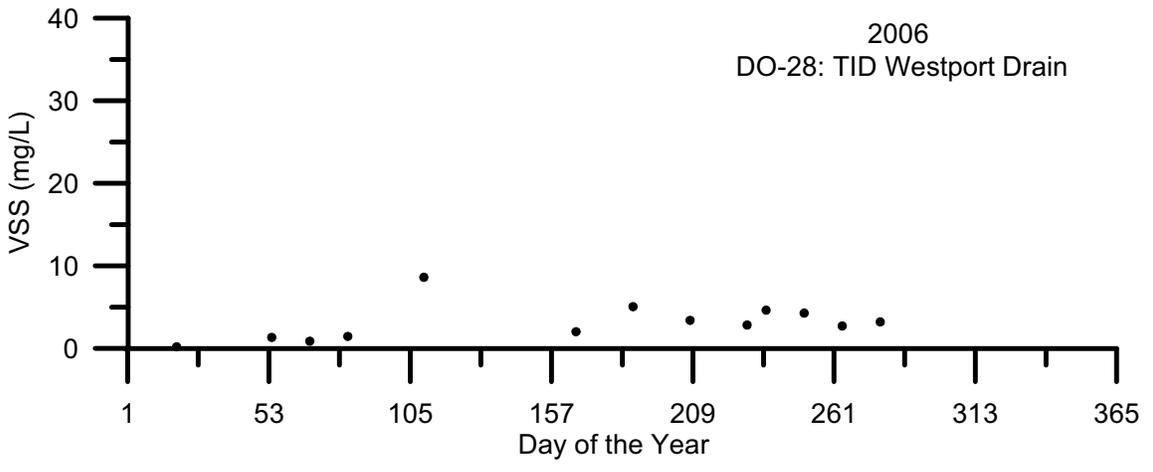
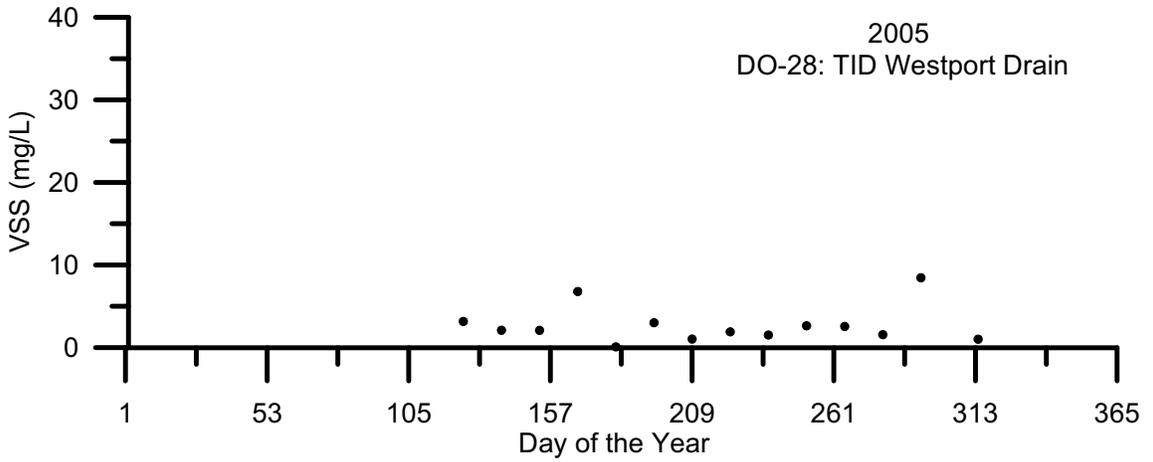


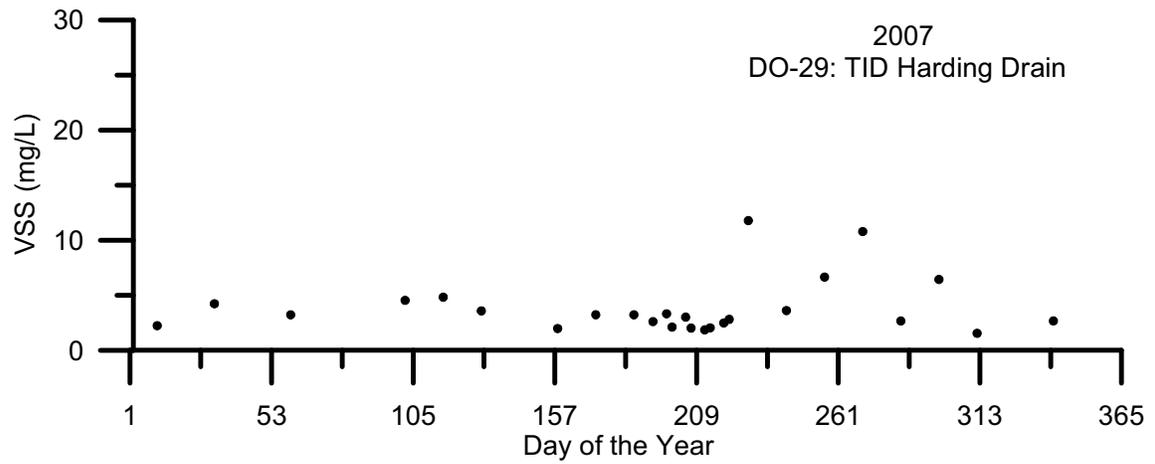
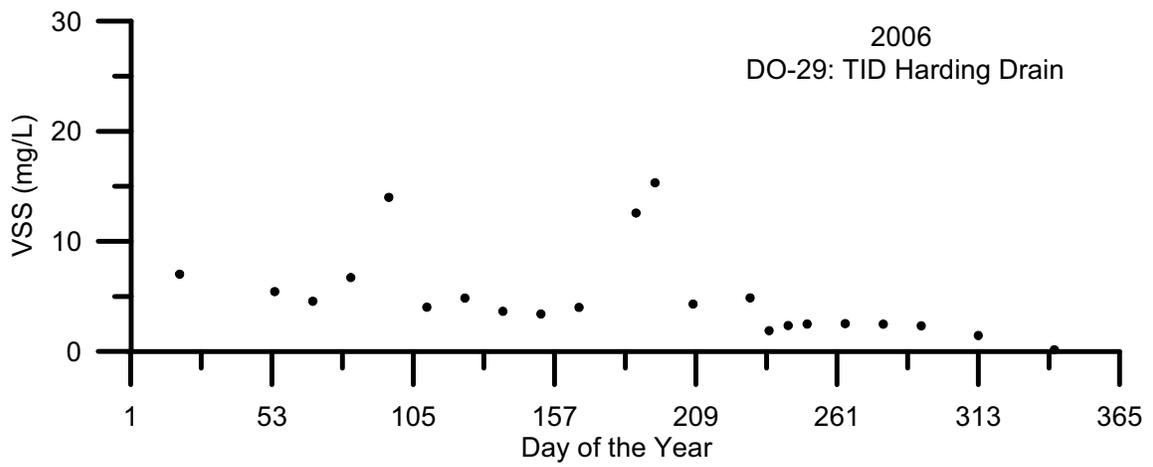
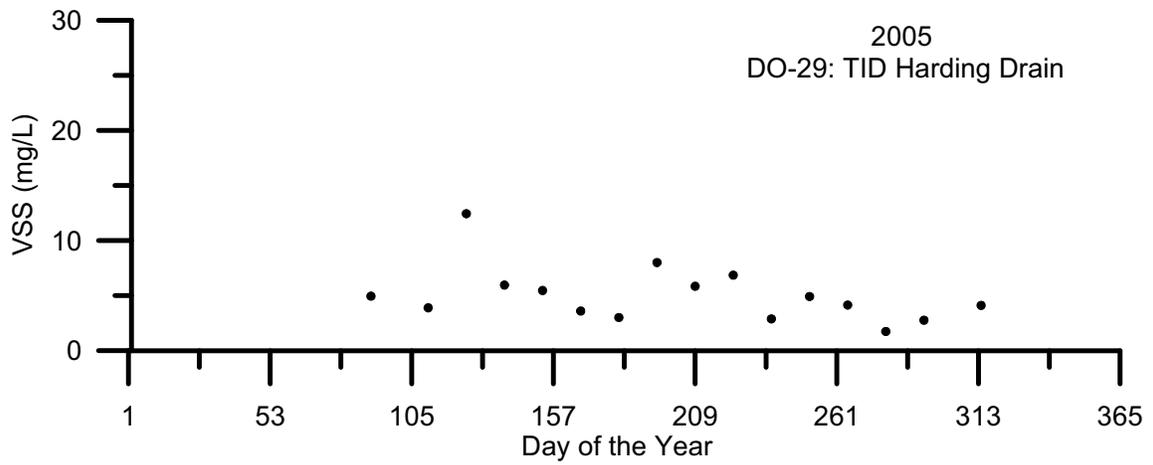


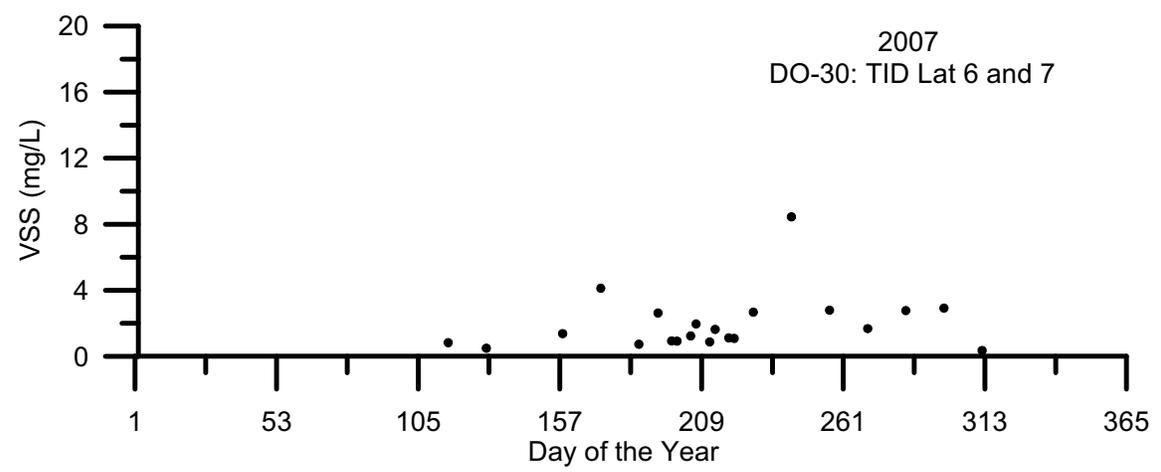
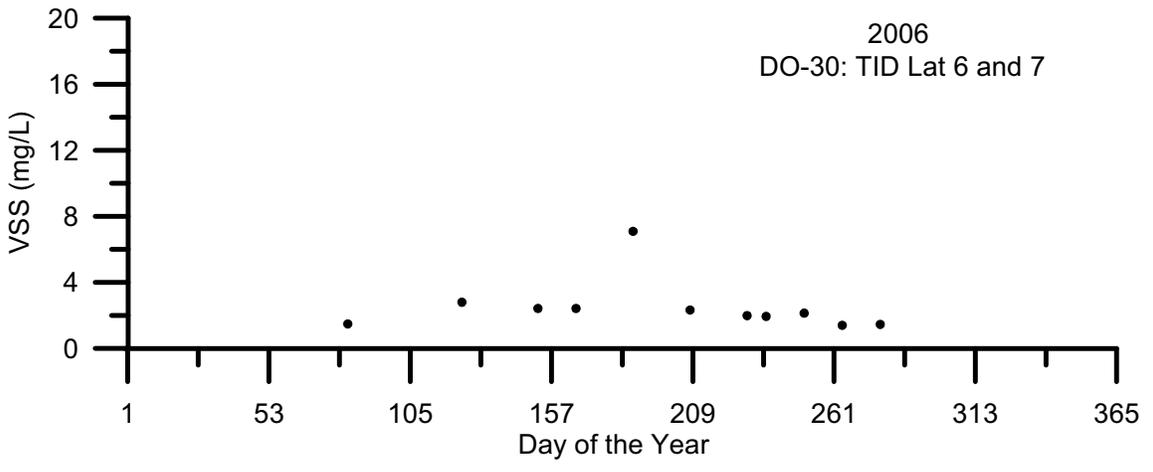
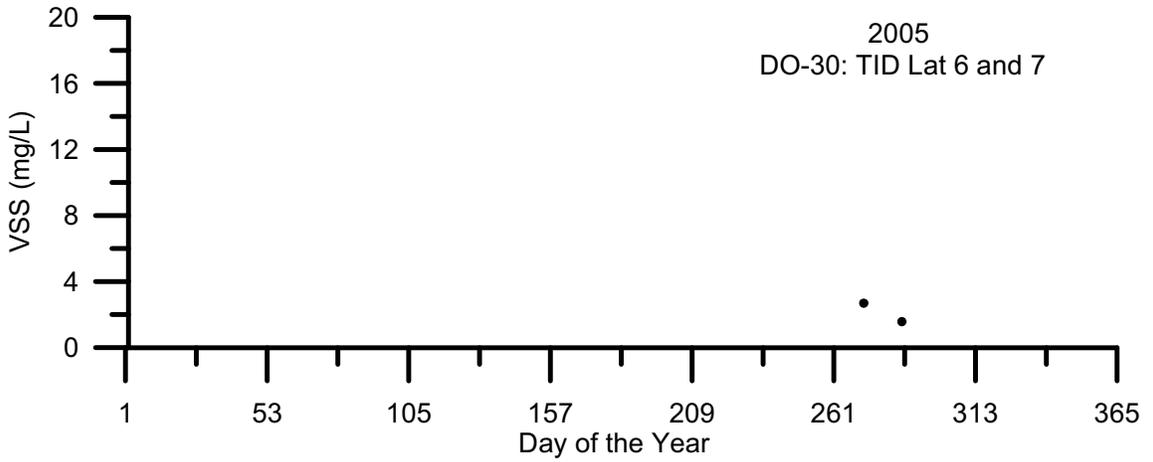


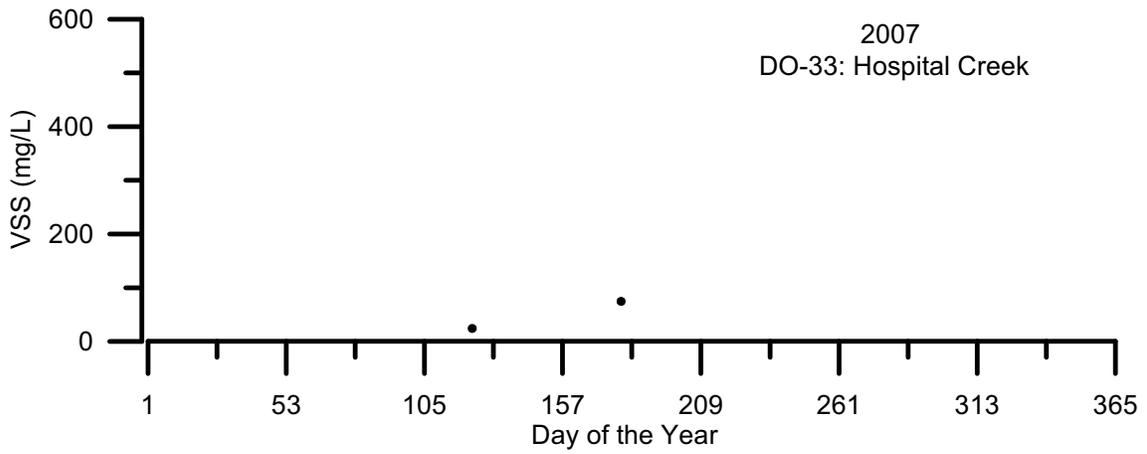
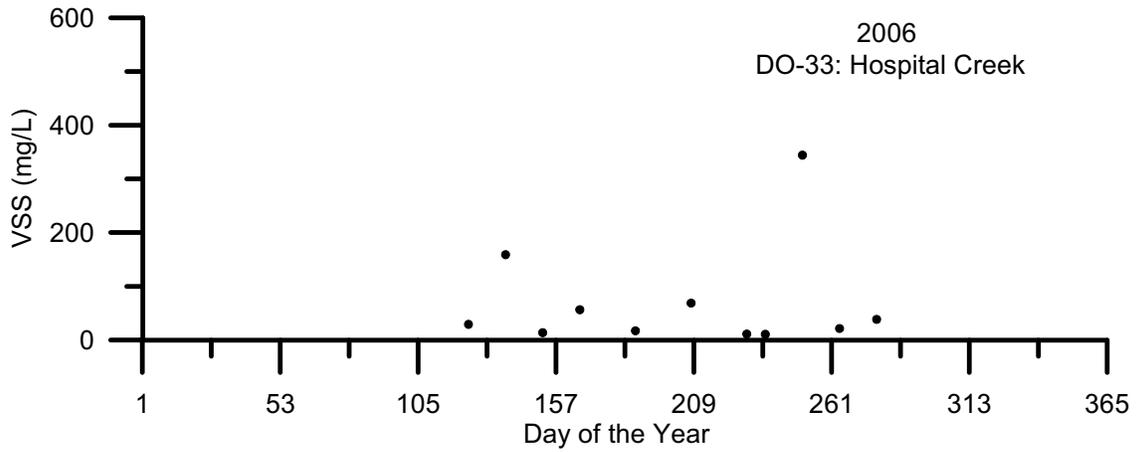
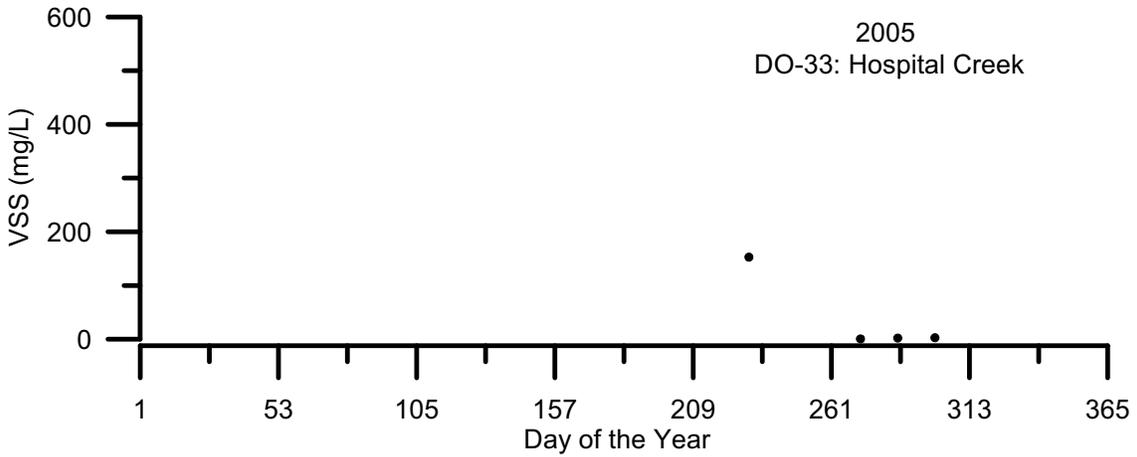


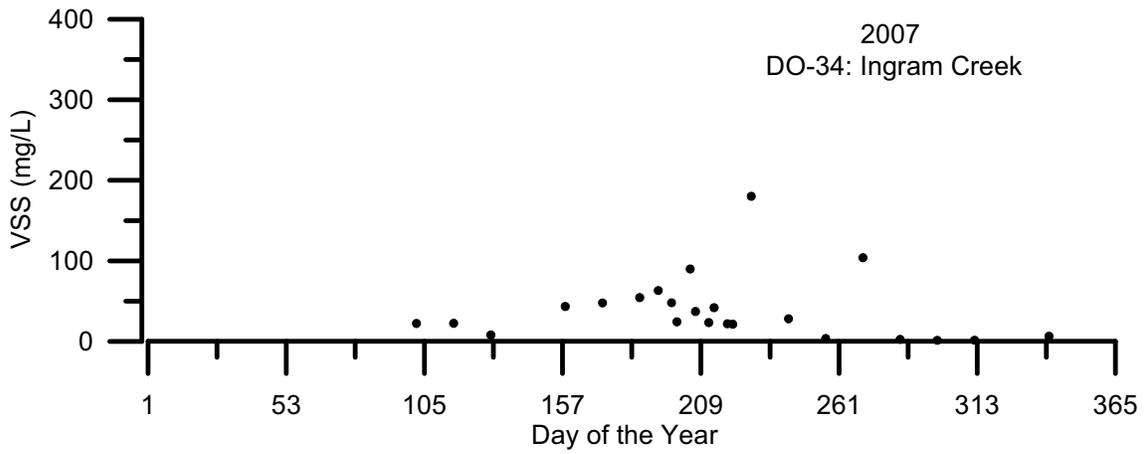
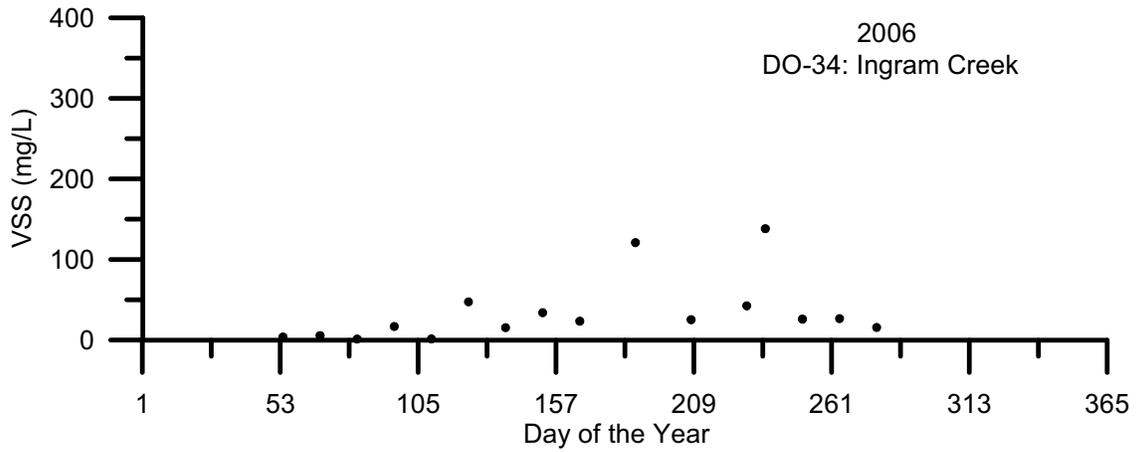
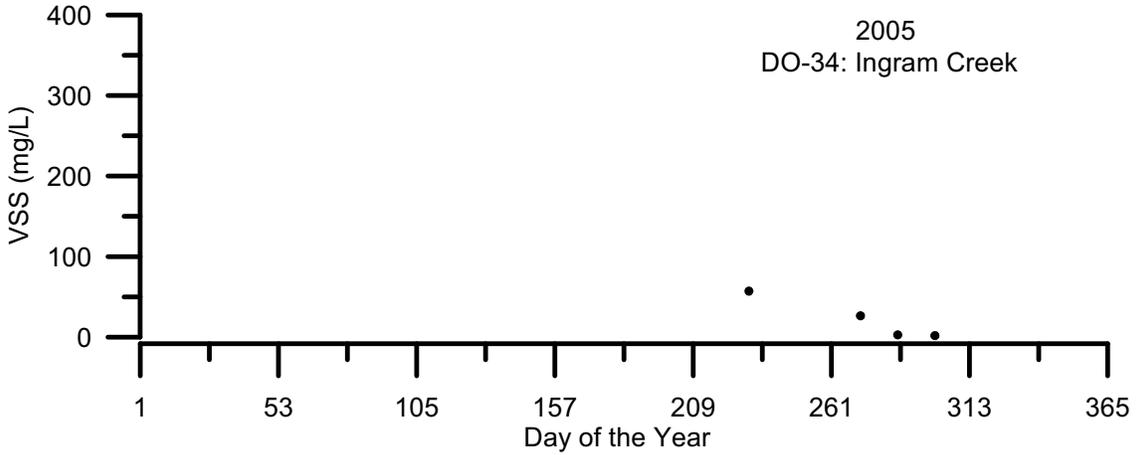


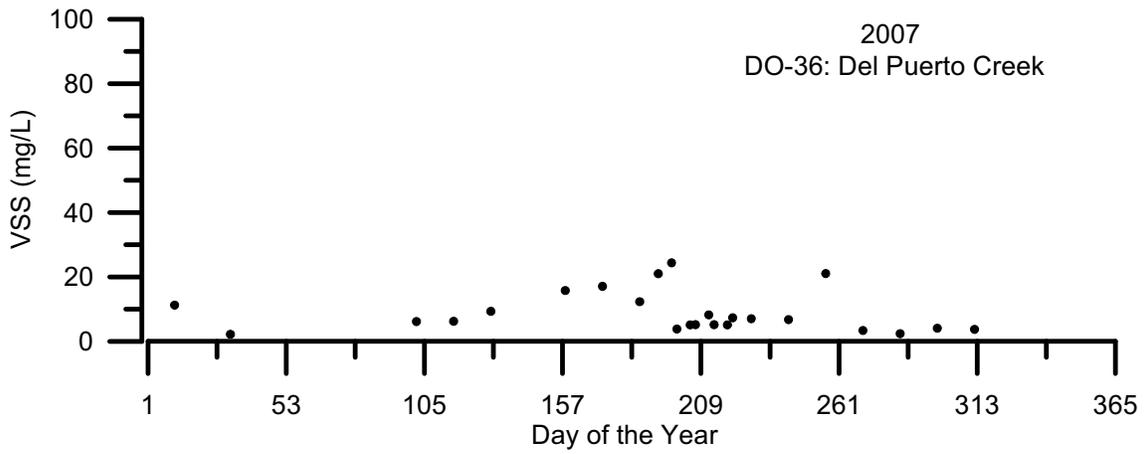
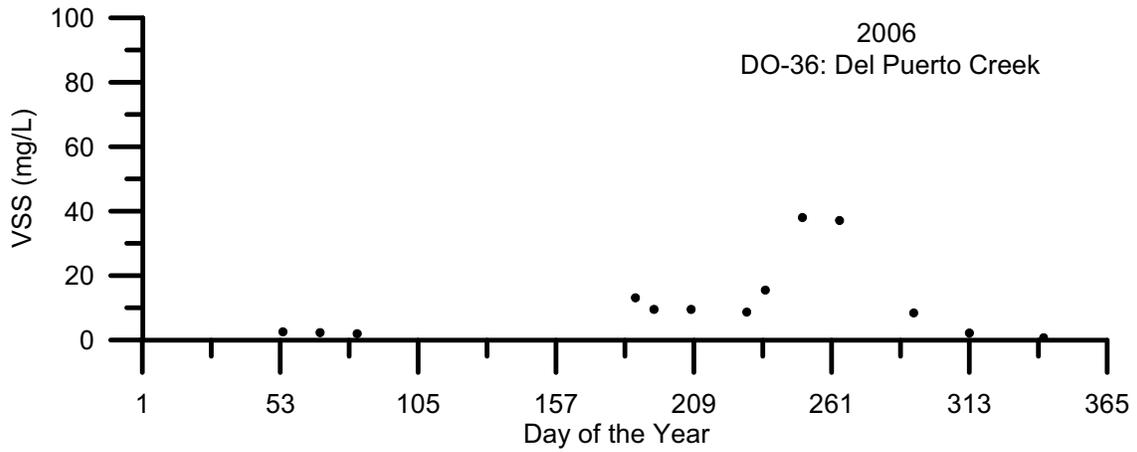
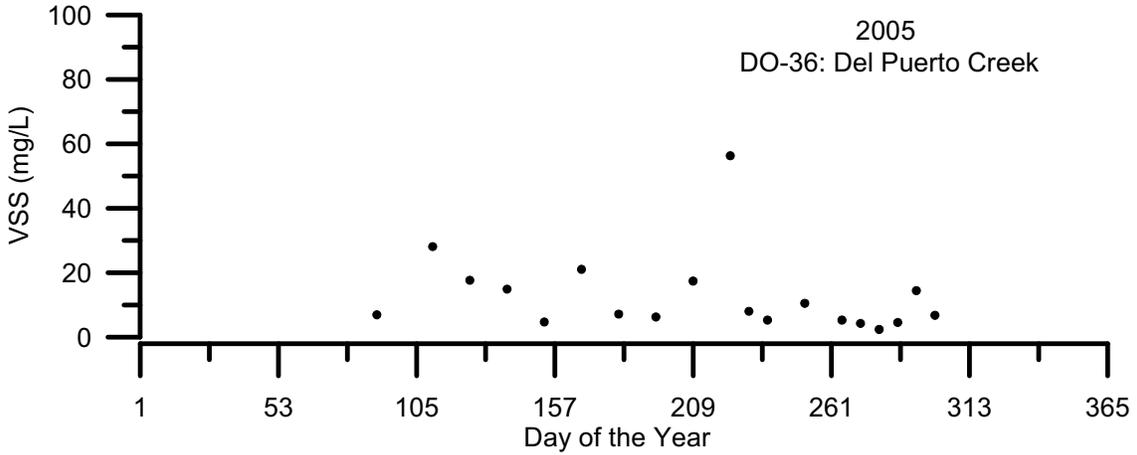


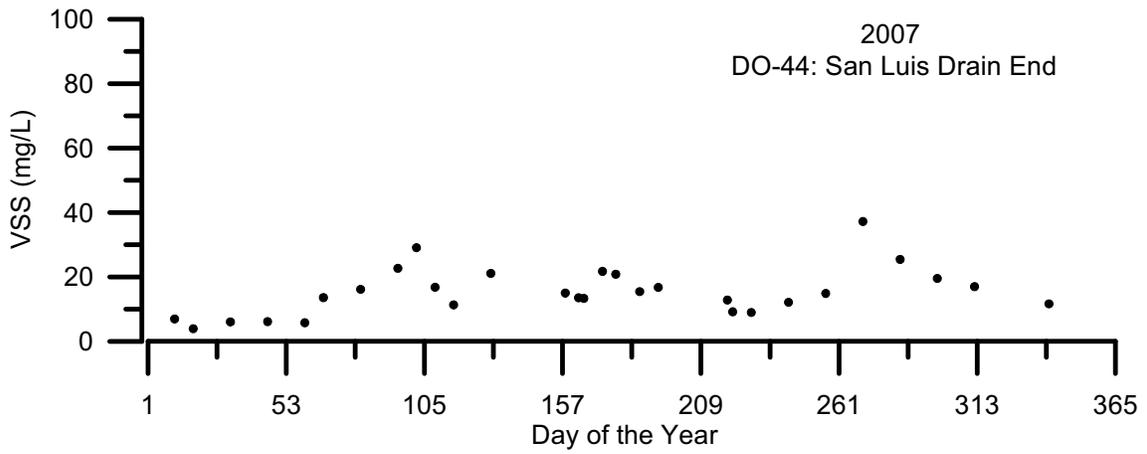
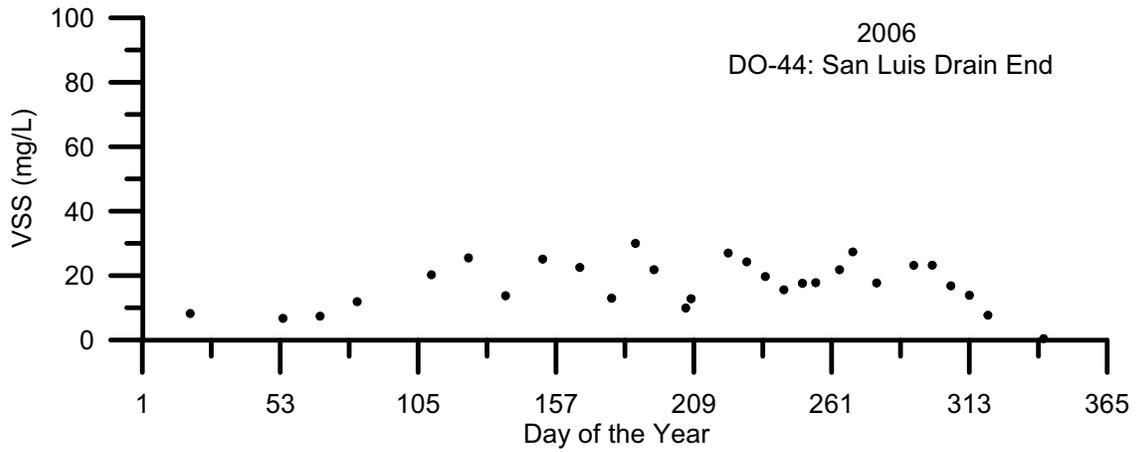
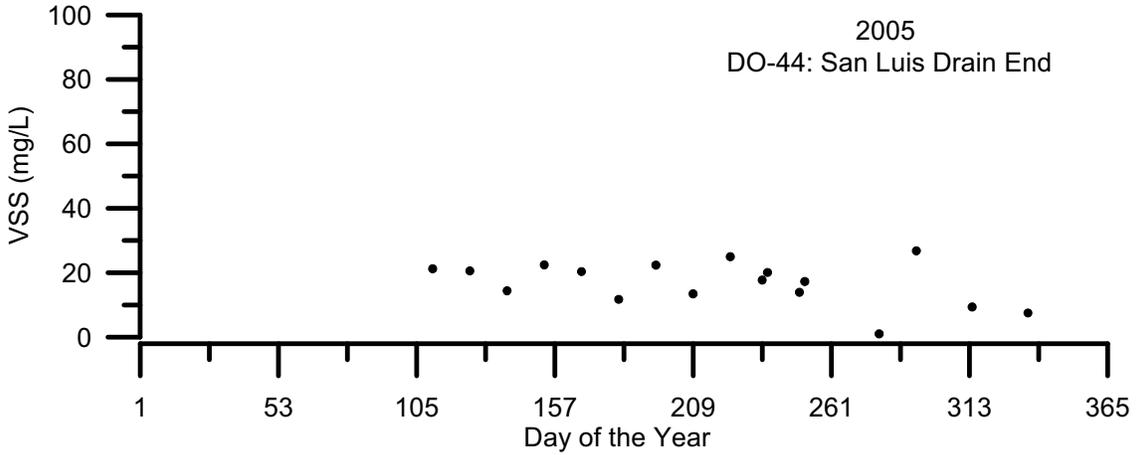


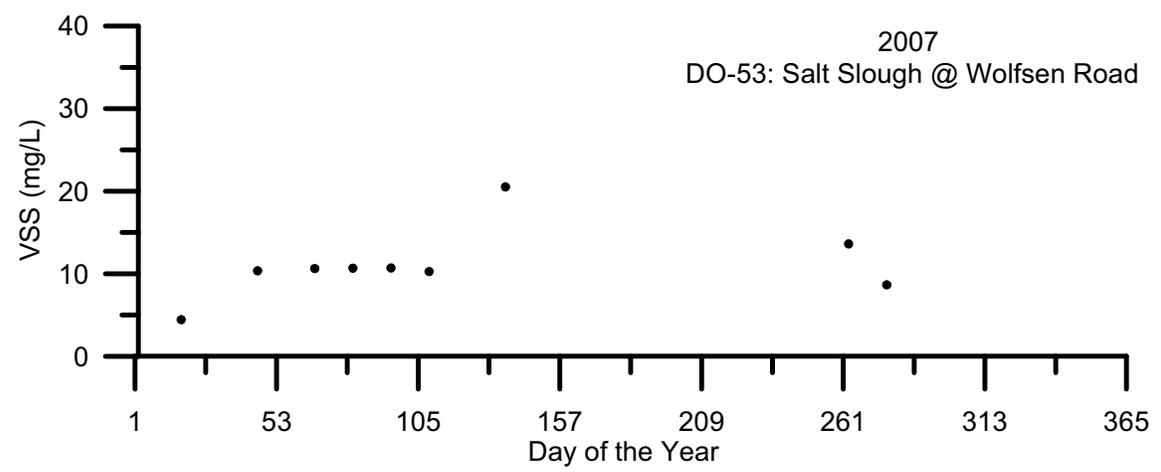
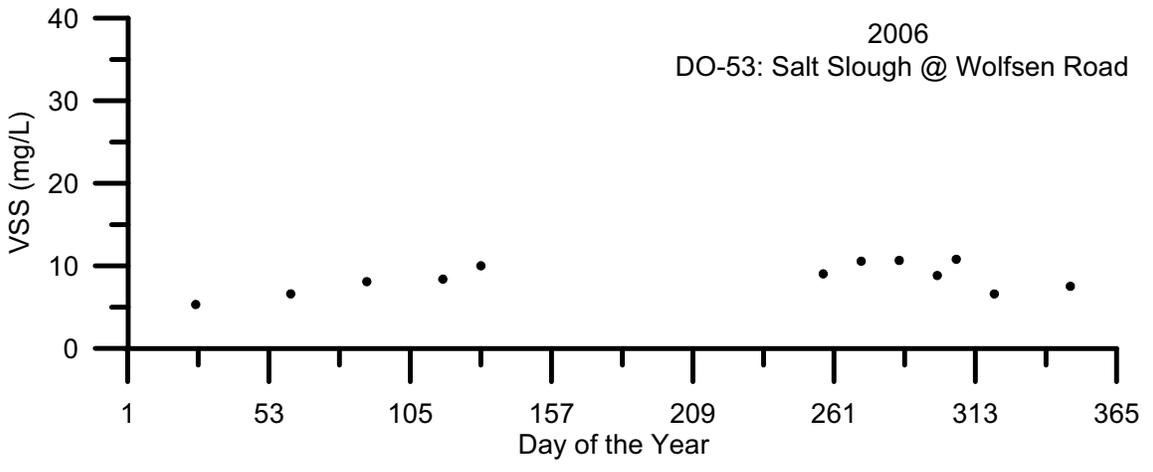
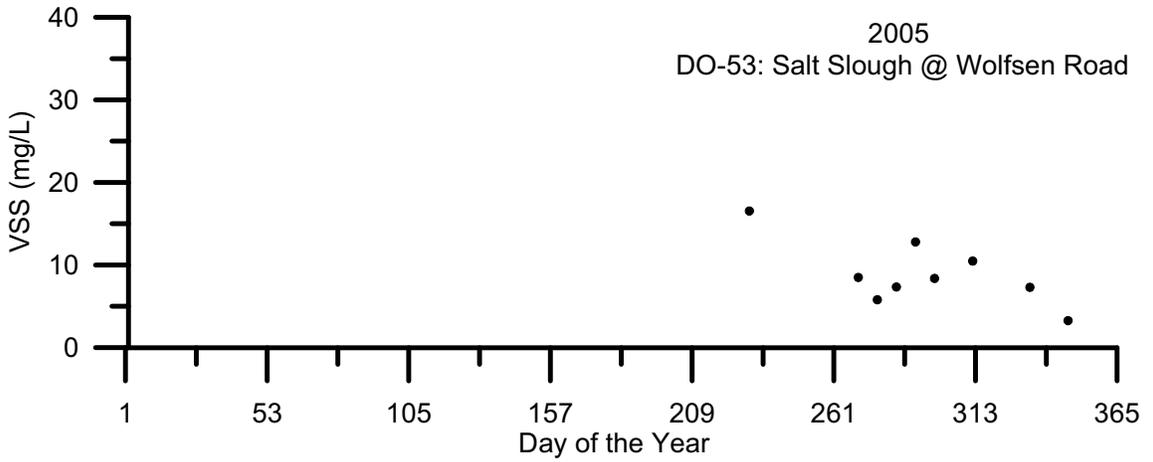


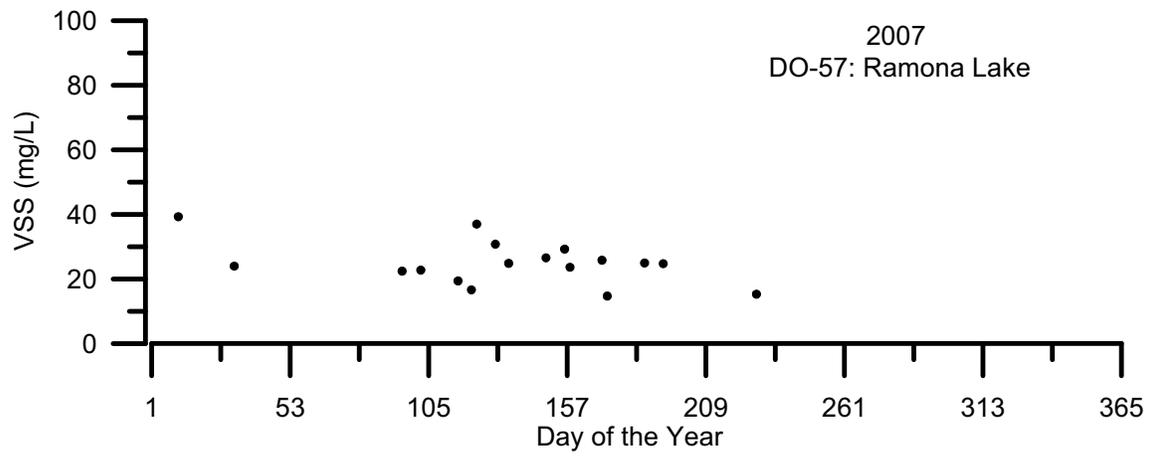
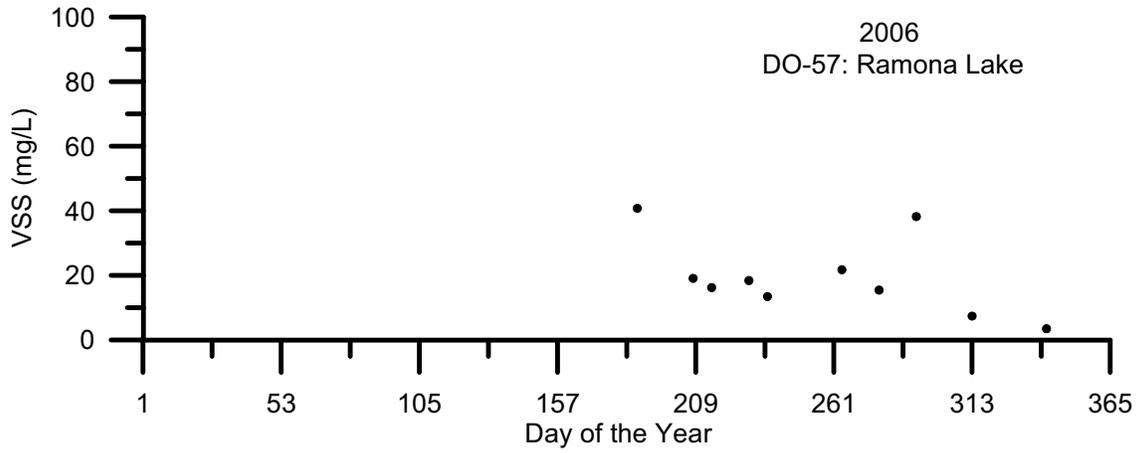


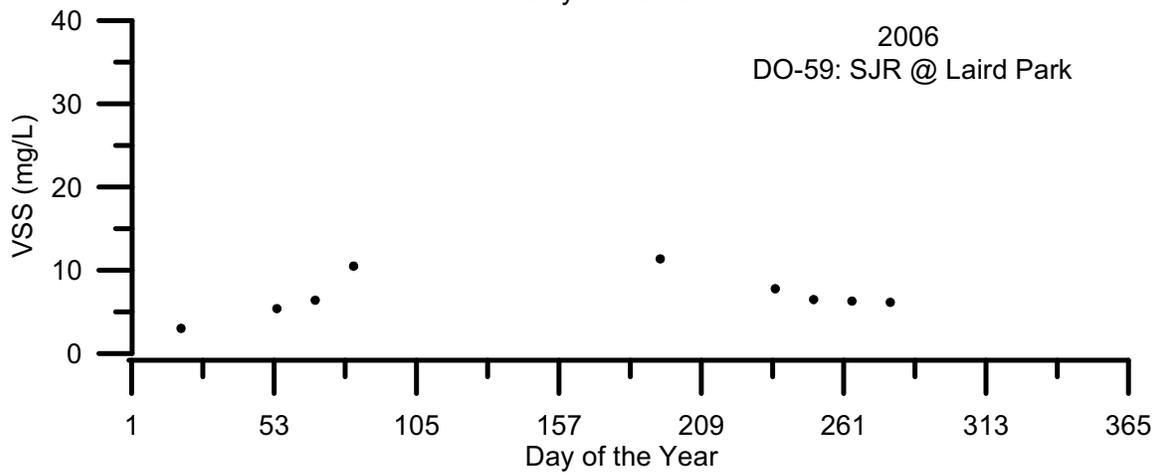
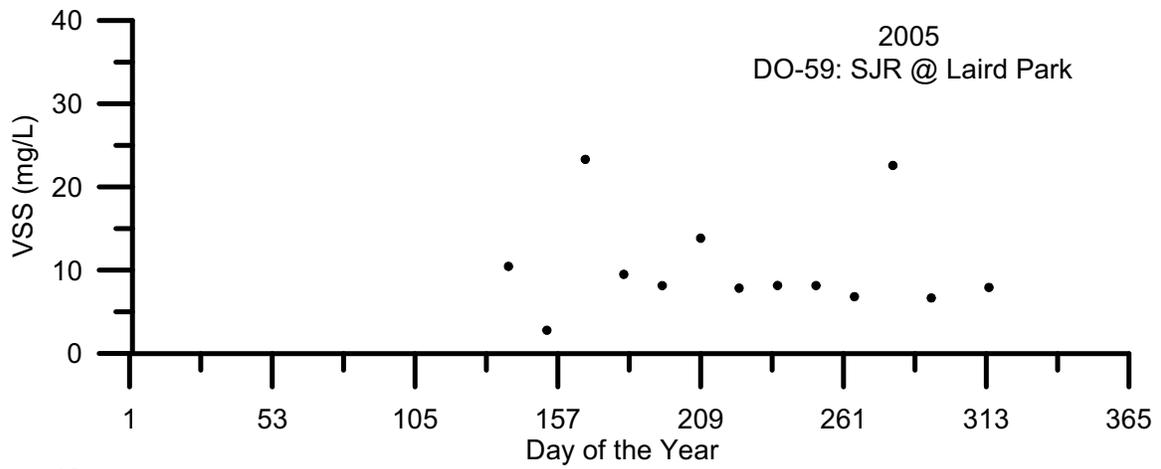


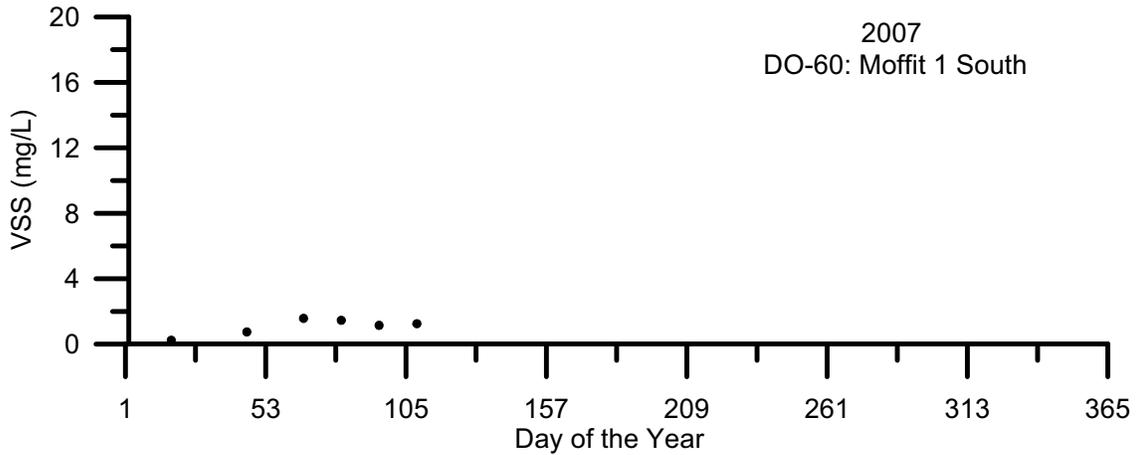
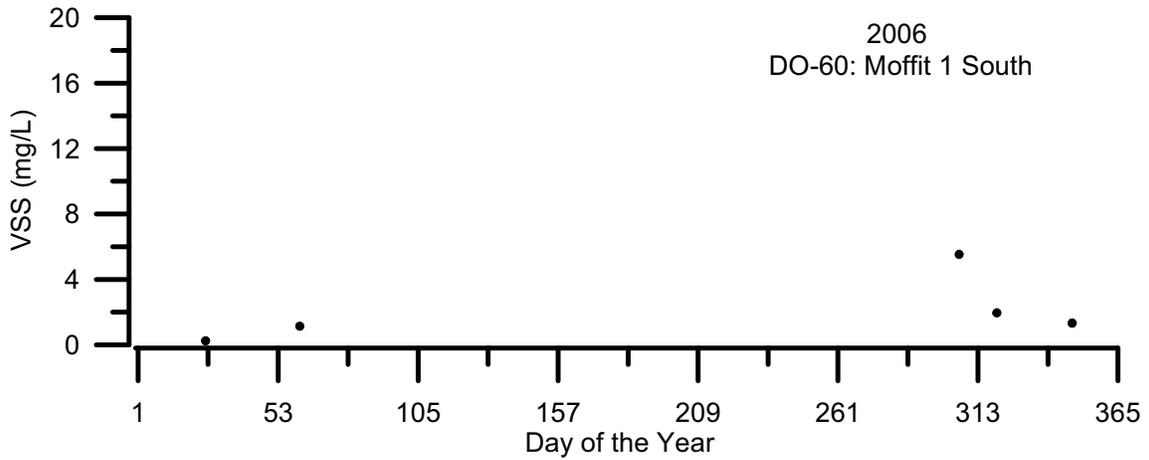
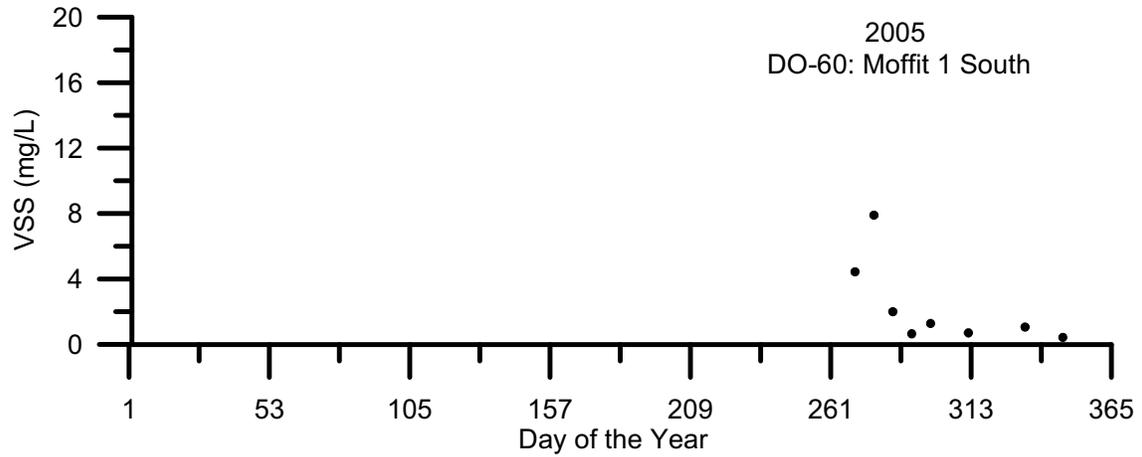


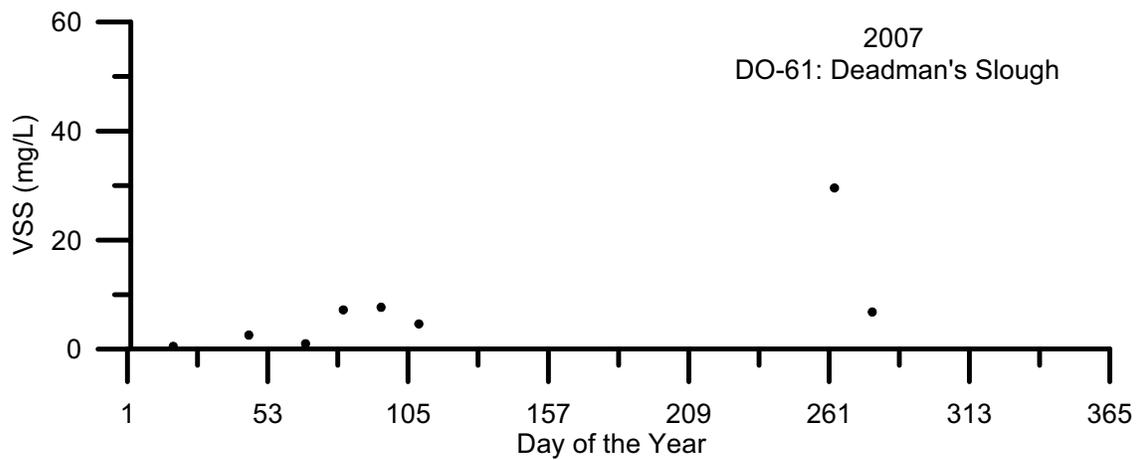
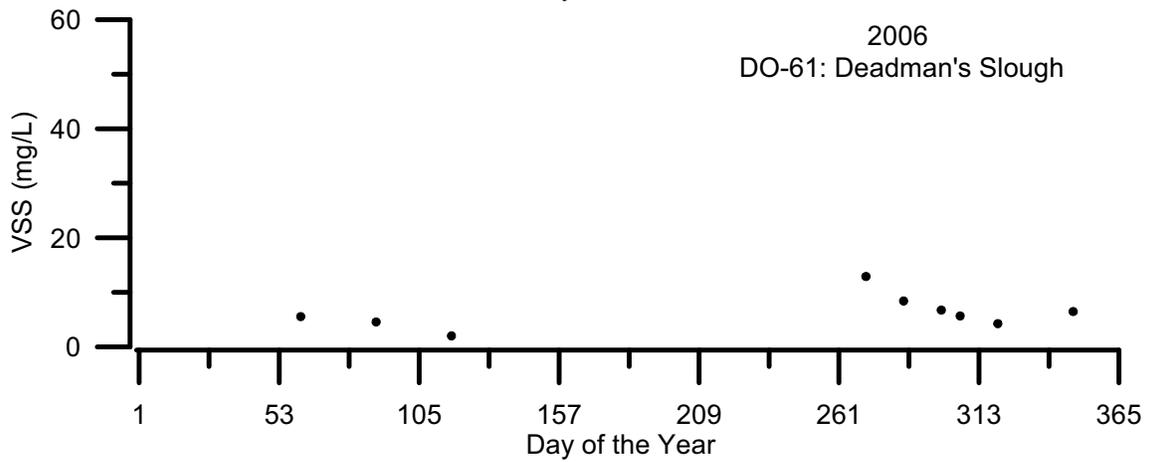
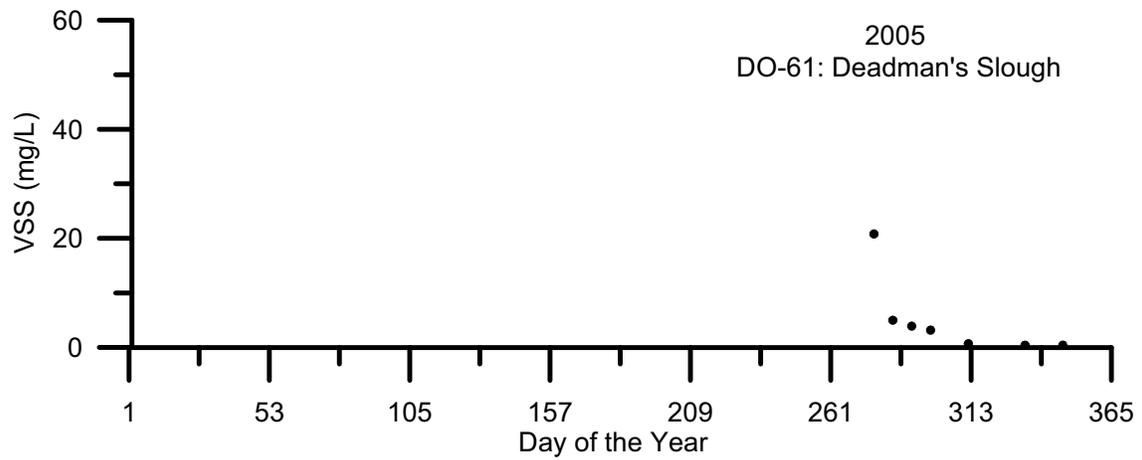


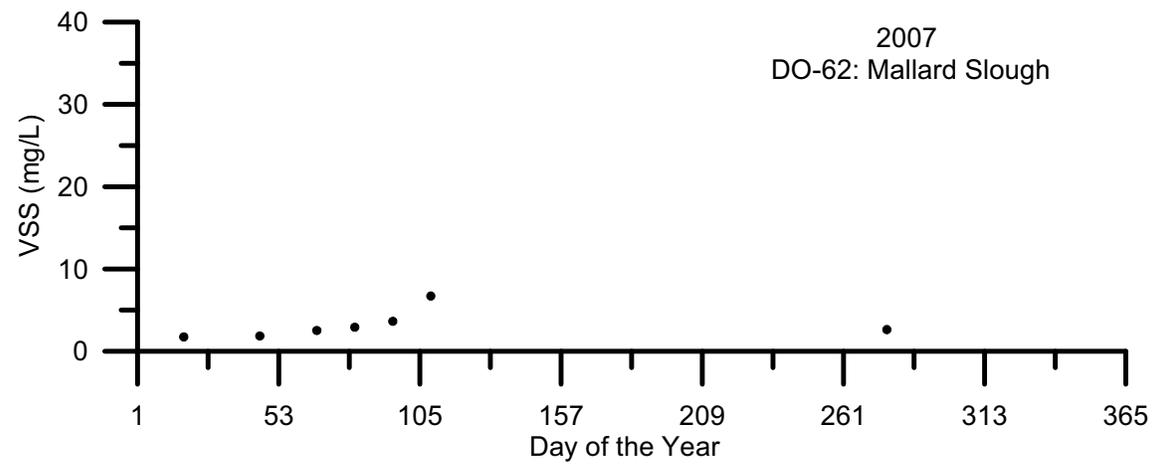
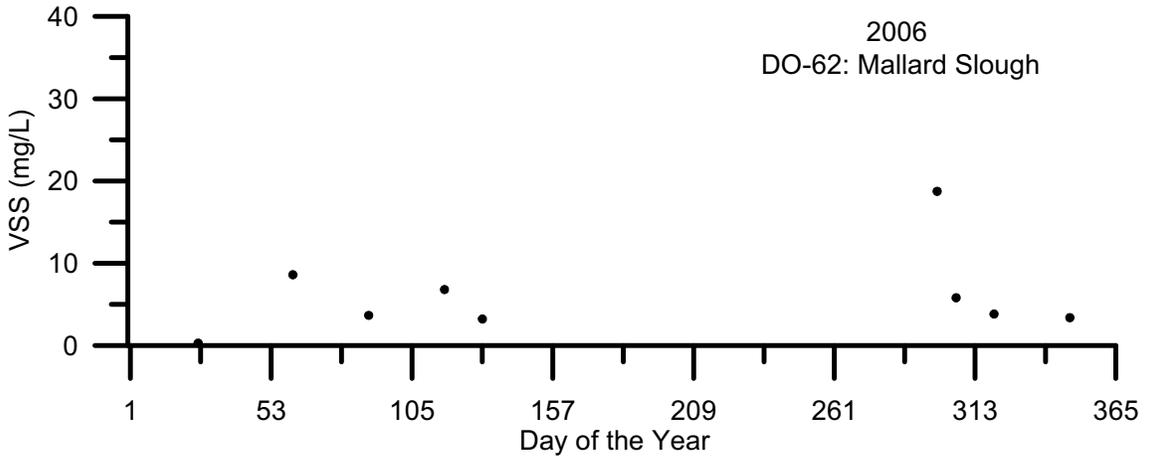
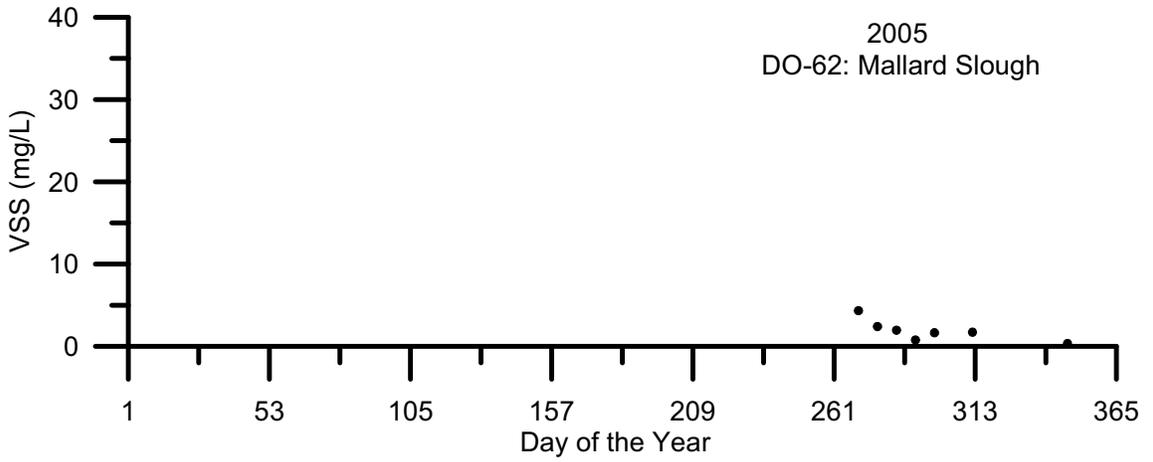


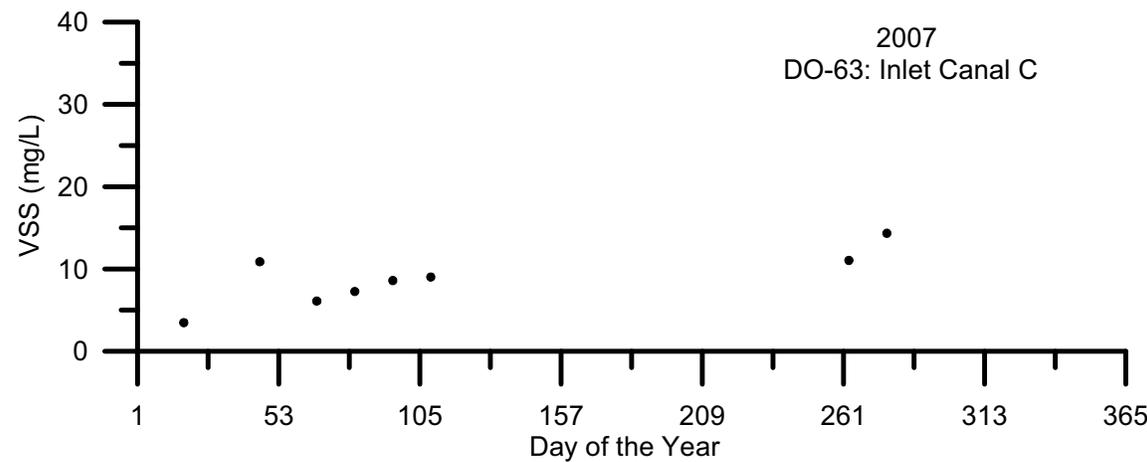
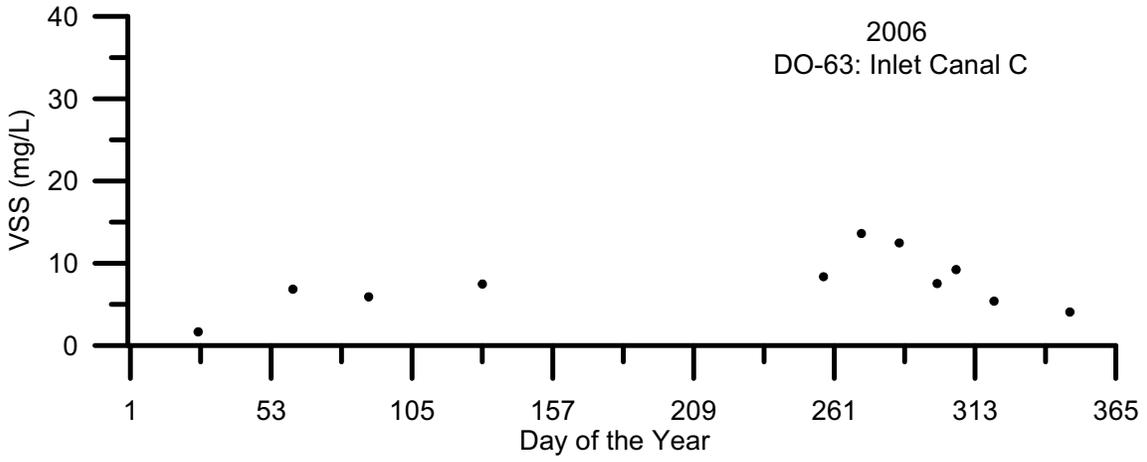
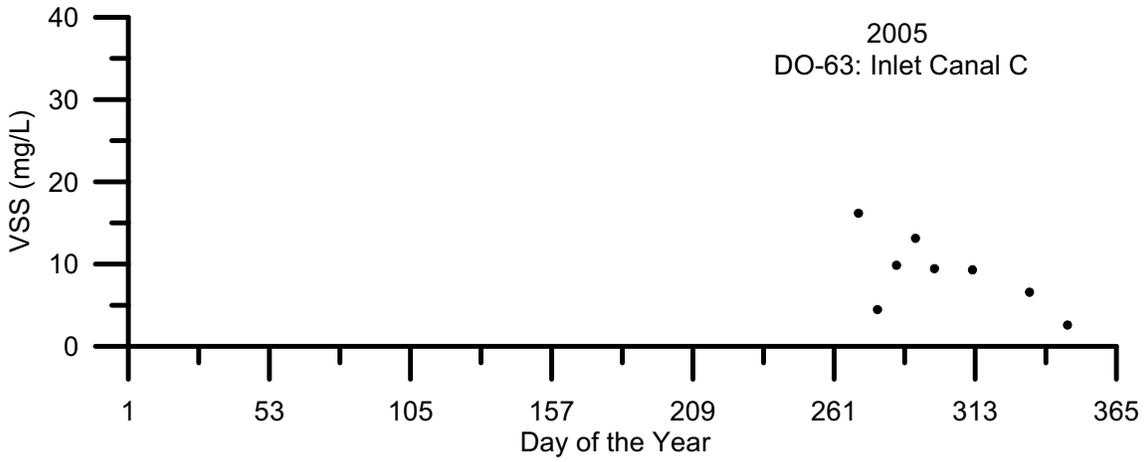


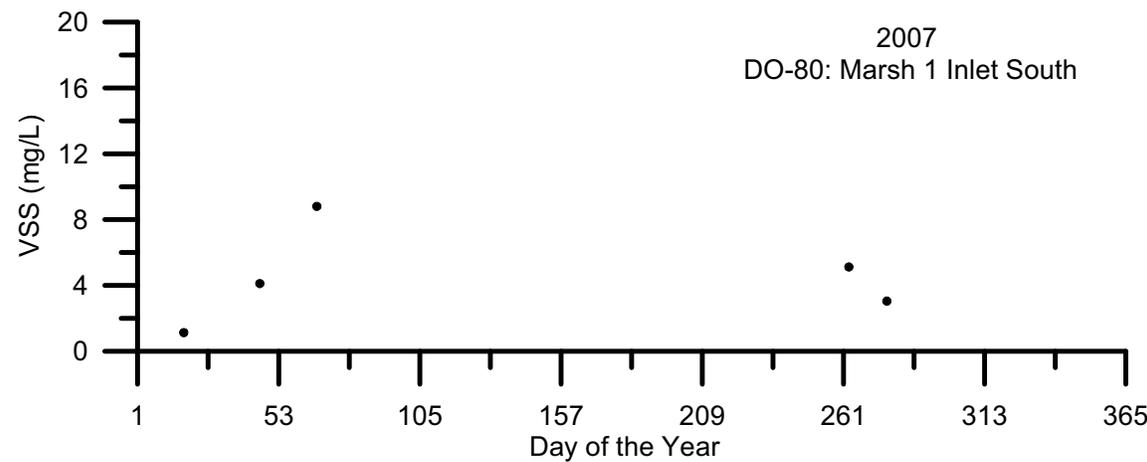
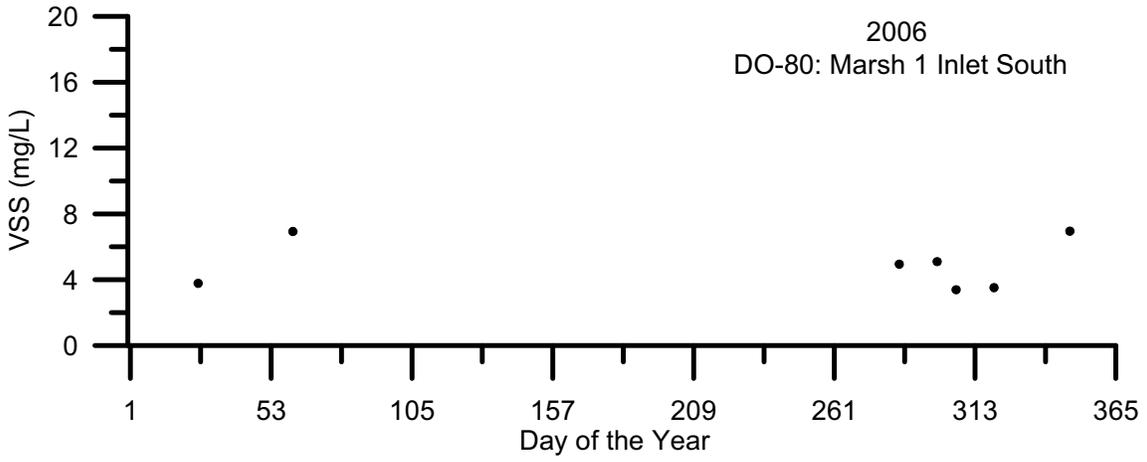
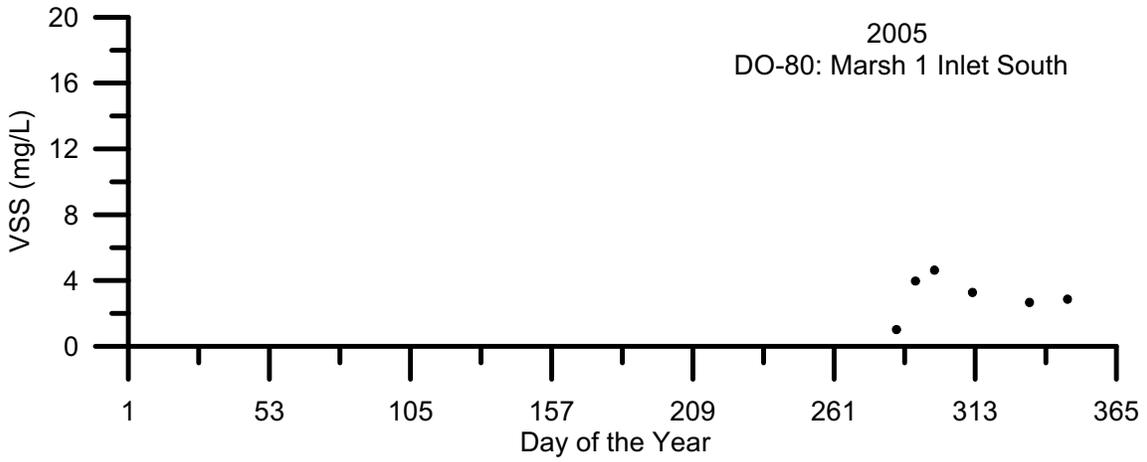


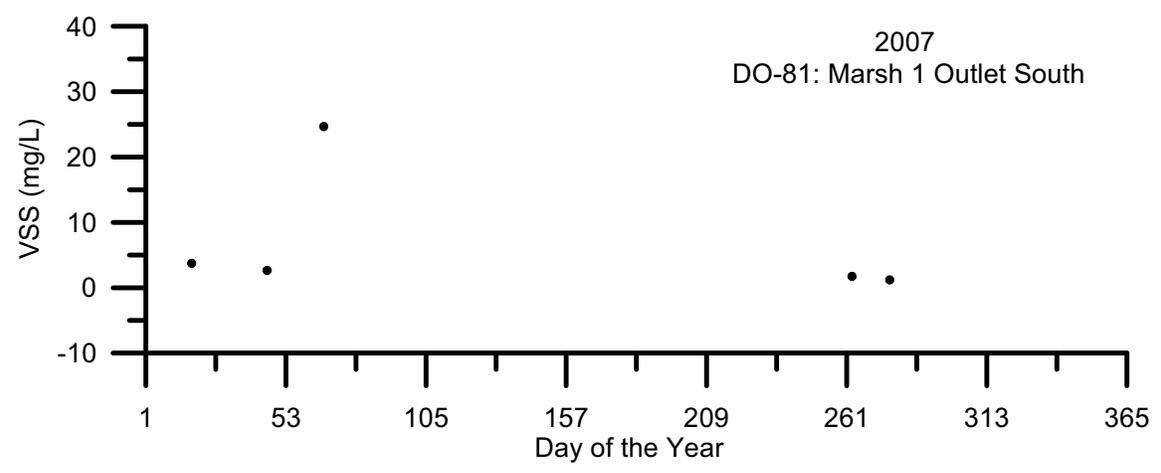
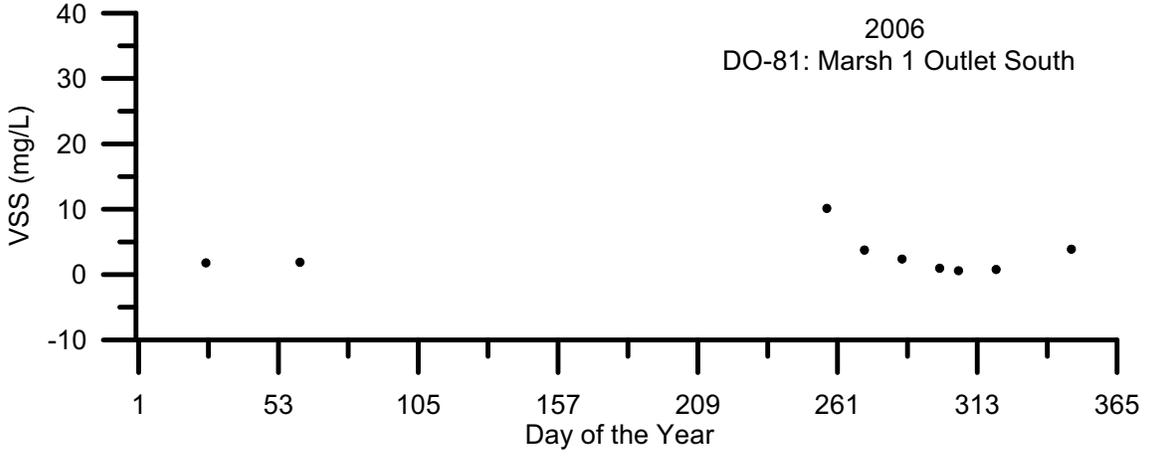
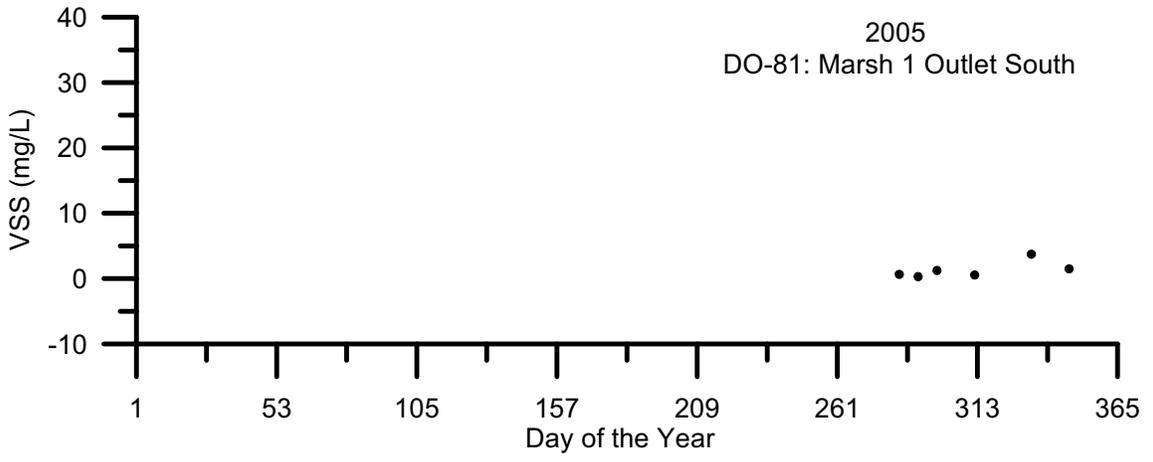


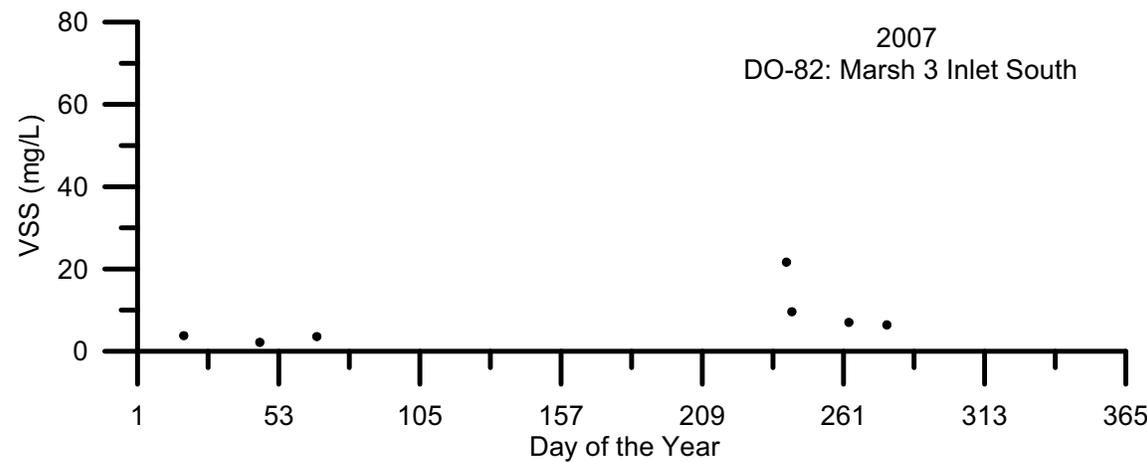
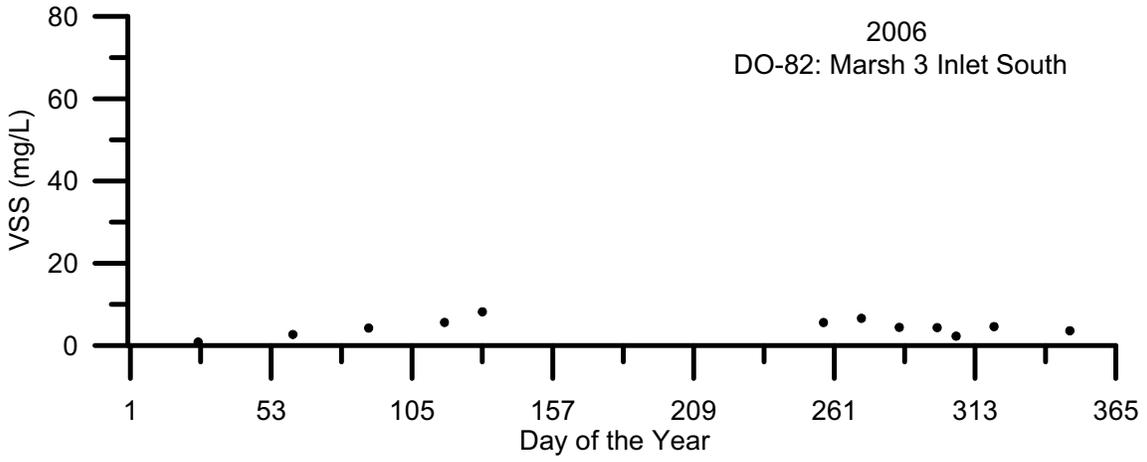
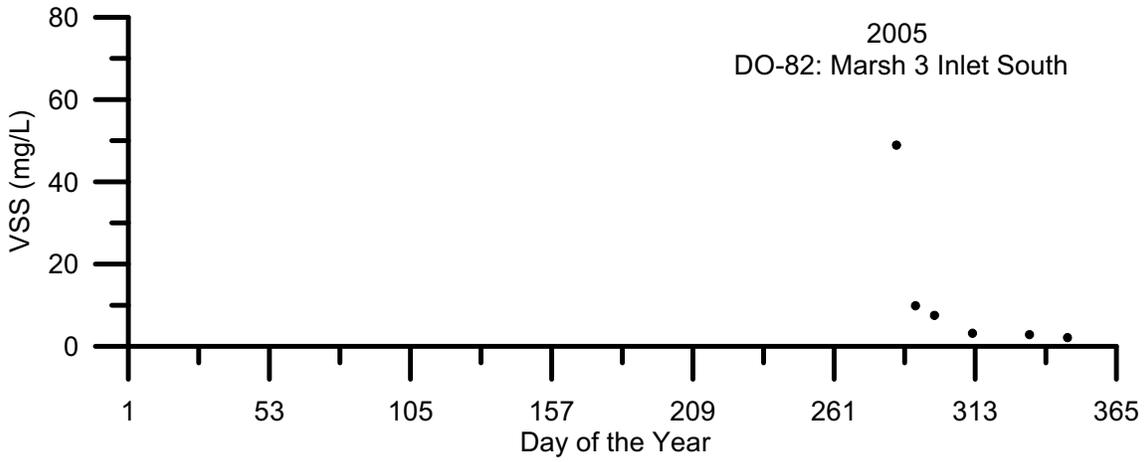


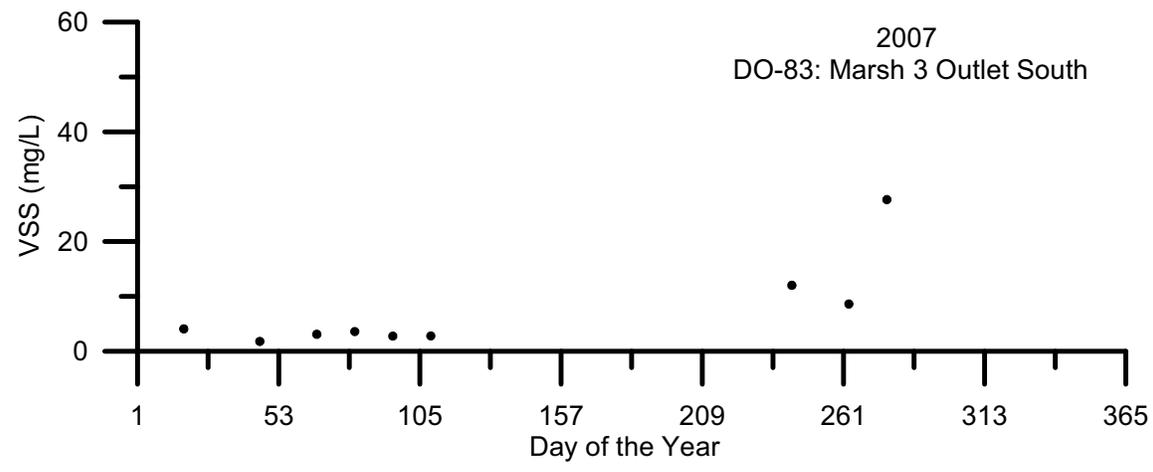
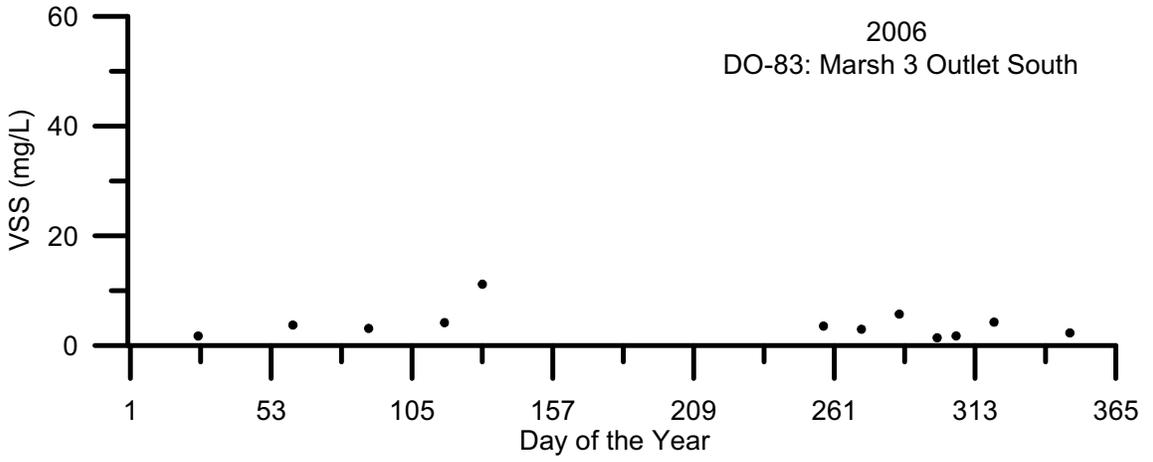
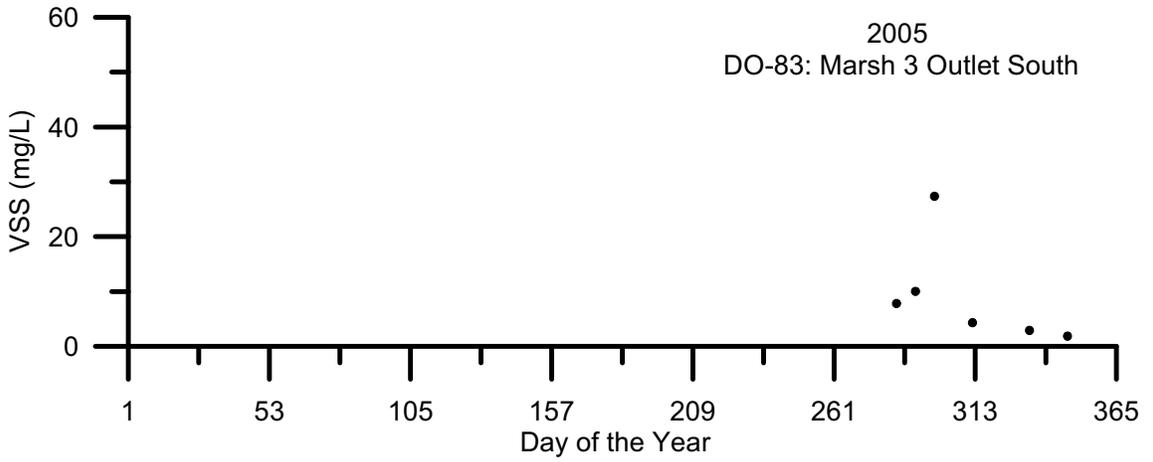


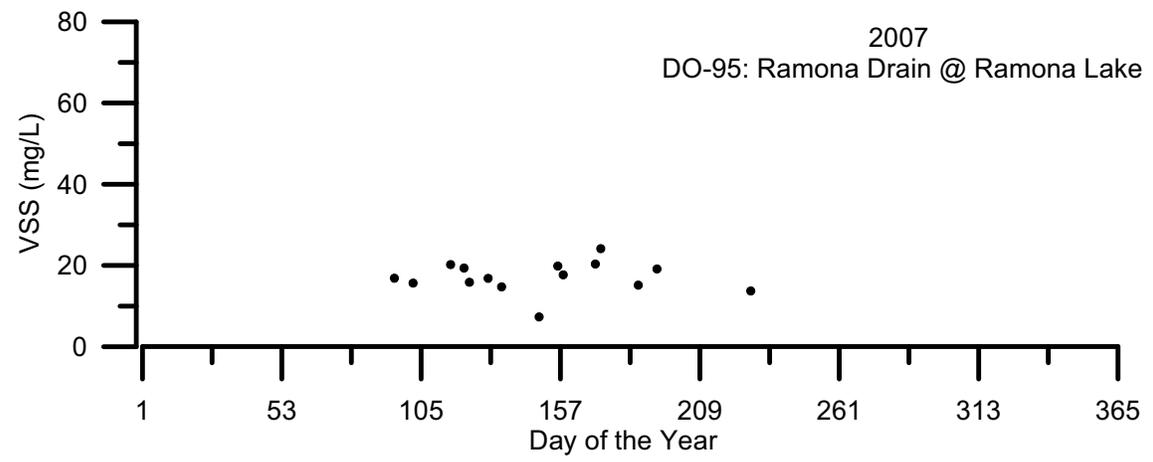
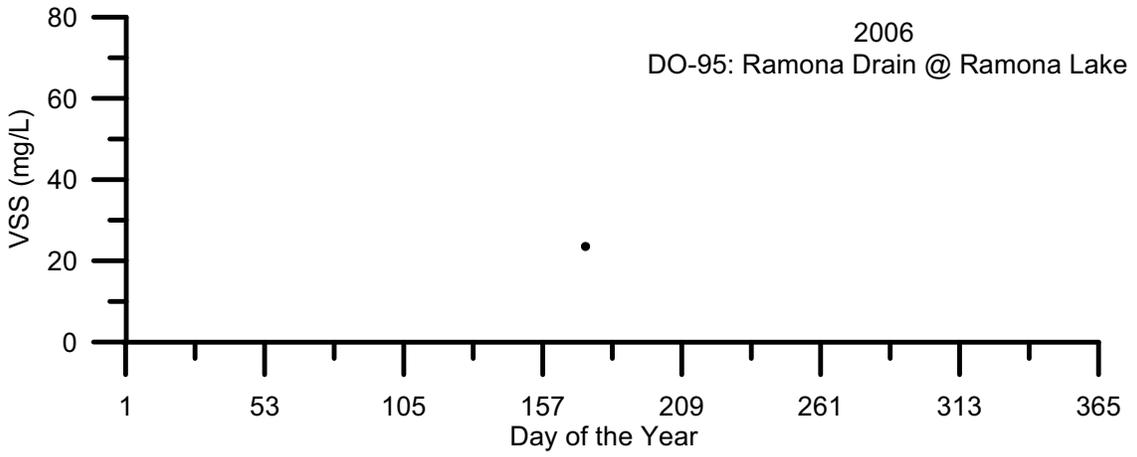














**Analysis of Dissolved Organic Carbon in Addition
to Volatile Suspended Solid (VSS + DOC)
Concentrations, as a Measure of Total Organic
Solids in the San Joaquin River Watershed
2005-2007**

*Remie Burks
Chelsea Spier
Sharon Borglin
Jeremy Hanlon
Justin Graham
William Stringfellow*

February 2008

*Environmental Engineering Research Program
University of the Pacific
3601 Pacific Avenue, Sears Hall
Stockton CA 95211*

Introduction

The San Joaquin River (SJR) supports one of the most productive agricultural regions in the world and its productivity is heavily dependant on irrigated agriculture. A consequence of irrigated agriculture is the production of return flows conveyed down gradient drains that eventually discharge to surface waters. Agricultural drainage may have significant nutrient load and can impact algae growth and general water quality in the SJR. Individual farmers and agricultural organizations, such as drainage authorities, are in need of tools to manage the environmental impacts of agricultural activities (Stringfellow, 2008).

For the years 2005 through 2007, sites throughout the San Joaquin Valley watershed were sampled to assess the overall water quality in the region. One thousand nine hundred and ninety-six (1996) individual surface water samples were collected and analyzed and WQ was assessed at 113 sampling locations in the SJR basin (Borglin et al., 2008). Samples were processed and analyzed by the Environmental Engineering Research Program (EERP) laboratory at the University of the Pacific as well as at the University of California, Davis, Dahlgren Lab. This report presents temporal plots of volatile suspended solids in addition to dissolved organic carbon as a measure of total organic solids for all sites sampled in the SJR from 2005-2007.

Methods

Field sampling consisted of collecting water samples, measuring water quality with a YSI Sonde 6600 with MDS650 hand-held display, and recording of field conditions at sites within the study area per *EERP Field Protocol Book* (Graham, 2008).

Water samples were collected in glass 1000 mL bottles (Wheaton Science Products, Millville, NJ), 1000 mL HDPE Trace-Clean narrow mouth plastic bottles (VWR International), 250 mL HDPE Trace-Clean wide mouth plastic bottles (VWR International) as well as 40 mL trace clean vials with PTFE septa (IChem, Rockwood, TN) in accordance with requirements for different lab analysis and volume requirements.

Samples were immediately stored at 4 °C after sampling (cooler temperature was recorded in the lab upon delivery) and transported to the lab on the day of sampling.

Samples were received by the laboratory the same day they were sampled and stored at 4 °C until filtering and analysis. Samples were collected, preserved, stored, and analyzed by methods outlined in *Standard Methods for the Analysis of Water and Wastewater*, (APHA, 2005, 1998). Filtered water samples were analyzed for dissolved organic carbon (DOC) on a Teledyne-Tekmar Apollo 9000 (Mason, OH) by high temperature combustion according to SM 5310 B (APHA, 2005) and quantified on using a NDIR detector. DOC samples were preserved < pH 2 with concentrated H₃PO₄ and stored at 4 °C until analysis. The limit of detection for DOC is 1.00mg/L C.

Total suspended solids (TSS) and volatile suspended solids (VSS) were analyzed by SM 2540 D and E (APHA, 2005). Typically 1000 mL of sample was filtered on pre-weighed, pre-combusted, Whatman GF/F filters. The filters were placed in an aluminum dish and dried at 105 °C under vacuum to constant weight. After drying the filter and dish were allowed to cool in a dessicator and were weighed for TSS determination. The dried and

weighted filters were subsequently combusted at 550°C for 6 hours and reweighed for VSS determination. Mineral suspended solids (MSS) concentration was calculated by subtracting VSS from TSS.

Dissolved organic carbon results (mg/L C) were then added to volatile suspended solid values (mg/L) for the following samples as a measure of total organic solids for each sample.

Results and Discussion

Both volatile suspended solid (VSS) analysis and dissolved organic carbon analysis were performed routinely over the years 2005-2007 although using two different Apollo 9000HS instruments for DOC analysis. VSS analysis had a 94.44% passage rate while DOC had a 98.94% passage rate for all QA parameters over the three year period (Borglin et. al., 2008). Apollo 9000 high sensitivity instruments are challenging to maintain and on a few occasions sample sets were analyzed by outside laboratories due to malfunctions of the instrument that could not be repaired within the holding time for these samples.

References

Stringfellow, W.T., et al., (2008), *Evaluation of Vegetated Ditches, Ponds, and Wetlands as BMPs for Mitigating the Water Quality Impact of Irrigated Agriculture in the San Joaquin Valley*, University of the Pacific, Stockton, CA

Graham, J., Hanlon, J.S., Stringfellow, W.T., (2008), *EERP Field Protocol Book*, University of the Pacific, Stockton, CA.

Borglin, S., W. Stringfellow, J. Hanlon. (2005), *Standard Operating Procedures for the Up-Stream Dissolved Oxygen TMDL Project*, LBNL/Pub-937.

Borglin, S., Burks, R., Hanlon, J., Graham, J., Spier, C., Stringfellow, W., and Dahlgren, R., (2008) *Methods overview, quality assurance, and quality control*, University of the Pacific, Stockton, CA

American Public Health Association, (2005), *Standard Methods for the Examination of Water and Wastewater*, American Public Health Association, Washington, D.C.

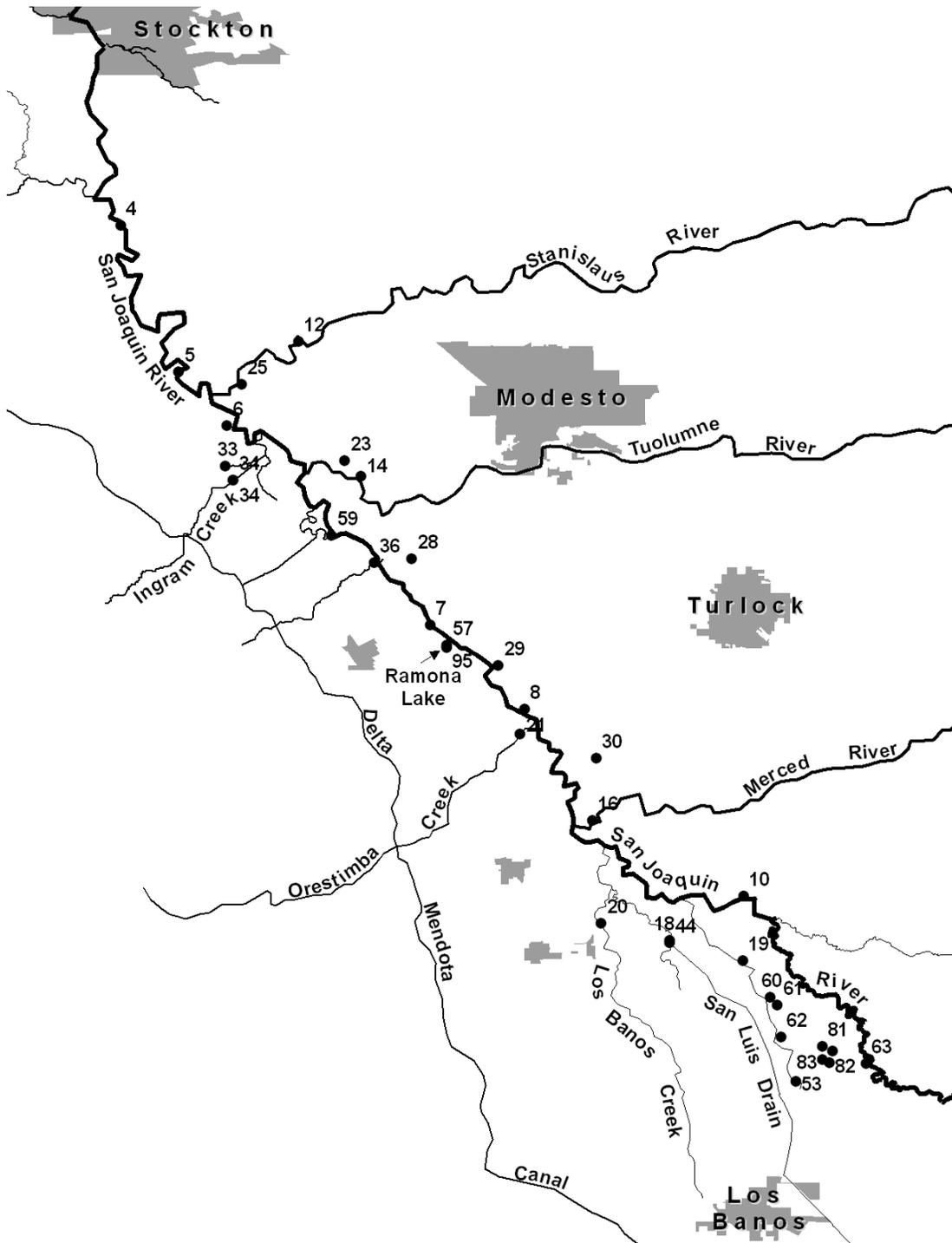
Borglin, S.E., Burks, R.D., Hanlon, J.S., Stringfellow, W.T. (2008) *EERP Lab Protocol Book*, University of the Pacific, Stockton, CA.

YSI Environmental Operations Manual, (2005), 6-Series Environmental Monitoring Systems, Yellow Springs, OH.

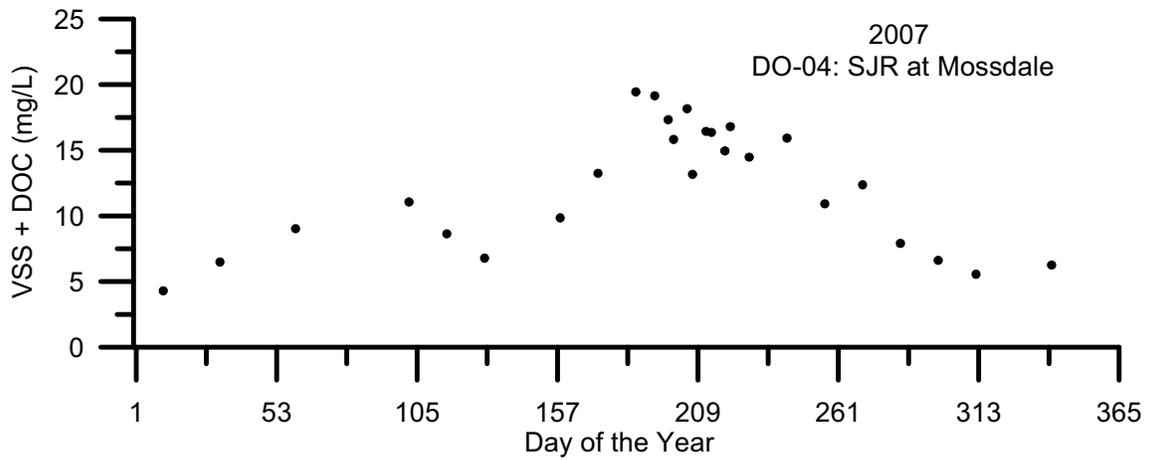
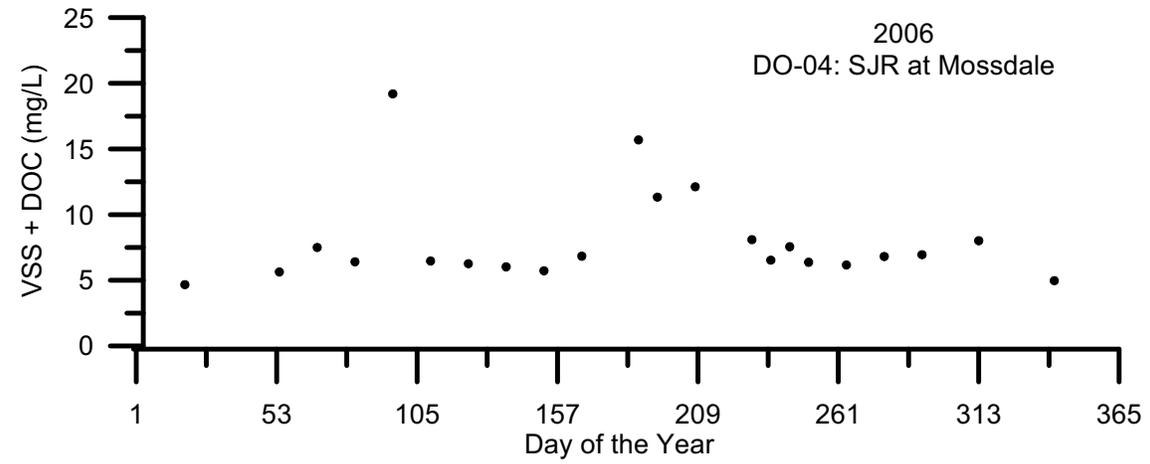
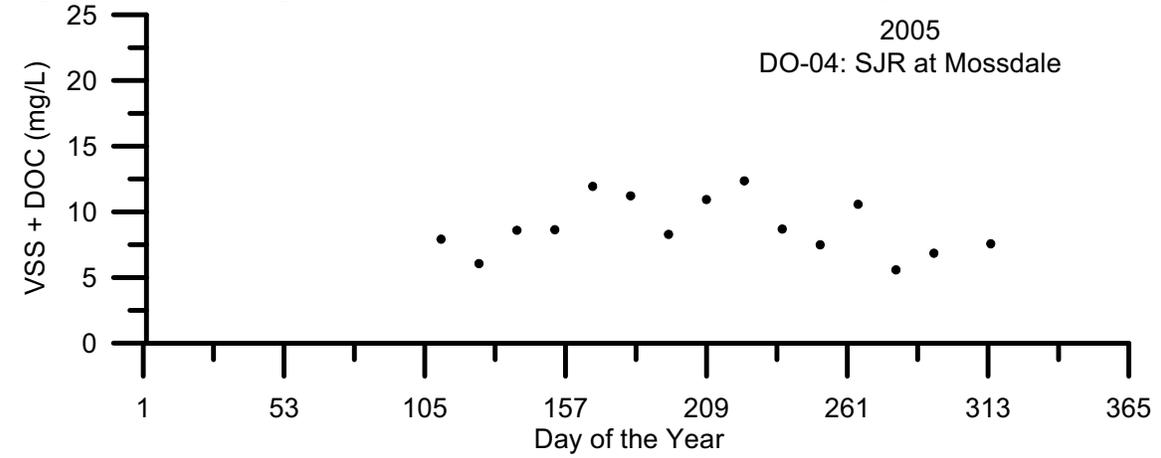
Table 1: EERP Sampling Site List

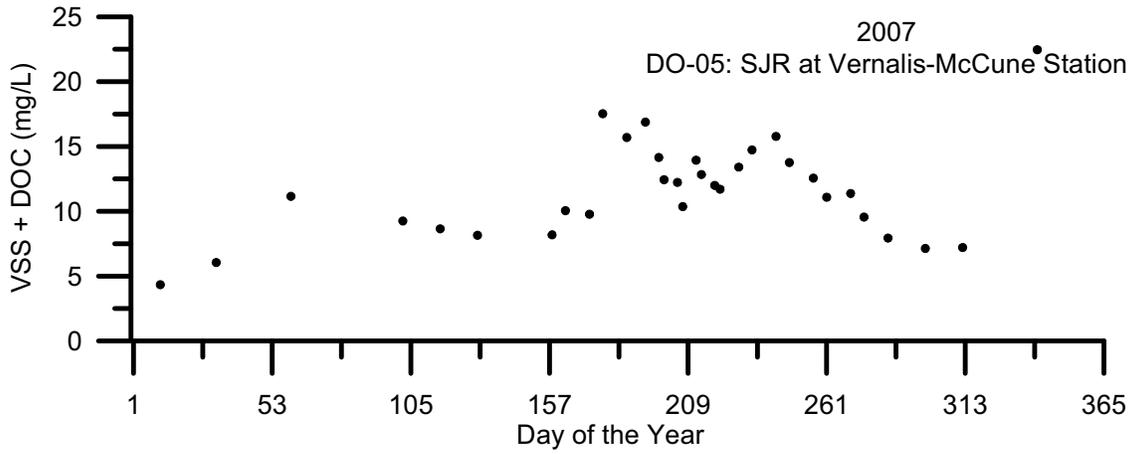
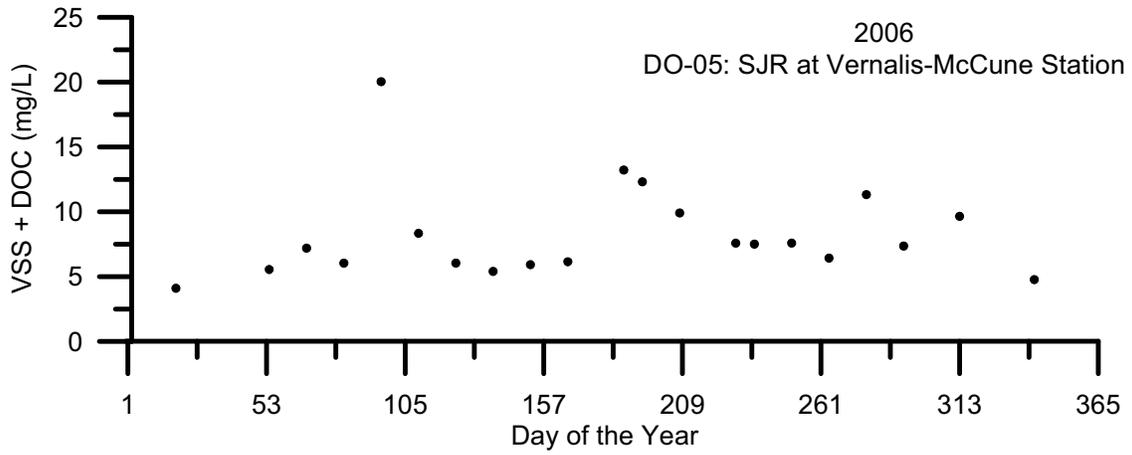
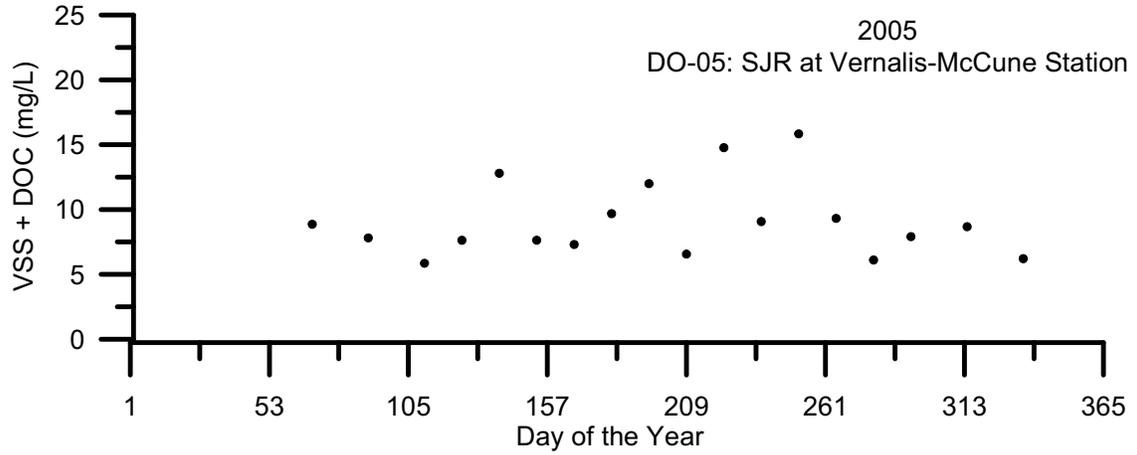
DO Number	Site Name	Type
4	SJR at Mossdale	Core sites
5	SJR at Vernalis-McCune Station (River Club)	Core sites, BMP
6	SJR at Maze	Core sites, BMP
7	SJR at Patterson	Core sites, BMP
8	SJR at Crows Landing	Core sites, BMP
10	SJR at Lander Avenue	Core sites
12	Stanislaus River at Caswell Park	Core sites
14	Tuolumne River at Shiloh Bridge	Core sites
16	Merced River at River Road	Core sites
18	Mud Slough near Gustine	Core sites, Wetland
19	Salt Slough at Lander Avenue	Core sites, Wetland
20	Los Banos Creek Flow Station	Core sites, Wetland
21	Orestimba Creek at River Road	Core sites, BMP
23	Modesto ID Lateral 5 to Tuolumne	Core sites
25	Modesto ID Main Drain to Stan. R. via Miller Lake	Core sites
28	Turlock ID Westport Drain Flow station	Core sites
29	Turlock ID Harding Drain	Core sites
30	Turlock ID Lateral 6 & 7 at Levee	Core sites
33	Hospital Creek	Intermittent, BMP
34	Ingram Creek	Core sites, BMP
36	Del Puerto Creek Flow Station	Core sites, BMP
44	San Luis Drain End	Core sites
53	Salt Slough at Wolfsen Road	Wetland
57	Ramona Lake Drain	Core sites, BMP
59	SJR Laird Park	Core sites
60	Moffit 1 South	Wetland
61	Deadman's Slough	Wetland
62	Mallard Slough	Wetland
63	Inlet C Canal	Wetland
80	South Marsh-1-Inlet	Wetland
81	South Marsh-1-Outlet	Wetland
82	South Marsh-3-Inlet	Wetland
83	South Marsh-3-Outlet	Wetland
95	Ramona drain at Ramona Lake	BMP, Intermittent

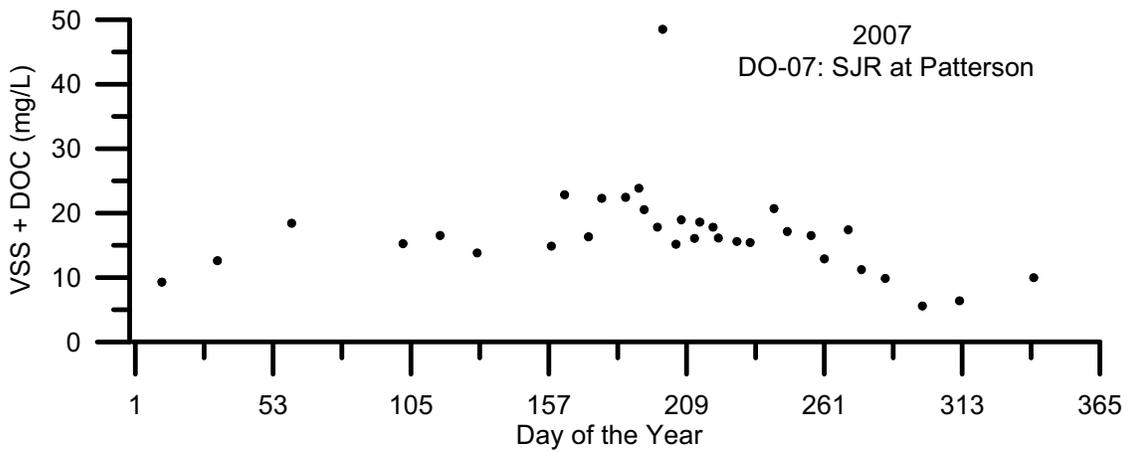
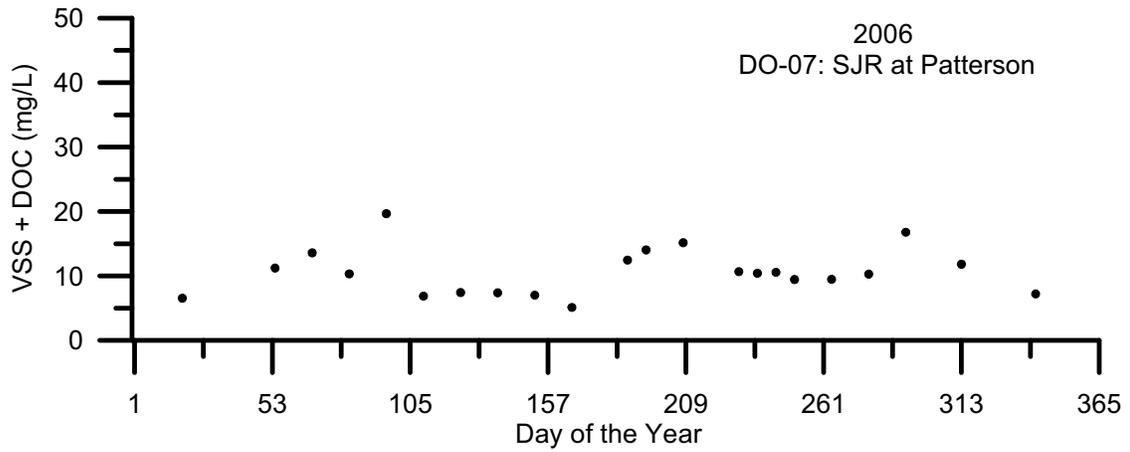
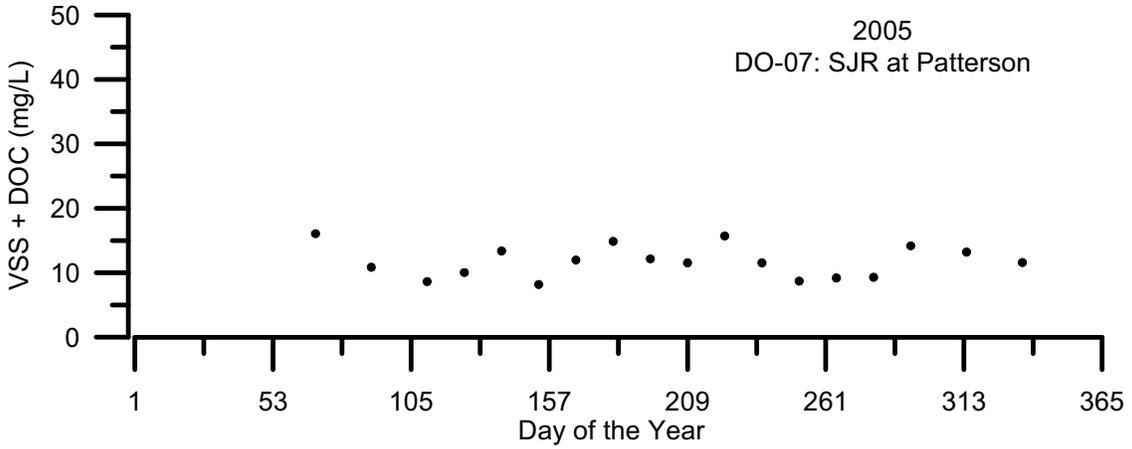
Figure 1: EERP Sampling Site Map of SJR Watershed and Tributaries

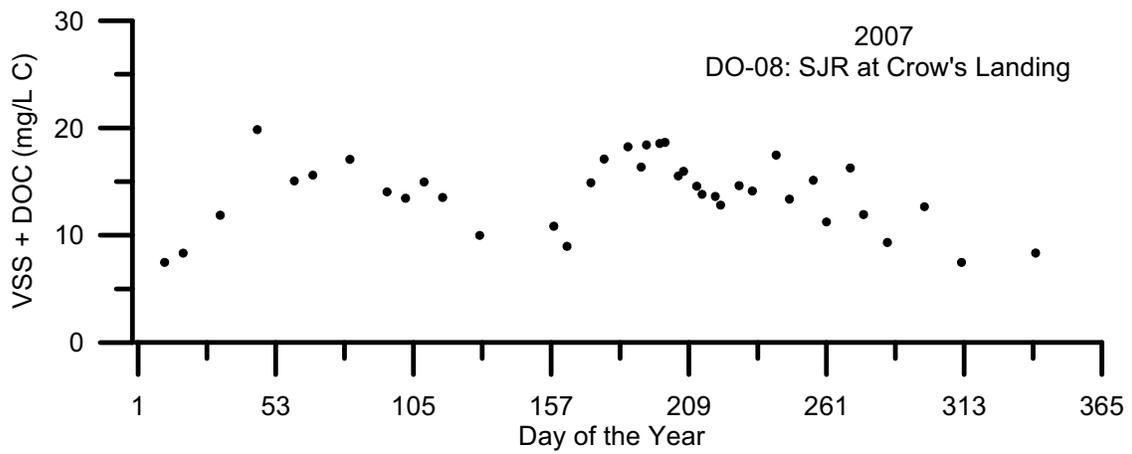
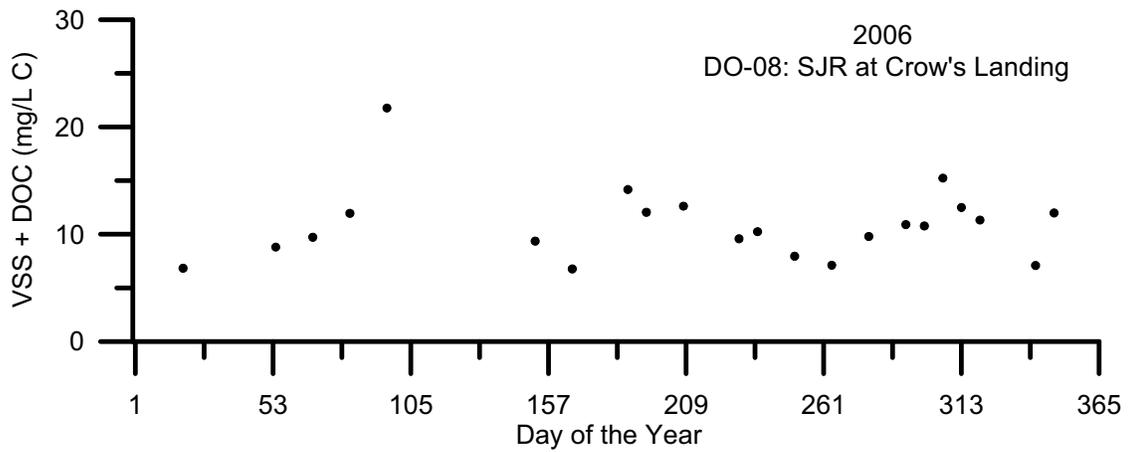
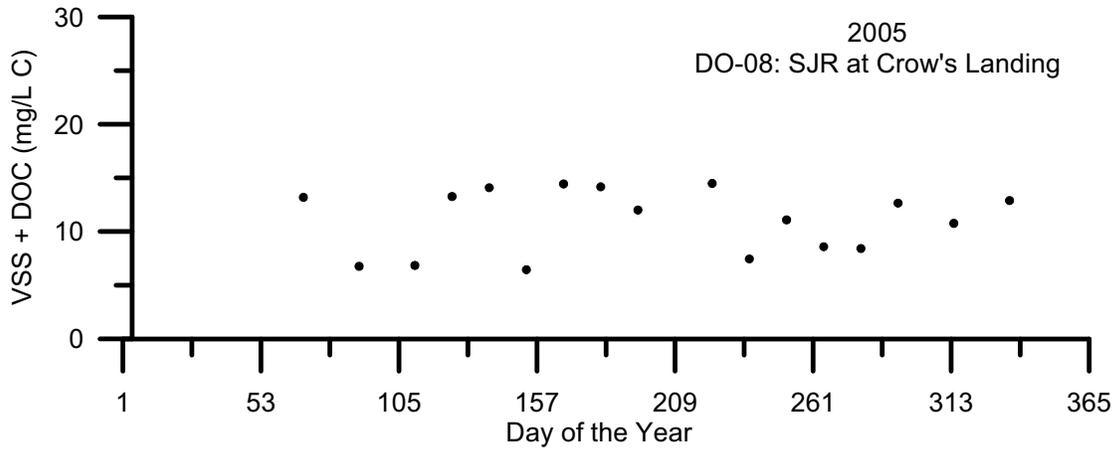


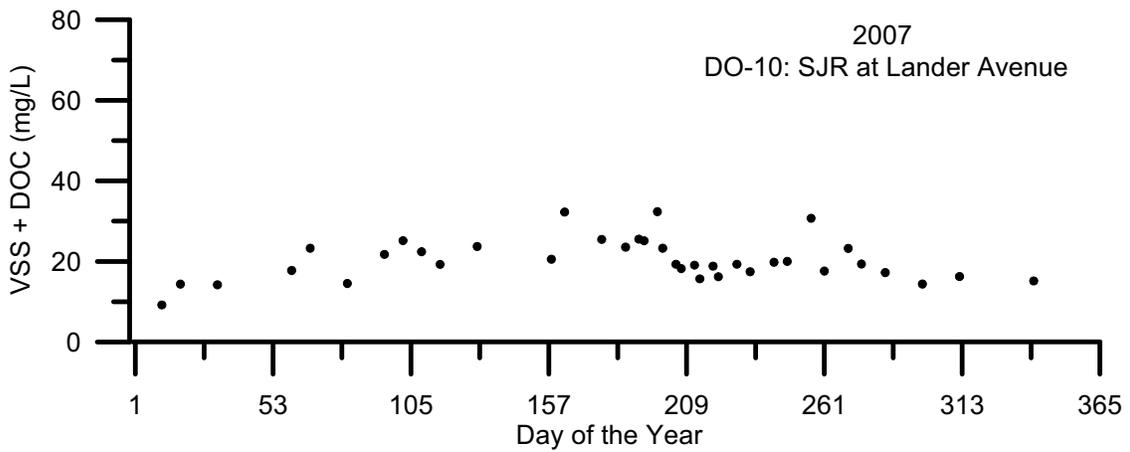
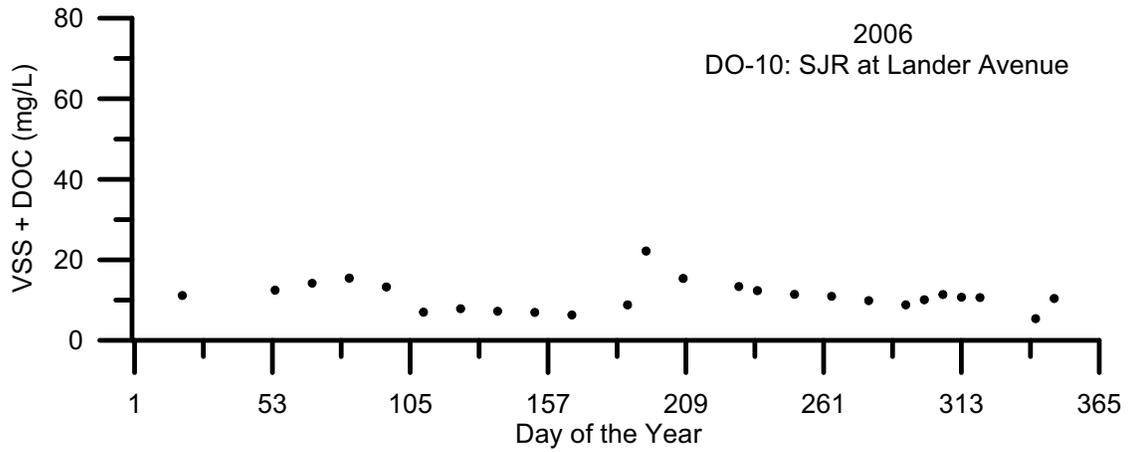
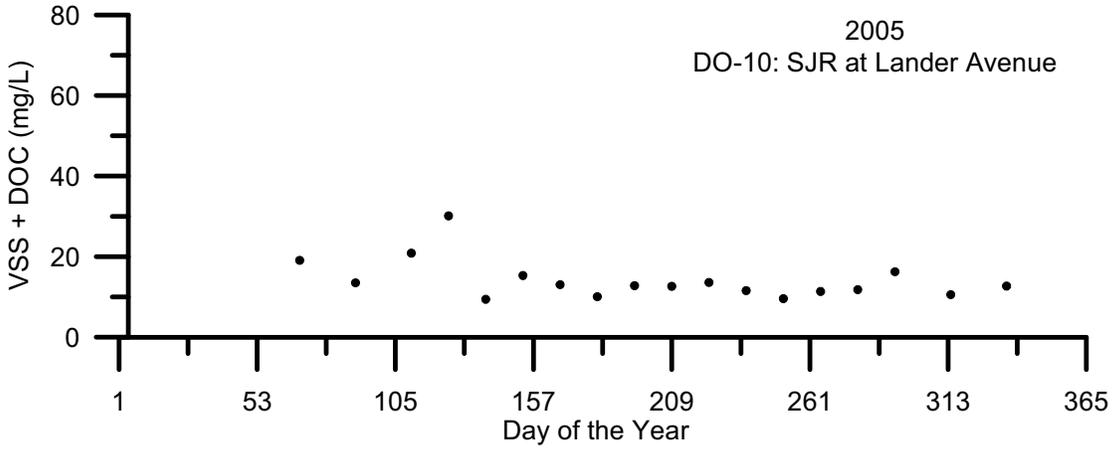
Figures 2 -101: Temporal Plots of VSS + DOC Concentrations By Site ID

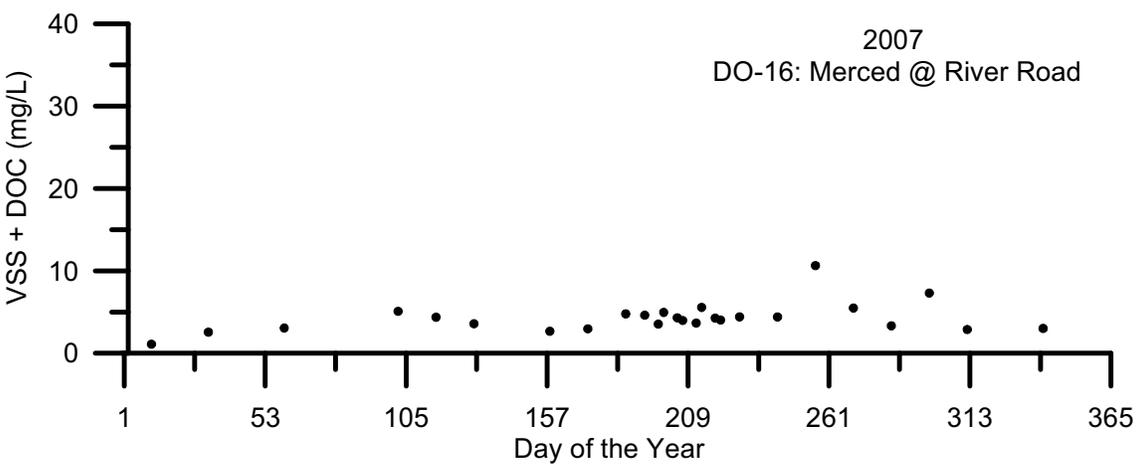
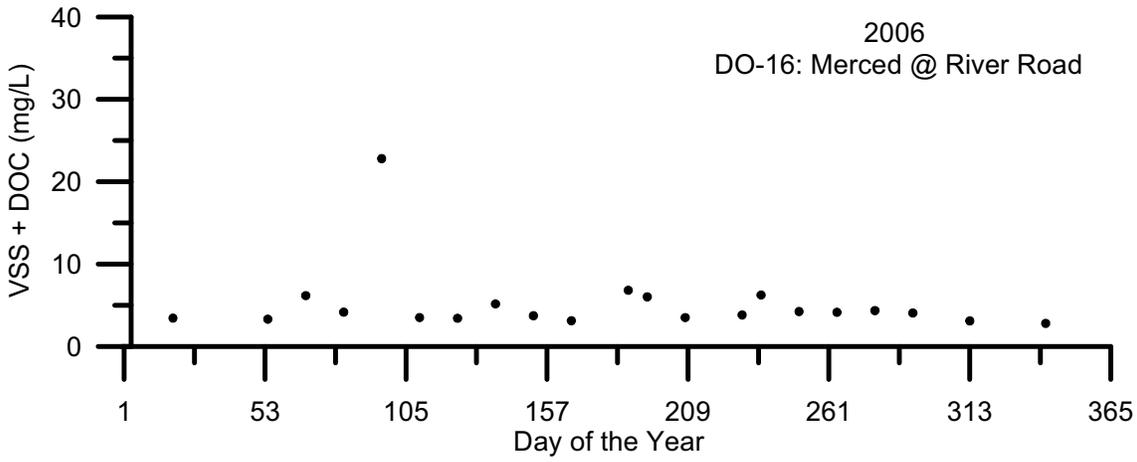
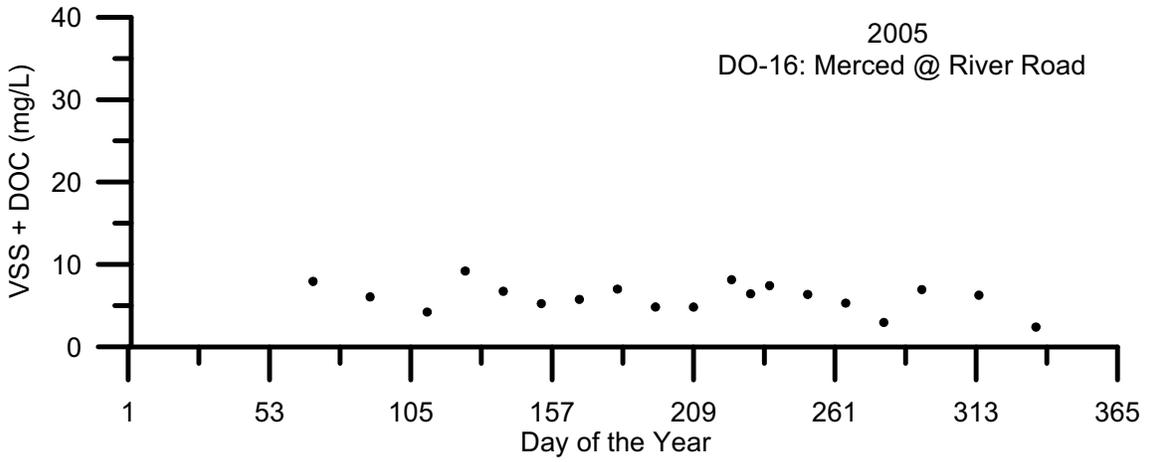


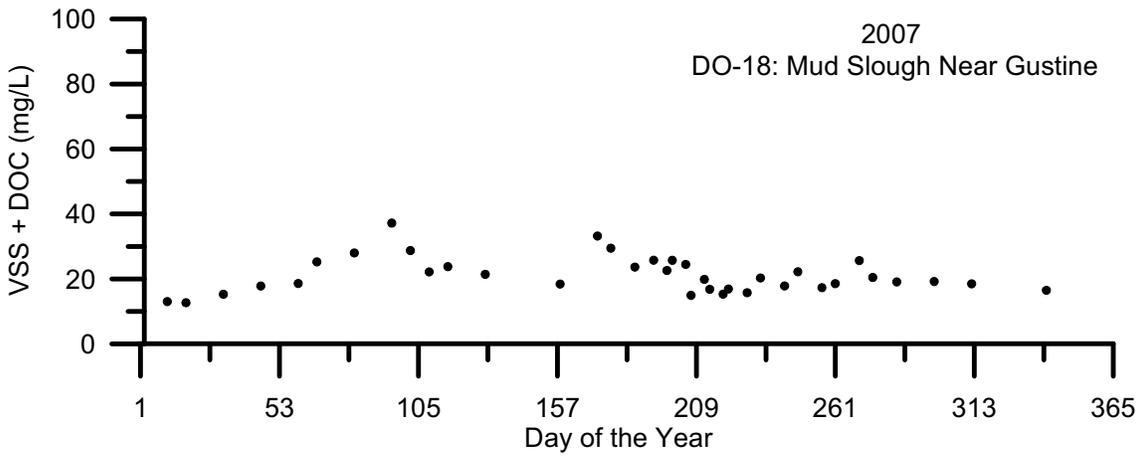
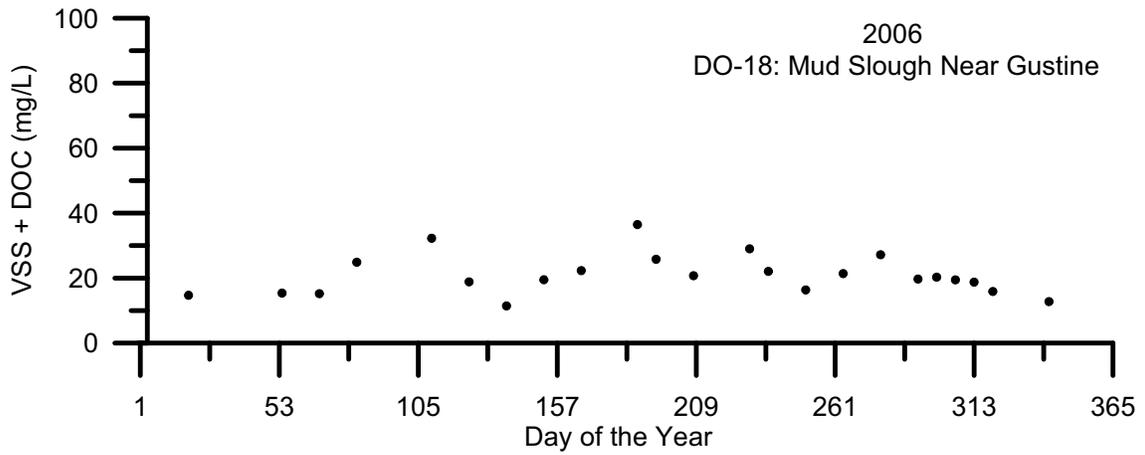
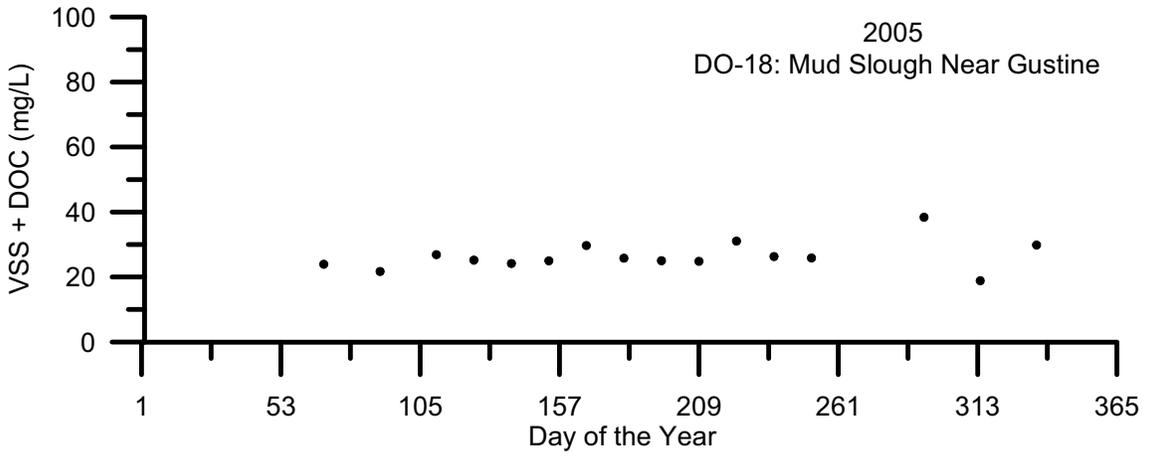


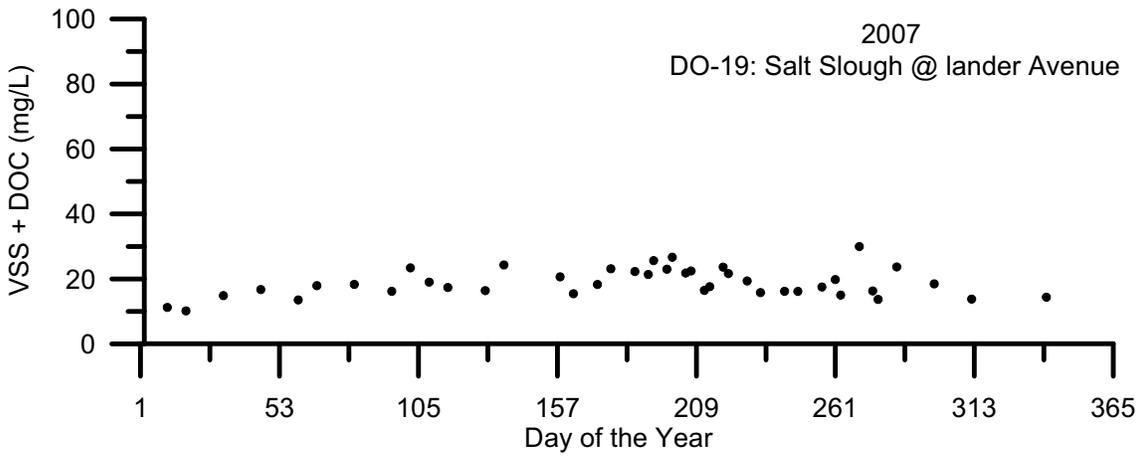
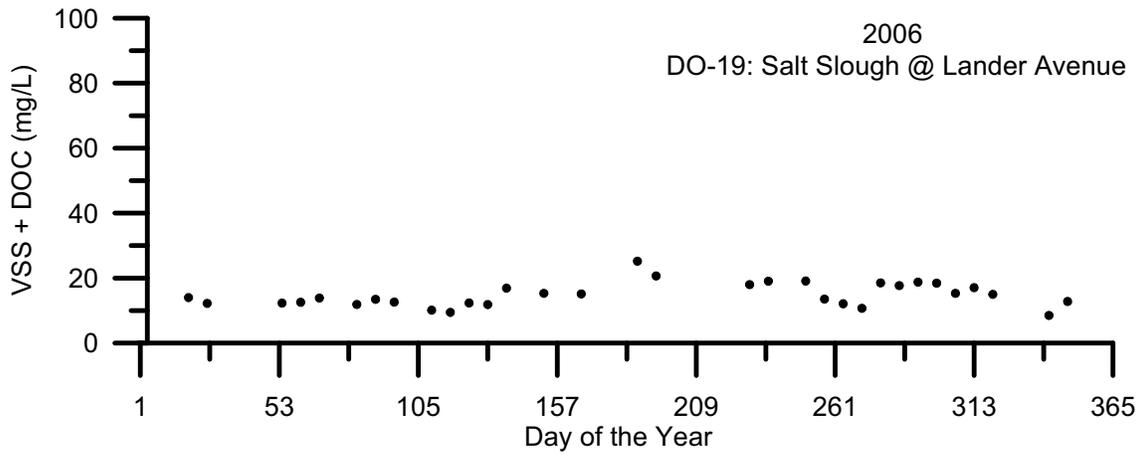
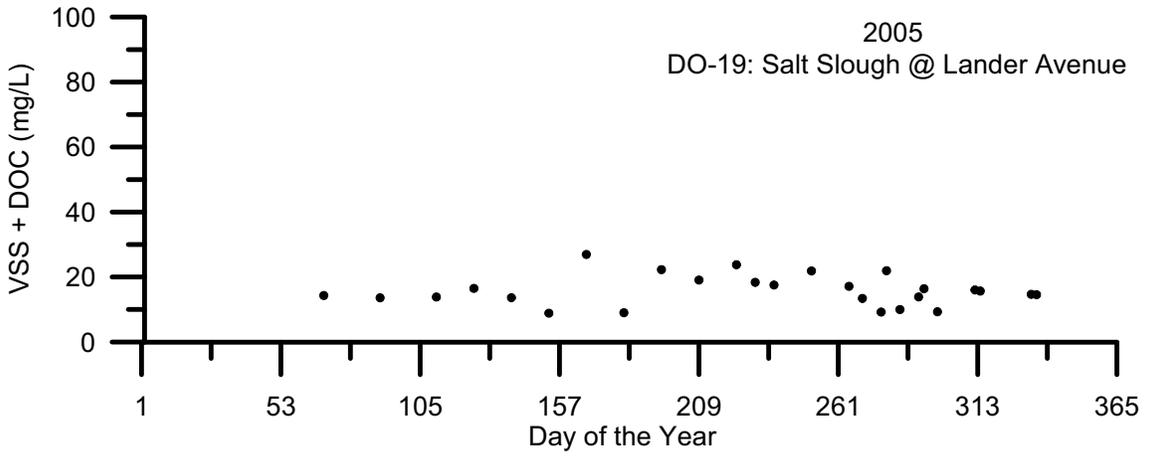


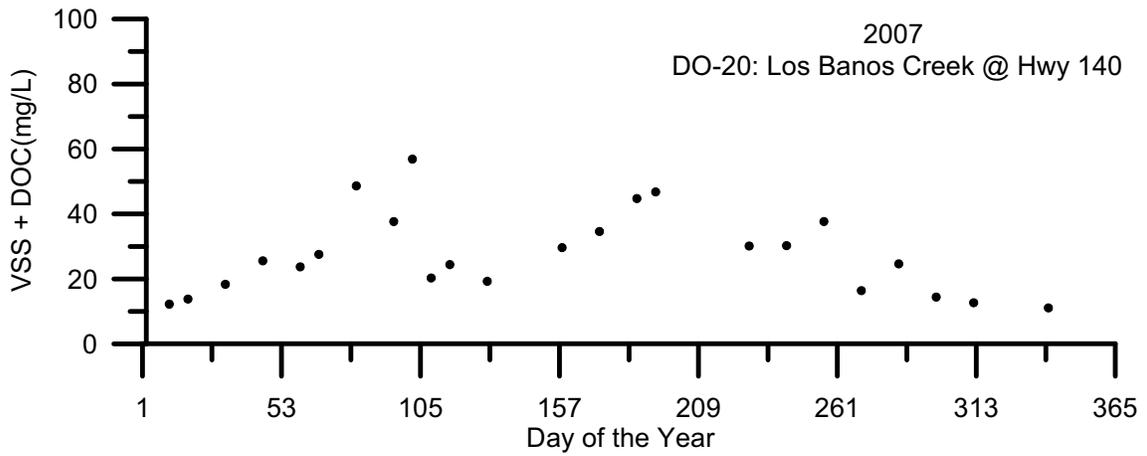
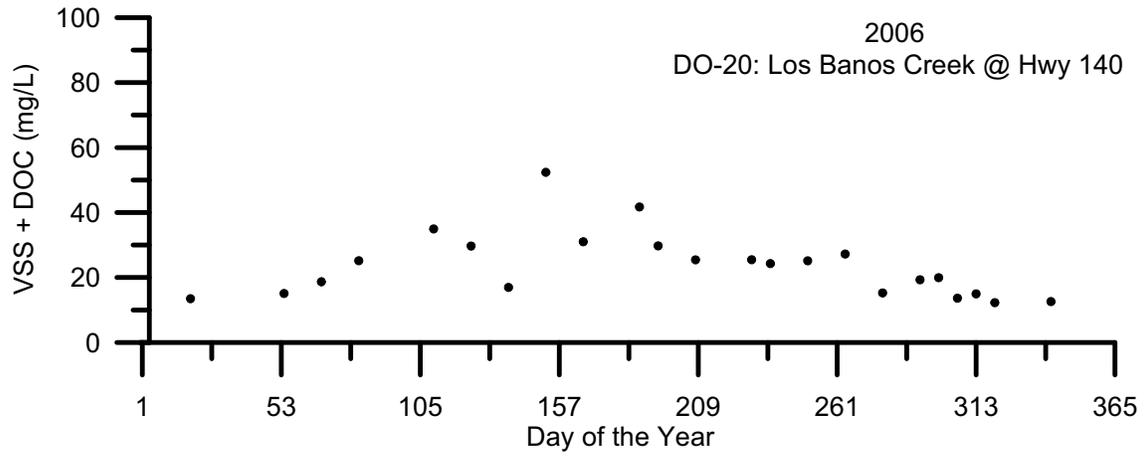
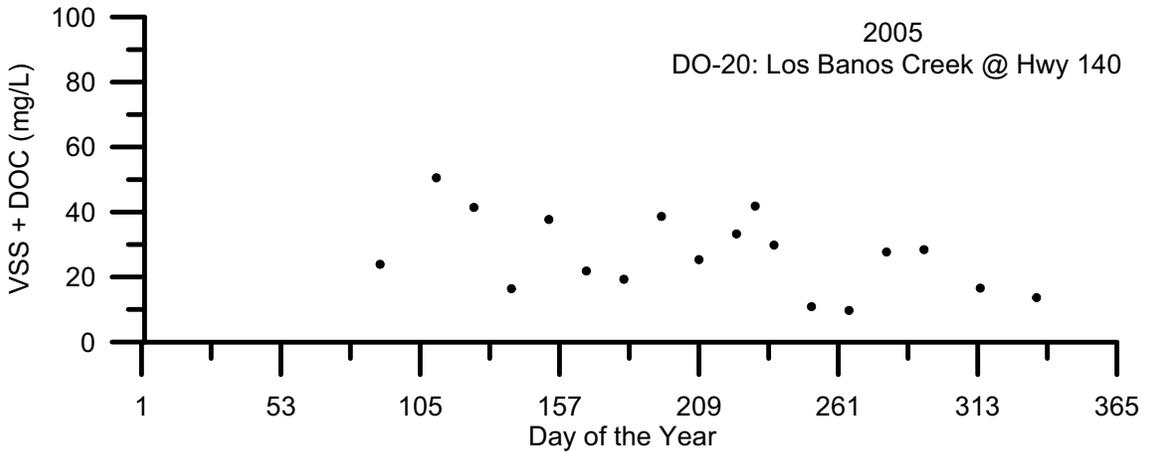


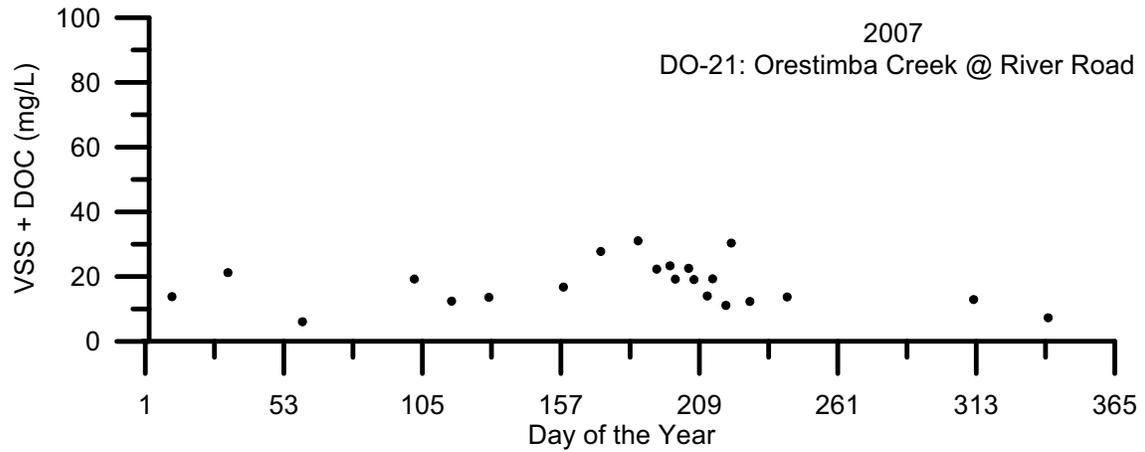
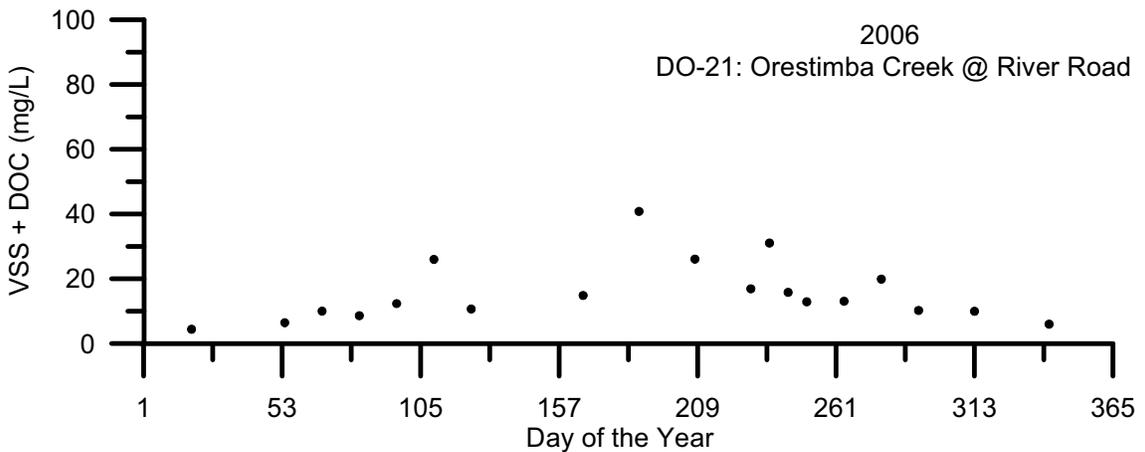
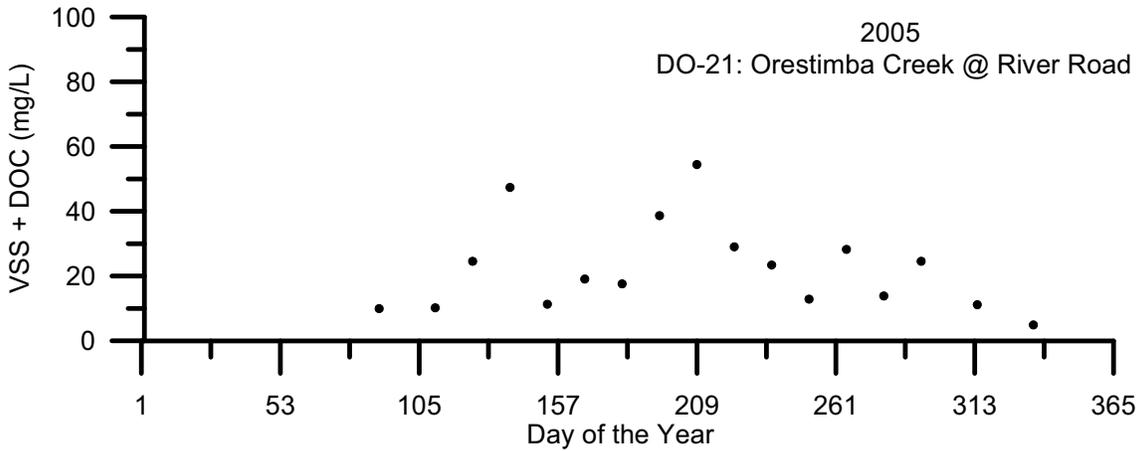


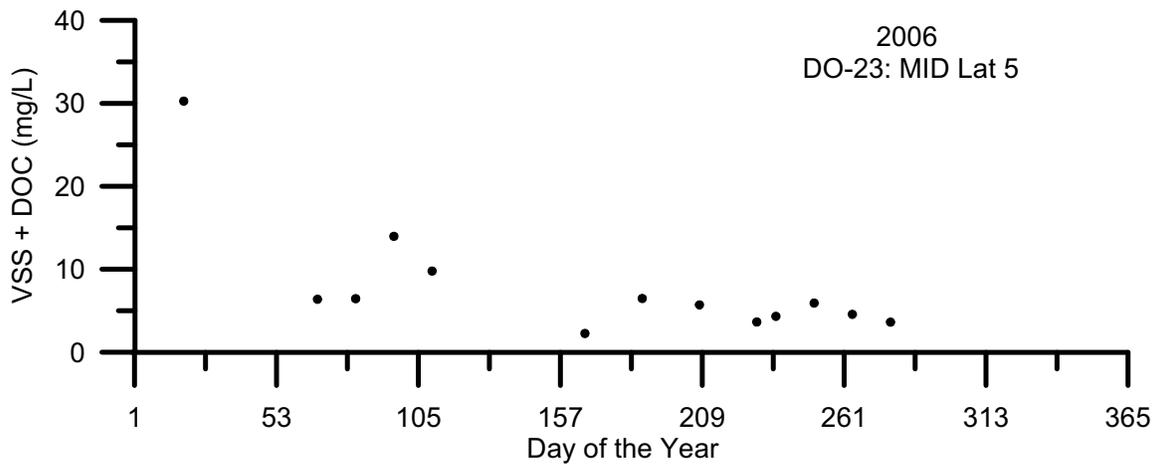
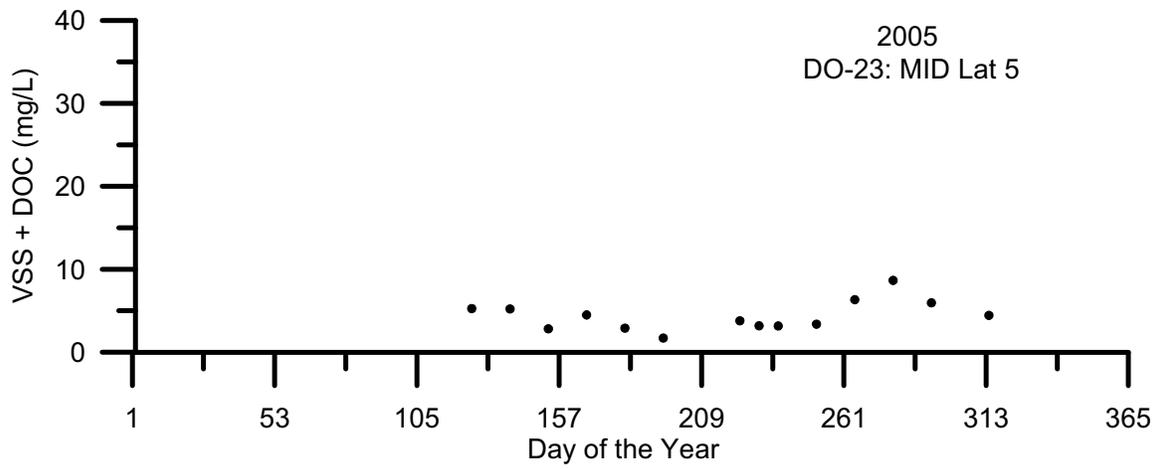


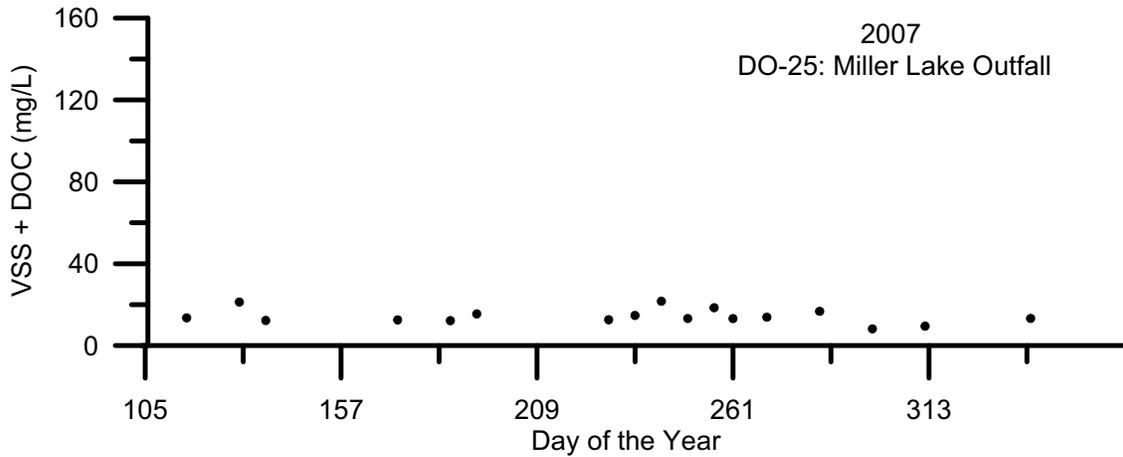
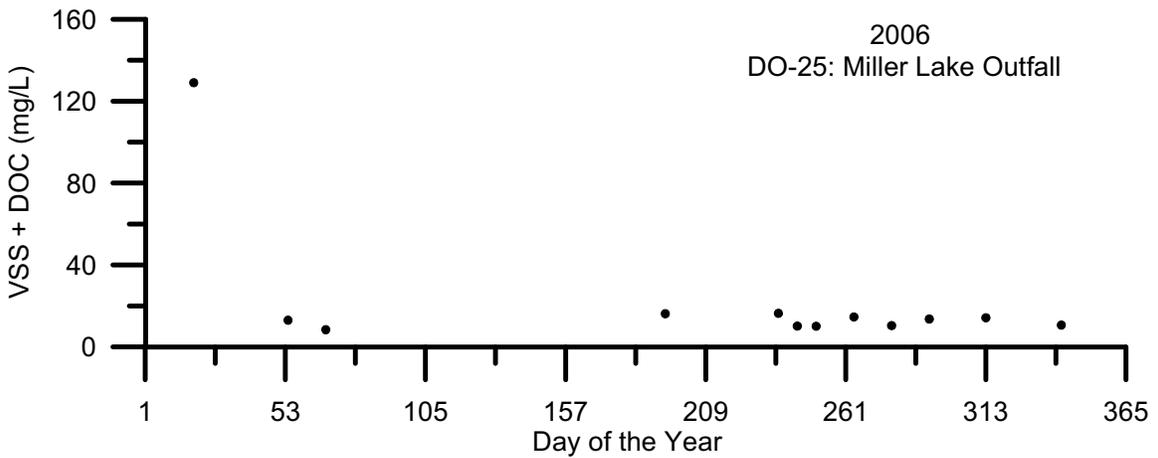
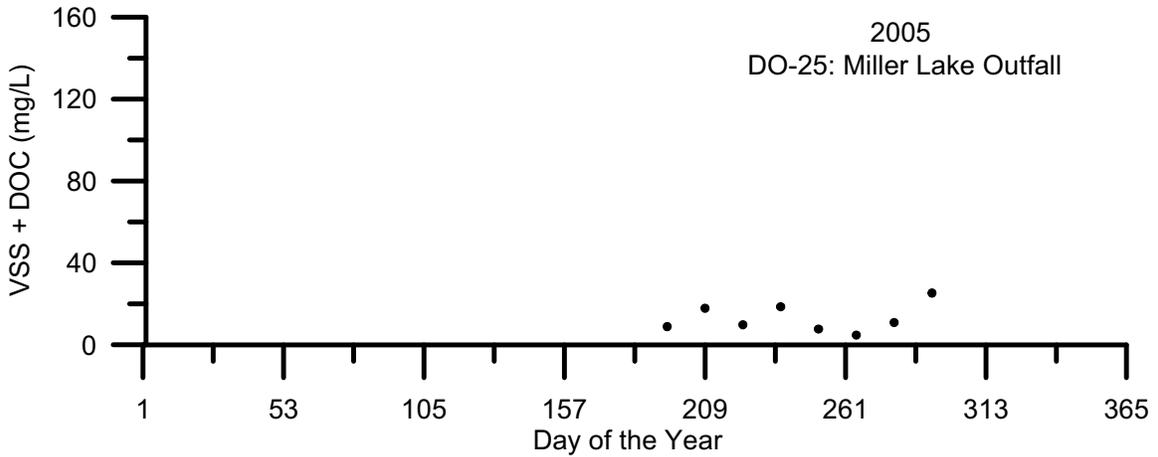


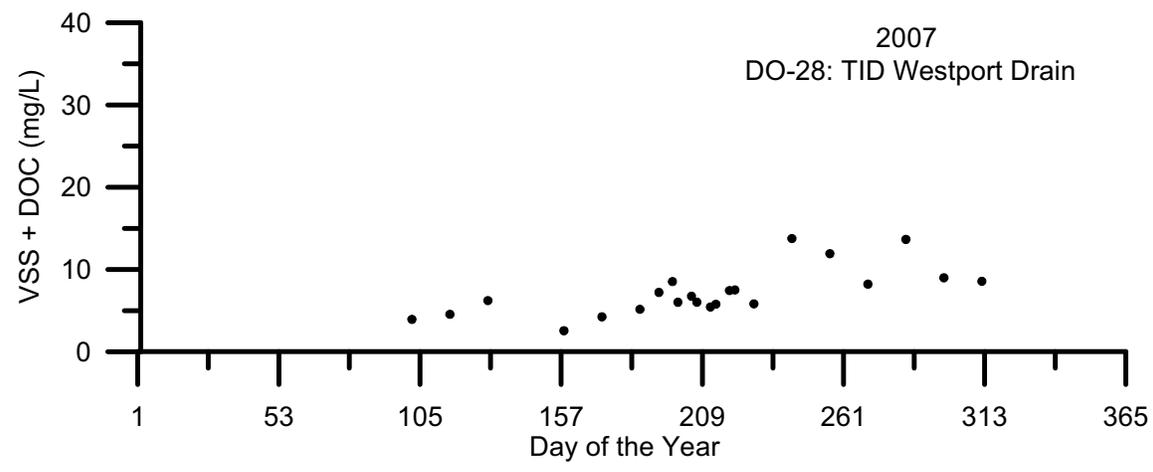
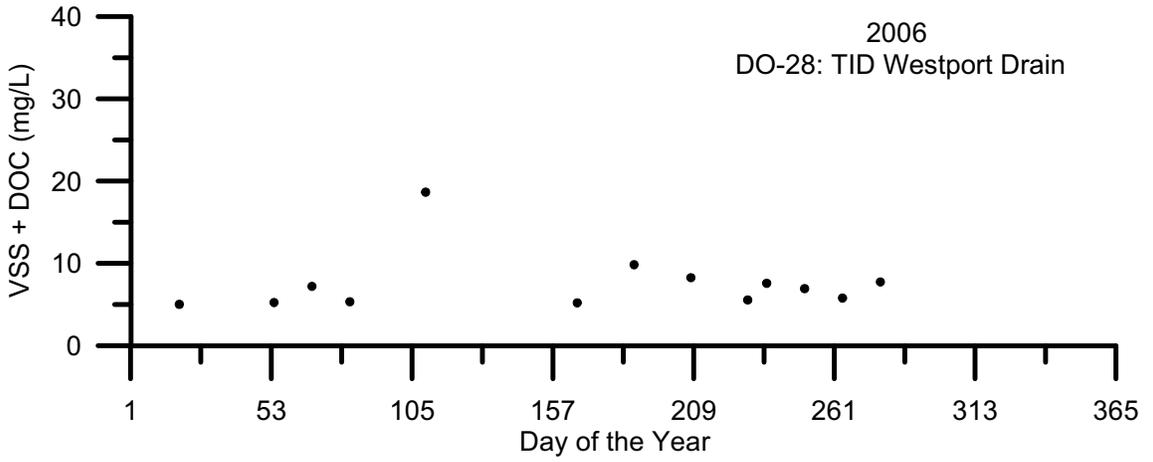
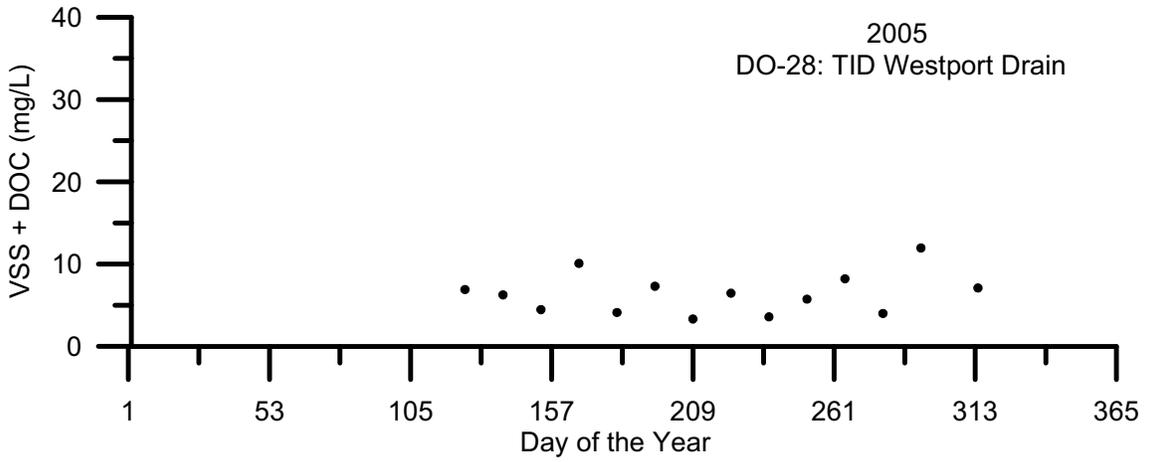


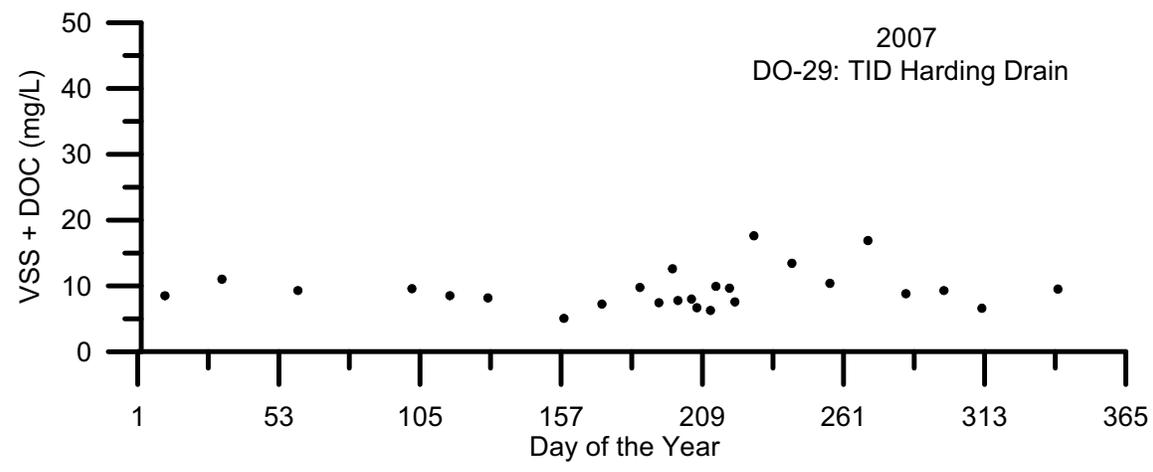
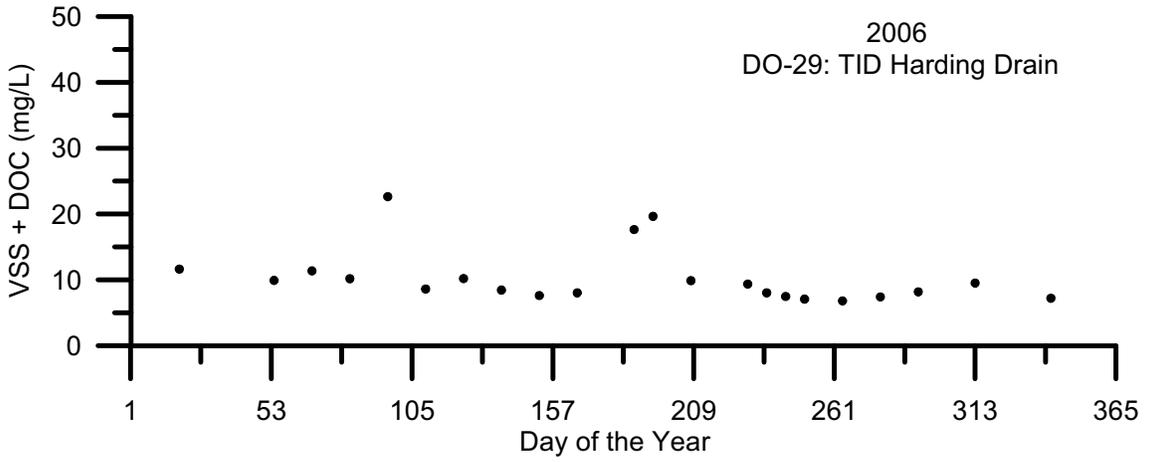
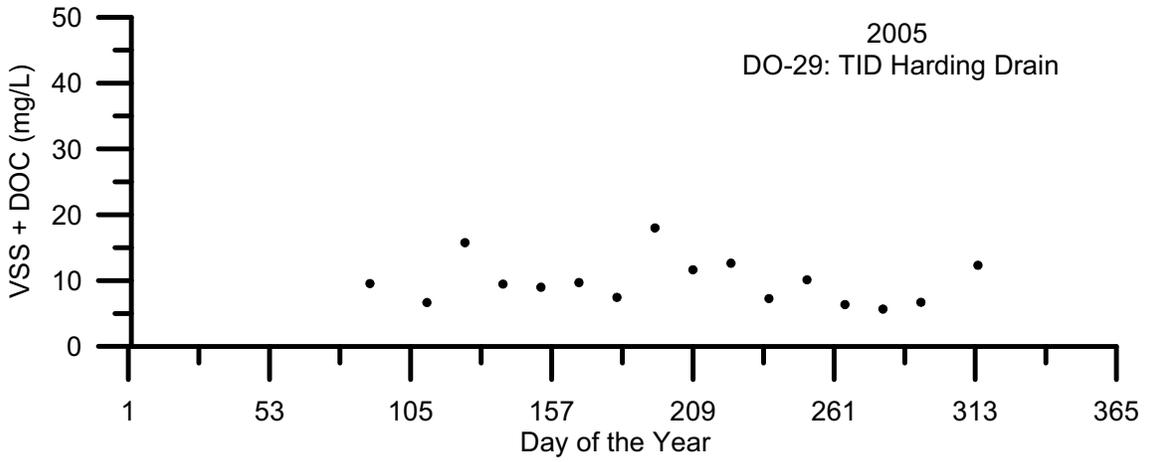


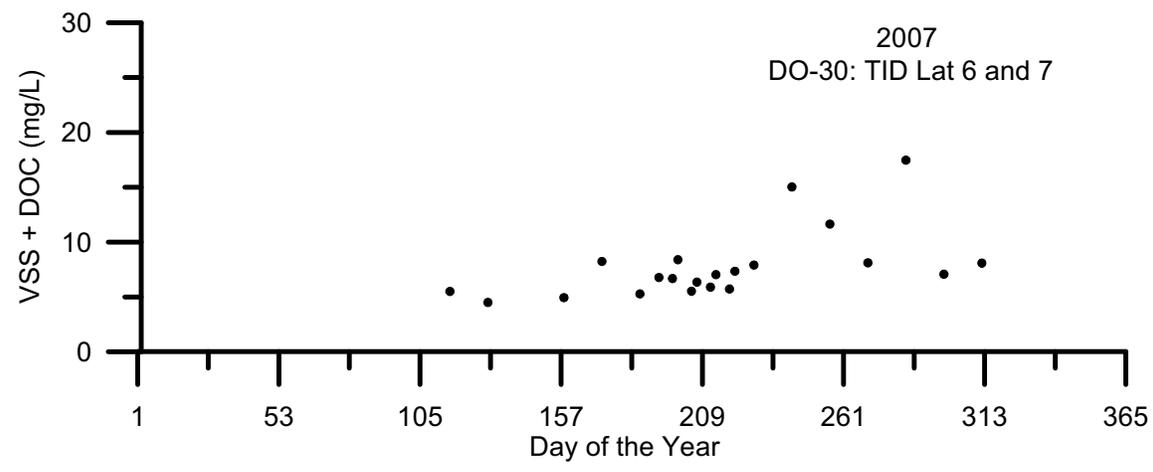
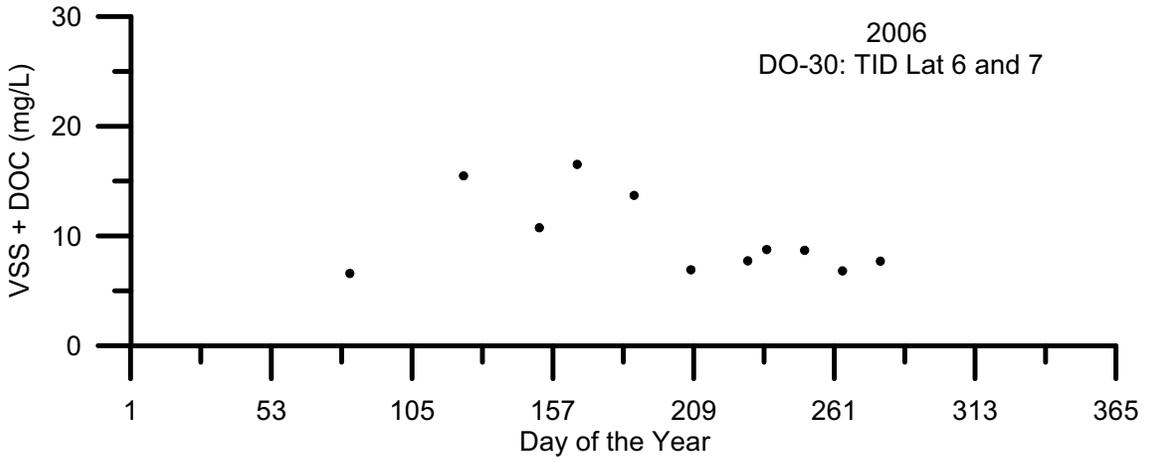
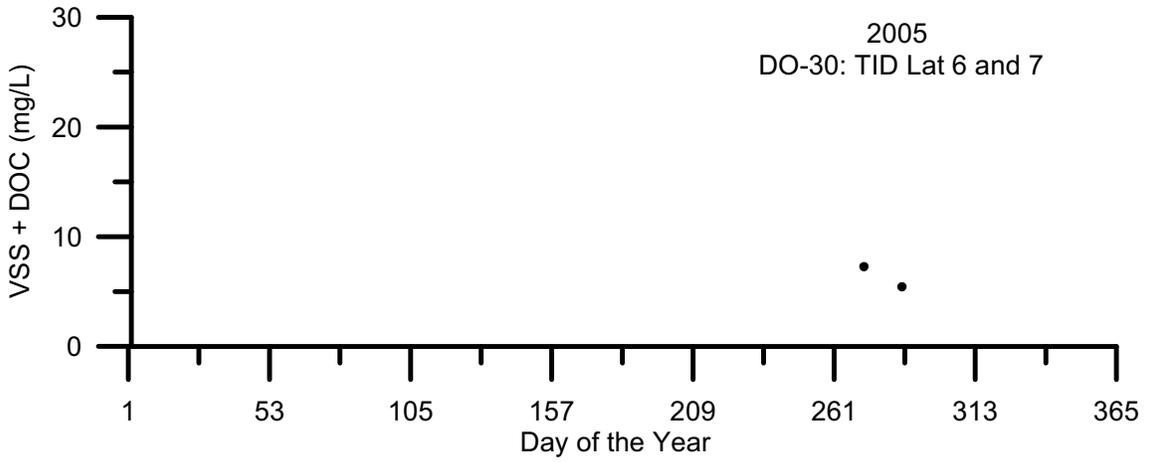


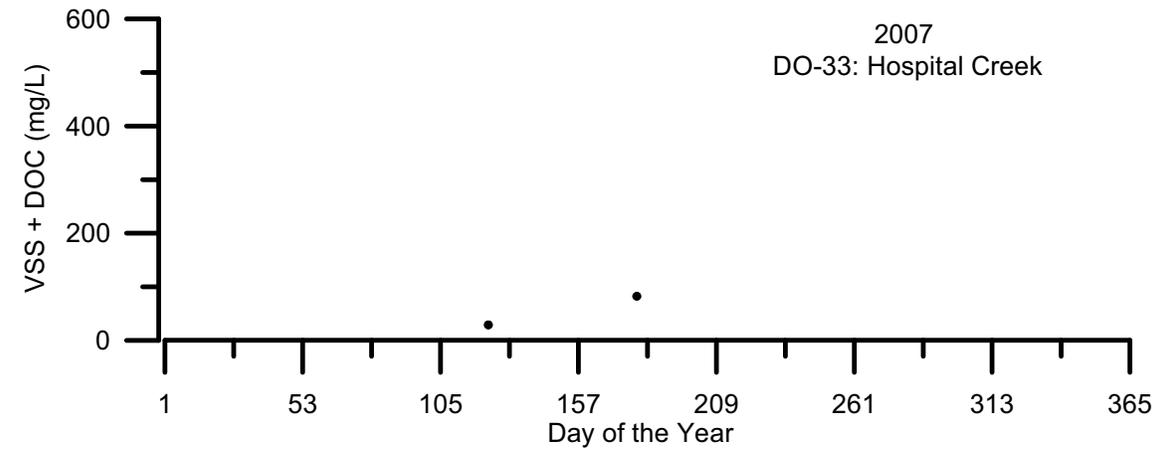
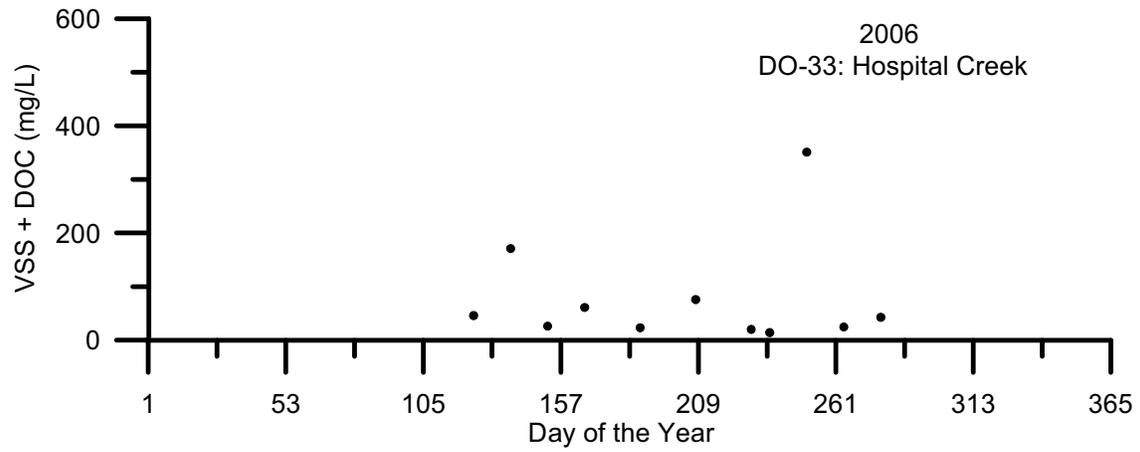
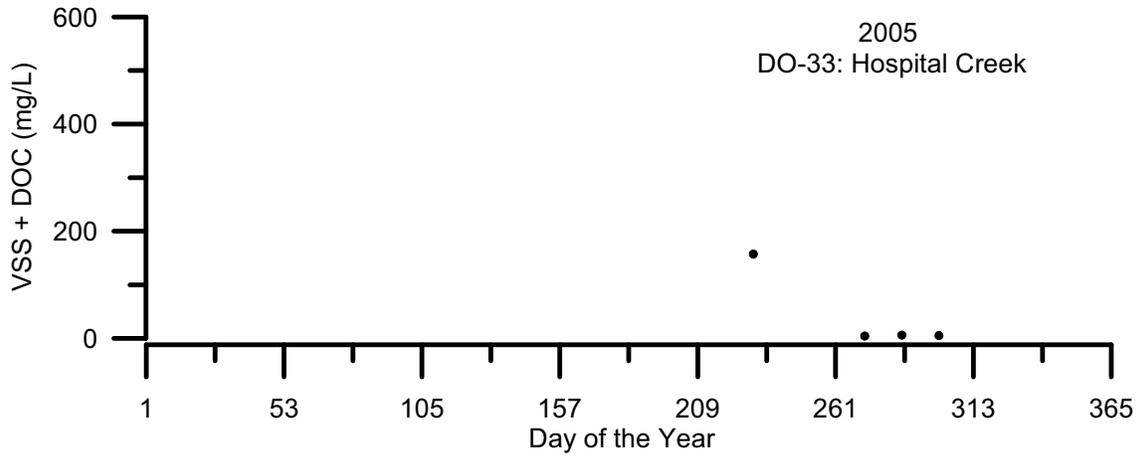


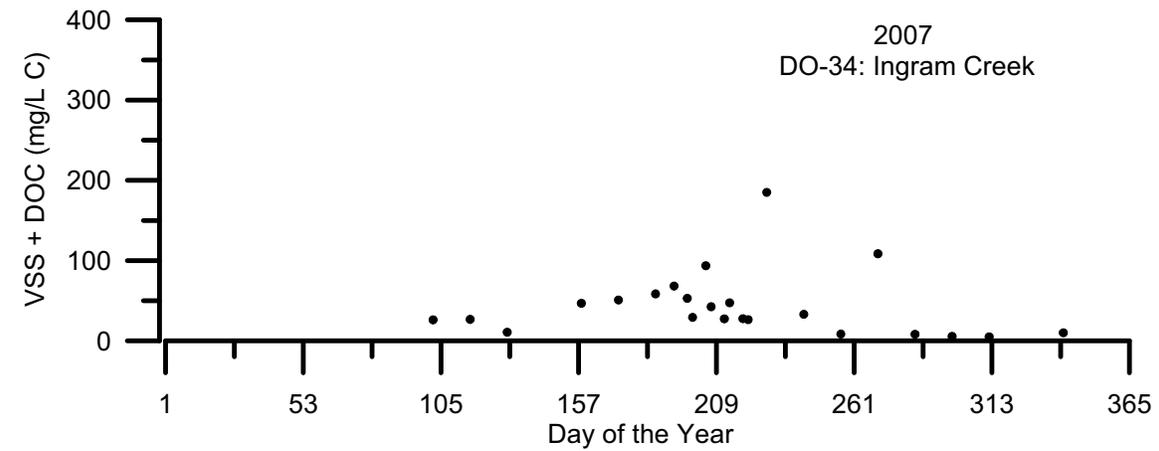
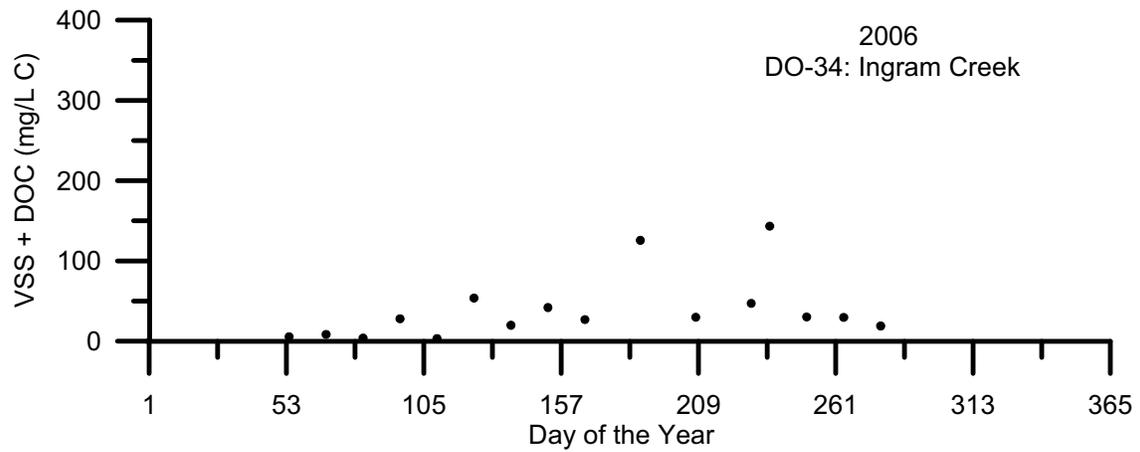
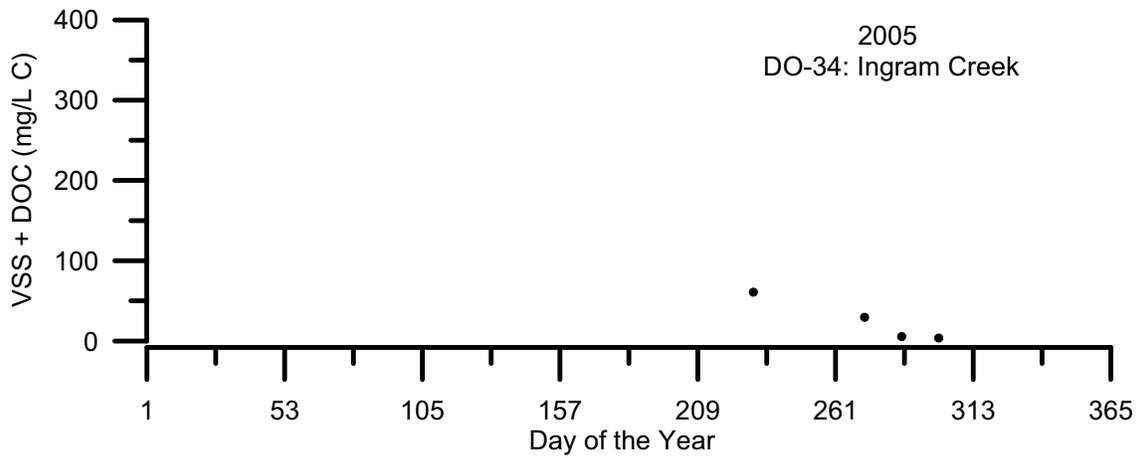


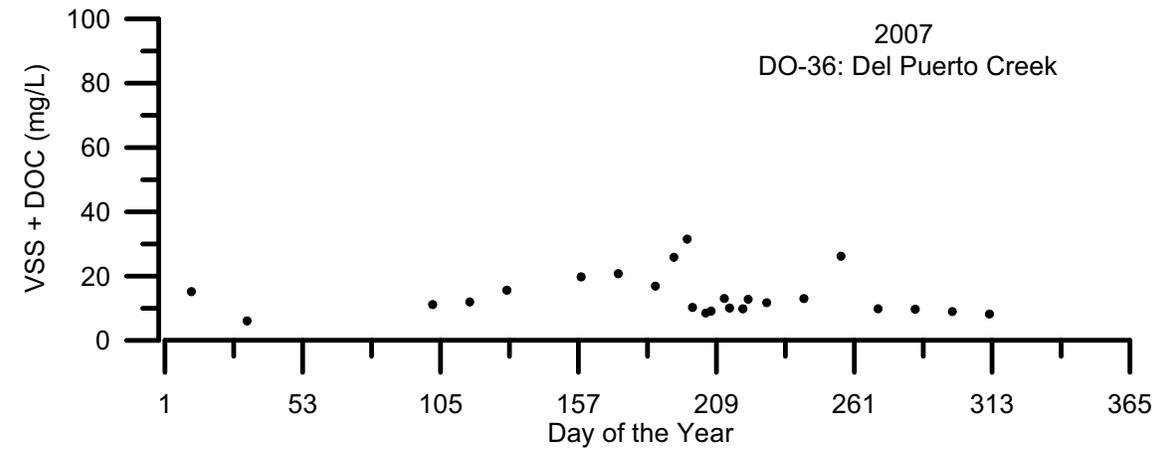
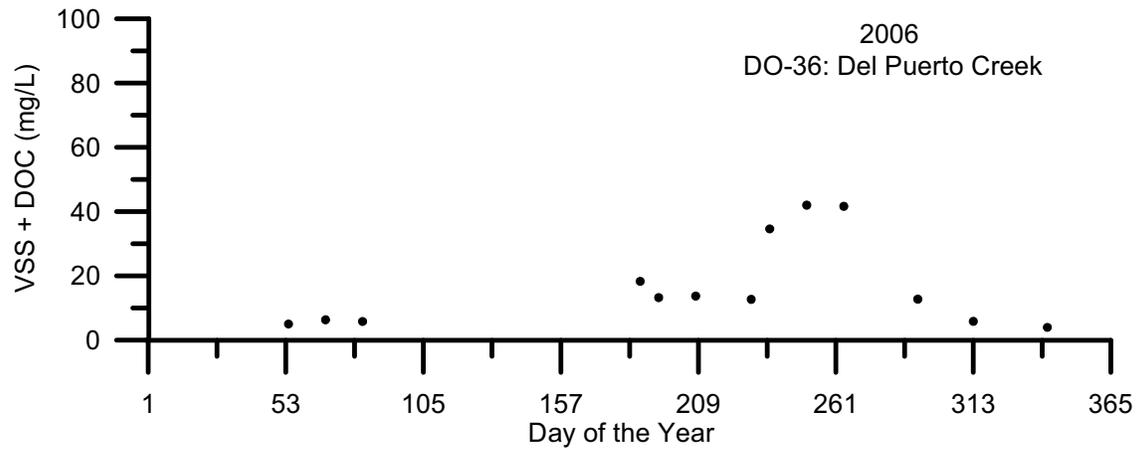
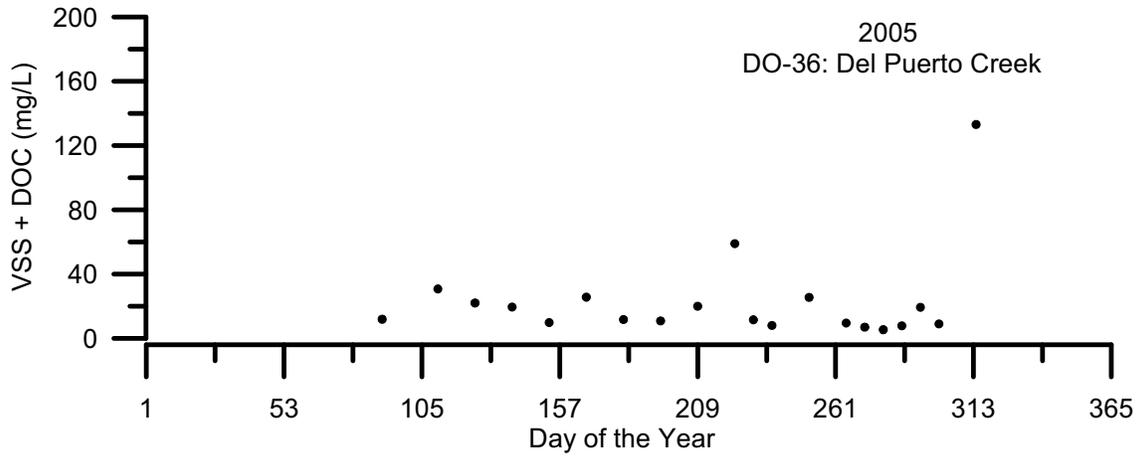


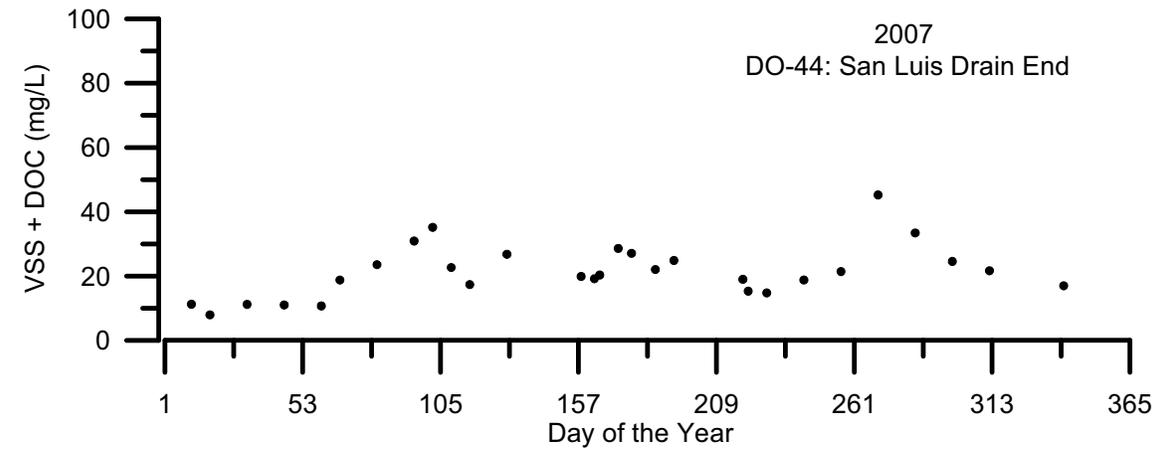
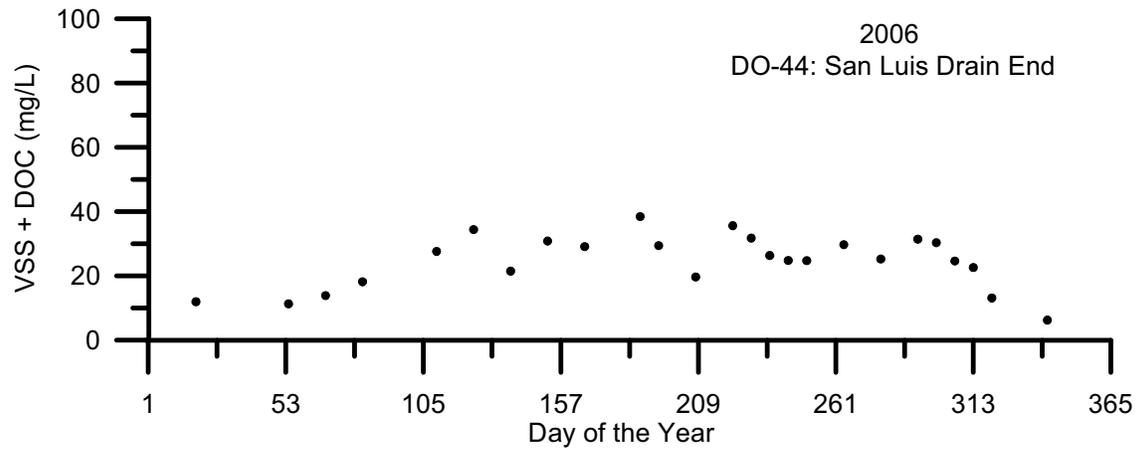
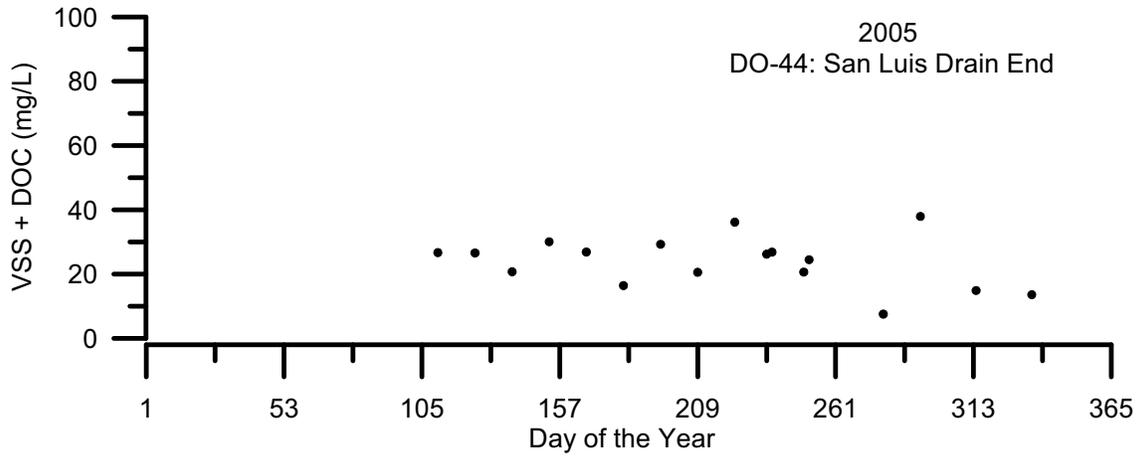


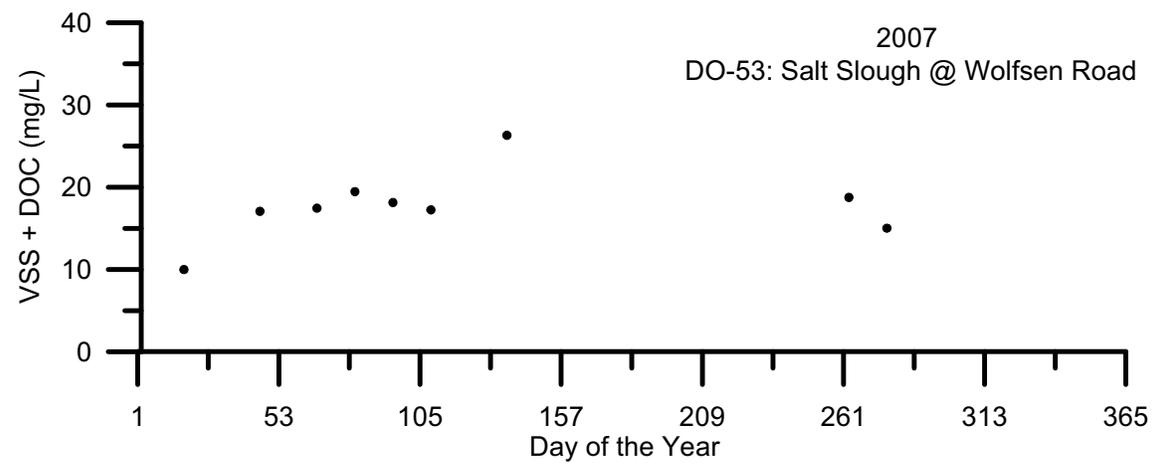
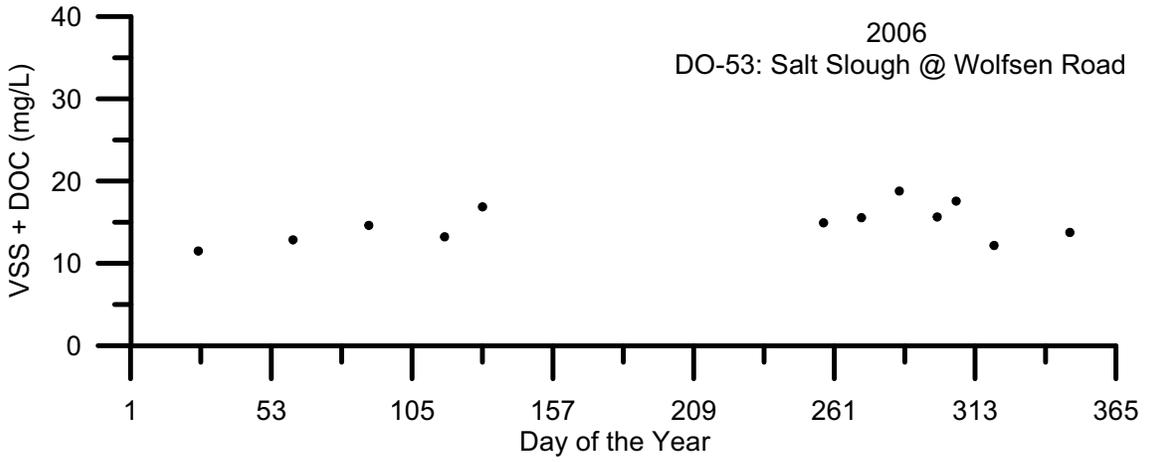
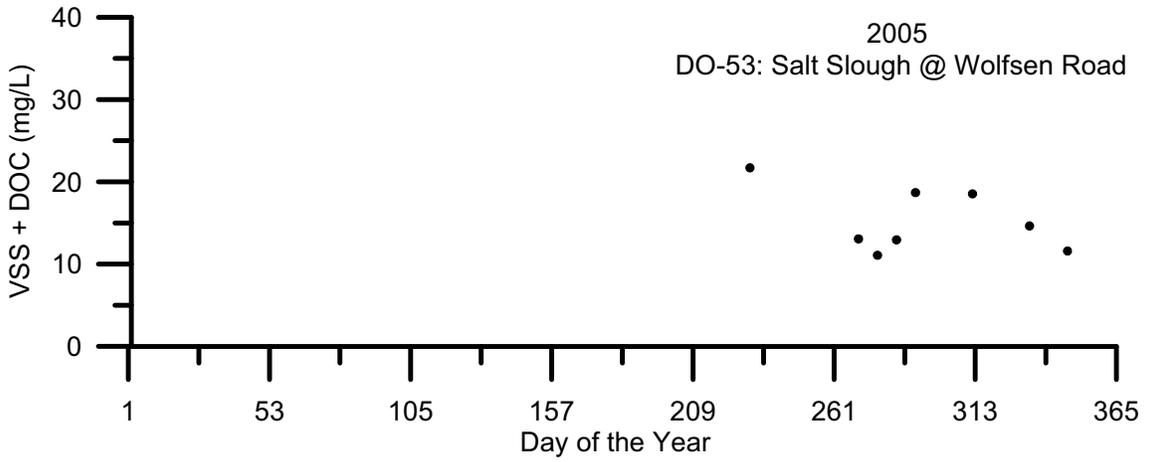


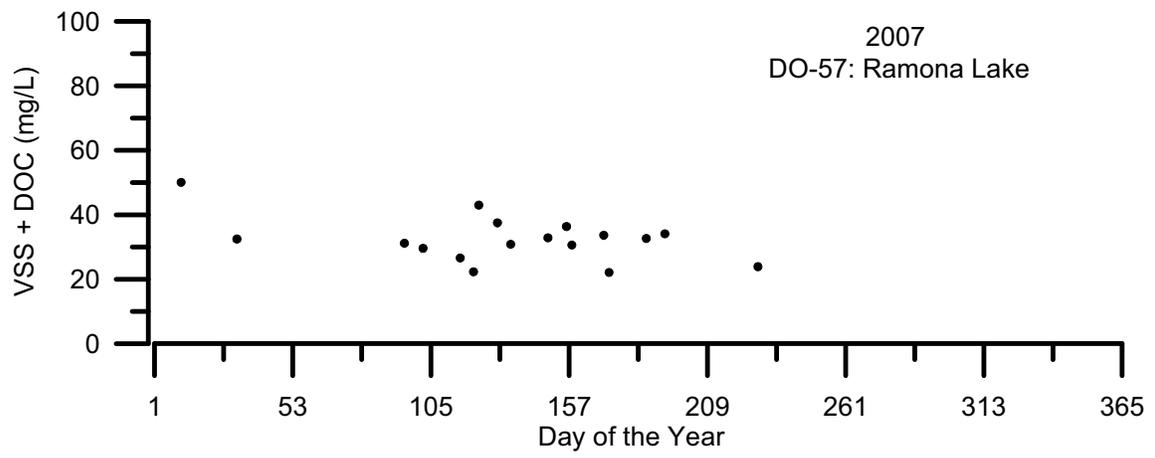
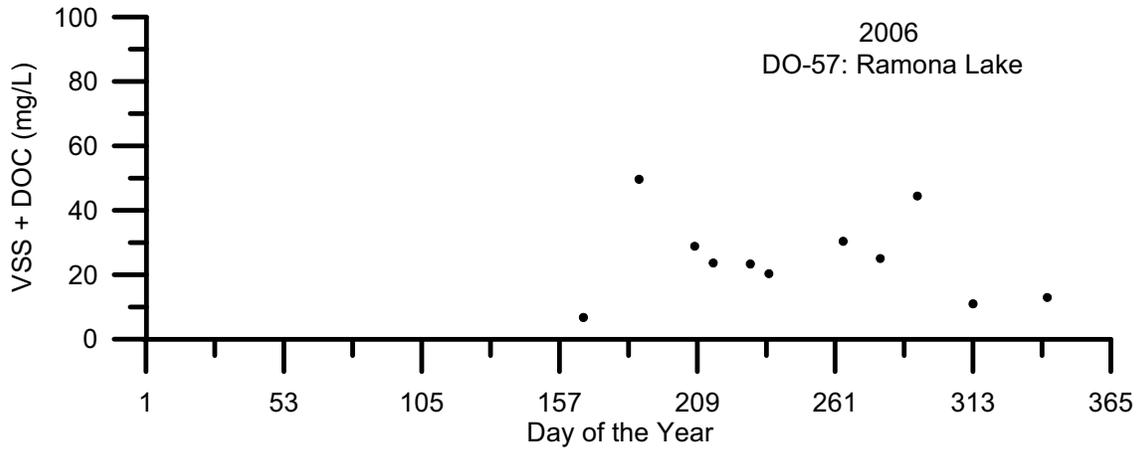


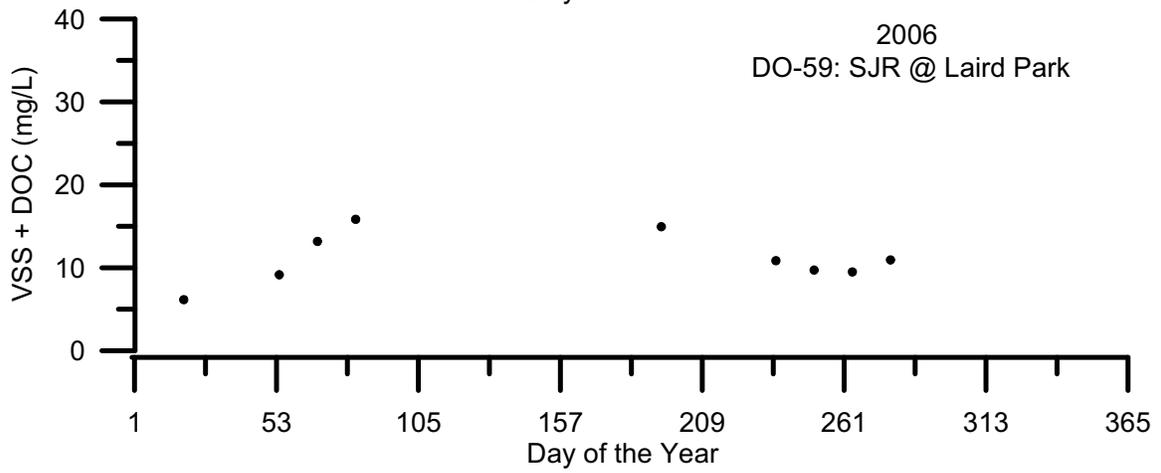
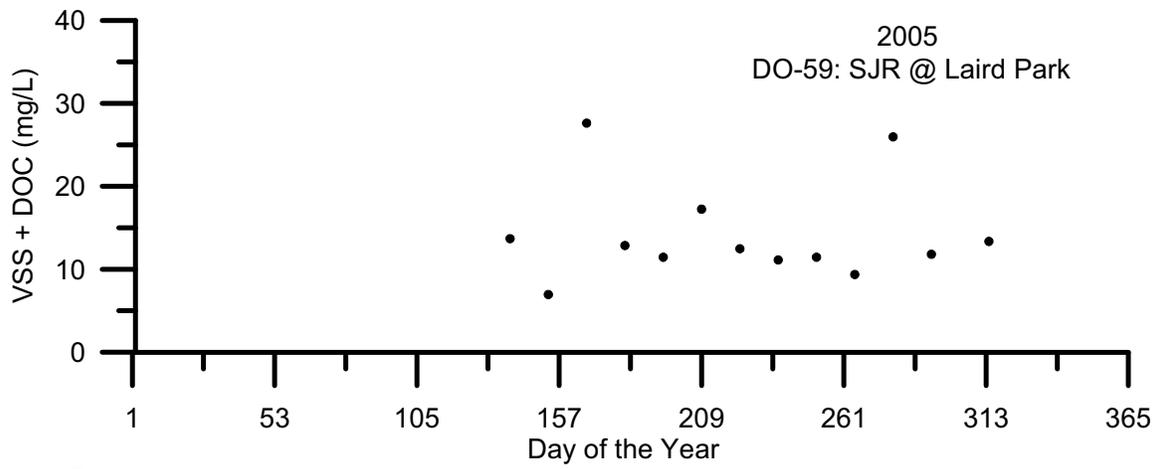


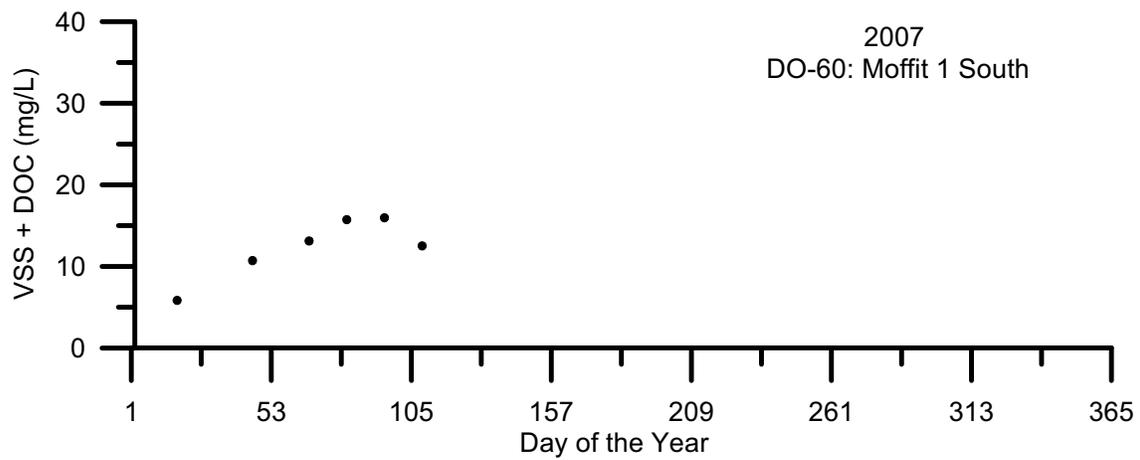
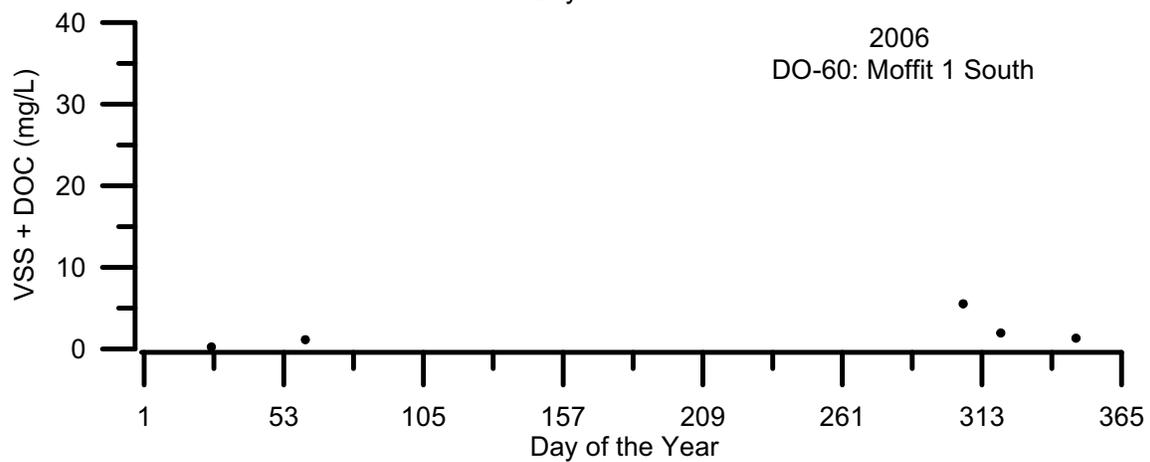
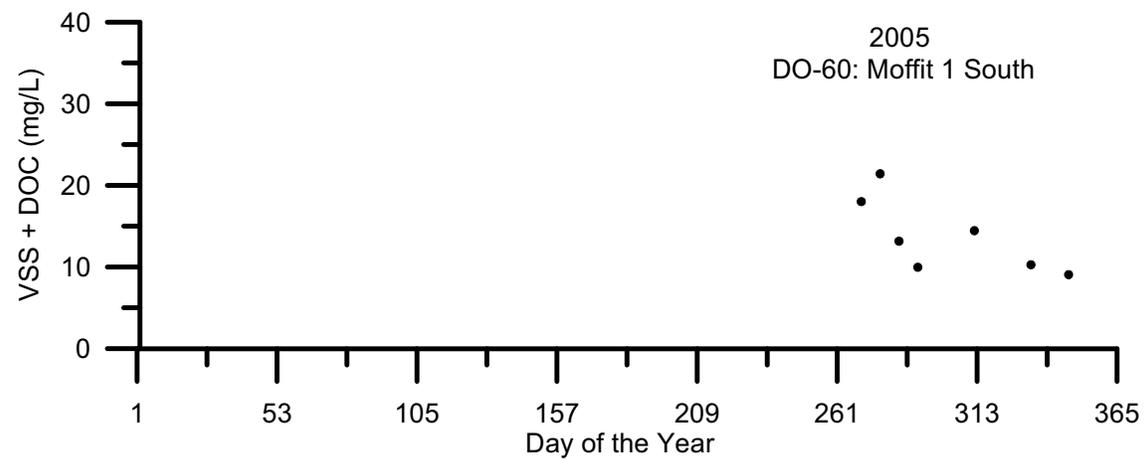


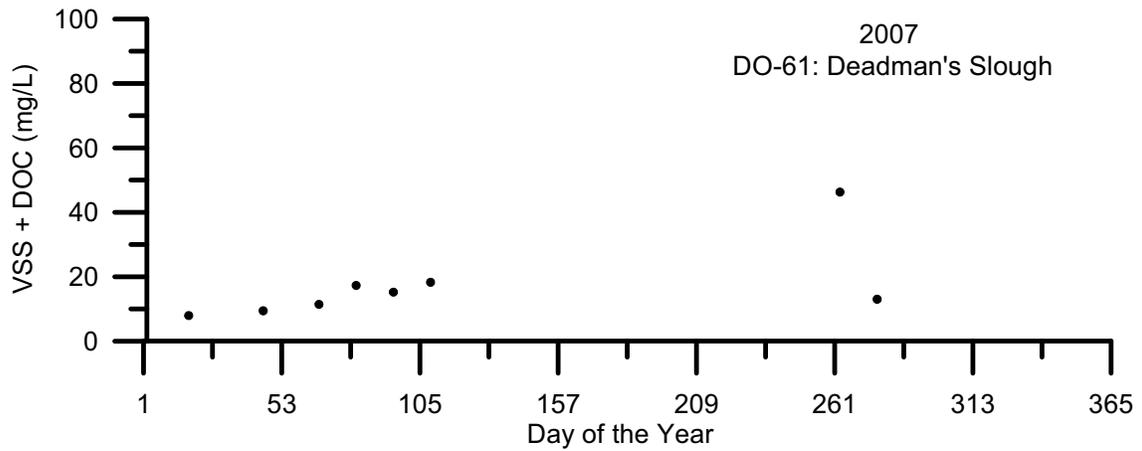
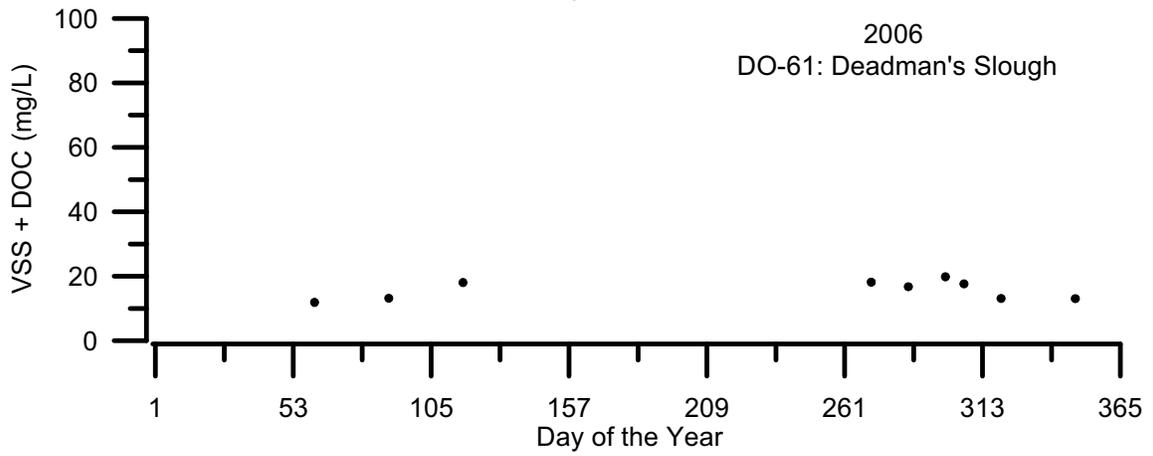
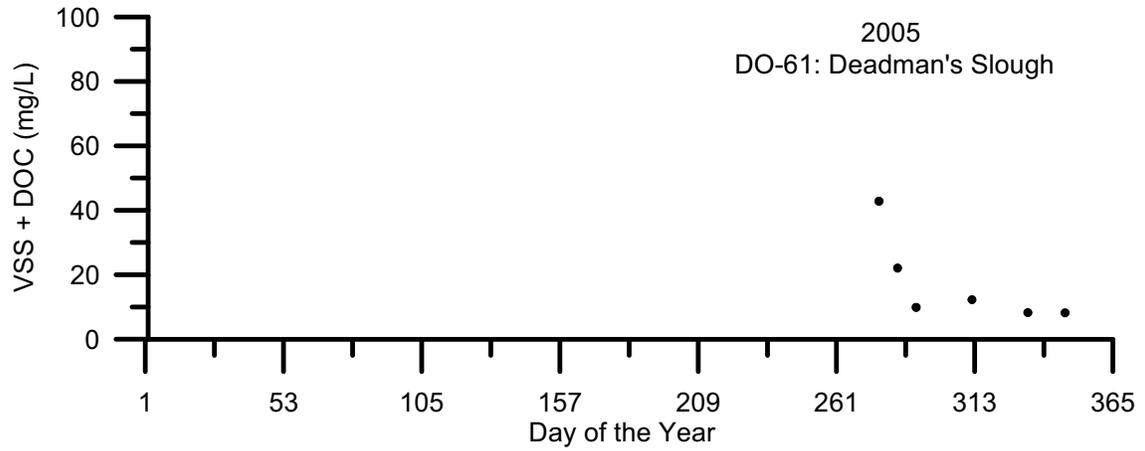


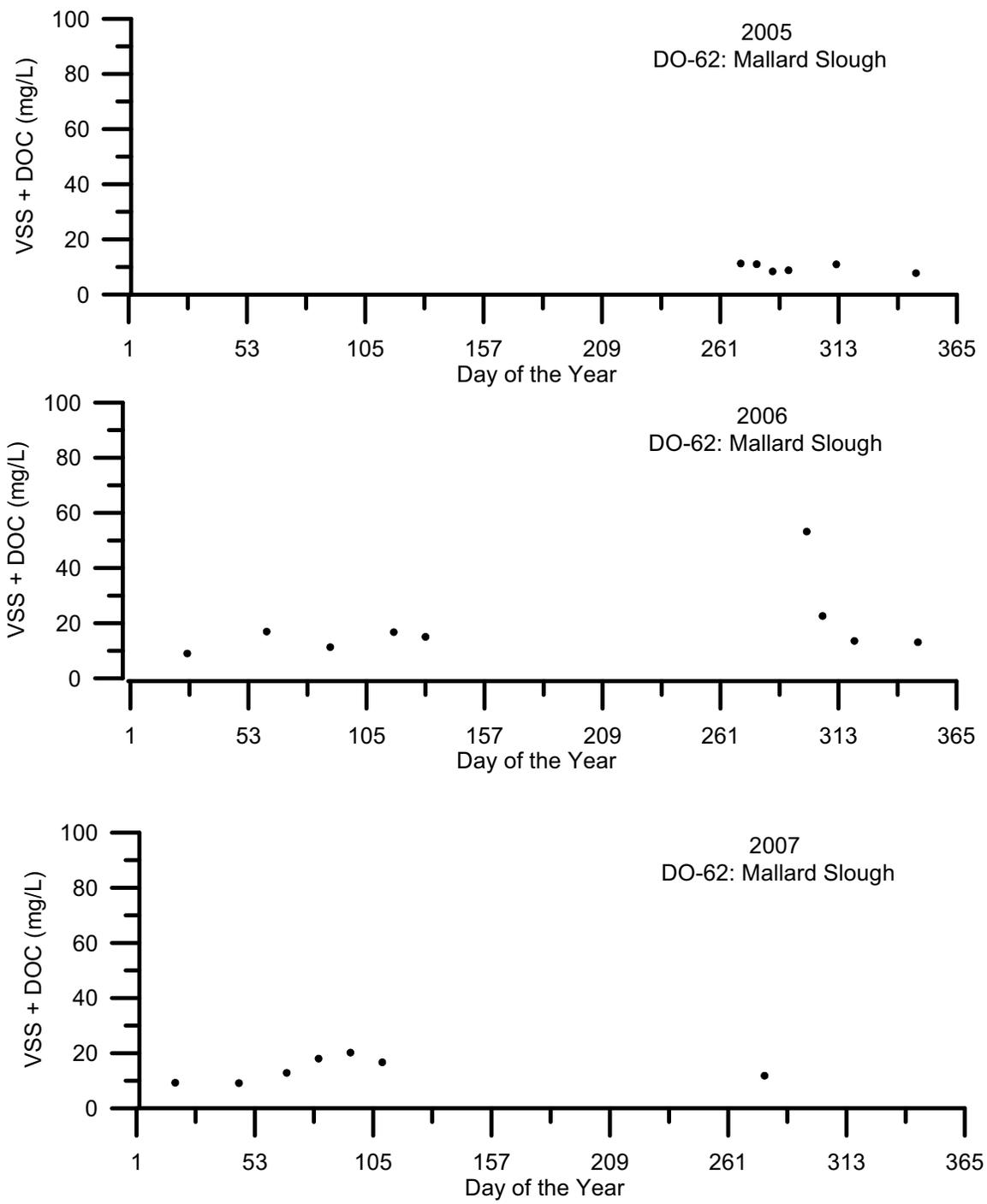


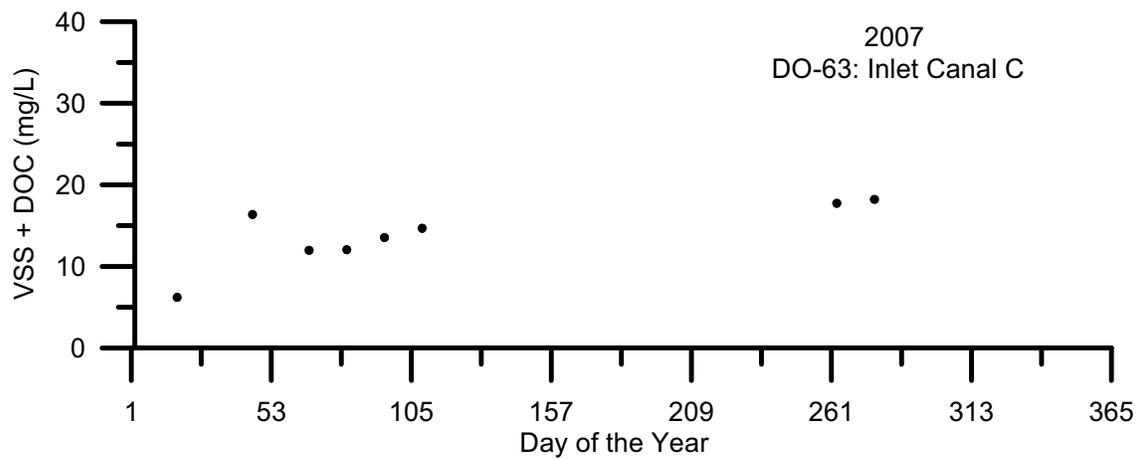
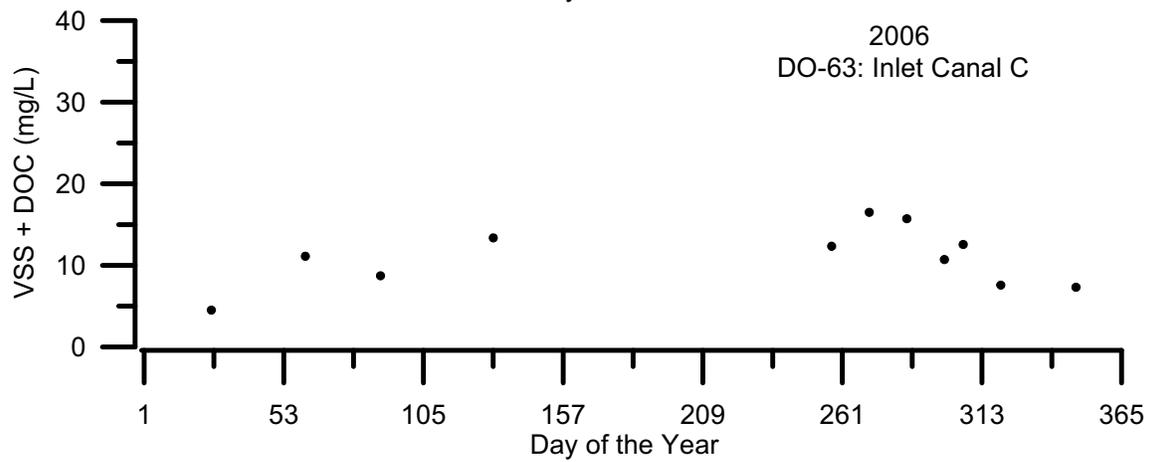
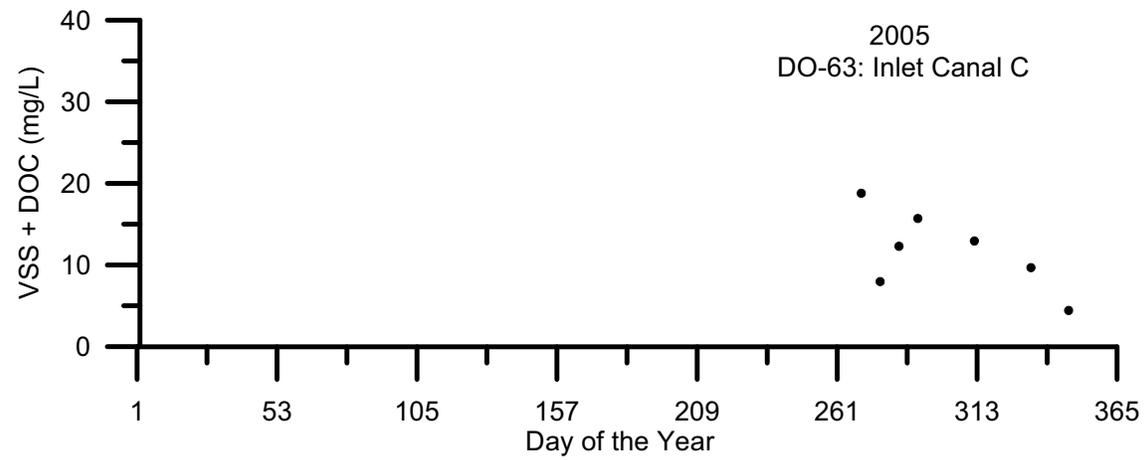


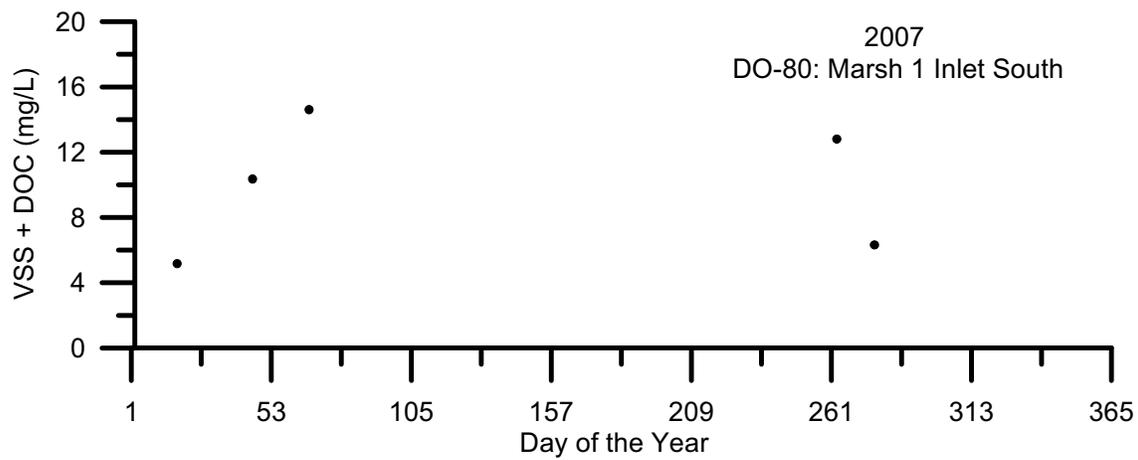
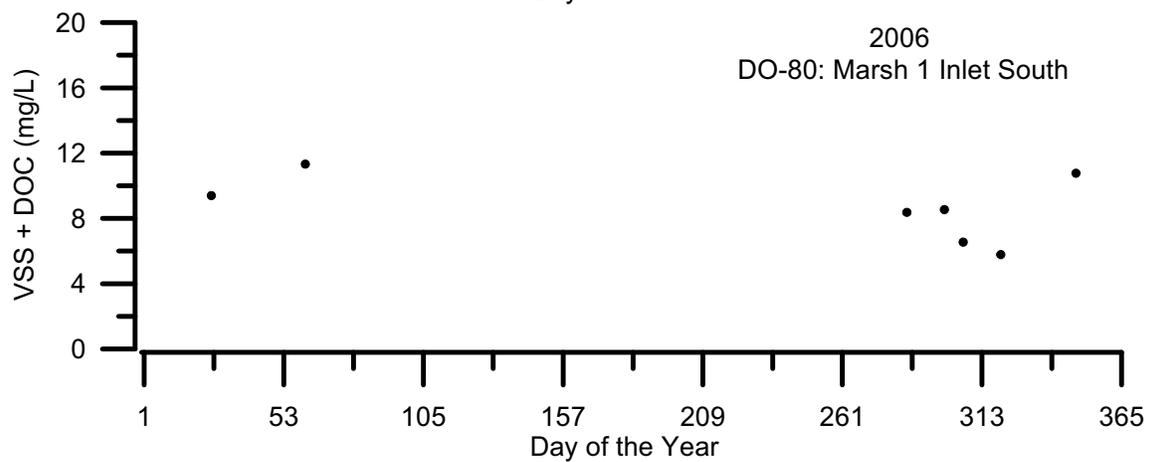
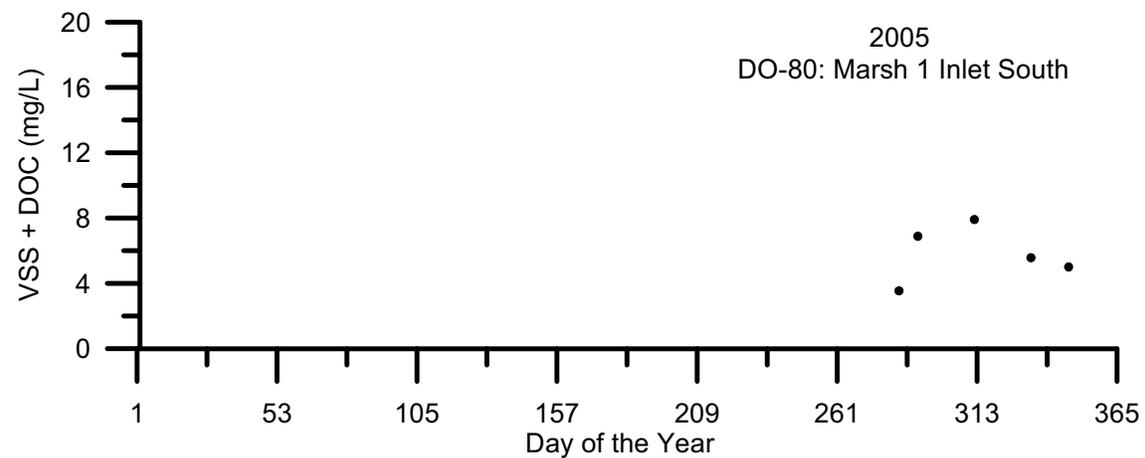


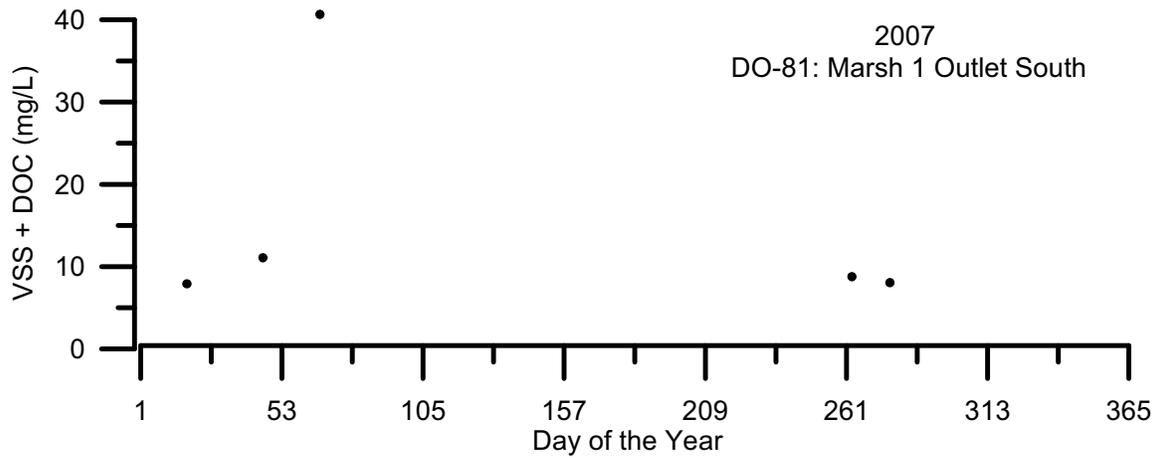
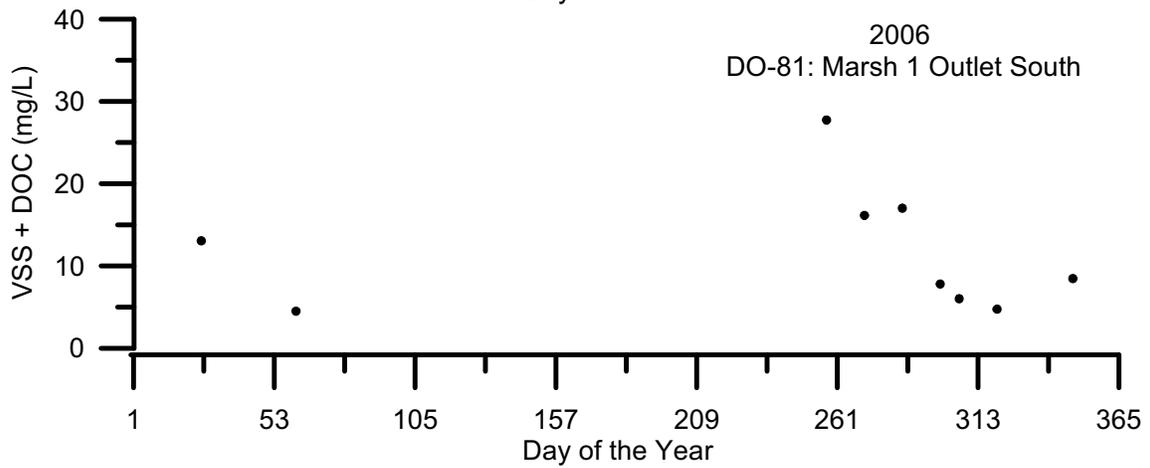
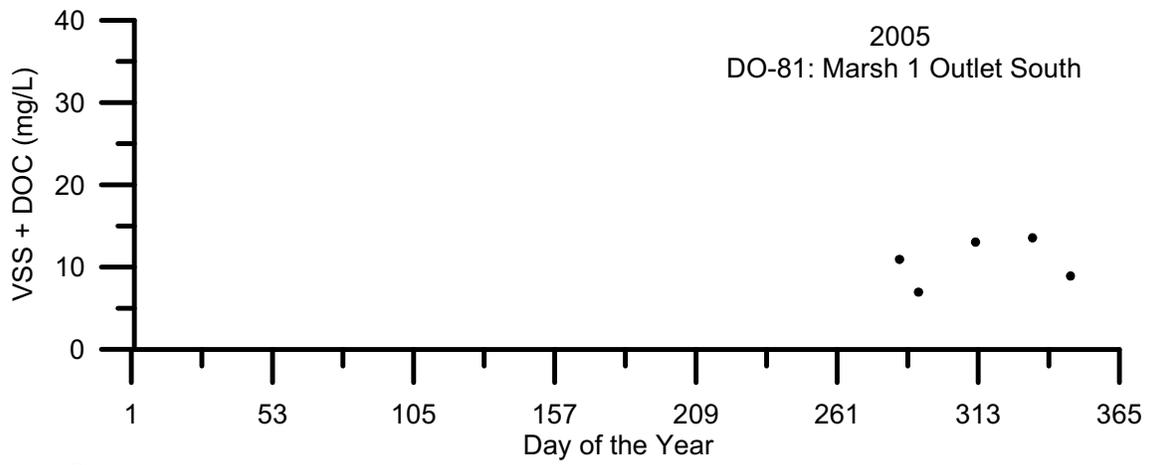


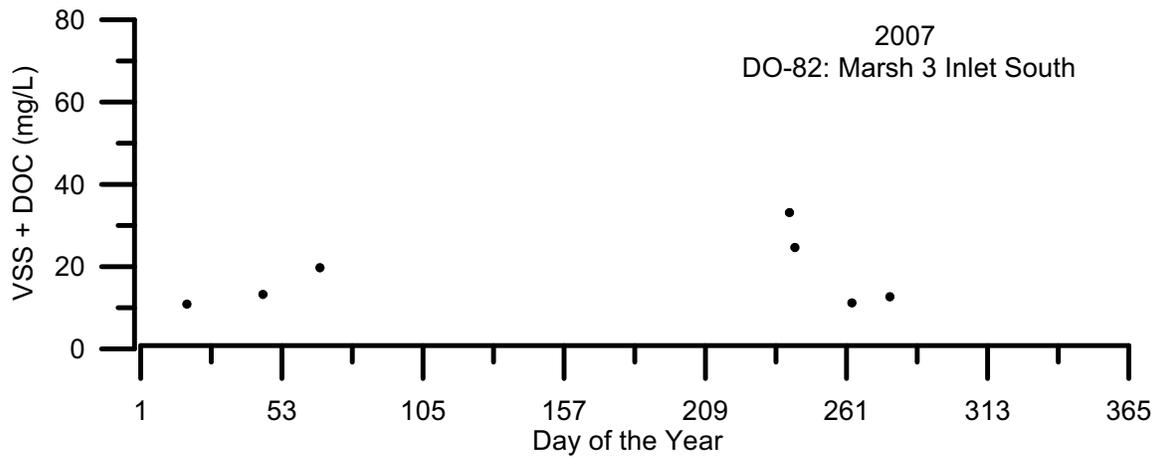
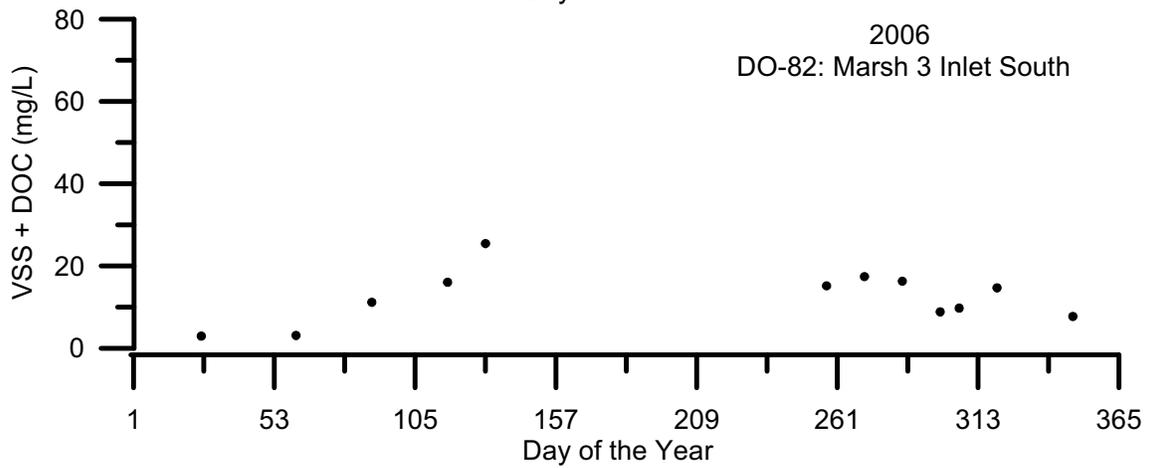
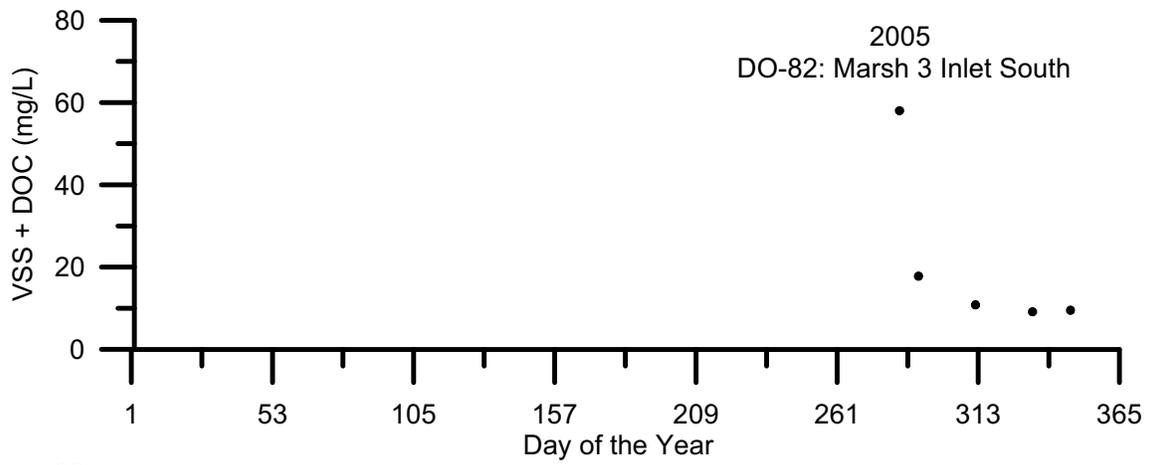


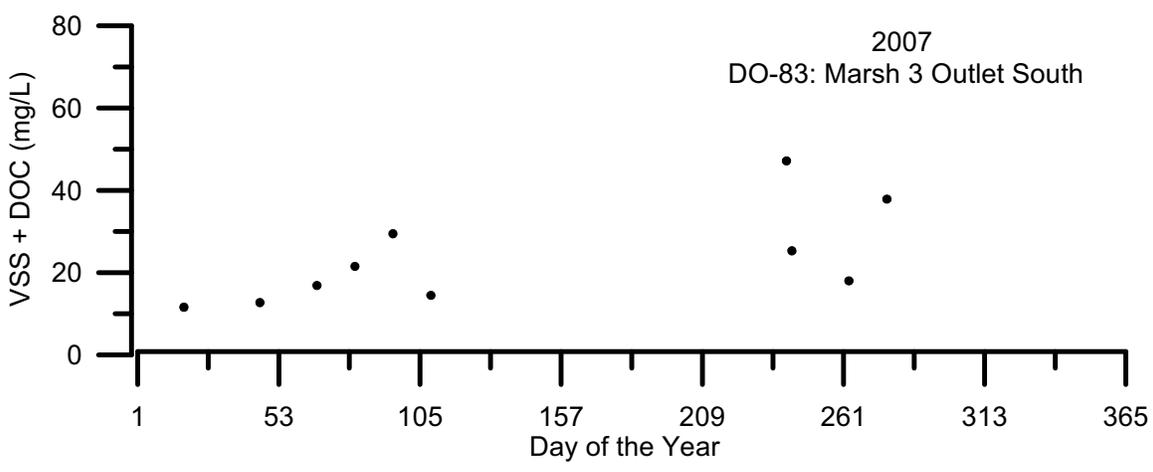
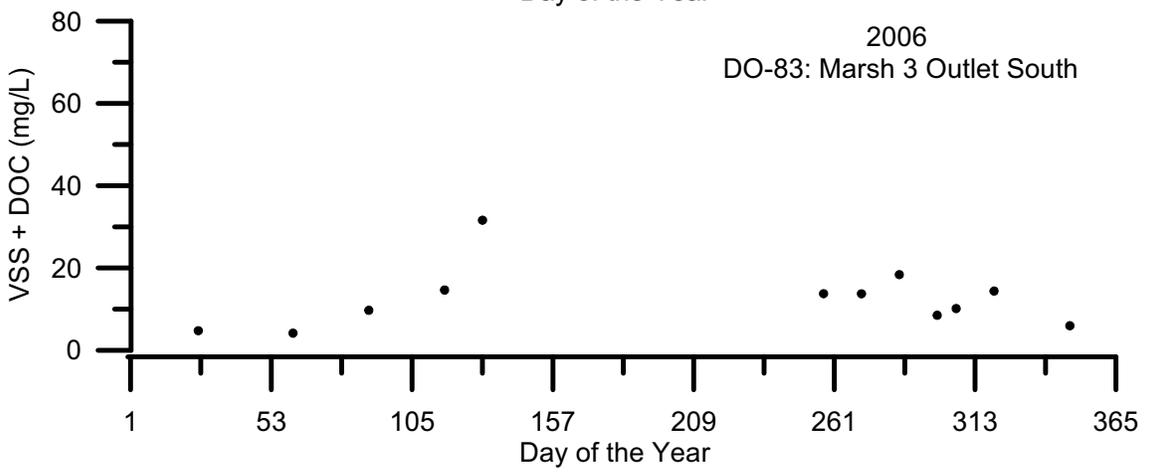
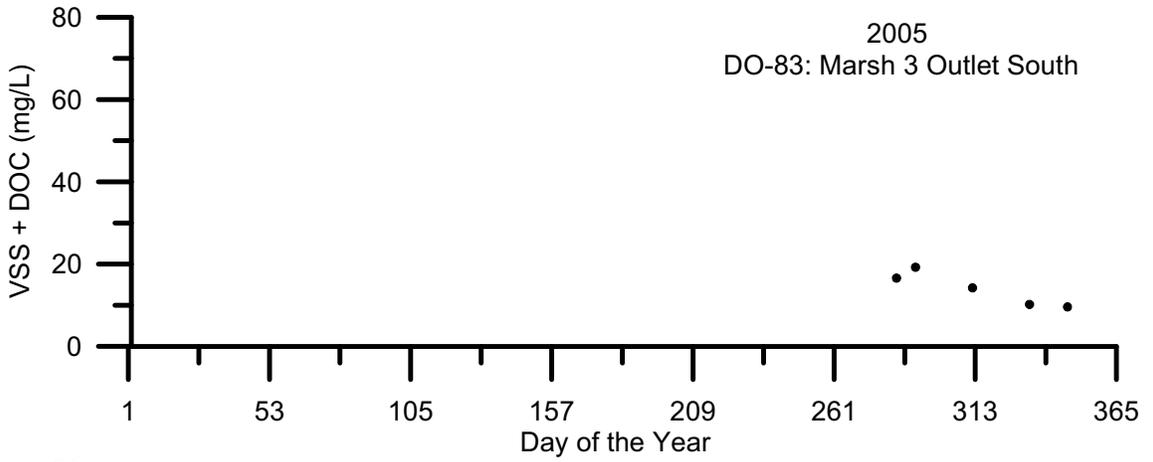


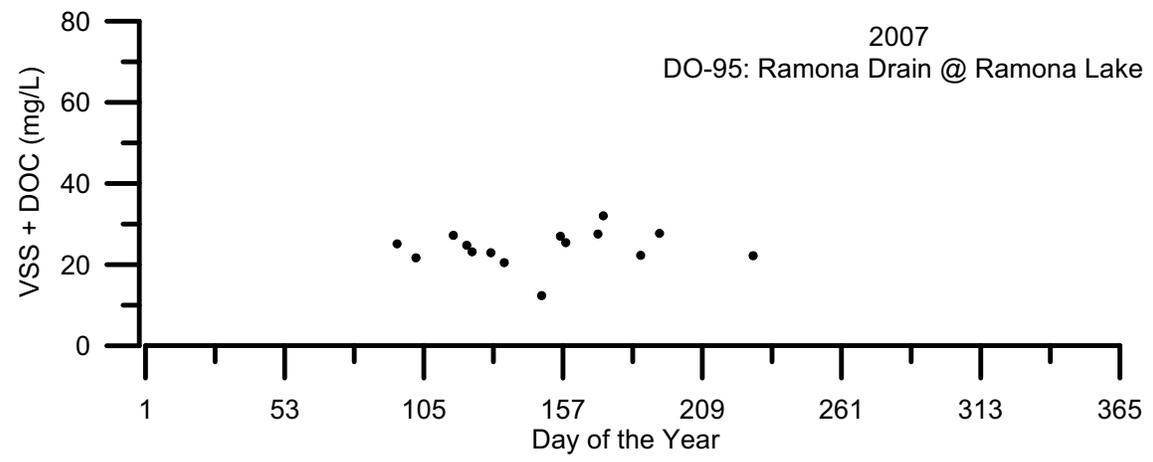
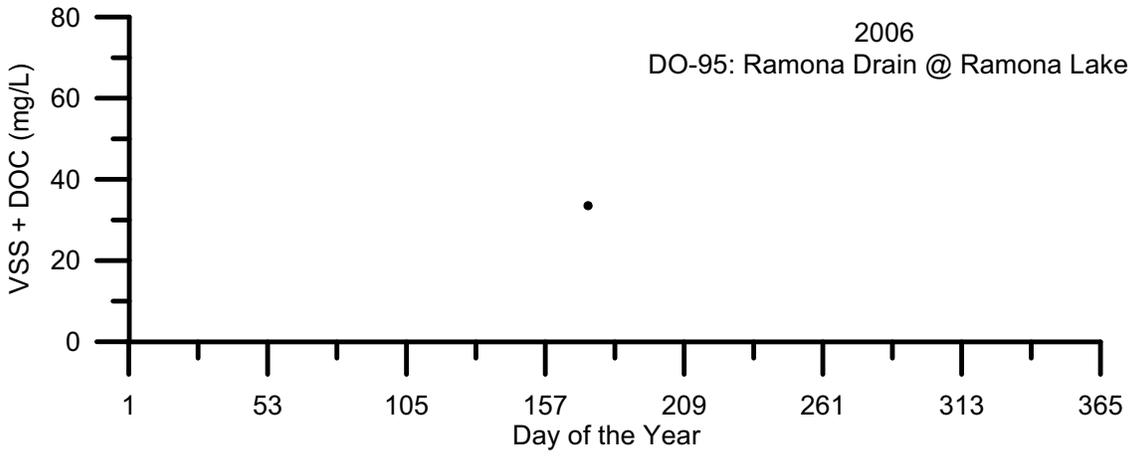














**Analysis of Total Suspended Solid Concentrations
in the San Joaquin River Watershed
2005-2007**

*Remie Burks
Chelsea Spier
Sharon Borglin
Jeremy Hanlon
Justin Graham
William Stringfellow*

February 2008

*Environmental Engineering Research Program
University of the Pacific
3601 Pacific Avenue, Sears Hall
Stockton CA 95211*

Introduction

The San Joaquin River (SJR) supports one of the most productive agricultural regions in the world and its productivity is heavily dependant on irrigated agriculture. A consequence of irrigated agriculture is the production of return flows conveyed down gradient drains that eventually discharge to surface waters. Agricultural drainage may have significant nutrient load and can impact algae growth and general water quality in the SJR. Individual farmers and agricultural organizations, such as drainage authorities, are in need of tools to manage the environmental impacts of agricultural activities (Stringfellow, 2008).

For the years 2005 through 2007, sites throughout the San Joaquin Valley watershed were sampled to assess the overall water quality in the region. One thousand nine hundred and ninety-six (1996) individual surface water samples were collected and analyzed and WQ was assessed at 113 sampling locations in the SJR basin (Borglin et al., 2008). Samples were processed and analyzed by the Environmental Engineering Research Program (EERP) laboratory at the University of the Pacific as well as at the University of California, Davis, Dahlgren Lab. This report presents temporal plots of total suspended solids (TSS) for all sites sampled in the SJR from 2005-2007.

Methods

Field sampling consisted of collecting water samples, measuring water quality with a YSI Sonde 6600 with MDS650 hand-held display, and recording of field conditions at sites within the study area per *EERP Field Protocol Book* (Graham, 2008).

Water samples were collected in glass 1000 mL bottles (Wheaton Science Products, Millville, NJ), 1000 mL HDPE Trace-Clean narrow mouth plastic bottles (VWR International), 250 mL HDPE Trace-Clean wide mouth plastic bottles (VWR International) as well as 40 mL trace clean vials with PTFE septa (IChem, Rockwood, TN) in accordance with requirements for different lab analysis and volume requirements.

Samples were immediately stored at 4°C after sampling (cooler temperature was recorded in the lab upon delivery) and transported to the lab on the day of sampling.

Samples were received by the laboratory the same day they were sampled and stored at 4°C until filtering and analysis. Samples were collected, preserved, stored, and analyzed by methods outlined in *Standard Methods for the Analysis of Water and Wastewater*, (APHA, 2005, 1998). Total suspended solids (TSS) and volatile suspended solids (VSS) were analyzed by SM 2540 D and E (APHA, 2005). Typically 1000 mL of sample was filtered on pre-weighed, pre-combusted, Whatman GF/F filters. The filters were placed in an aluminum dish and dried at 105°C under vacuum to constant weight. After drying the filter and dish were allowed to cool in a dessicator and were weighed for TSS determination. The dried and weighted filters were subsequently combusted at 550°C for 6 hours and reweighed for VSS determination. Mineral suspended solids (MSS) concentration was calculated by subtracting VSS from TSS.

Results and Discussion

Total Suspended Solids (TSS) was analyzed routinely over the years 2005-2007 with no modification to the standard method (APHA, 2005). A 96.30% passage rate for all QA parameters over the three years was observed as well as a rate of 100% for proficiency check samples (Borglin et. al., 2008).

References

Stringfellow, W.T., et al., (2008), *Evaluation of Vegetated Ditches, Ponds, and Wetlands as BMPs for Mitigating the Water Quality Impact of Irrigated Agriculture in the San Joaquin Valley*, University of the Pacific, Stockton, CA

Graham, J., Hanlon, J.S., Stringfellow, W.T., (2008), *EERP Field Protocol Book*, University of the Pacific, Stockton, CA.

Borglin, S., W. Stringfellow, J. Hanlon. (2005), *Standard Operating Procedures for the Up-Stream Dissolved Oxygen TMDL Project*, LBNL/Pub-937.

Borglin, S., Burks, R., Hanlon, J., Graham, J., Spier, C., Stringfellow, W., and Dahlgren, R., (2008) Methods overview, quality assurance, and quality control, University of the Pacific, Stockton, CA

American Public Health Association, (2005), *Standard Methods for the Examination of Water and Wastewater*, American Public Health Association, Washington, D.C.

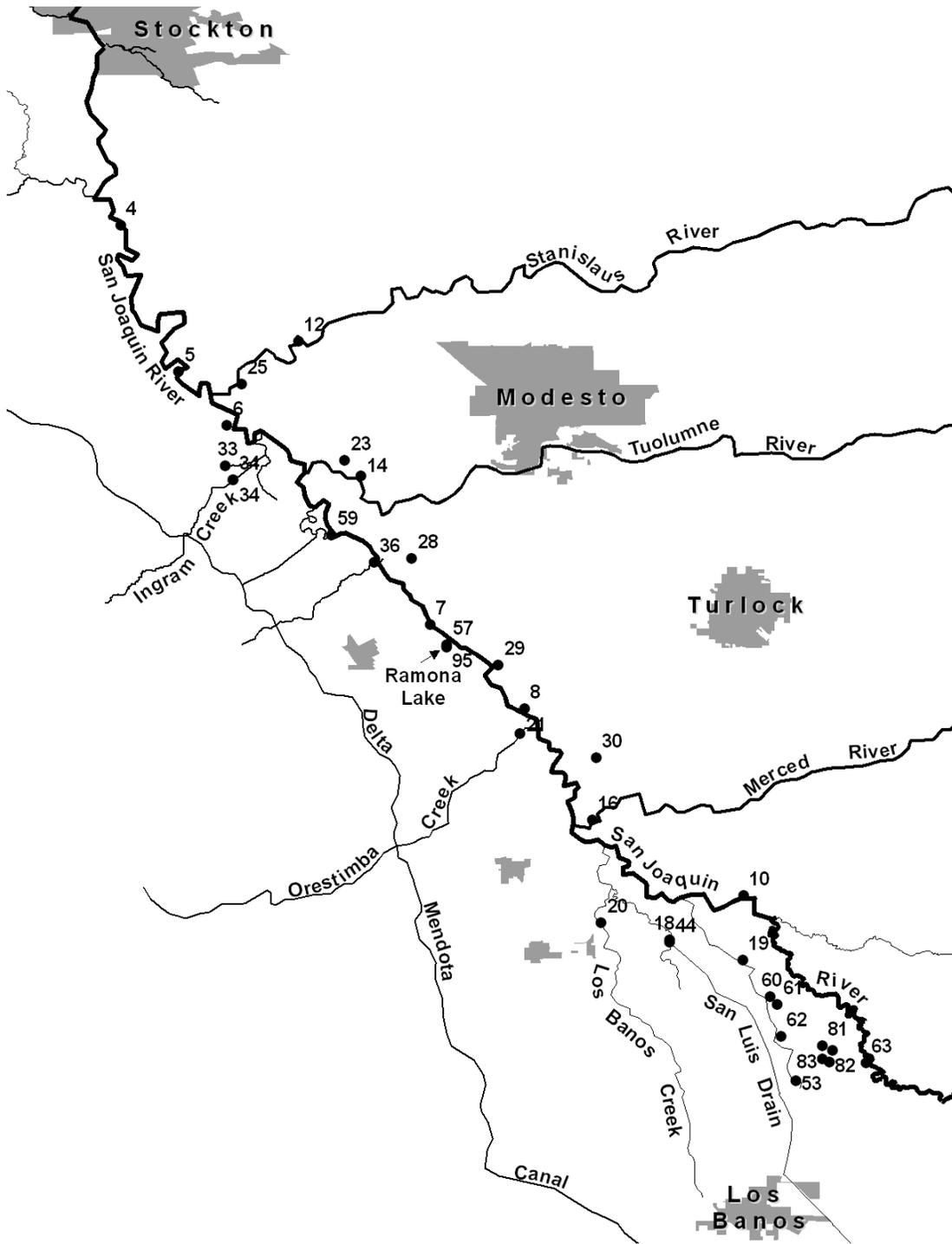
Borglin, S.E., Burks, R.D., Hanlon, J.S., Stringfellow, W.T. (2008) *EERP Lab Protocol Book*, University of the Pacific, Stockton, CA.

YSI Environmental Operations Manual, (2005), 6-Series Environmental Monitoring Systems, Yellow Springs, OH.

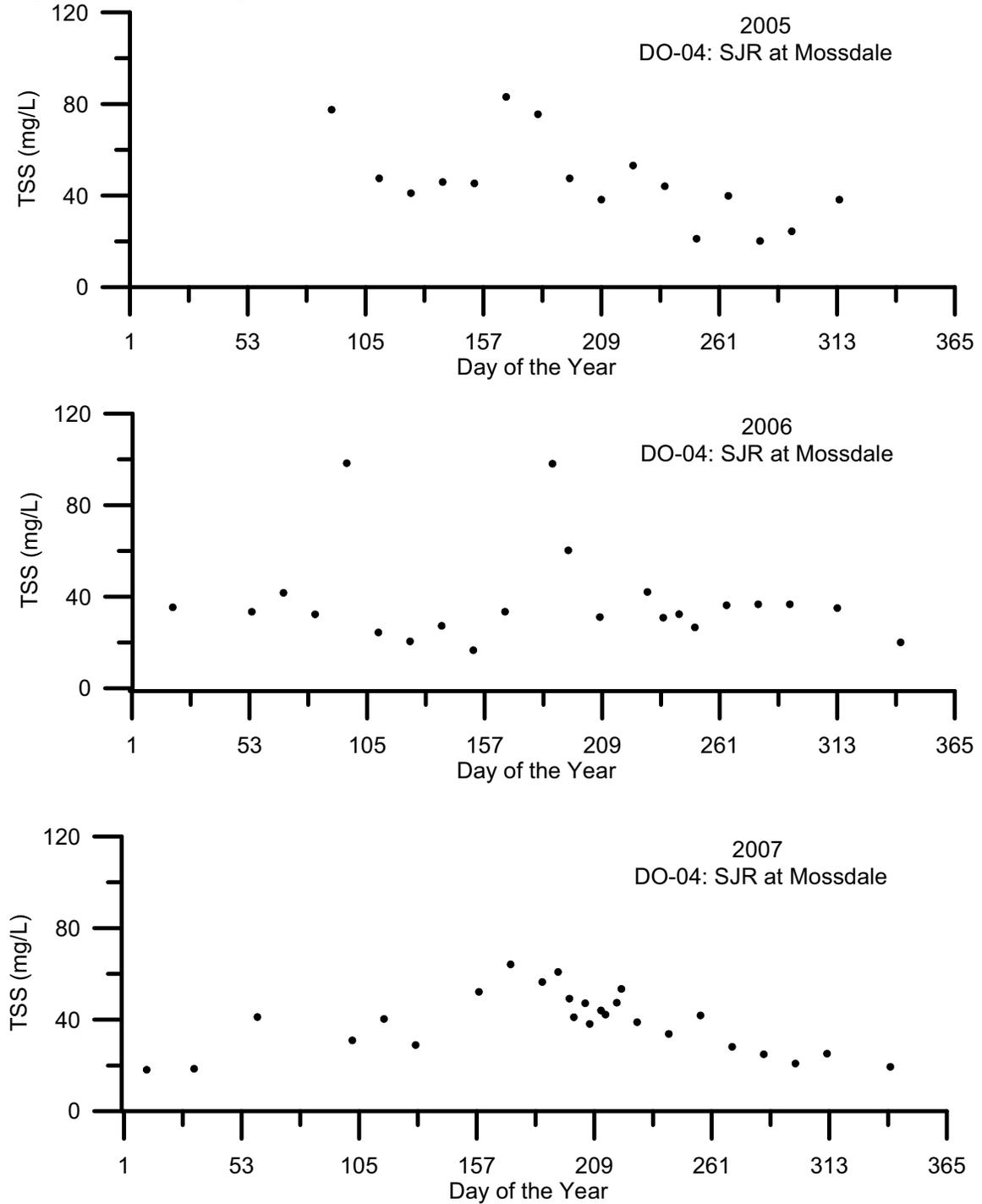
Table 1: EERP Sampling Site List

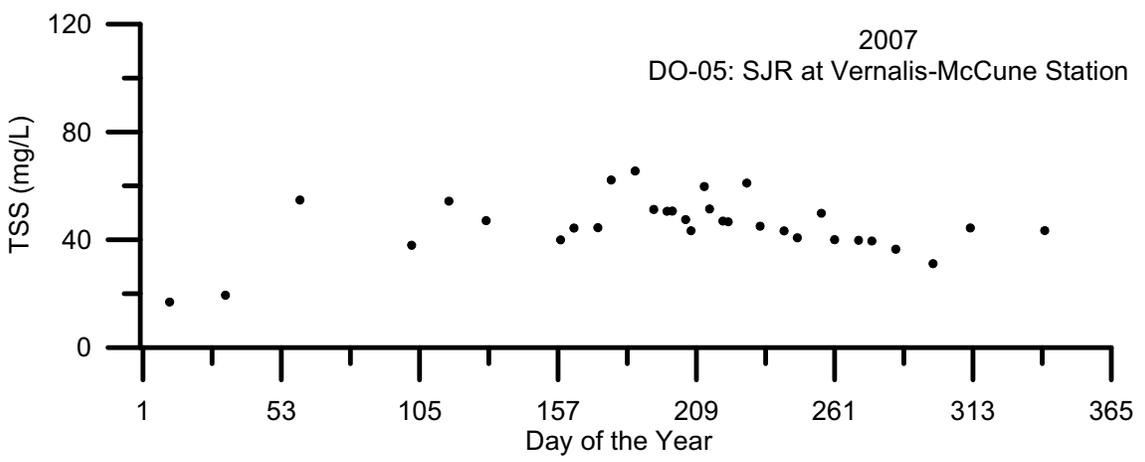
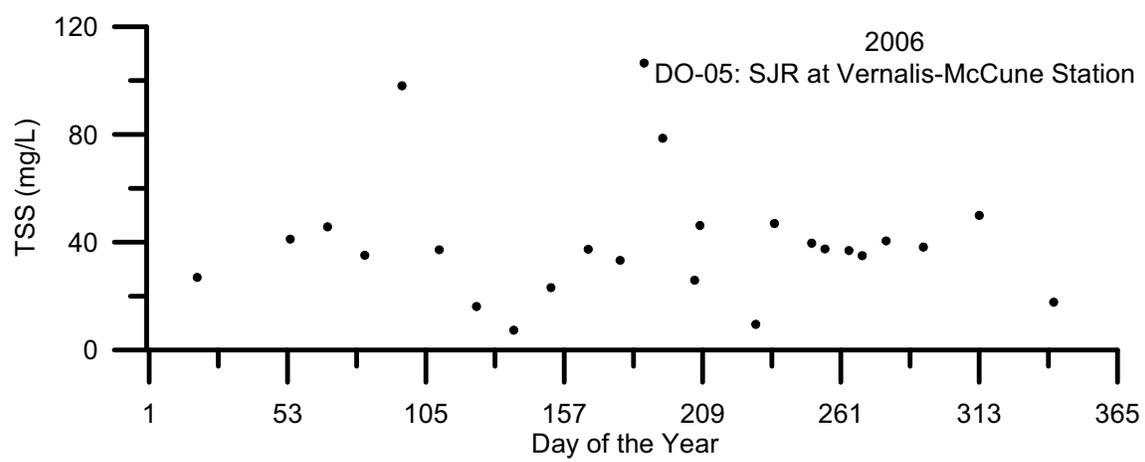
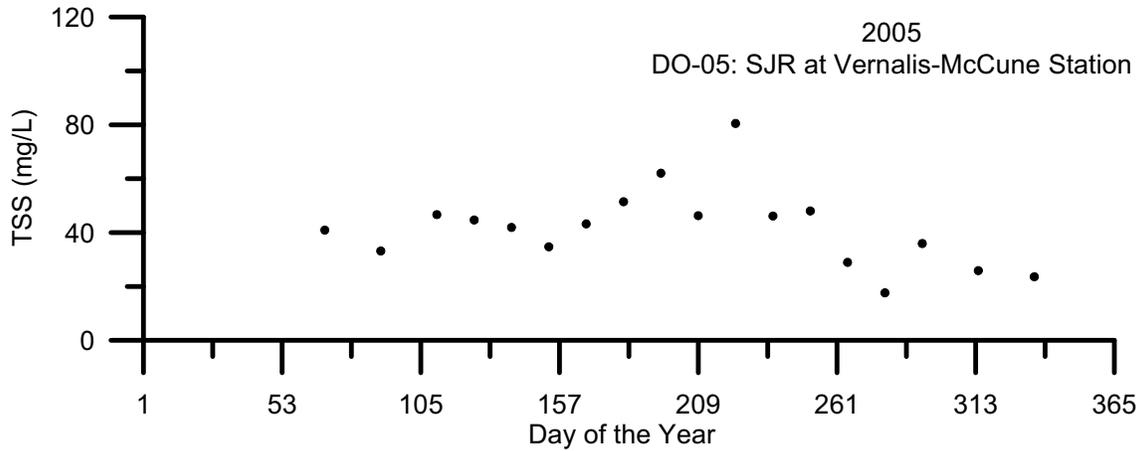
DO Number	Site Name	Type
4	SJR at Mossdale	Core sites
5	SJR at Vernalis-McCune Station (River Club)	Core sites, BMP
6	SJR at Maze	Core sites, BMP
7	SJR at Patterson	Core sites, BMP
8	SJR at Crows Landing	Core sites, BMP
10	SJR at Lander Avenue	Core sites
12	Stanislaus River at Caswell Park	Core sites
14	Tuolumne River at Shiloh Bridge	Core sites
16	Merced River at River Road	Core sites
18	Mud Slough near Gustine	Core sites, Wetland
19	Salt Slough at Lander Avenue	Core sites, Wetland
20	Los Banos Creek Flow Station	Core sites, Wetland
21	Orestimba Creek at River Road	Core sites, BMP
23	Modesto ID Lateral 5 to Tuolumne	Core sites
25	Modesto ID Main Drain to Stan. R. via Miller Lake	Core sites
28	Turlock ID Westport Drain Flow station	Core sites
29	Turlock ID Harding Drain	Core sites
30	Turlock ID Lateral 6 & 7 at Levee	Core sites
33	Hospital Creek	Intermittent, BMP
34	Ingram Creek	Core sites, BMP
36	Del Puerto Creek Flow Station	Core sites, BMP
44	San Luis Drain End	Core sites
53	Salt Slough at Wolfsen Road	Wetland
57	Ramona Lake Drain	Core sites, BMP
59	SJR Laird Park	Core sites
60	Moffit 1 South	Wetland
61	Deadman's Slough	Wetland
62	Mallard Slough	Wetland
63	Inlet C Canal	Wetland
80	South Marsh-1-Inlet	Wetland
81	South Marsh-1-Outlet	Wetland
82	South Marsh-3-Inlet	Wetland
83	South Marsh-3-Outlet	Wetland
95	Ramona drain at Ramona Lake	BMP, Intermittent

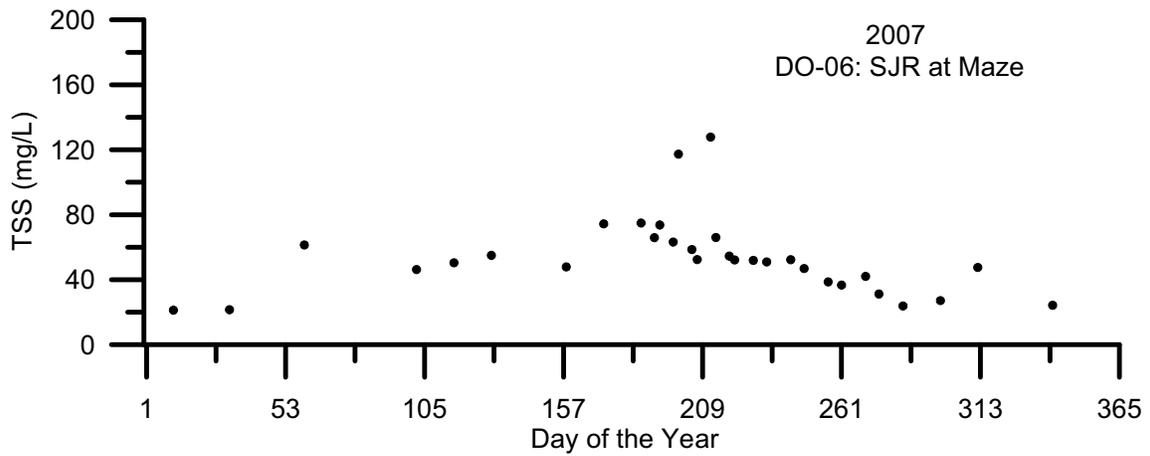
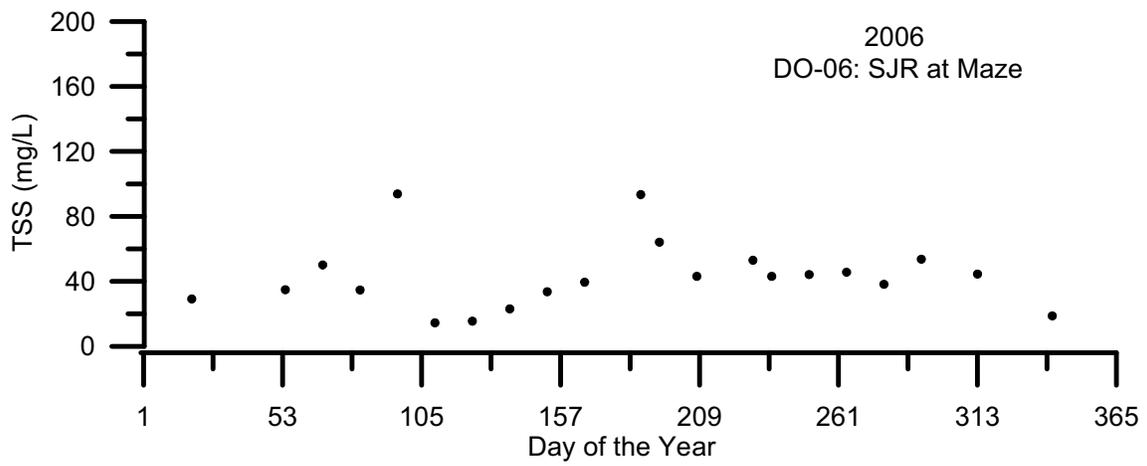
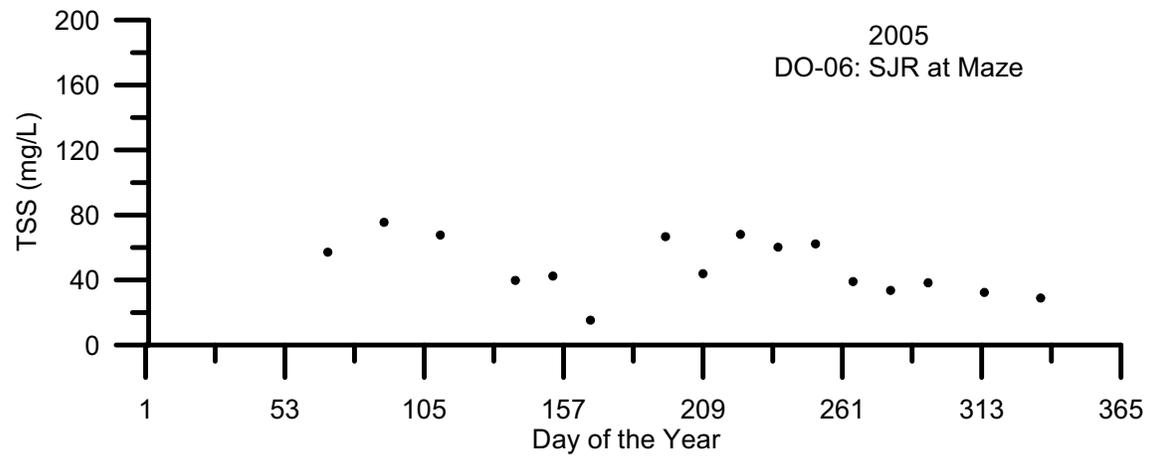
Figure 1: EERP Sampling Site Map of SJR Watershed and Tributaries

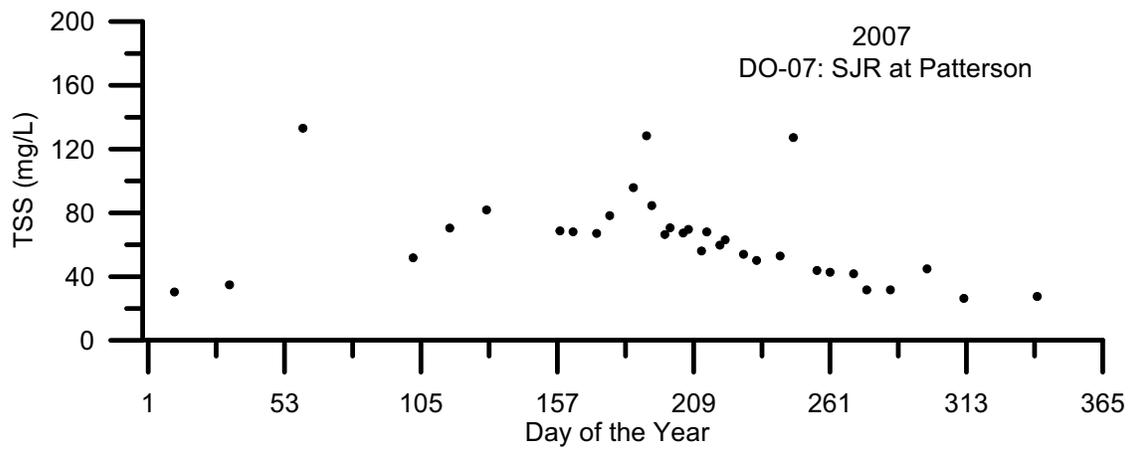
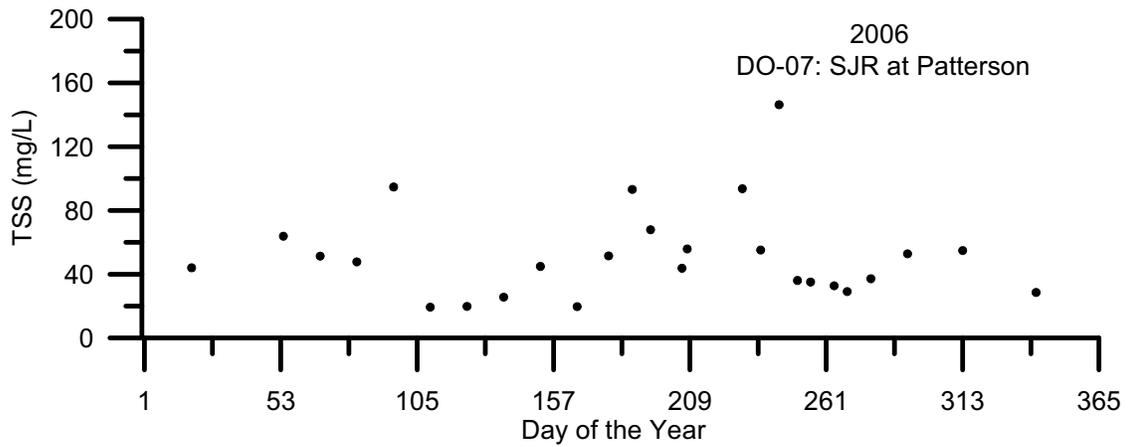
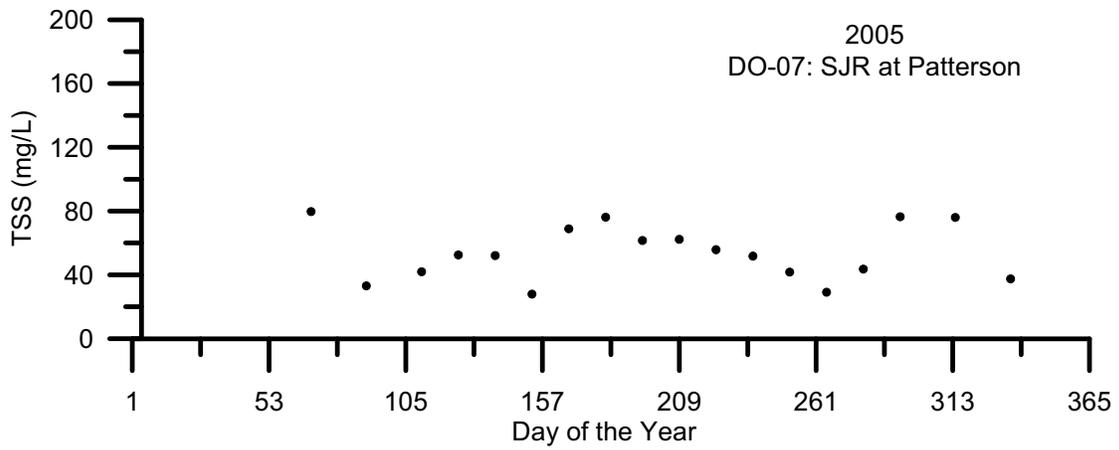


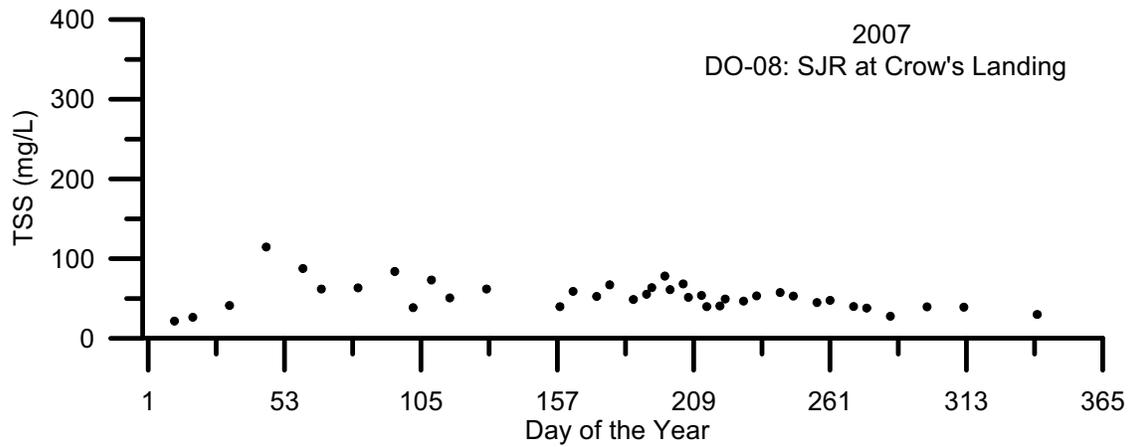
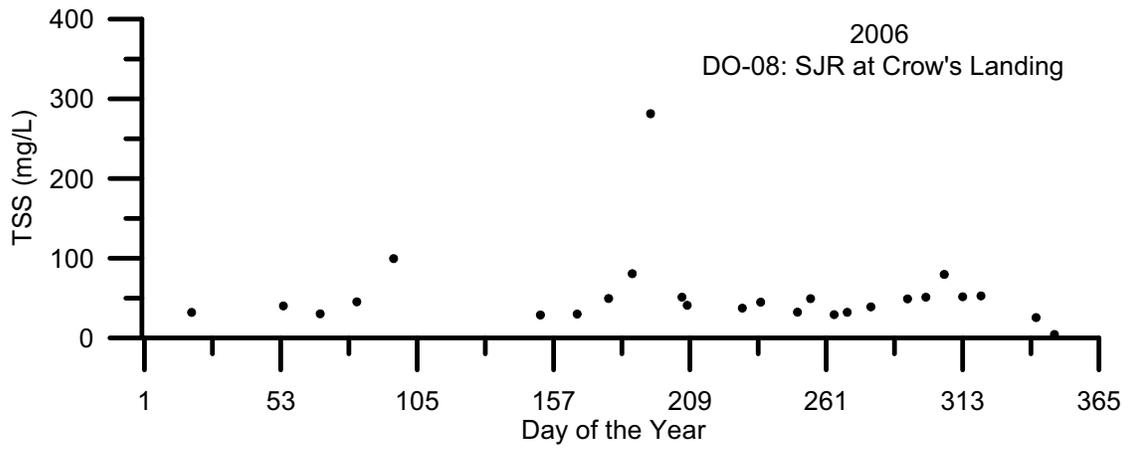
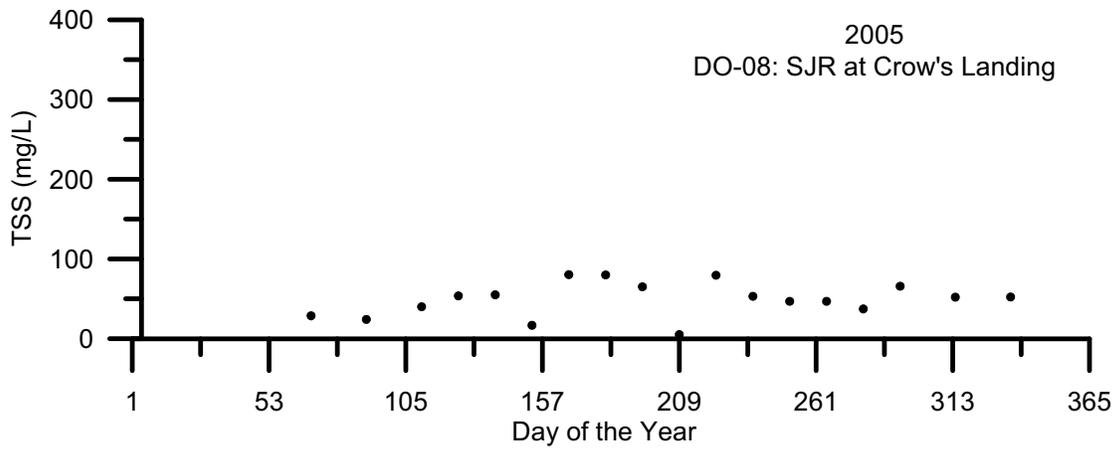
Figures 2 -101: Temporal Plots of TSS Concentrations By Site ID

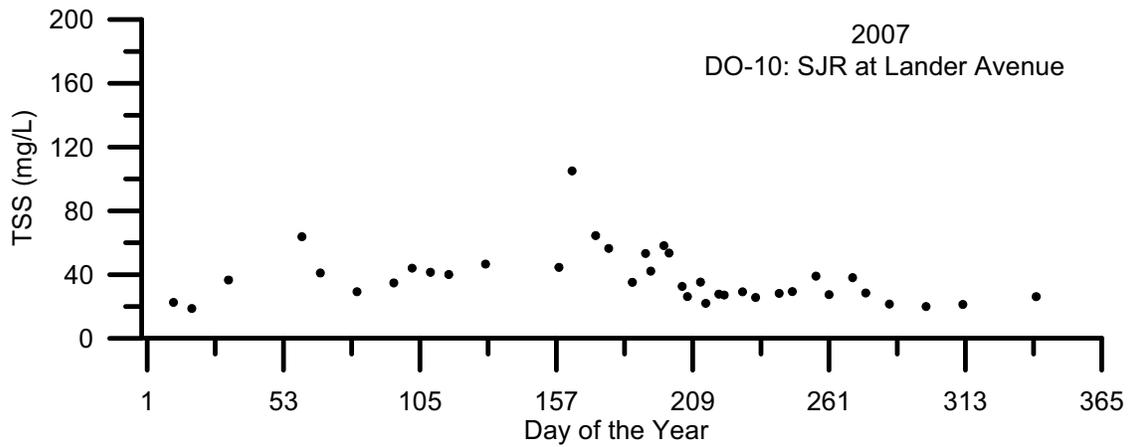
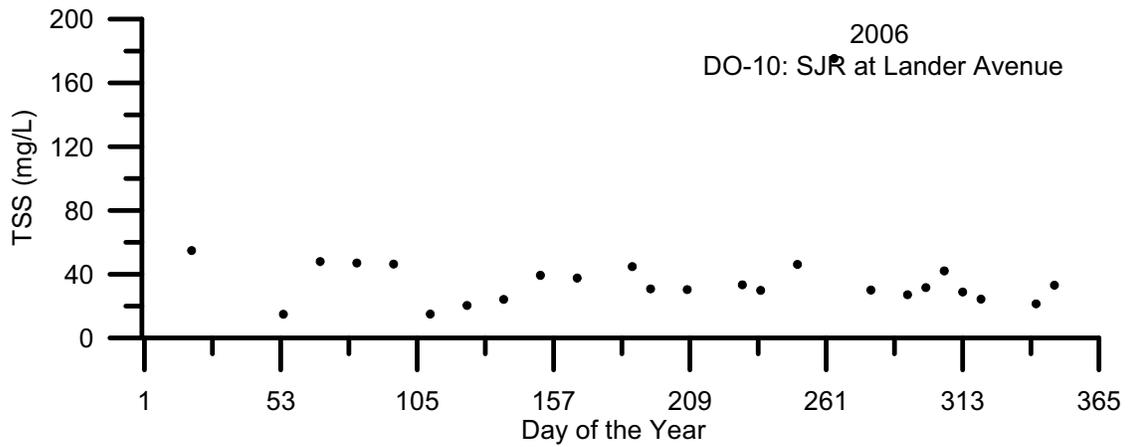
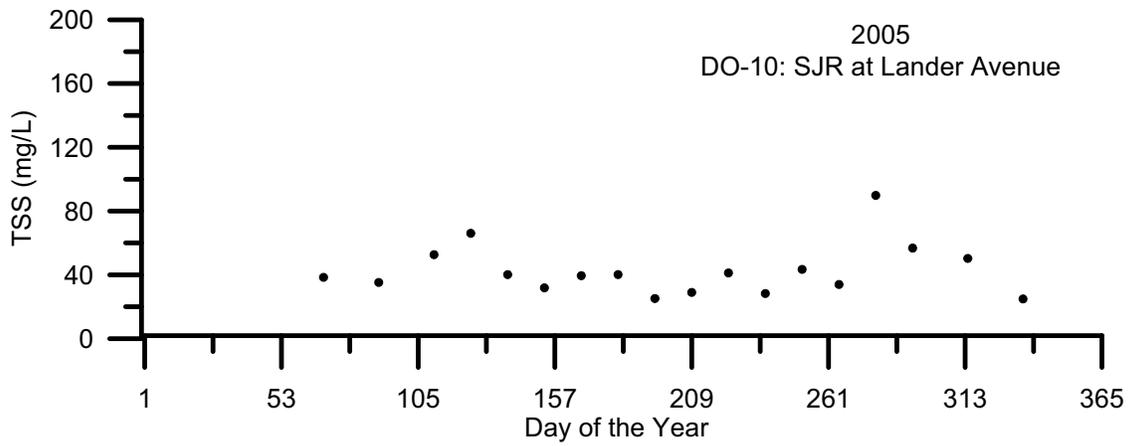


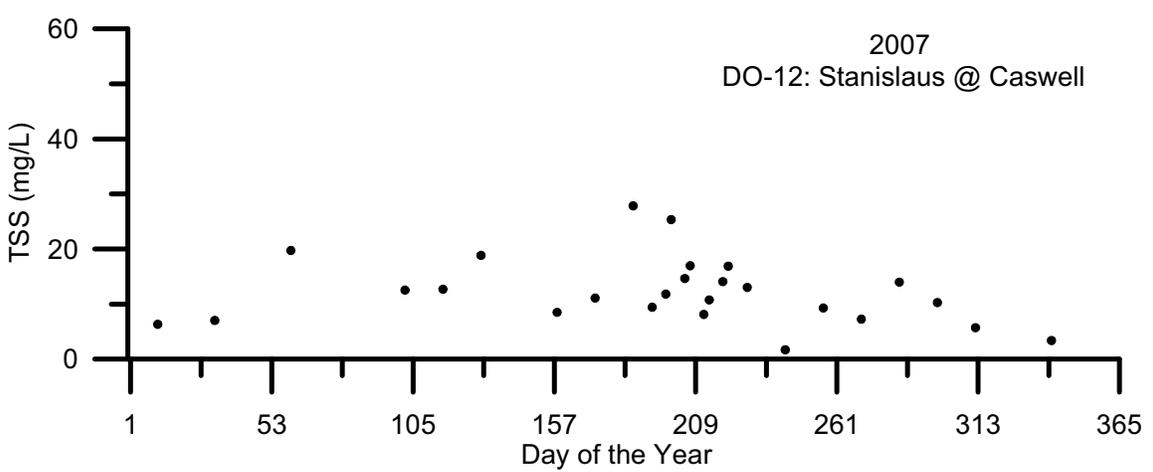
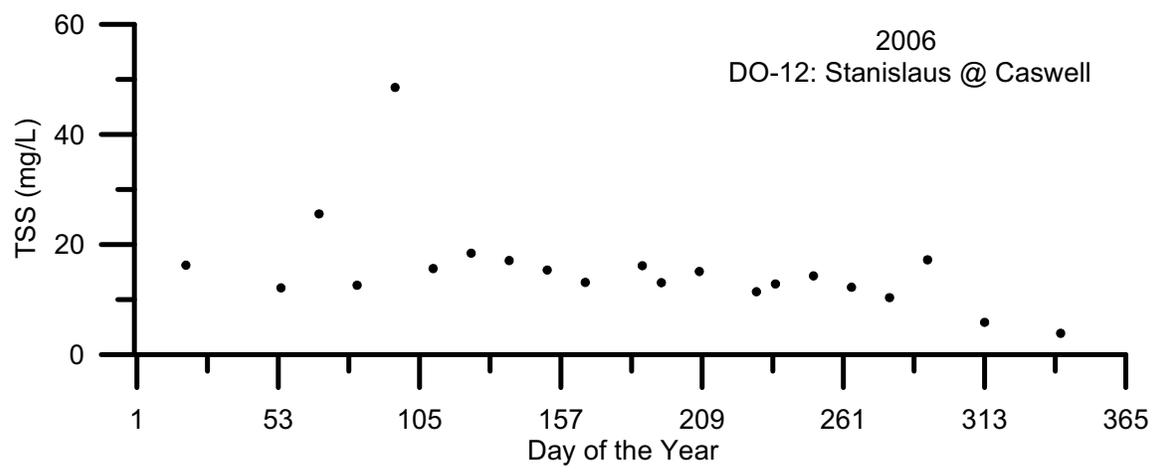
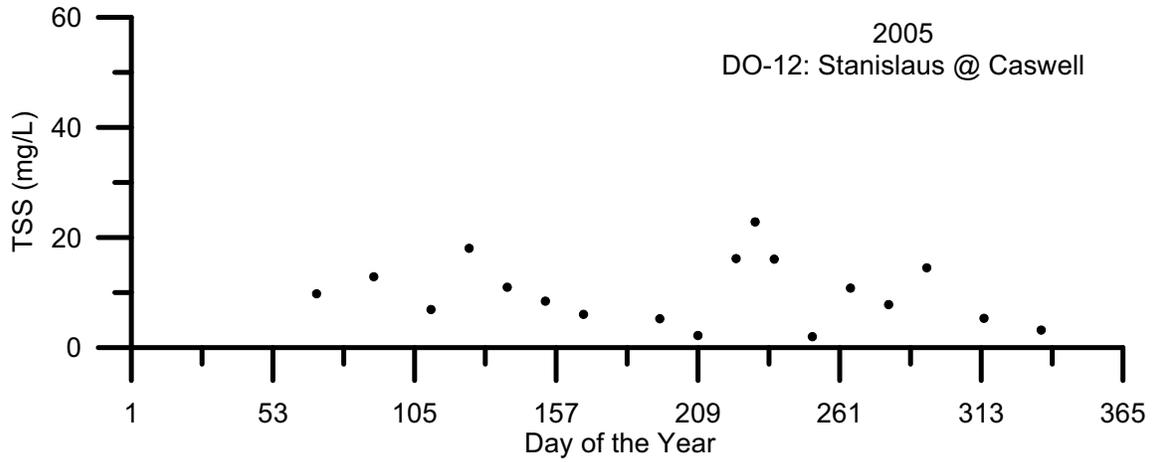


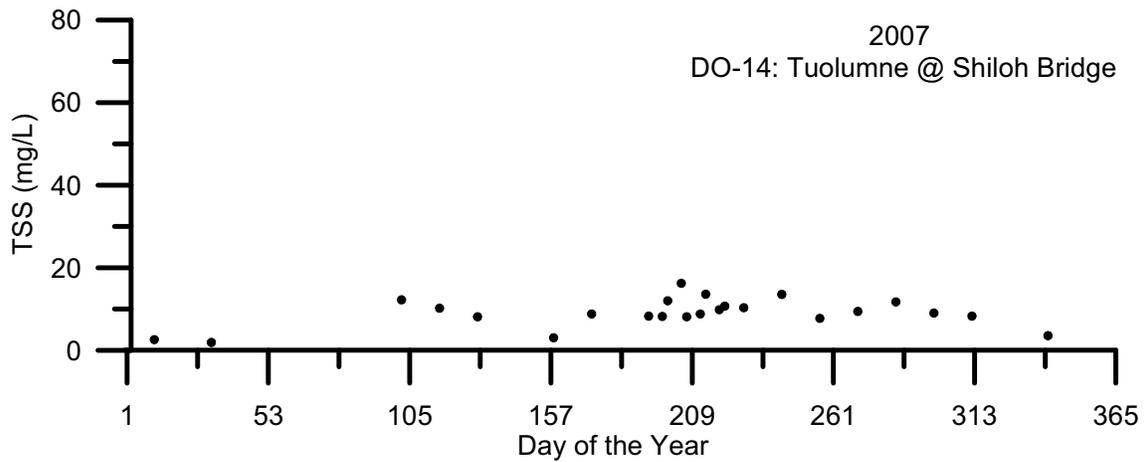
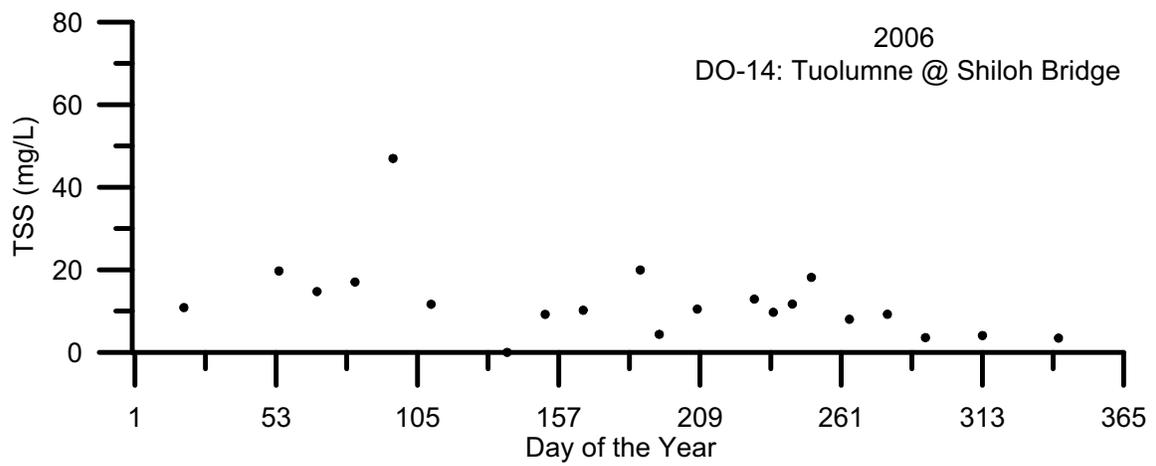
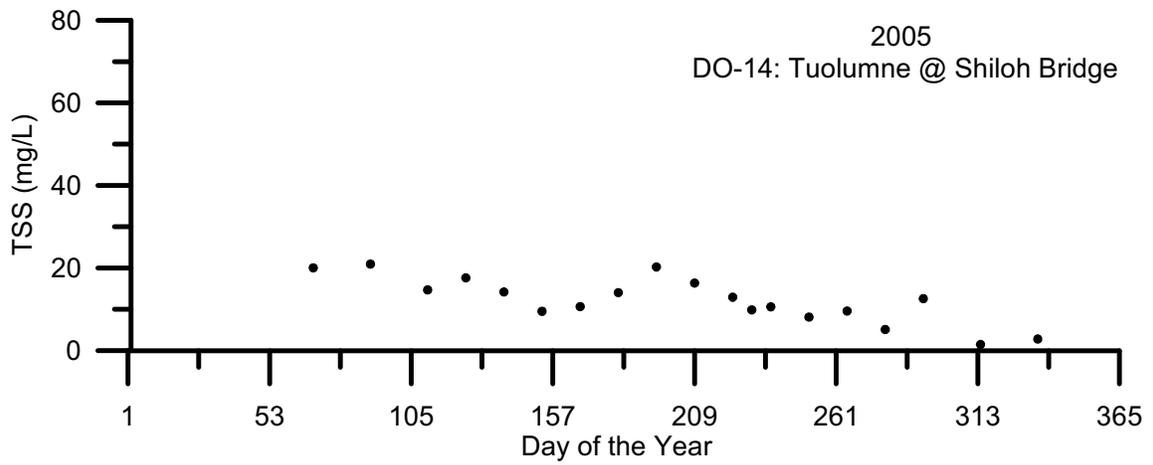


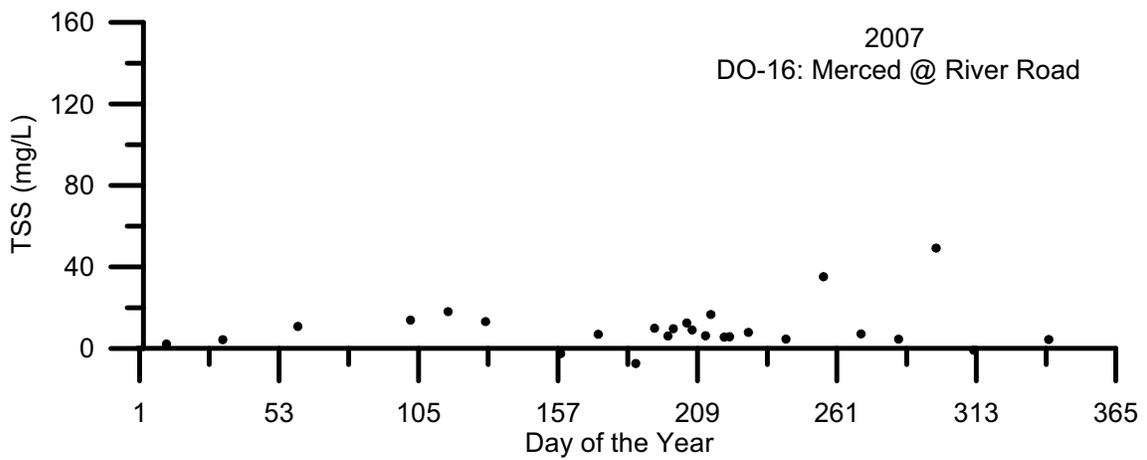
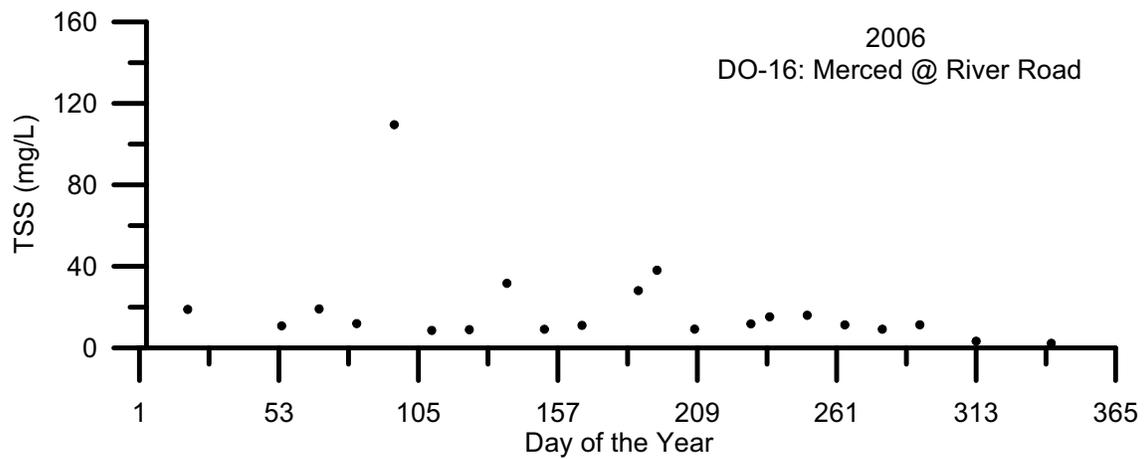
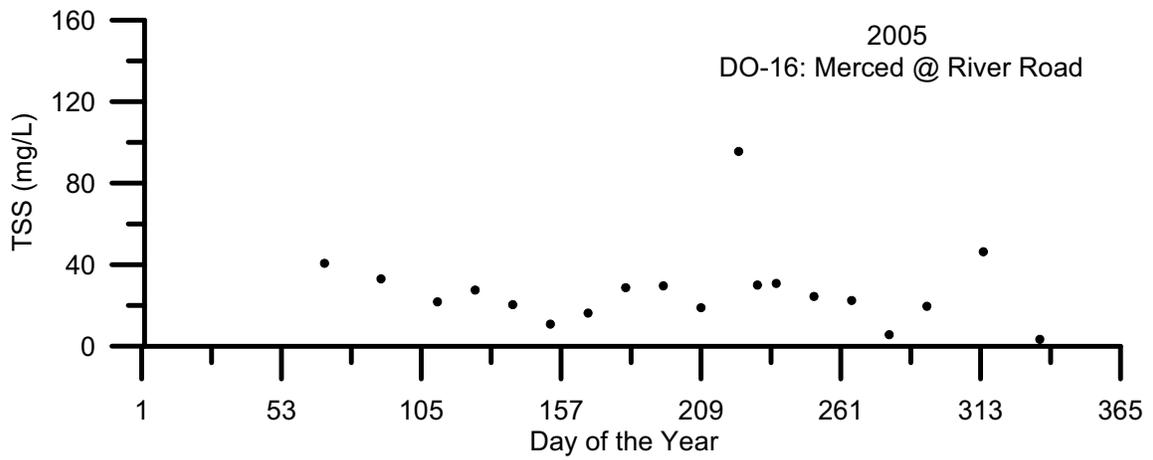


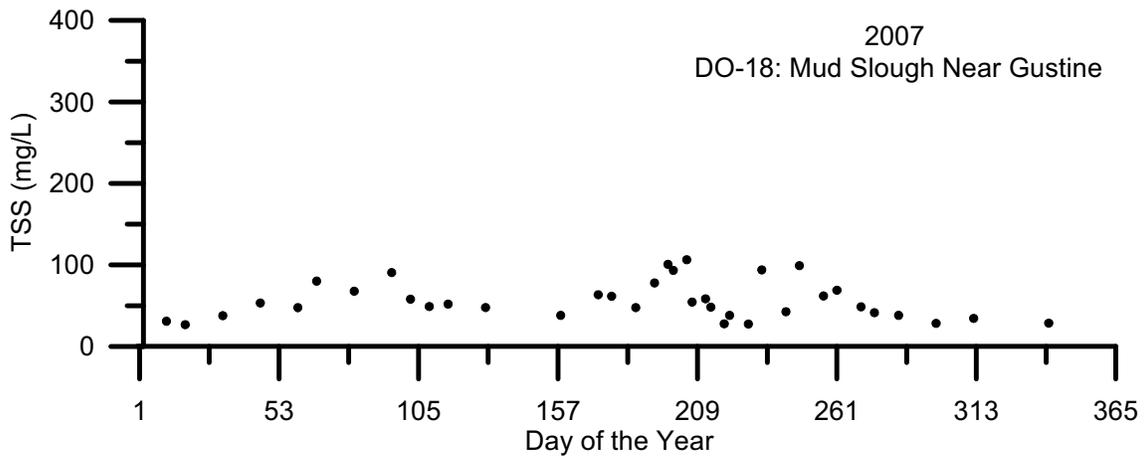
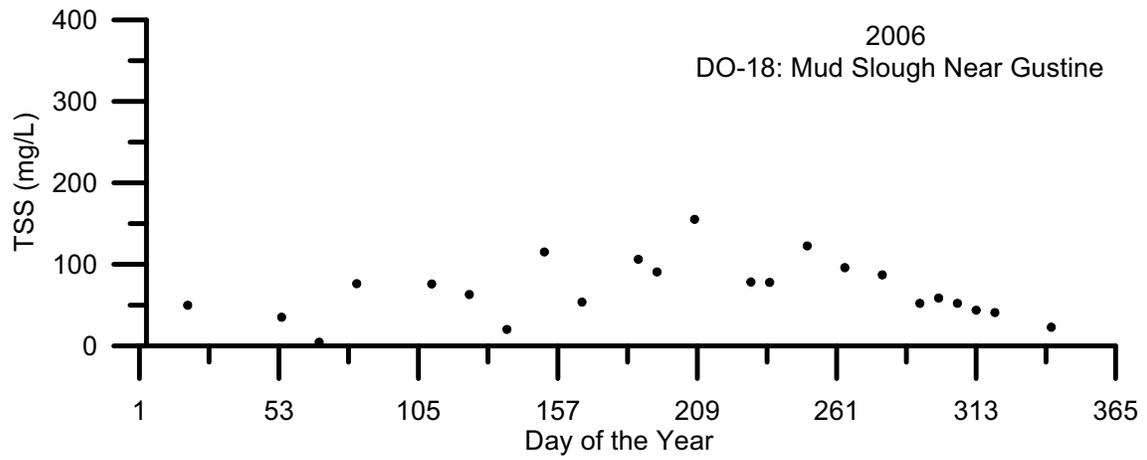
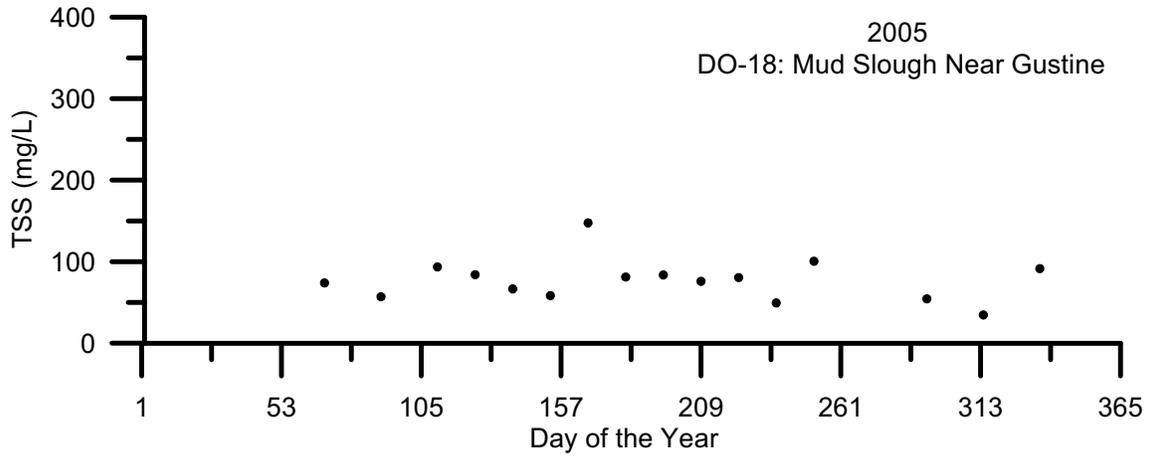


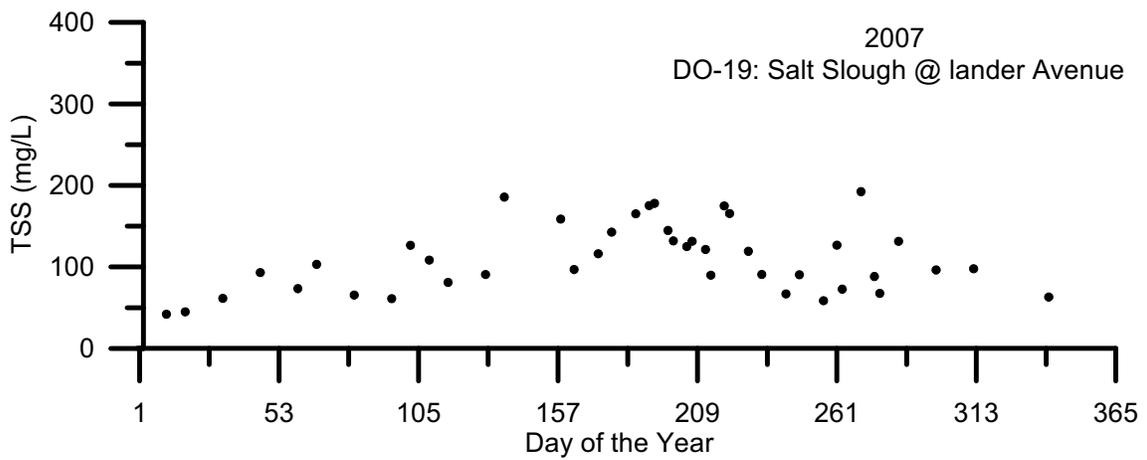
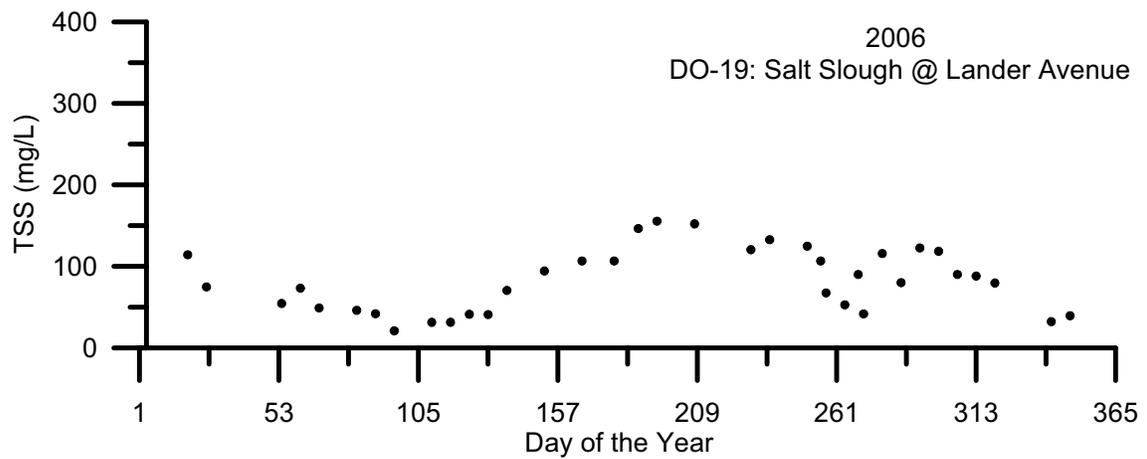
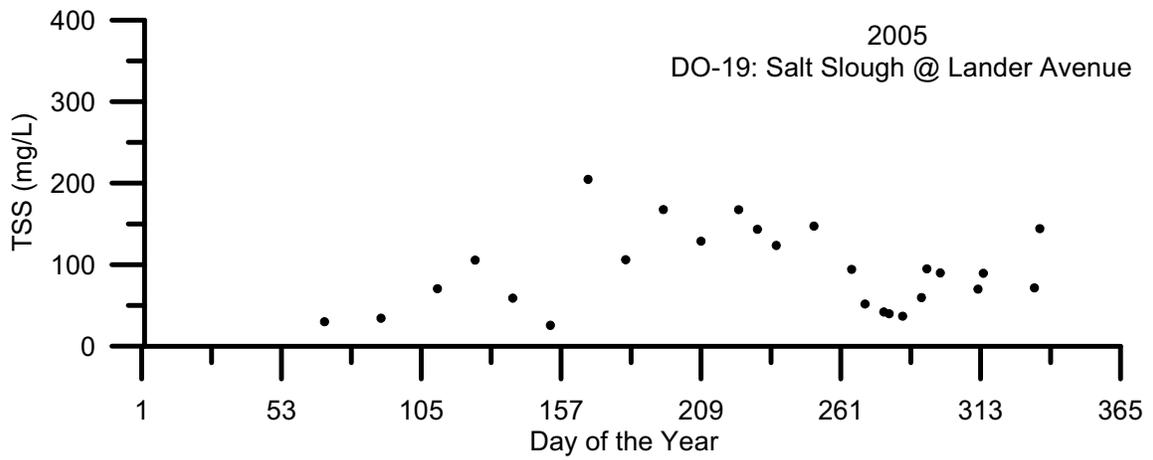


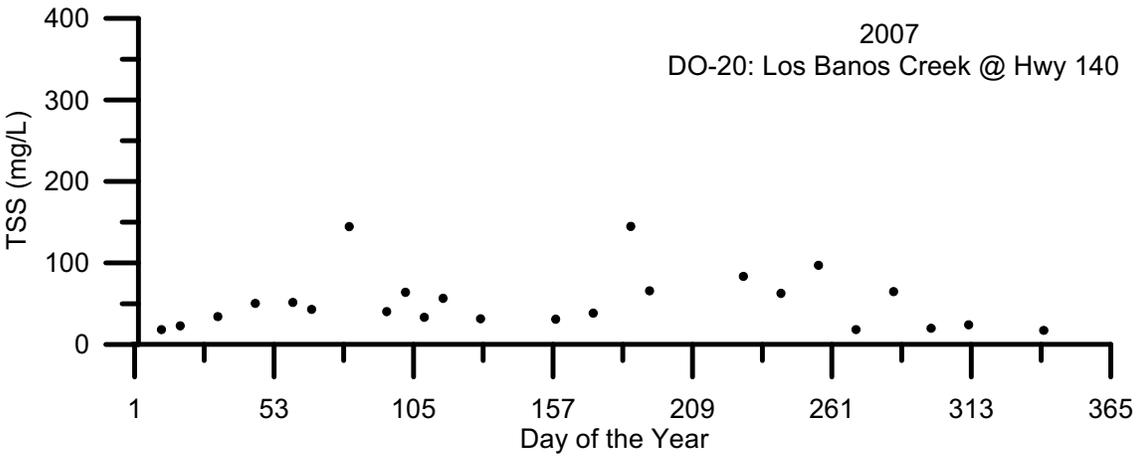
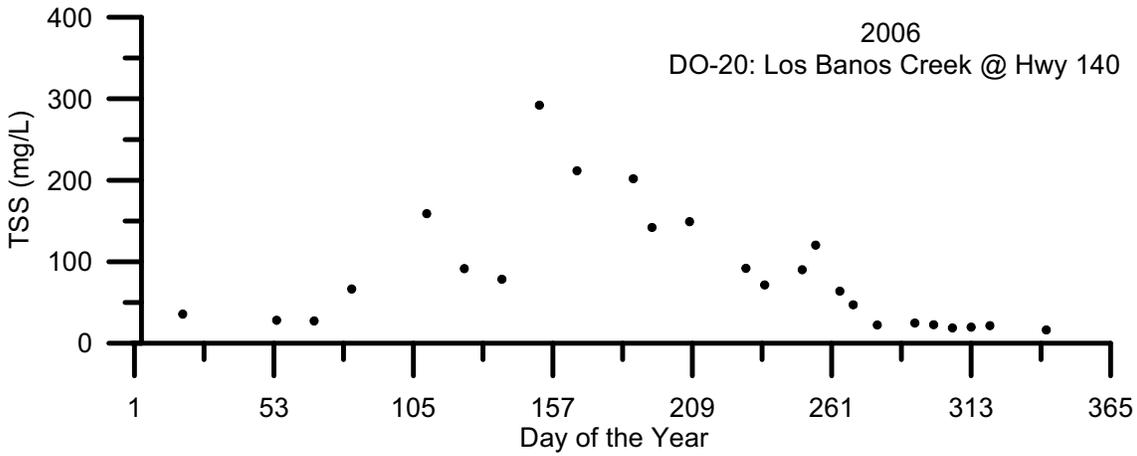
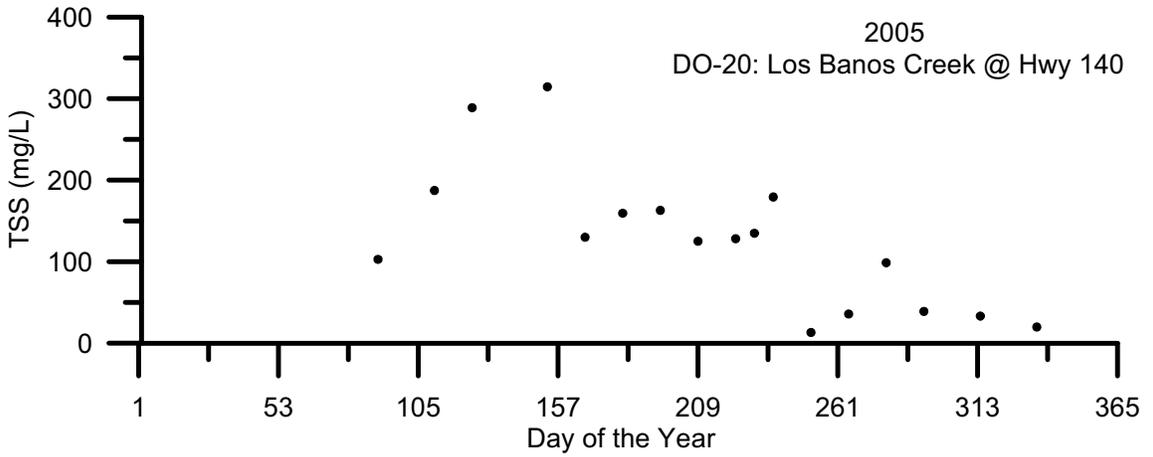


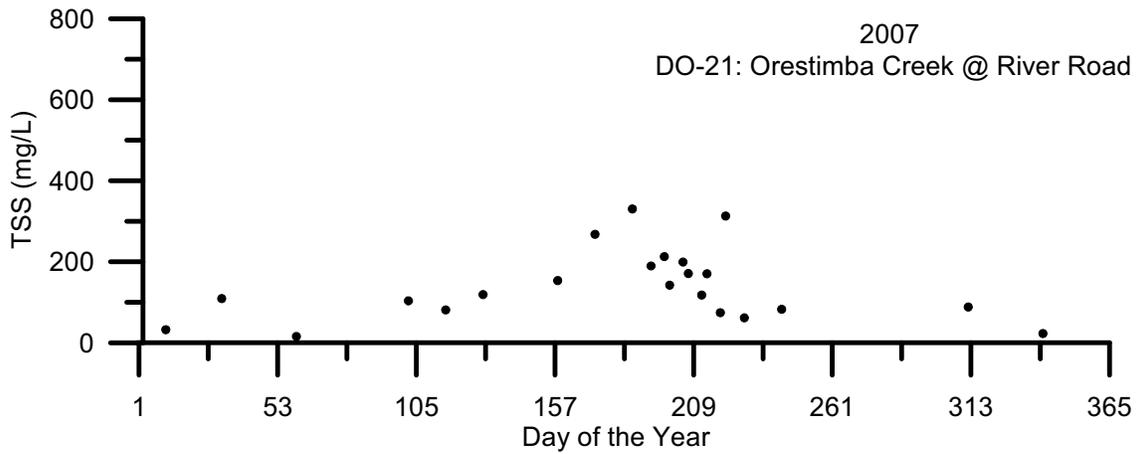
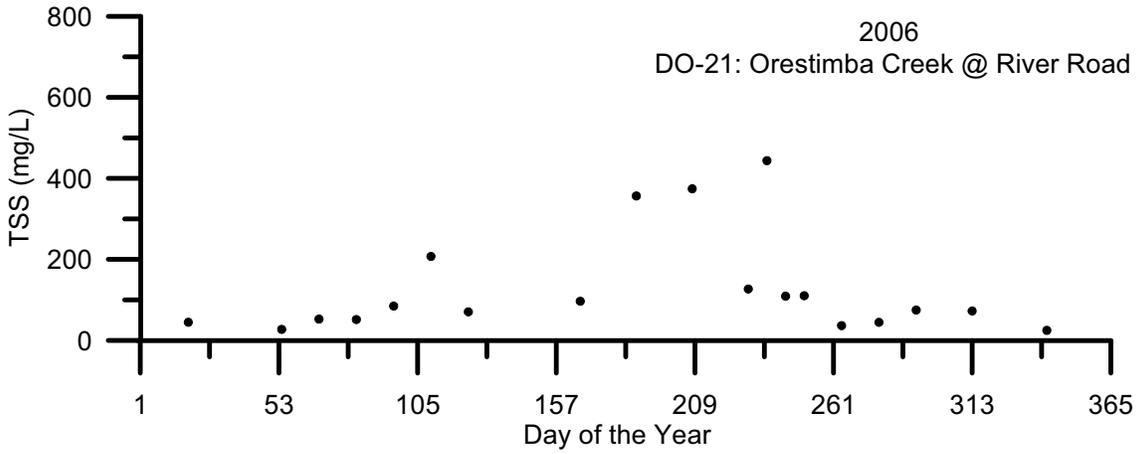
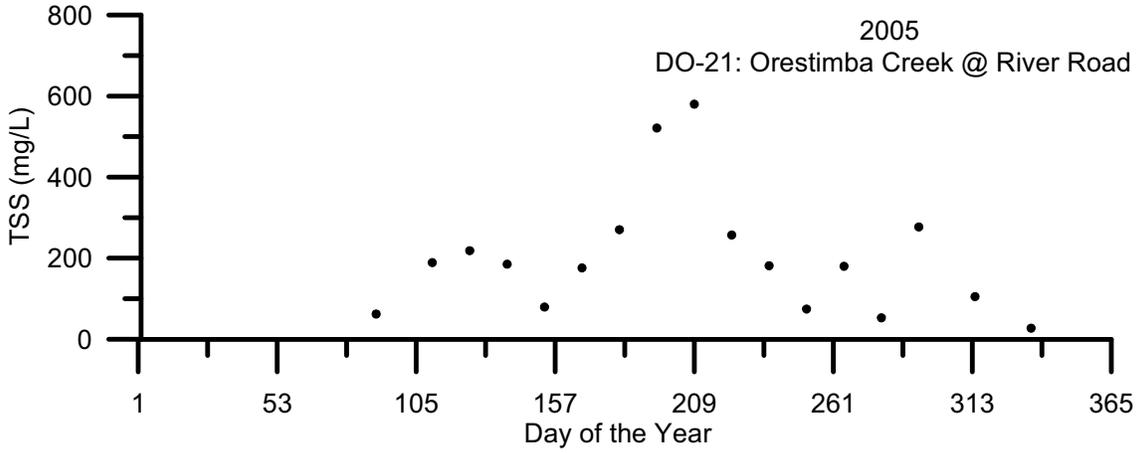


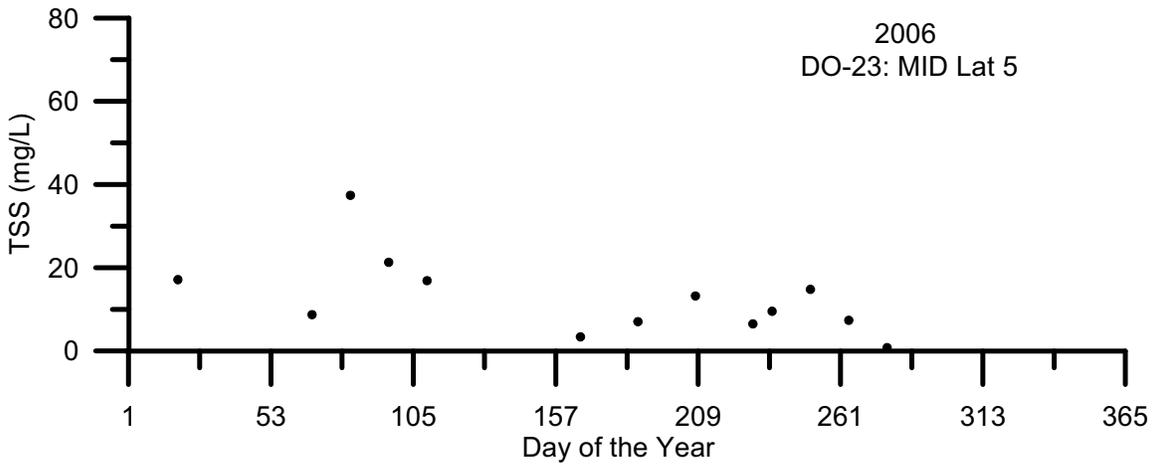
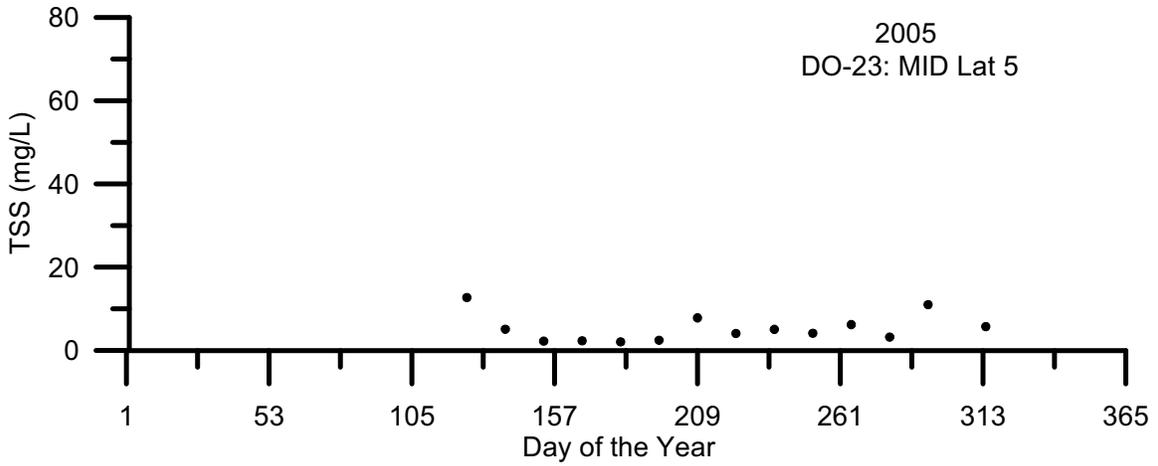


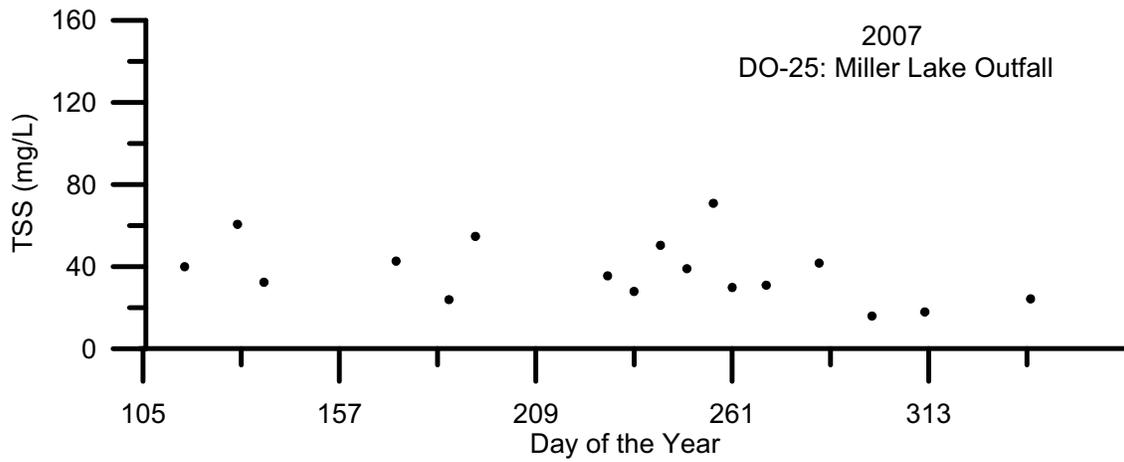
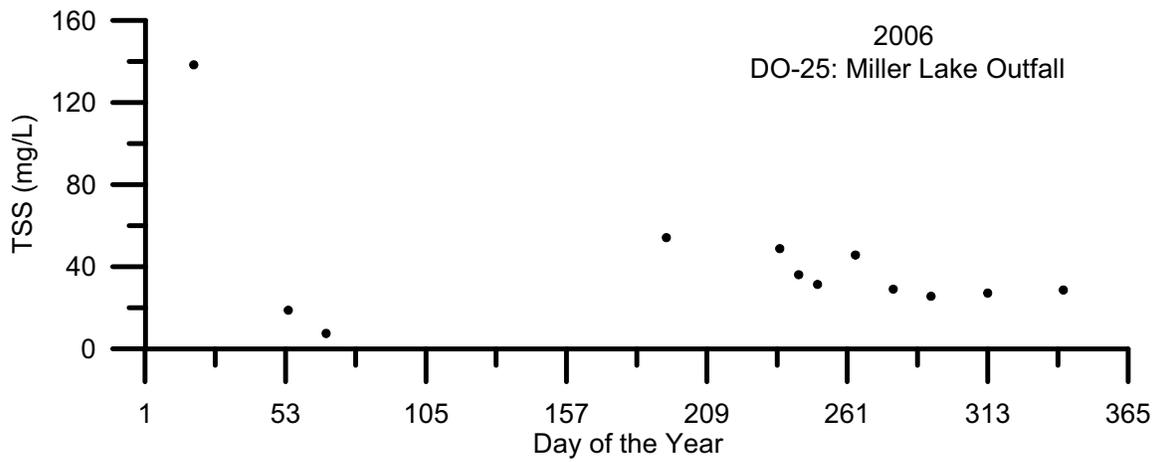
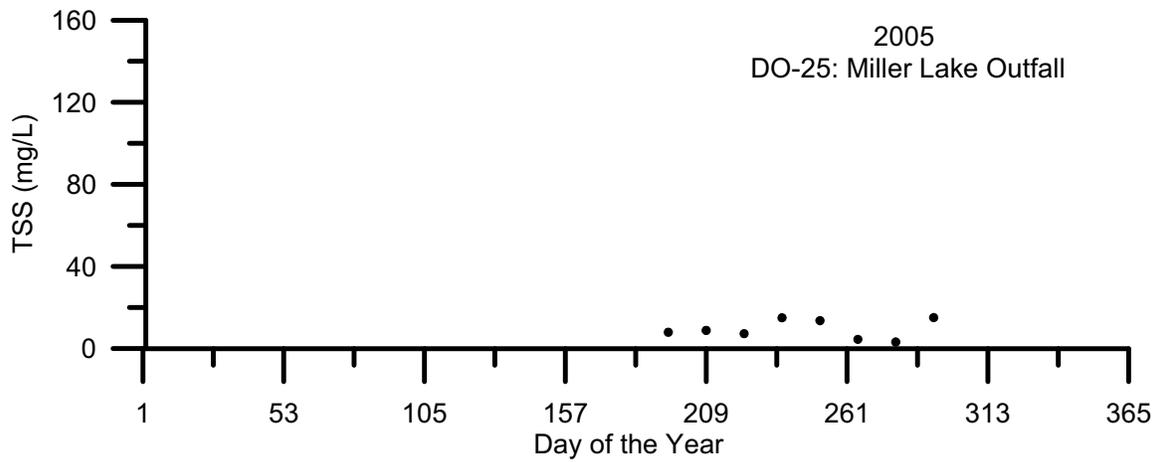


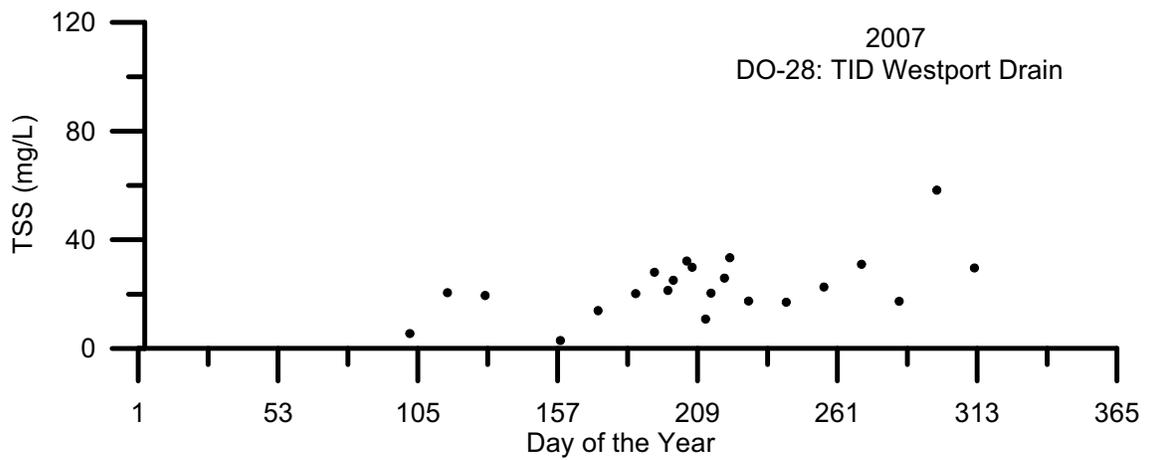
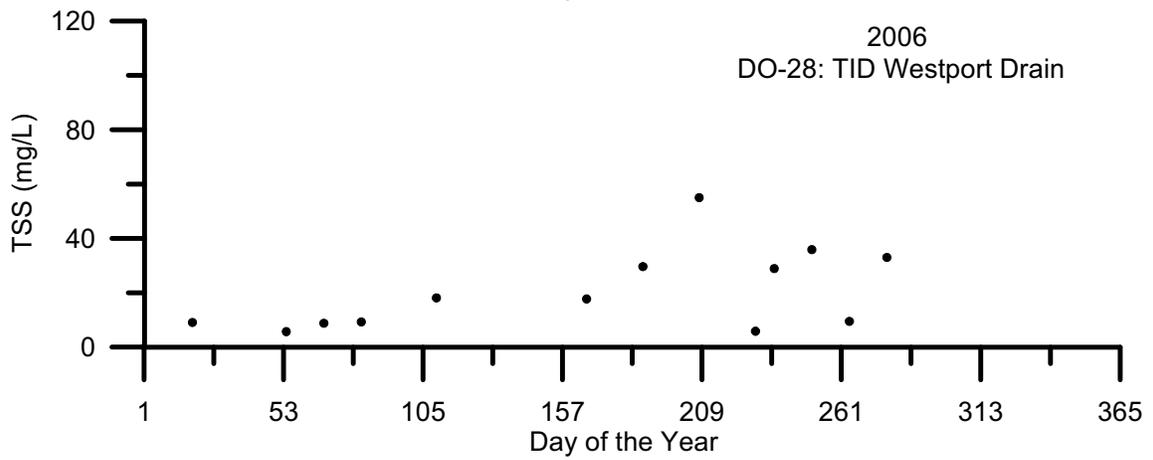
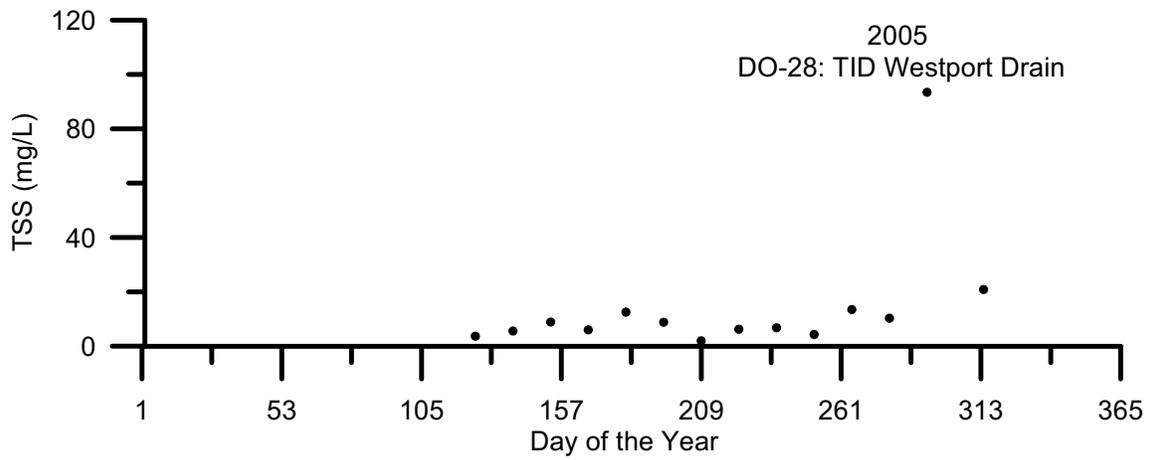


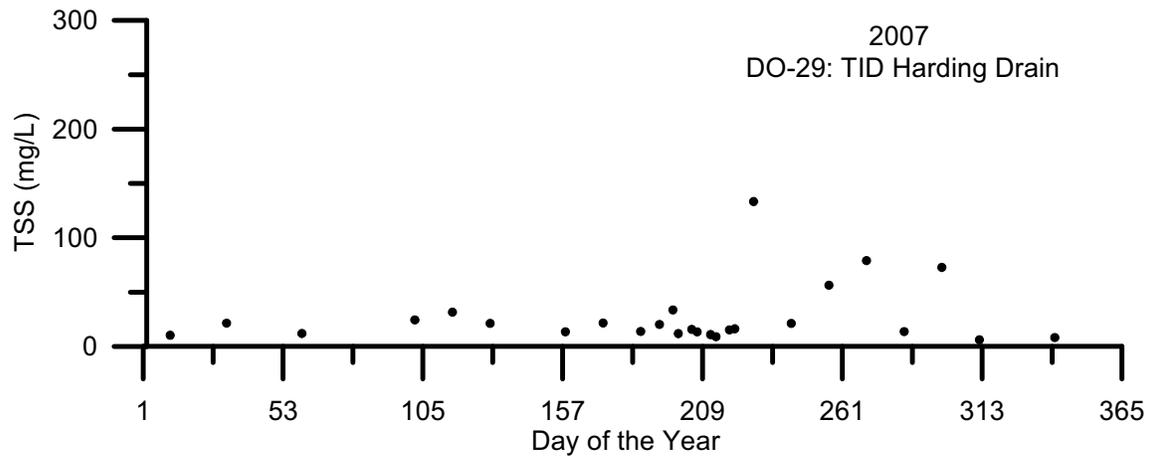
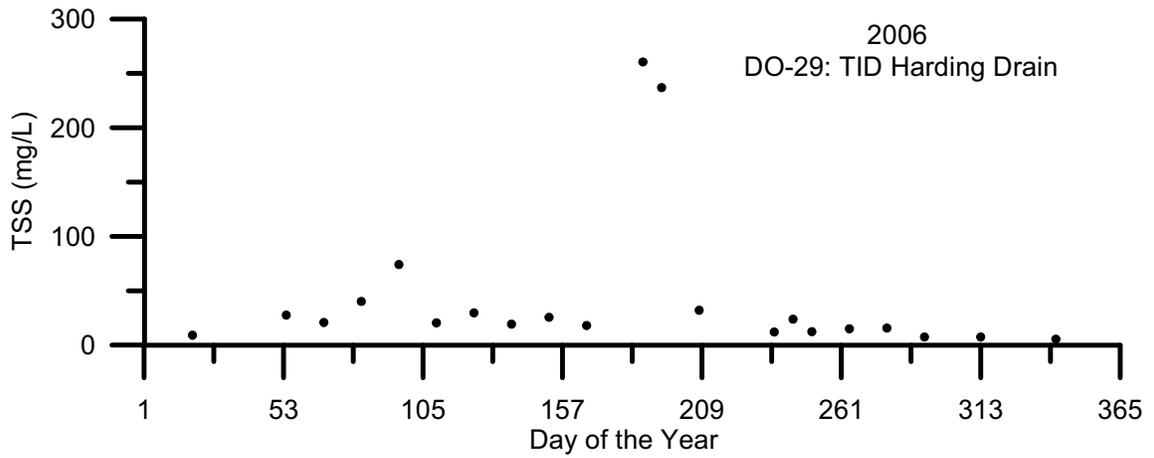
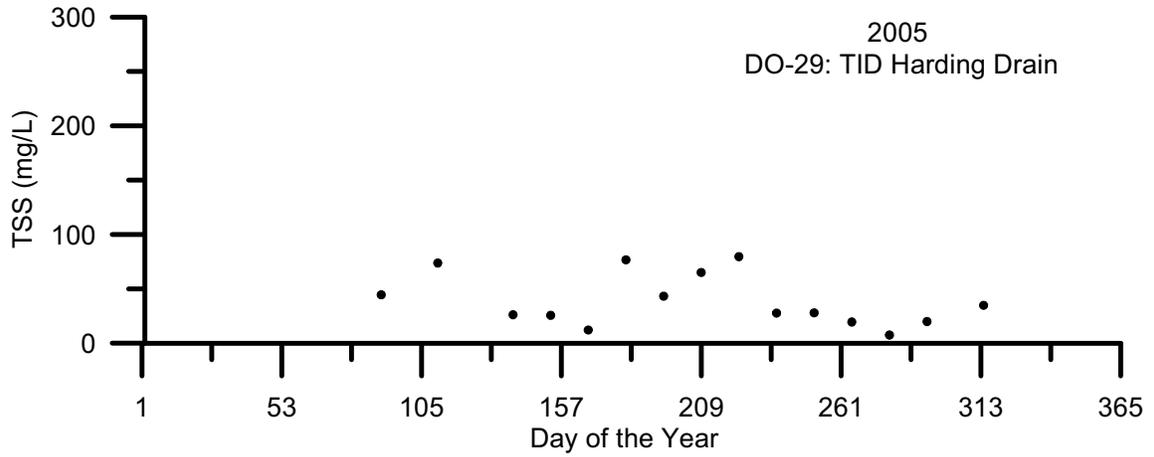


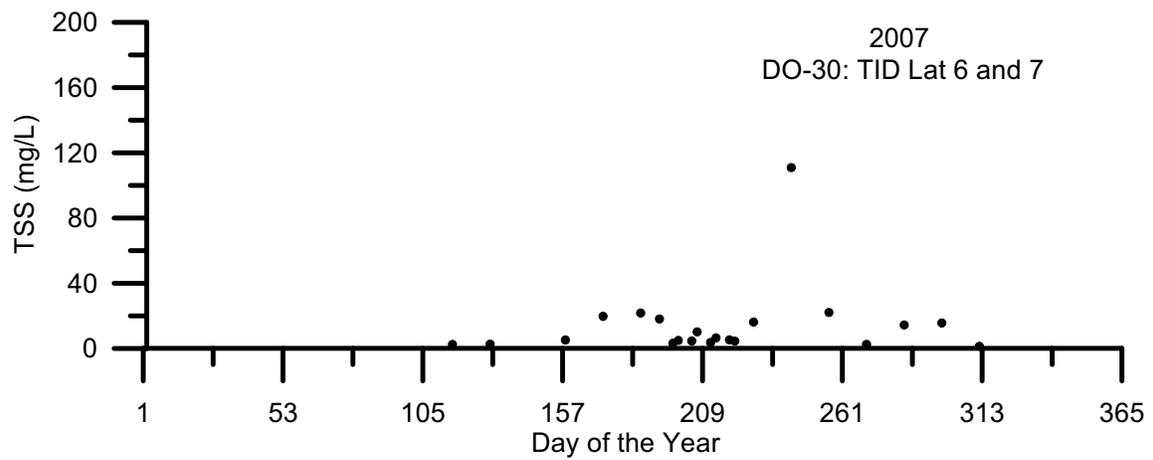
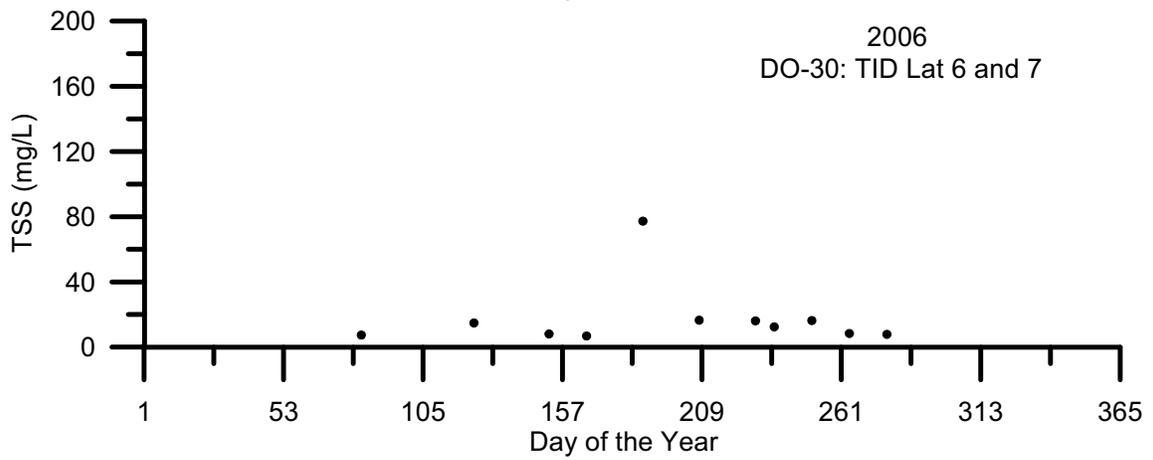
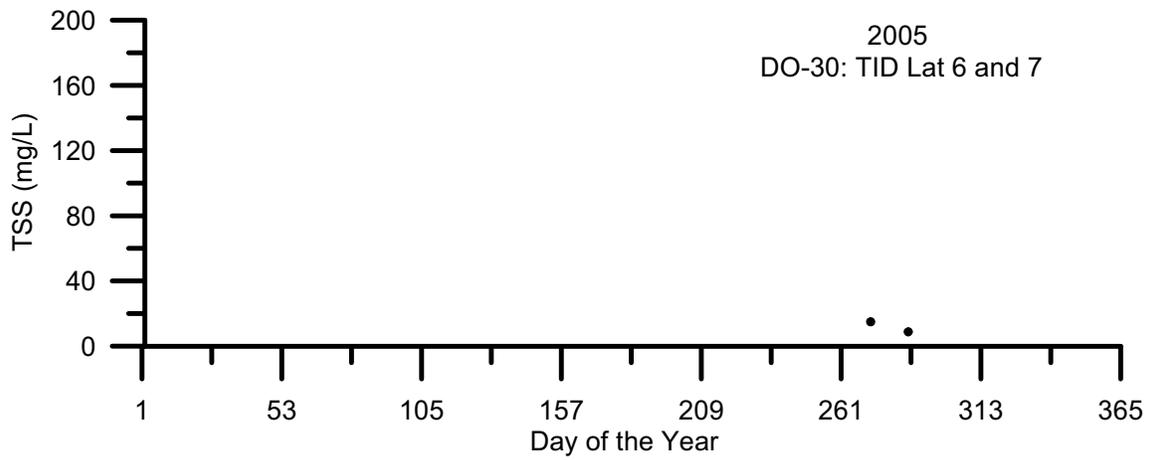


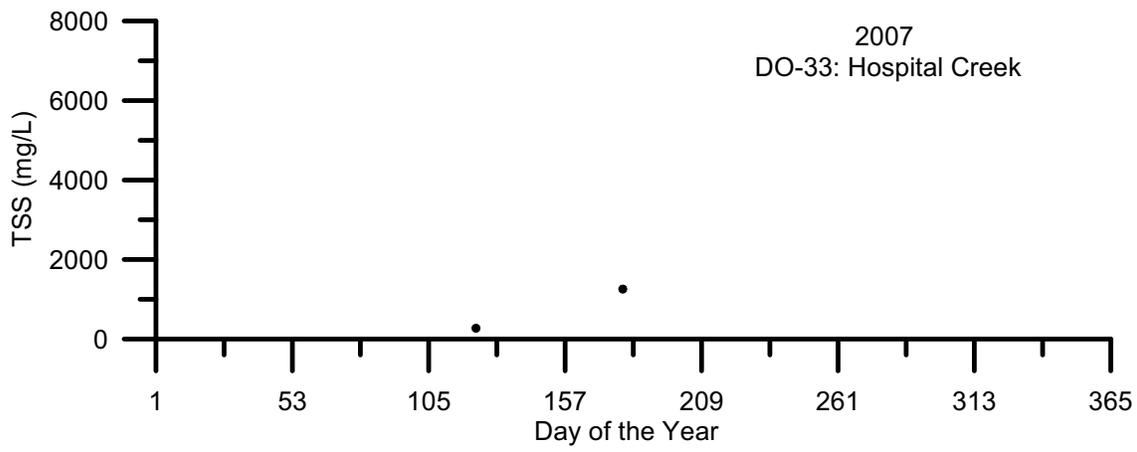
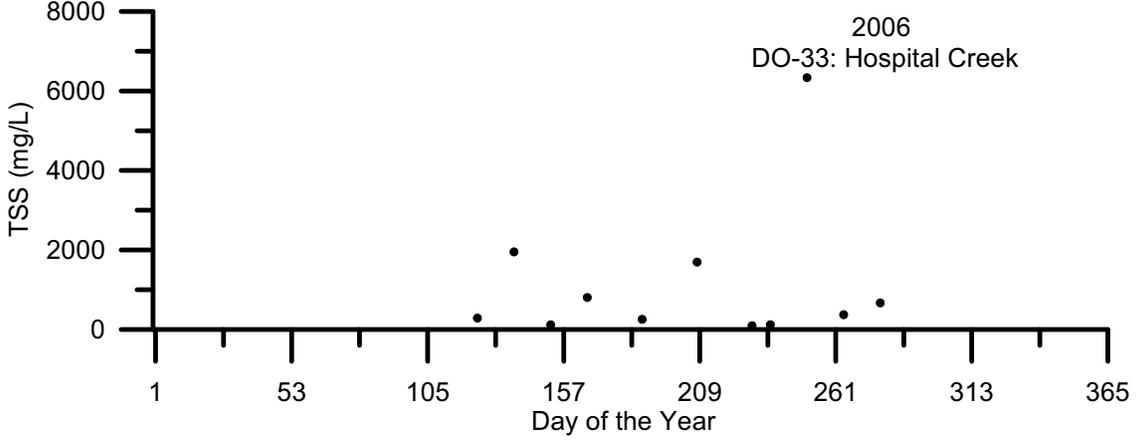
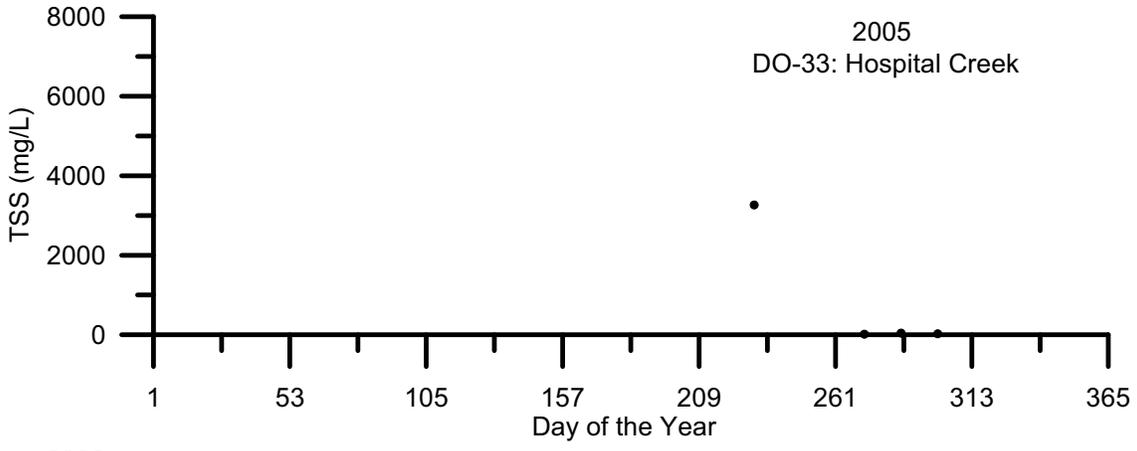


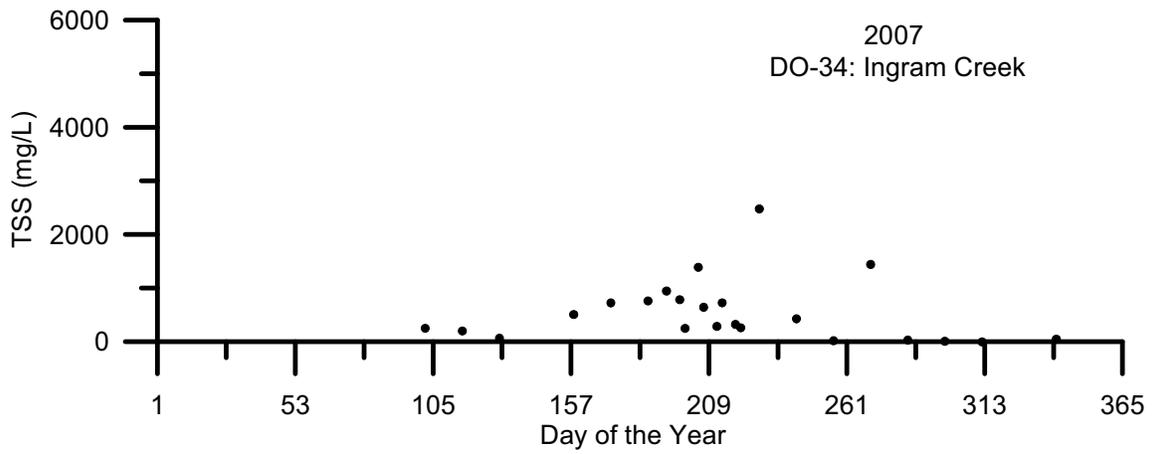
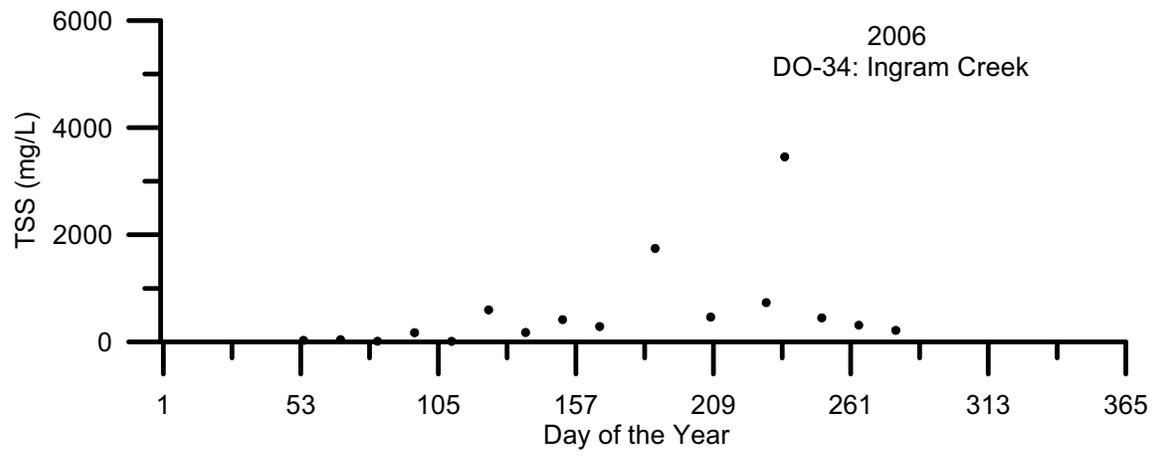
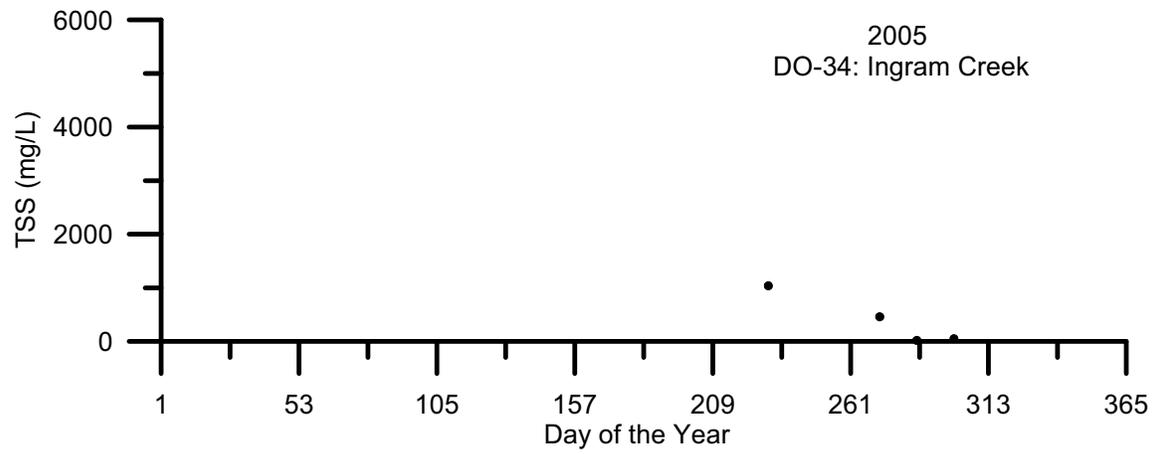


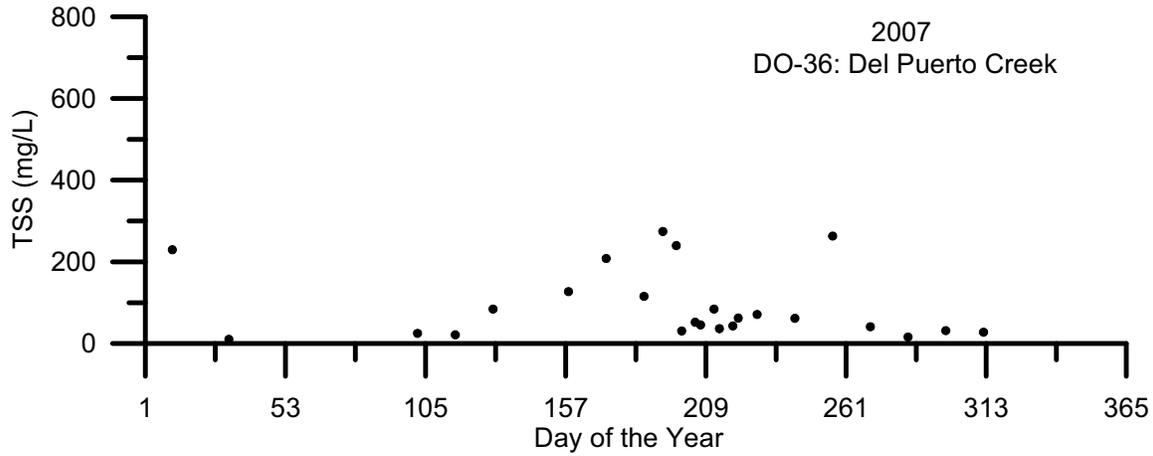
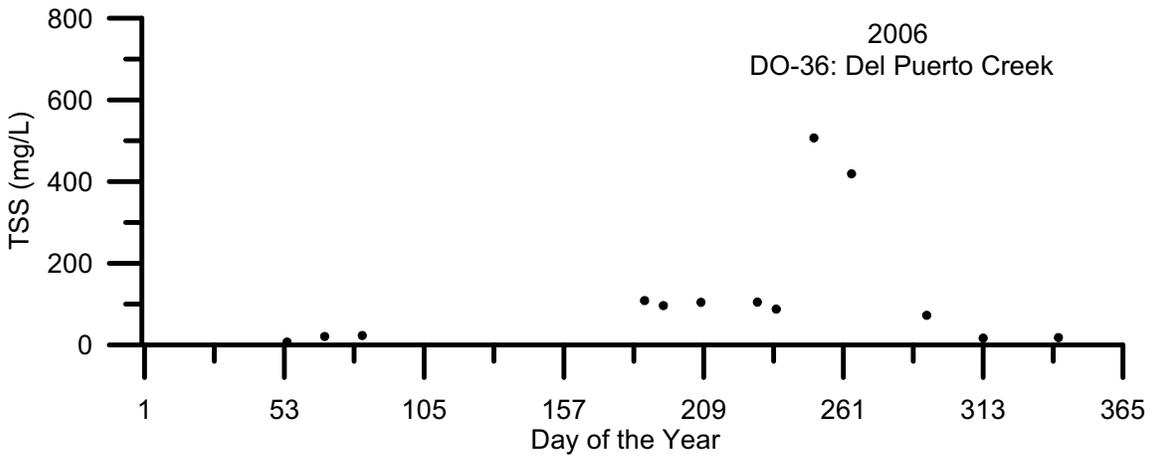
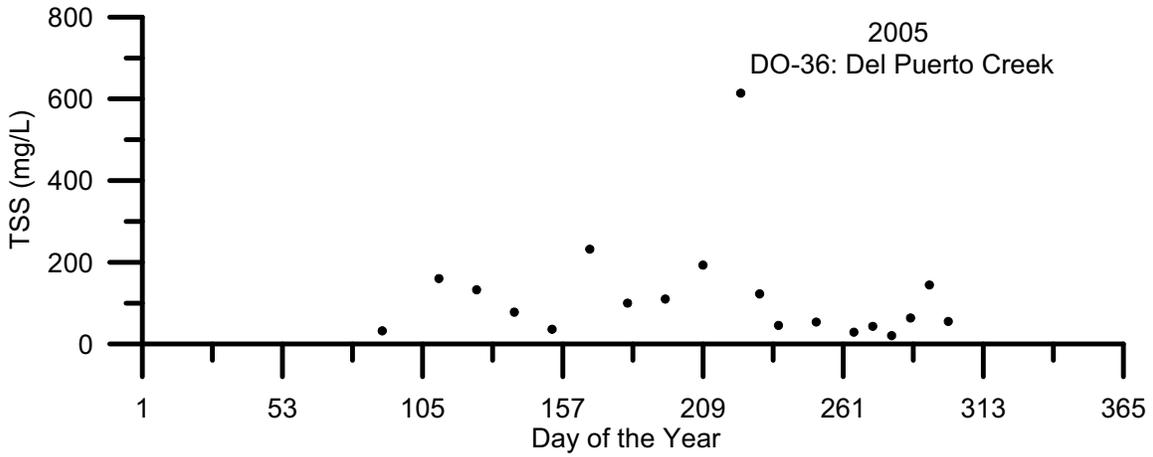


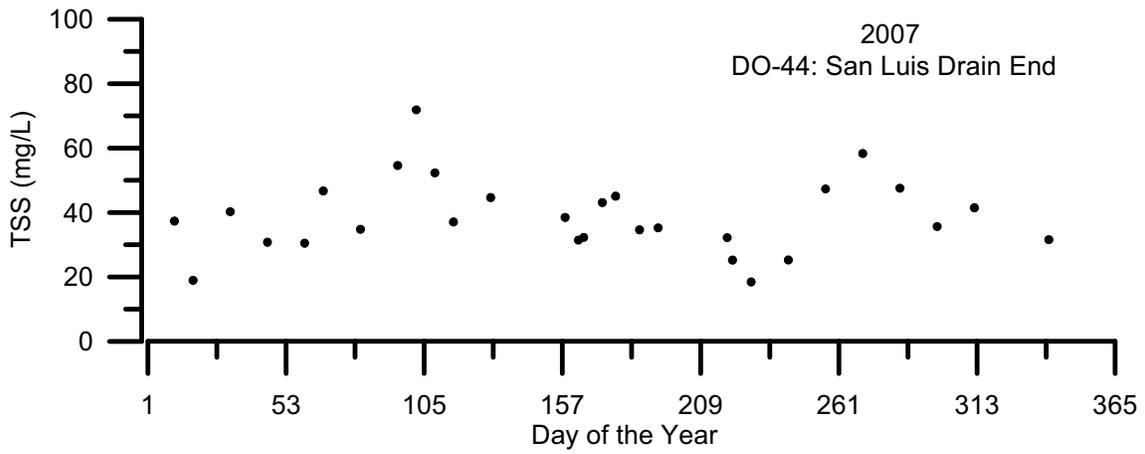
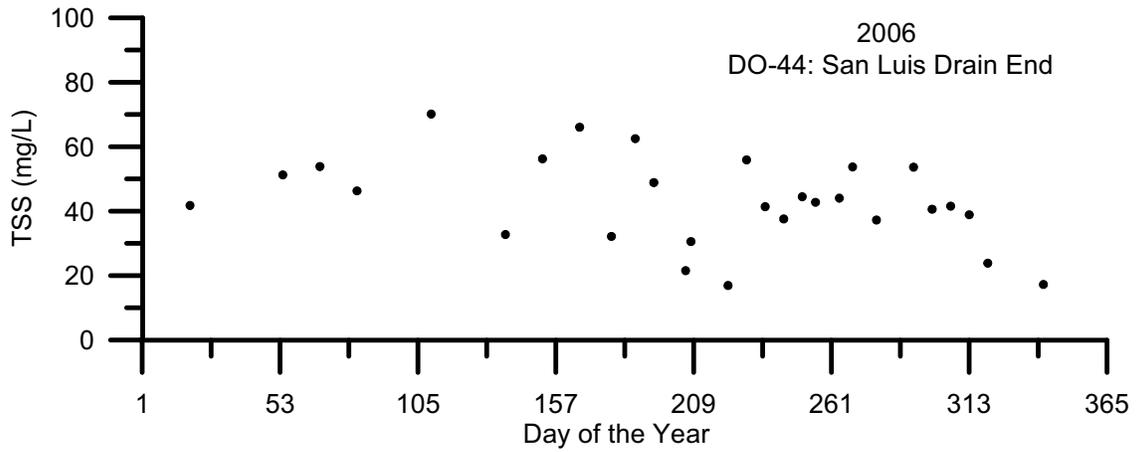
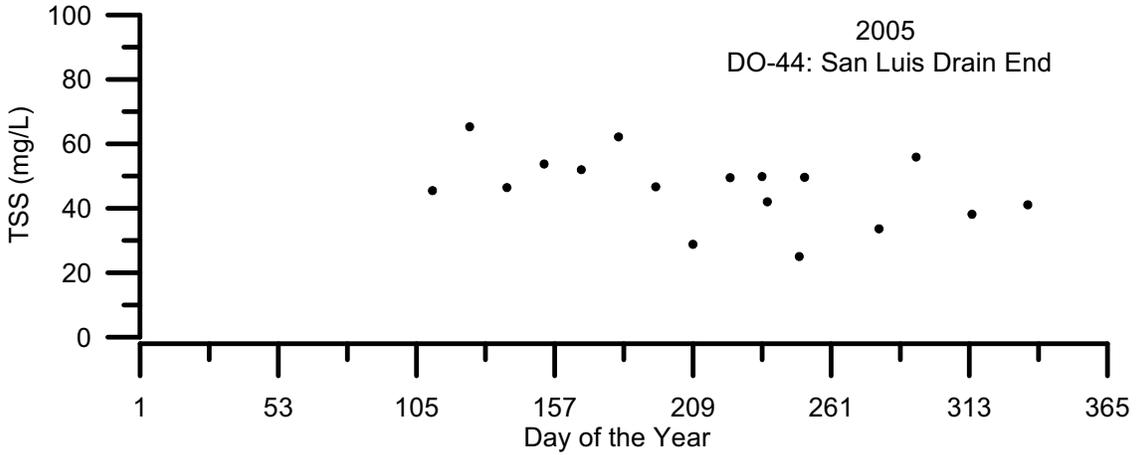


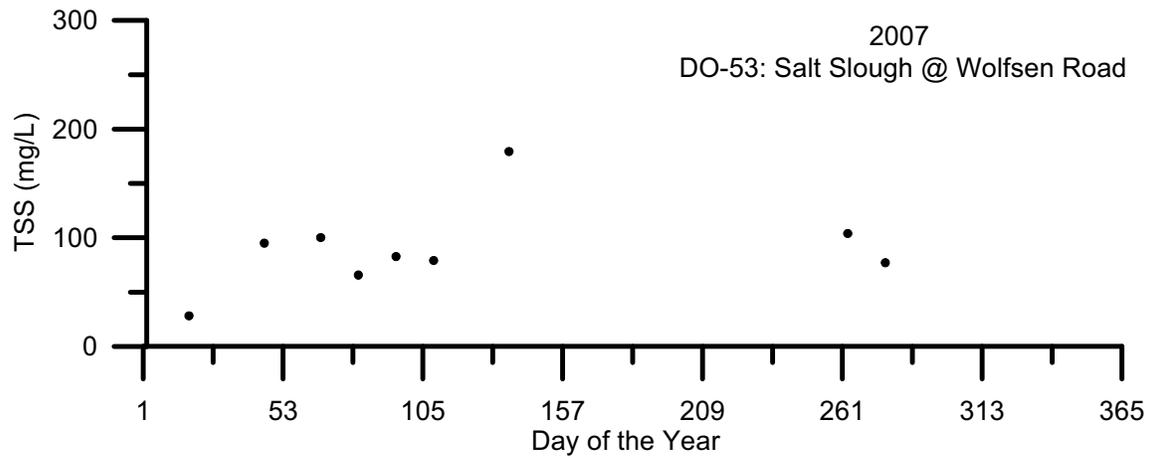
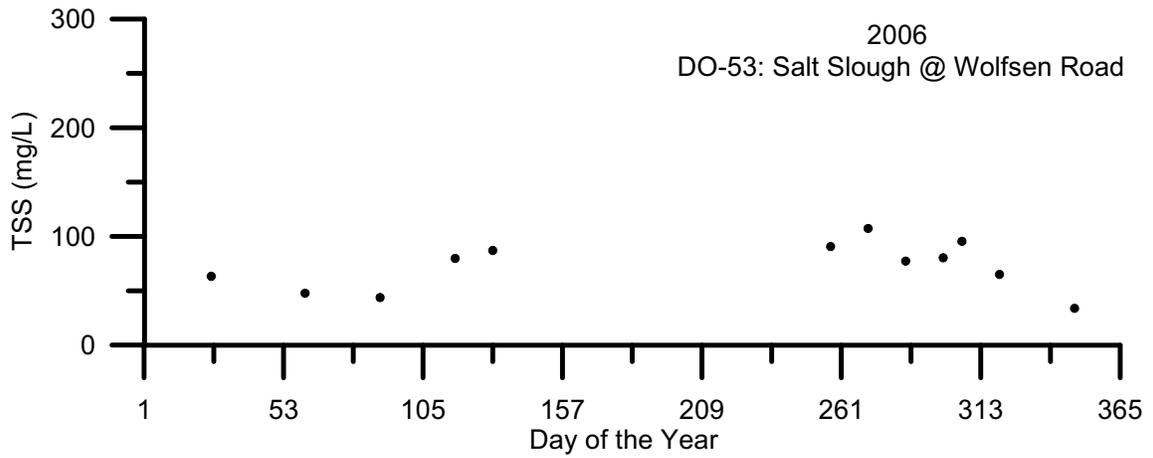
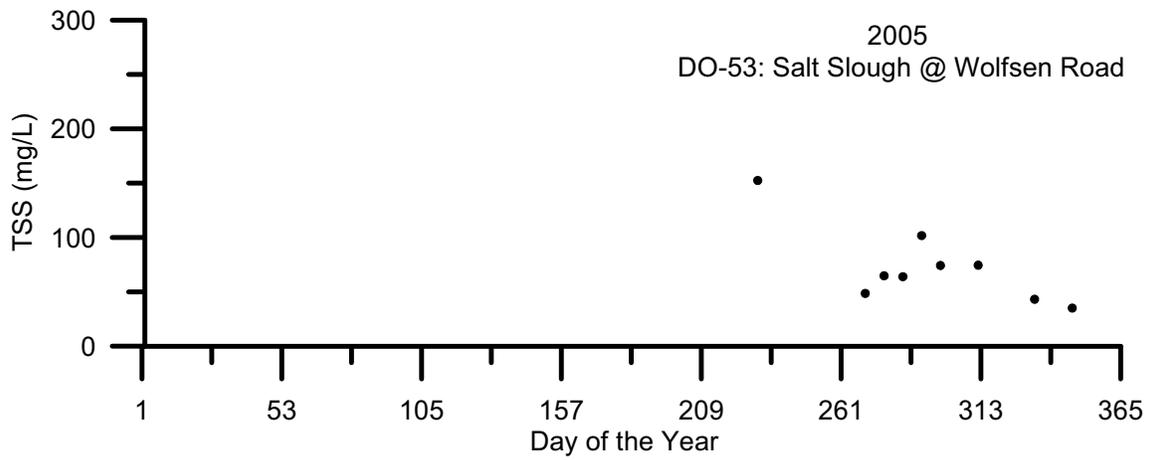


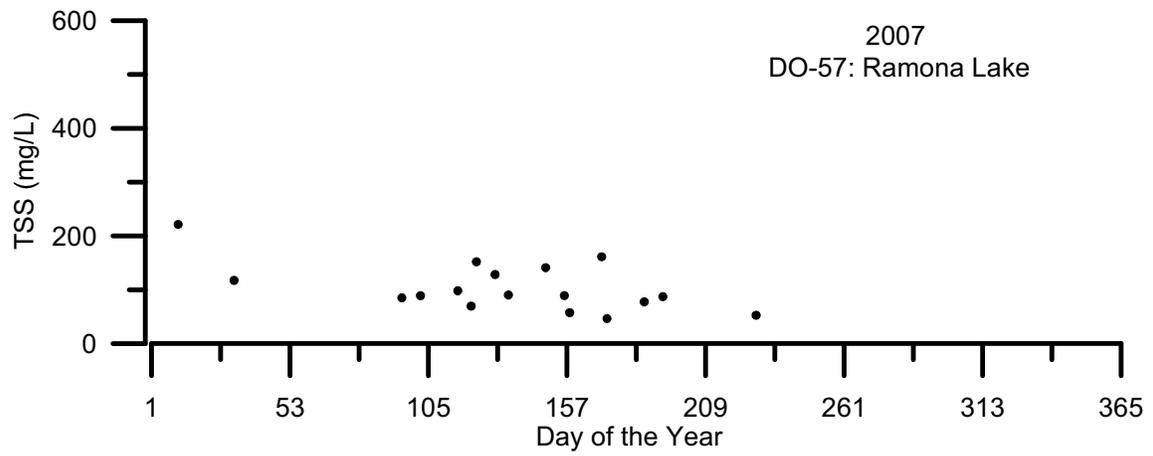
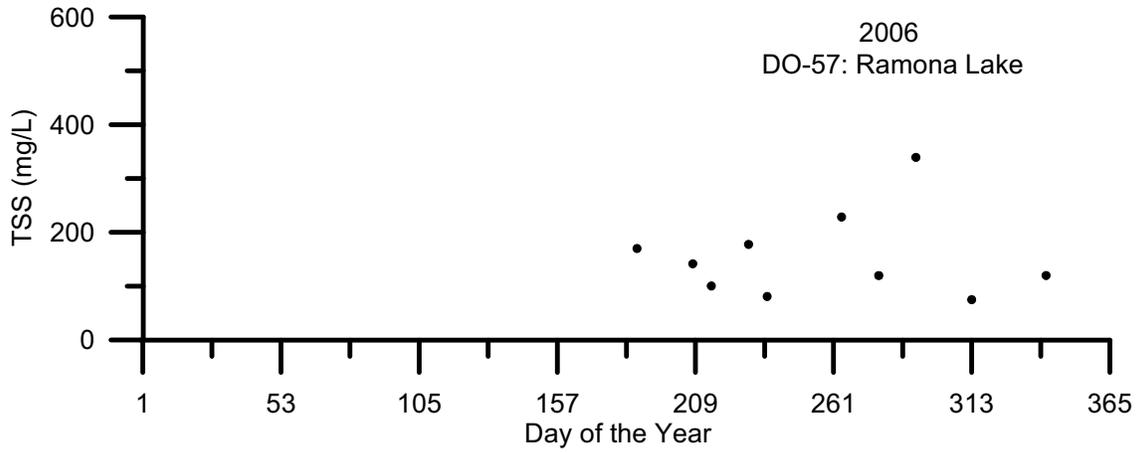


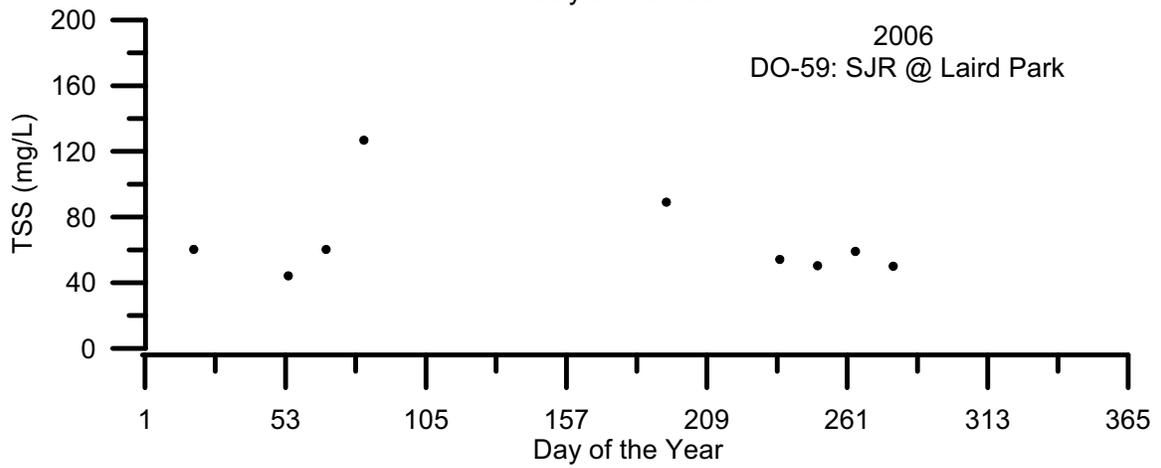
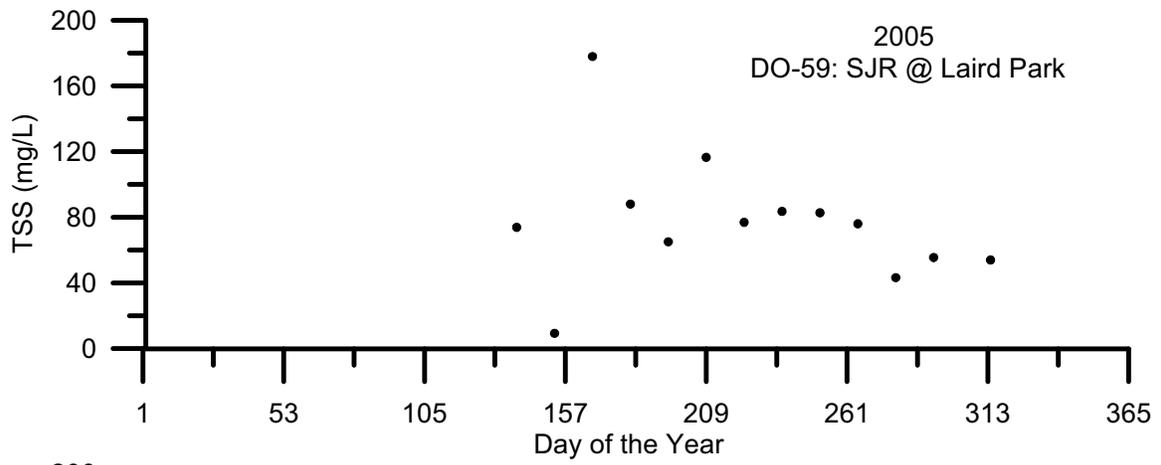


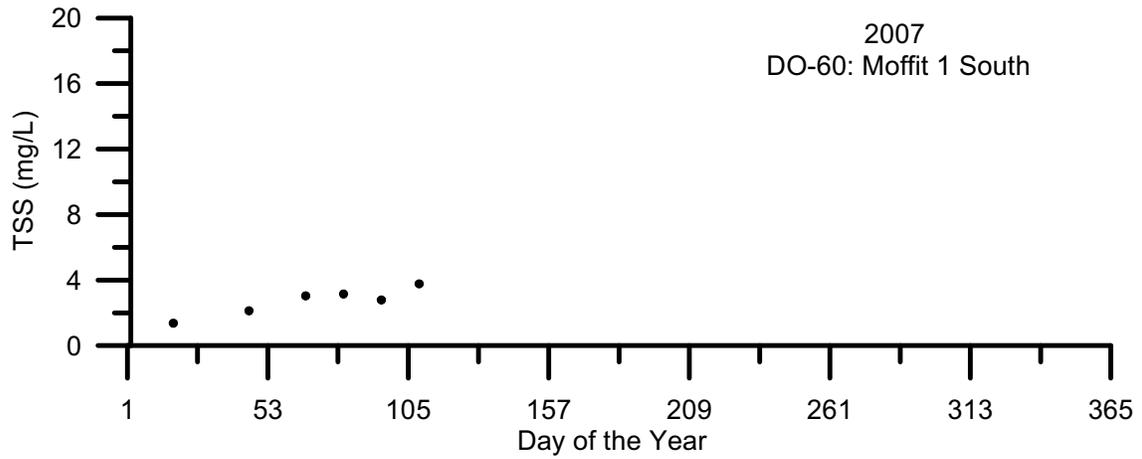
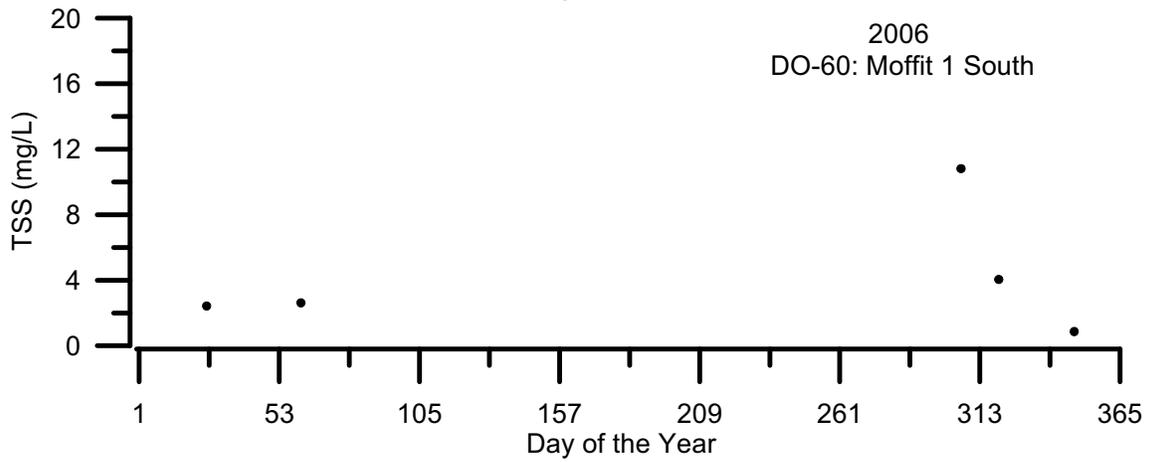
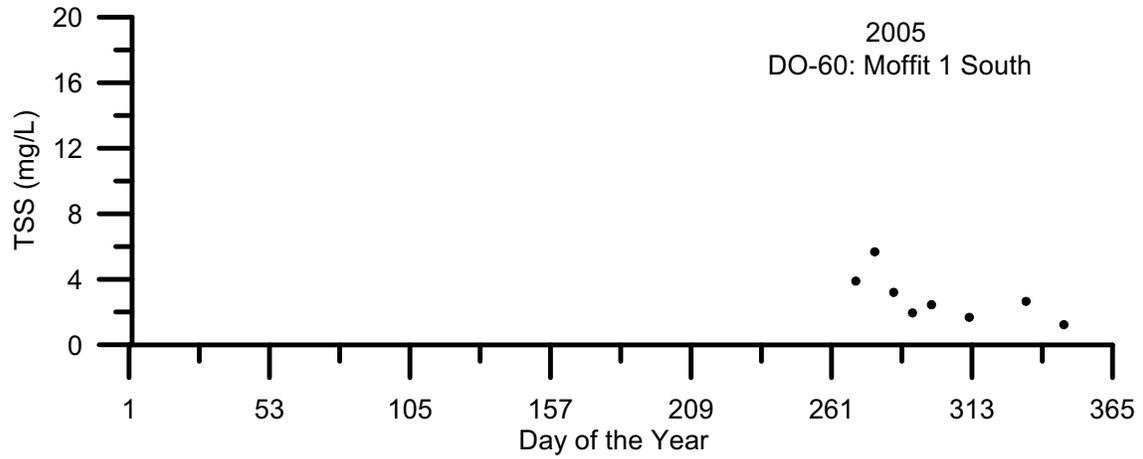


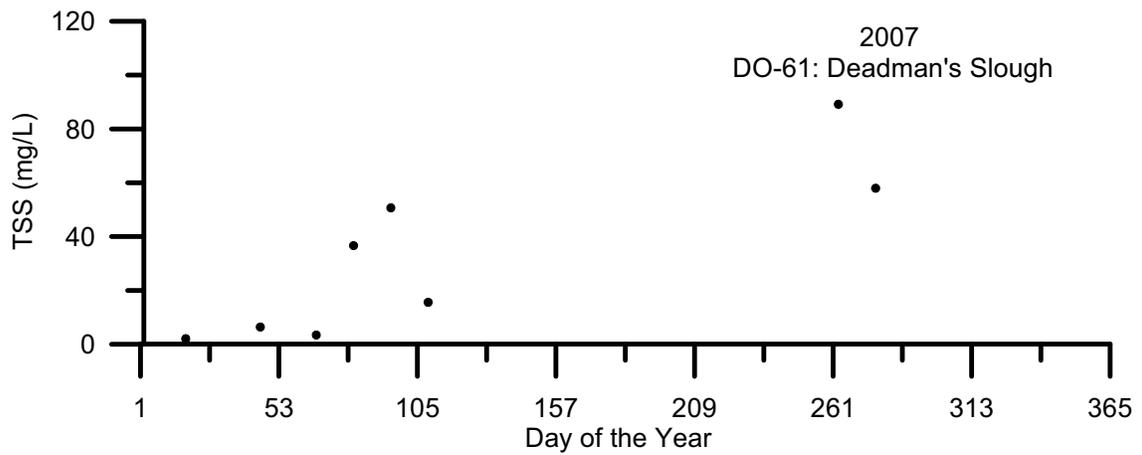
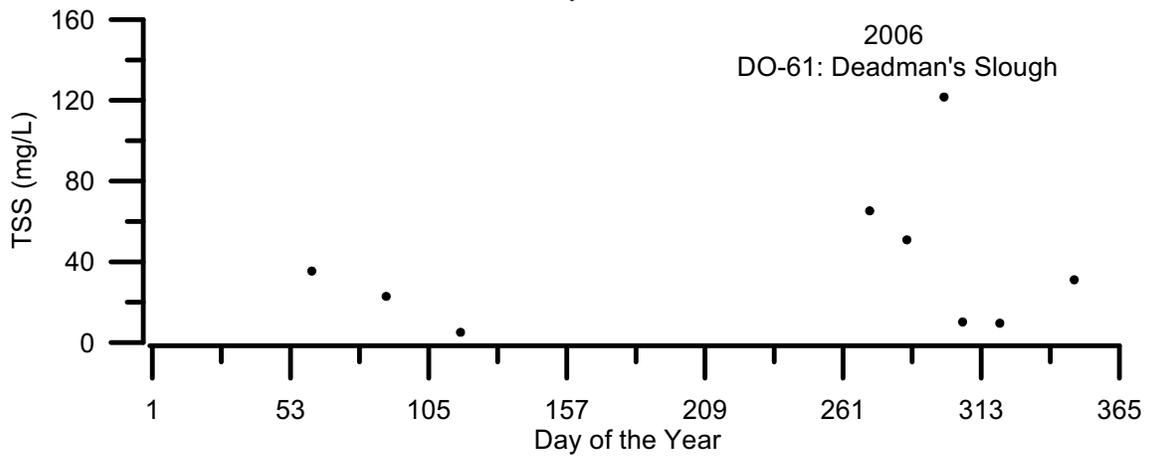
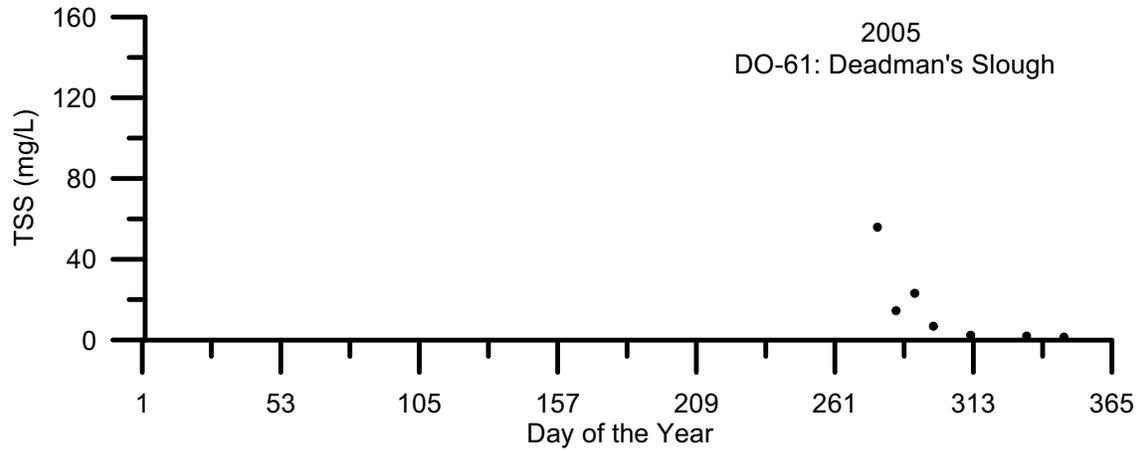


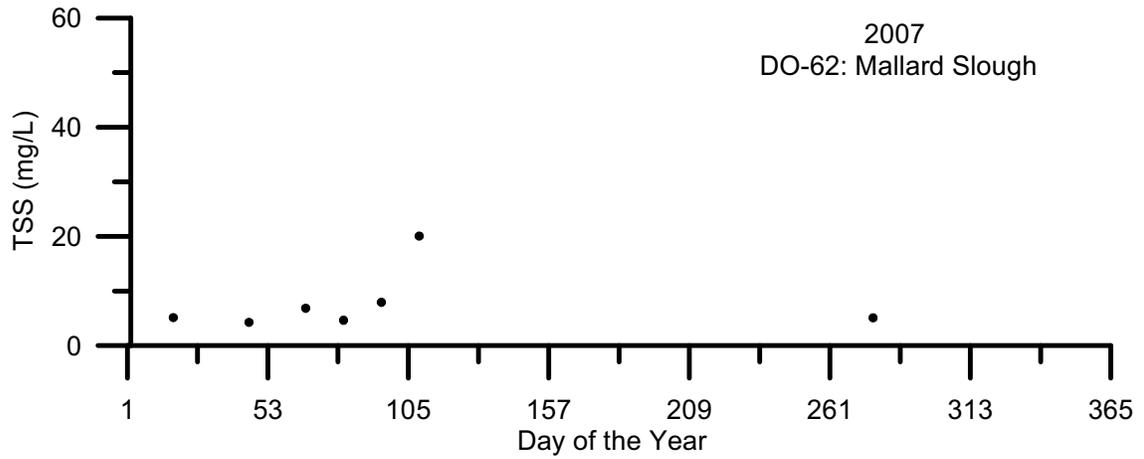
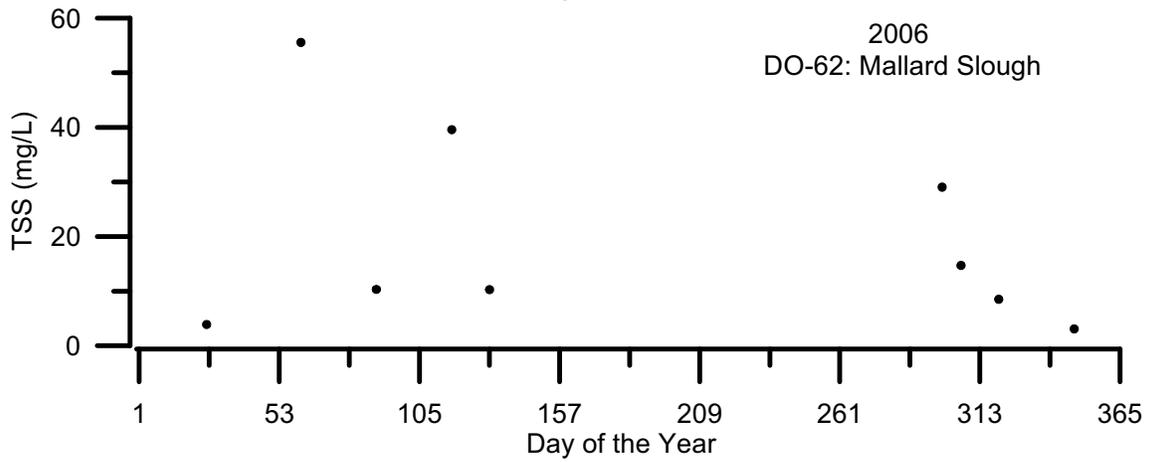
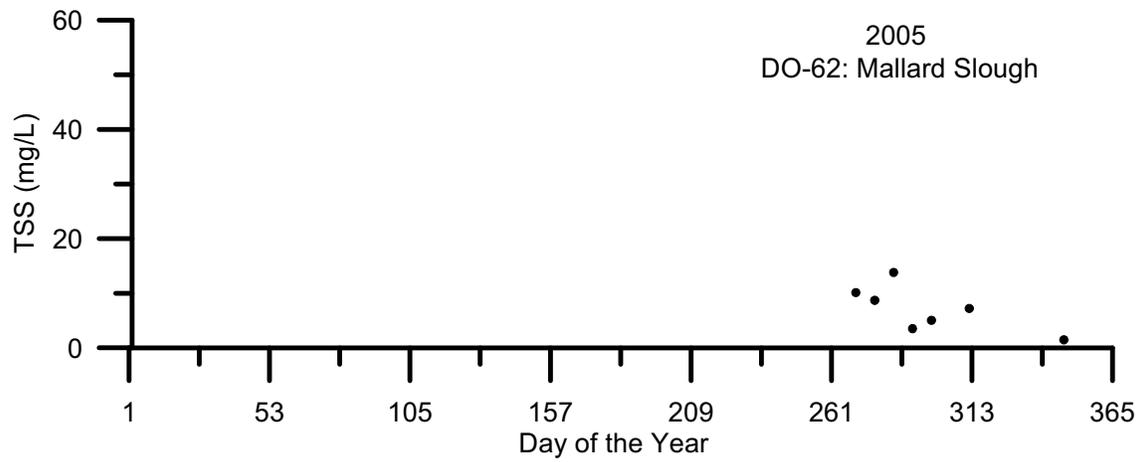


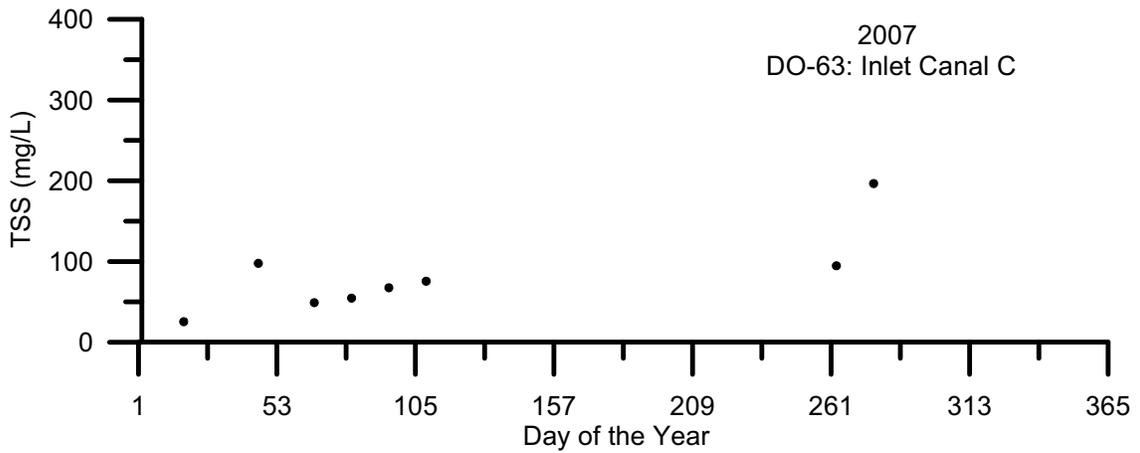
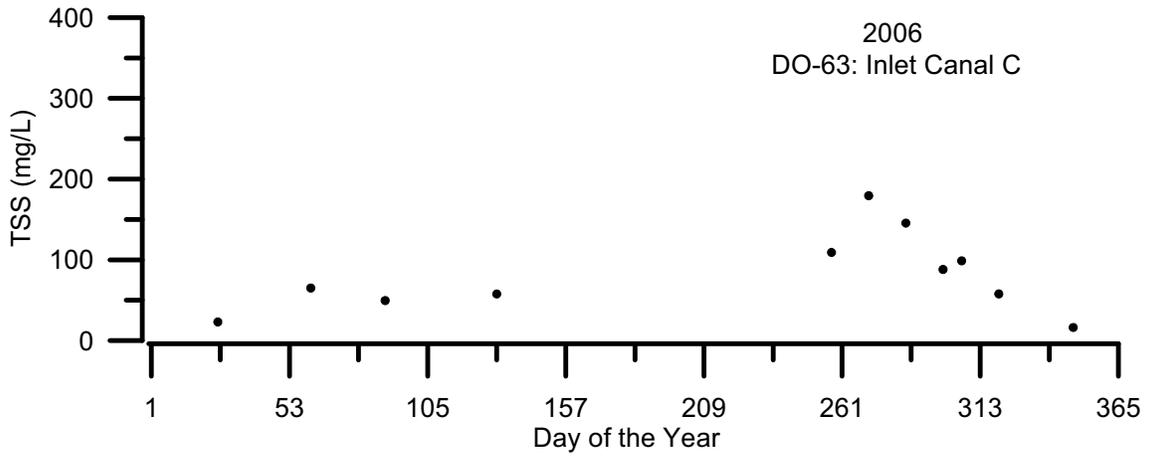
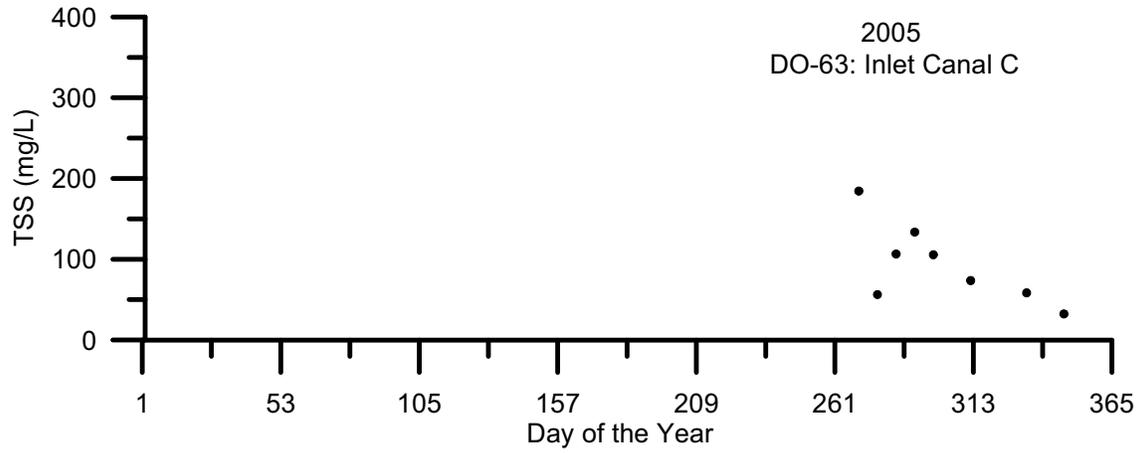


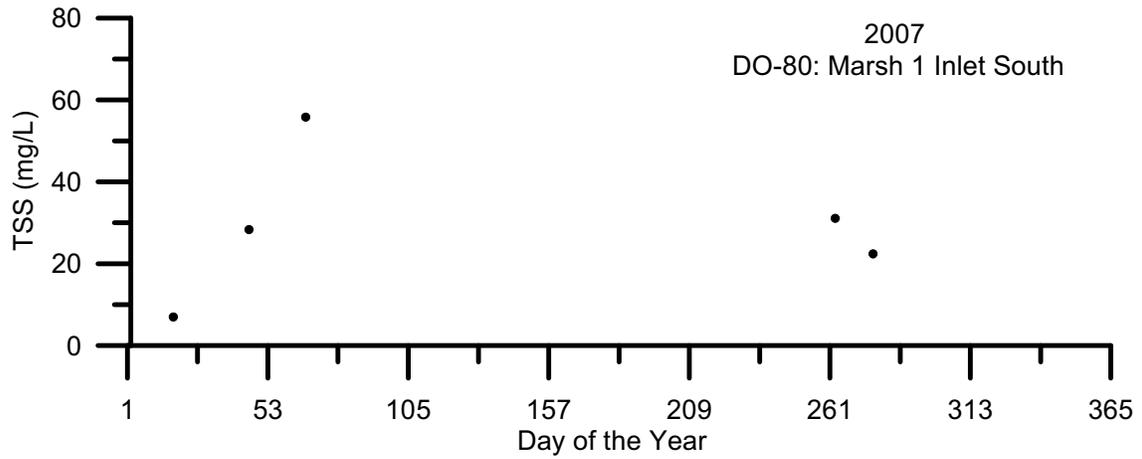
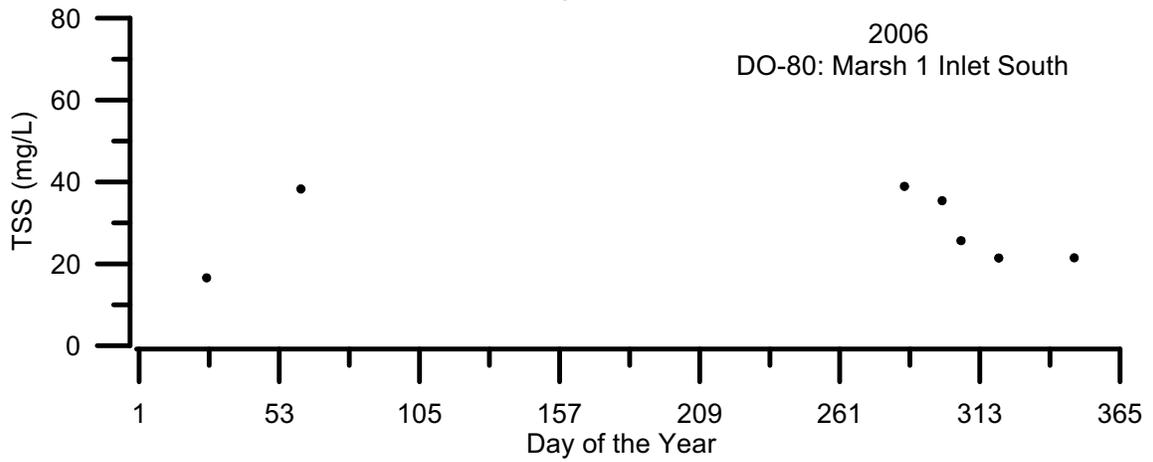
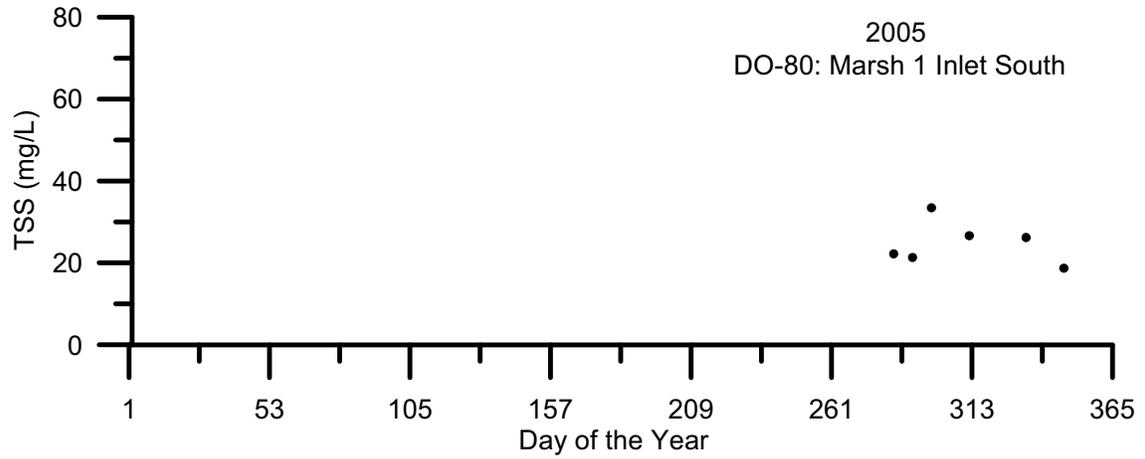


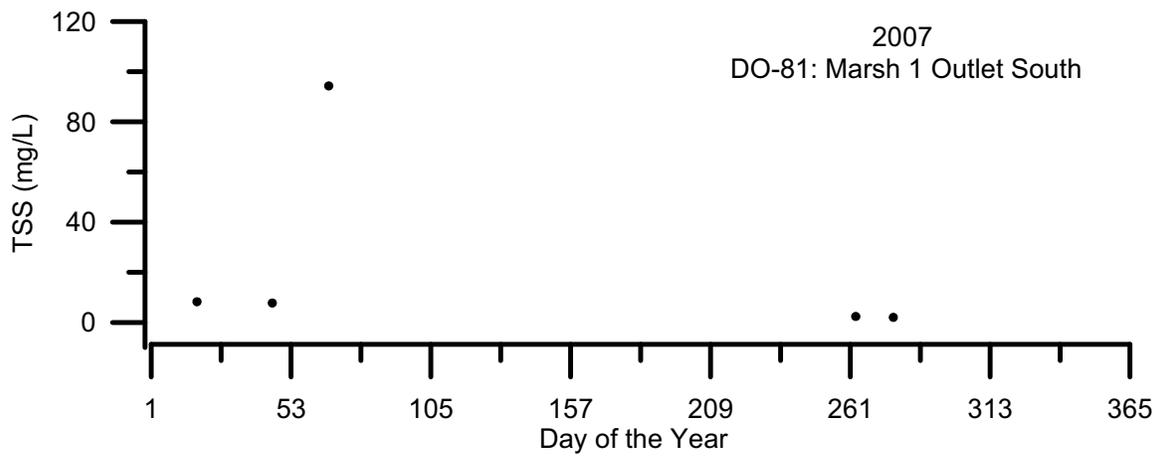
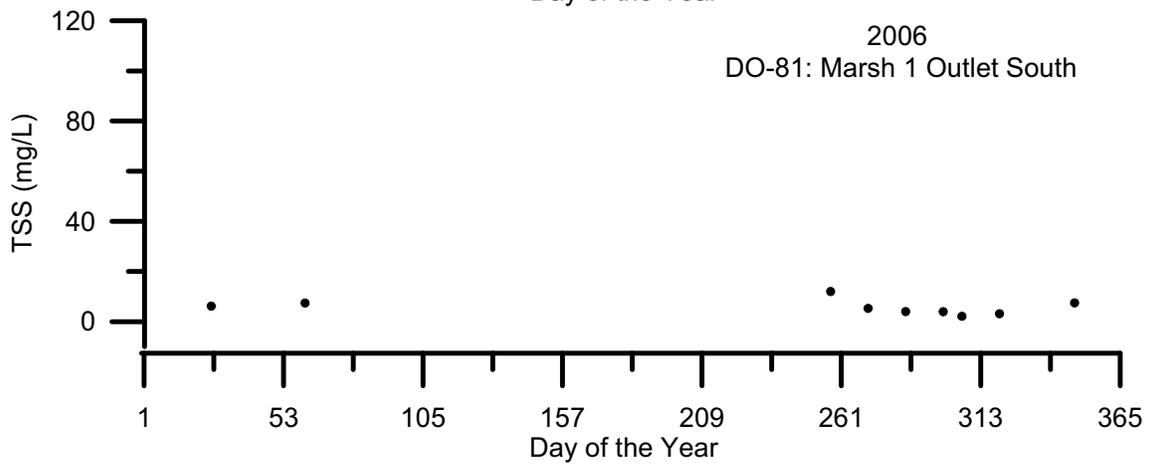
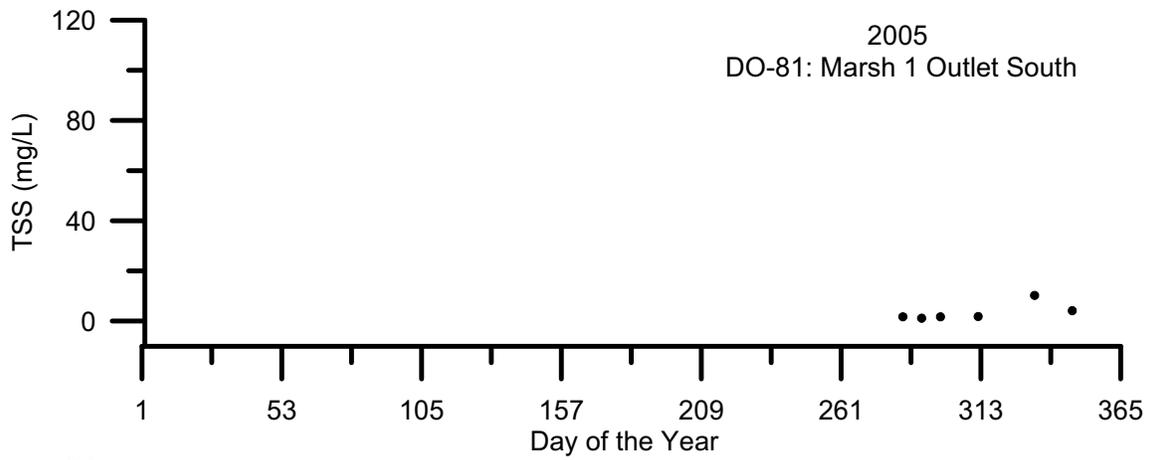


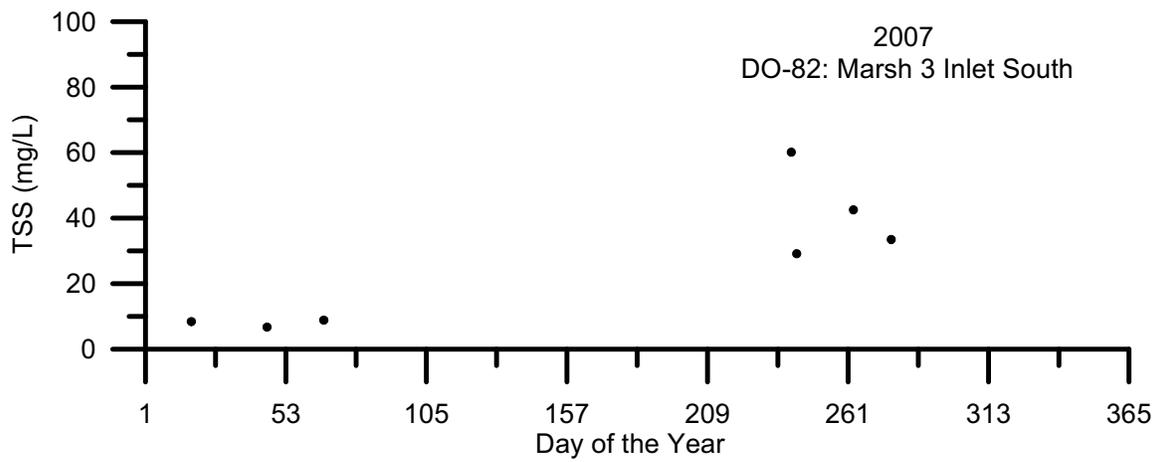
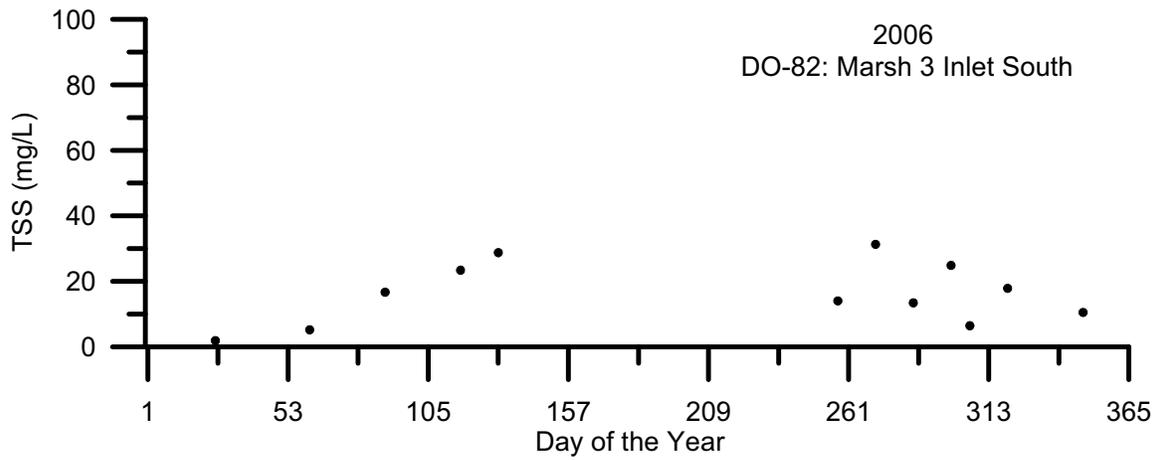
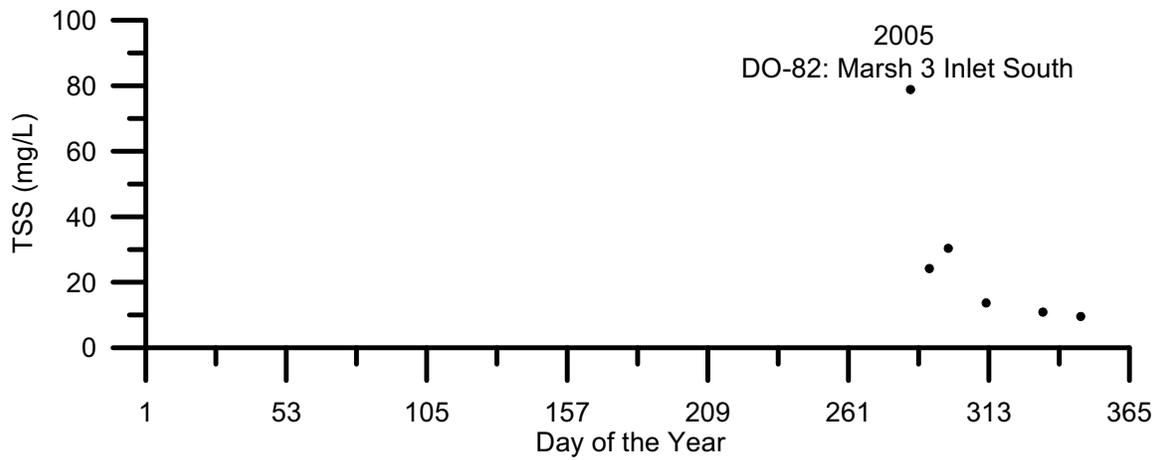


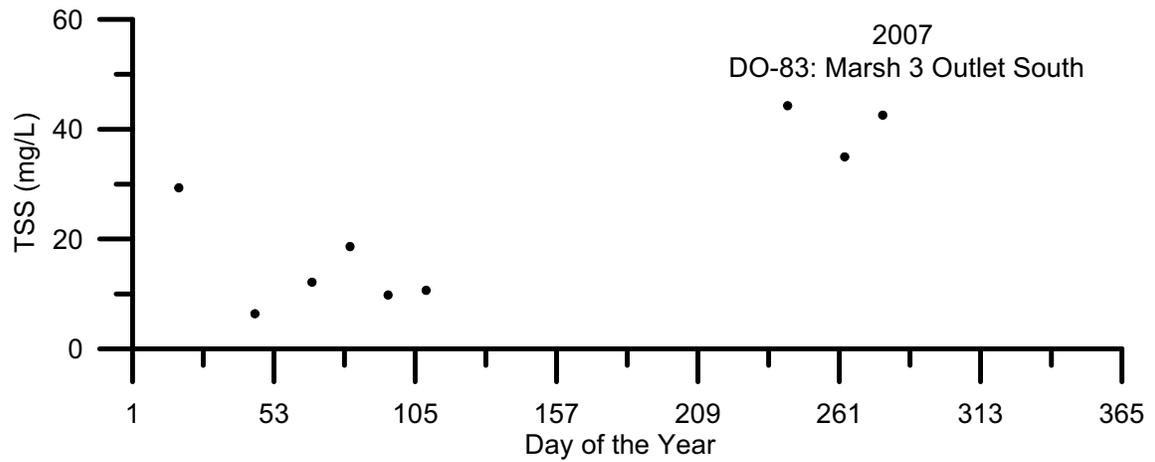
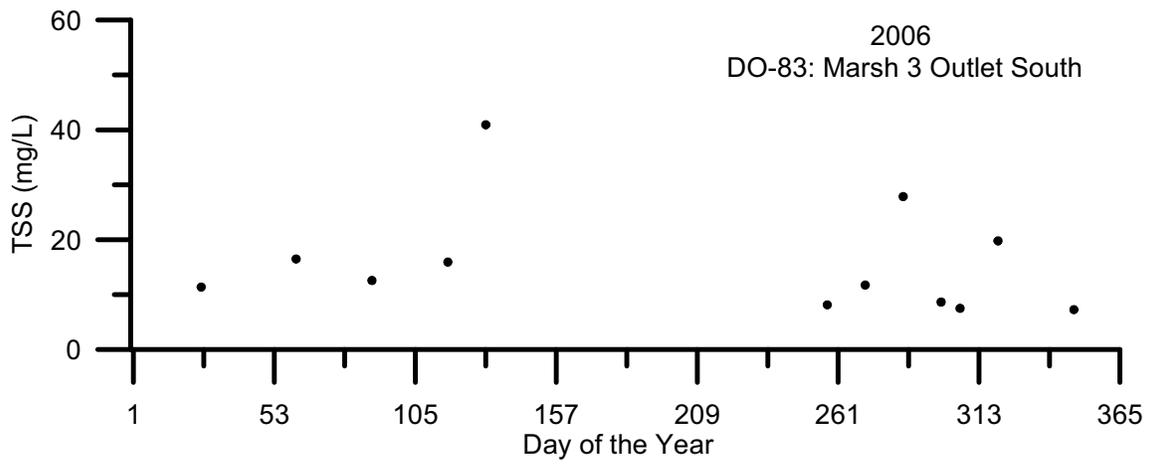
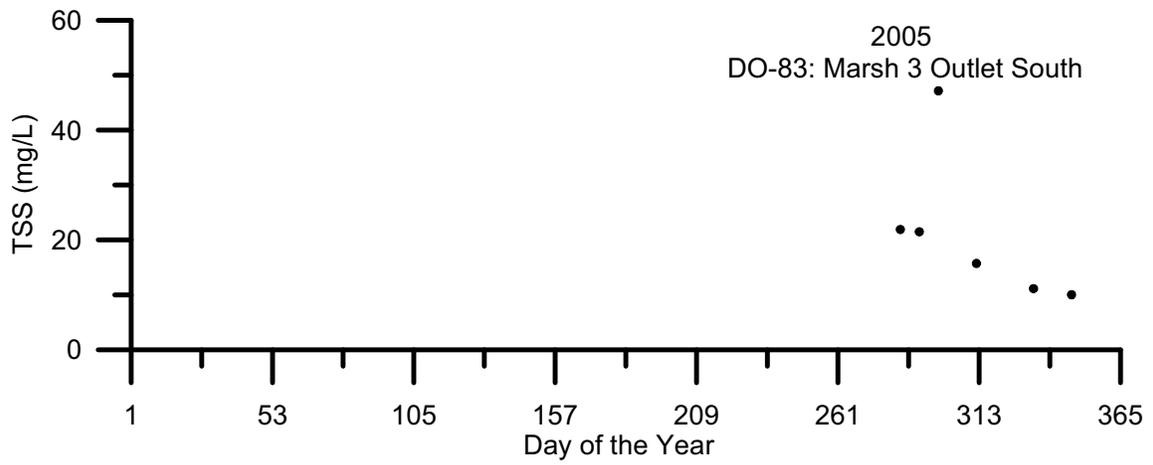


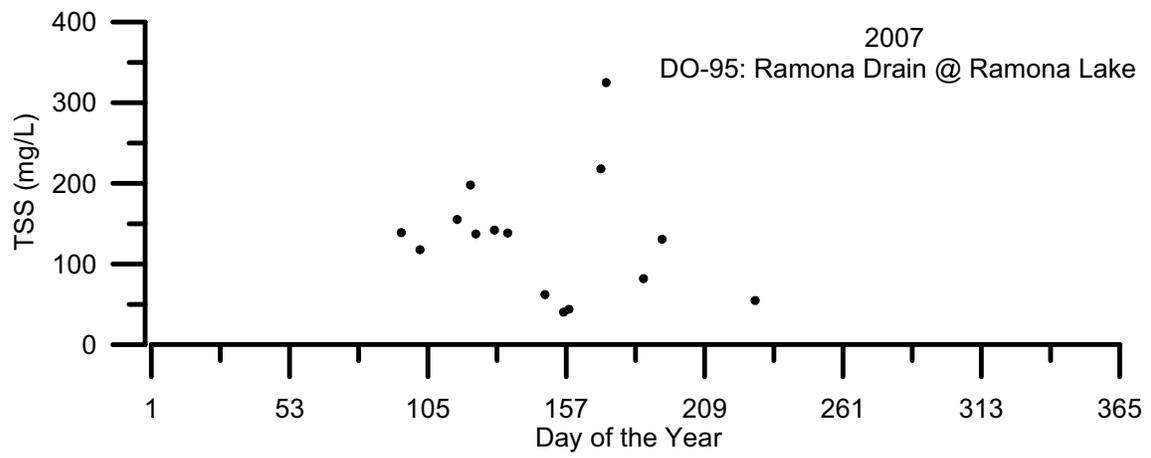
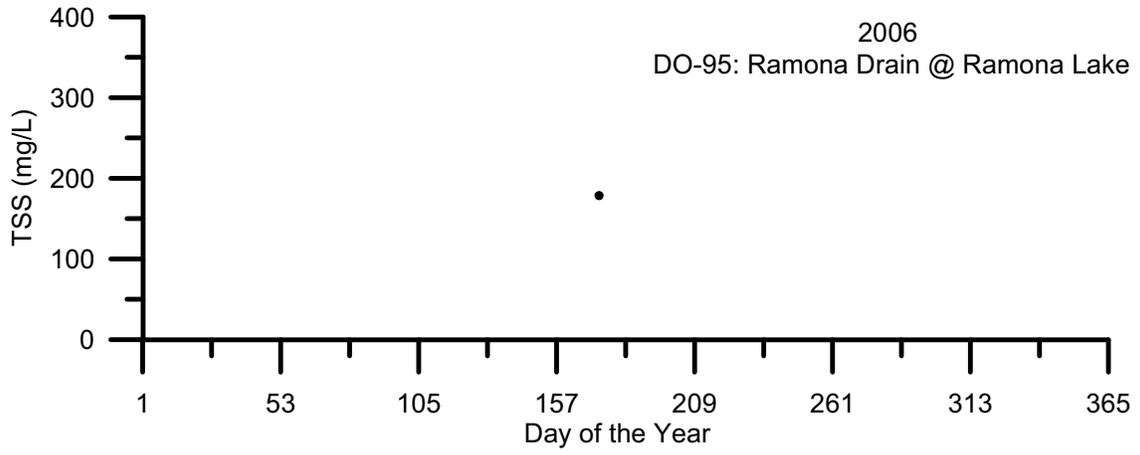














**Analysis of Total Protein Concentrations in the
San Joaquin River Watershed
2005-2007**

*Remie Burks
Chelsea Spier
Sharon Borglin
Jeremy Hanlon
Justin Graham
William Stringfellow*

February 2008

*Environmental Engineering Research Program
University of the Pacific
3601 Pacific Avenue, Sears Hall
Stockton CA 95211*

Introduction

The San Joaquin River (SJR) supports one of the most productive agricultural regions in the world and its productivity is heavily dependant on irrigated agriculture. A consequence of irrigated agriculture is the production of return flows conveyed down gradient drains that eventually discharge to surface waters. Agricultural drainage may have significant nutrient load and can impact algae growth and general water quality in the SJR. Individual farmers and agricultural organizations, such as drainage authorities, are in need of tools to manage the environmental impacts of agricultural activities (Stringfellow, 2008).

For the years 2005 through 2007, sites throughout the San Joaquin Valley watershed were sampled to assess the overall water quality in the region. One thousand nine hundred and ninety-six (1996) individual surface water samples were collected and analyzed and WQ was assessed at 113 sampling locations in the SJR basin (Borglin et al., 2008). Samples were processed and analyzed by the Environmental Engineering Research Program (EERP) laboratory at the University of the Pacific as well as at the University of California, Davis, Dahlgren Lab. This report presents temporal plots of total protein for all sites sampled in the SJR from 2005-2007.

Methods

Field sampling consisted of collecting water samples, measuring water quality with a YSI Sonde 6600 with MDS650 hand-held display, and recording of field conditions at sites within the study area per *EERP Field Protocol Book* (Graham, 2008).

Water samples were collected in glass 1000 mL bottles (Wheaton Science Products, Millville, NJ), 1000 mL HDPE Trace-Clean narrow mouth plastic bottles (VWR International), 250 mL HDPE Trace-Clean wide mouth plastic bottles (VWR International) as well as 40 mL trace clean vials with PTFE septa (ICChem, Rockwood, TN) in accordance with requirements for different lab analysis and volume requirements.

Samples were immediately stored at 4°C after sampling (cooler temperature was recorded in the lab upon delivery) and transported to the lab on the day of sampling.

Samples were received by the laboratory the same day they were sampled and stored at 4°C until filtering and analysis. Samples were collected, preserved, stored, and analyzed by methods outlined in *Standard Methods for the Analysis of Water and Wastewater*, (APHA, 2005, 1998). Total protein was quantified in all the samples using the Lowry method (Pierce Biosciences, Rockford, IL). The analysis was scaled up from the standard kit so the analysis was performed on 1 mL samples and analyzed in cuvettes with a 5 cm path length. Standard curves were made using bovine albumin from Pierce Biosciences (Rockford, IL). Samples were frozen within 24 hours of collection and defrosted prior to analysis. The limit of detection for this analysis is 0.5 mg/L Protein.

Results and Discussion

Total protein analysis was preformed intermittently over the three year period with the scale increase modification of the standard method.

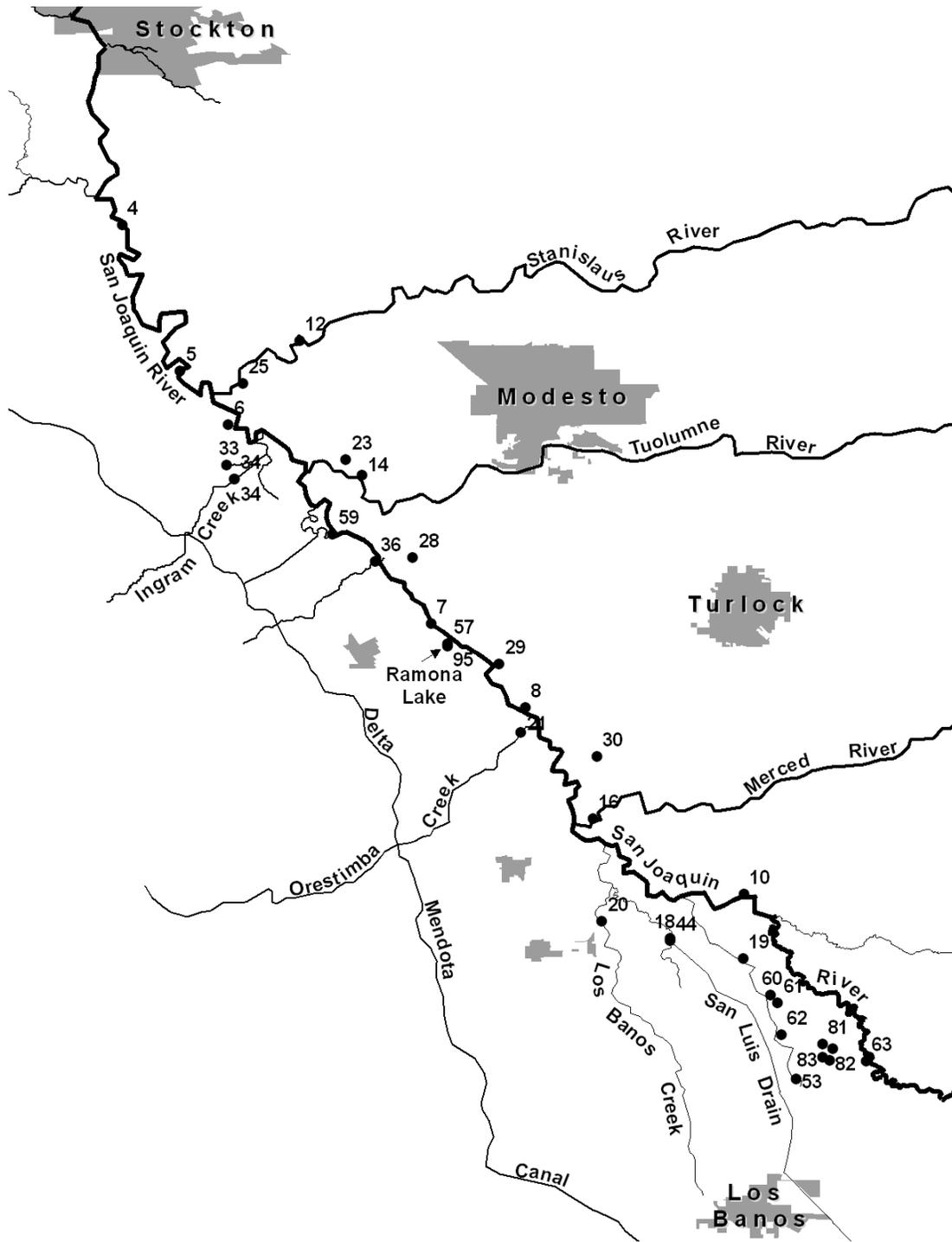
References

- Stringfellow, W.T., et al., (2008), *Evaluation of Vegetated Ditches, Ponds, and Wetlands as BMPs for Mitigating the Water Quality Impact of Irrigated Agriculture in the San Joaquin Valley*, University of the Pacific, Stockton, CA
- Graham, J., Hanlon, J.S., Stringfellow, W.T., (2008), *EERP Field Protocol Book*, University of the Pacific, Stockton, CA.
- Borglin, S., W. Stringfellow, J. Hanlon. (2005), *Standard Operating Procedures for the Up-Stream Dissolved Oxygen TMDL Project*, LBNL/Pub-937.
- Borglin, S., Burks, R., Hanlon, J., Graham, J., Spier, C., Stringfellow, W., and Dahlgren, R., (2008) *Methods overview, quality assurance, and quality control*, University of the Pacific, Stockton, CA
- American Public Health Association, (2005), *Standard Methods for the Examination of Water and Wastewater*, American Public Health Association, Washington, D.C.
- Borglin, S.E., Burks, R.D., Hanlon, J.S., Stringfellow, W.T. (2008) *EERP Lab Protocol Book*, University of the Pacific, Stockton, CA.
- YSI Environmental Operations Manual, (2005), 6-Series Environmental Monitoring Systems, Yellow Springs, OH.

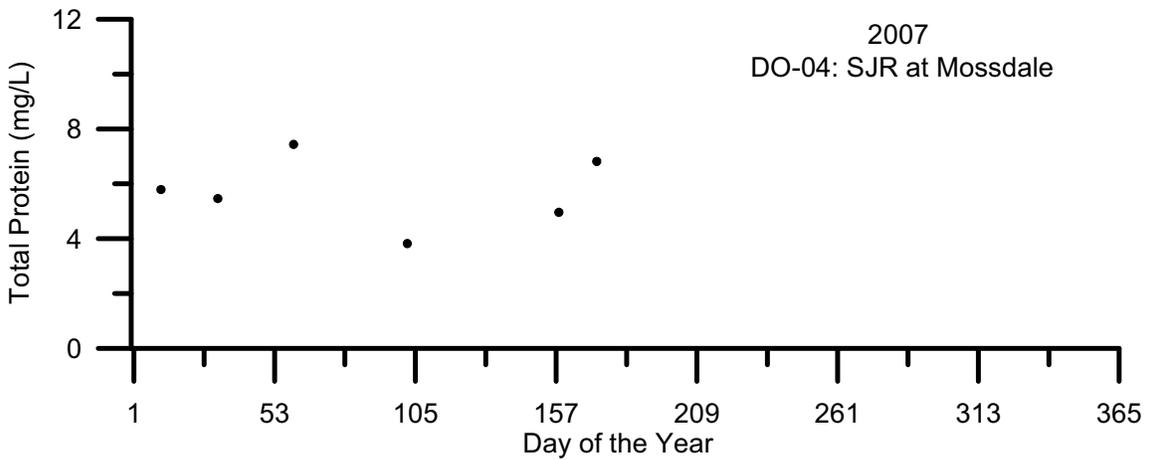
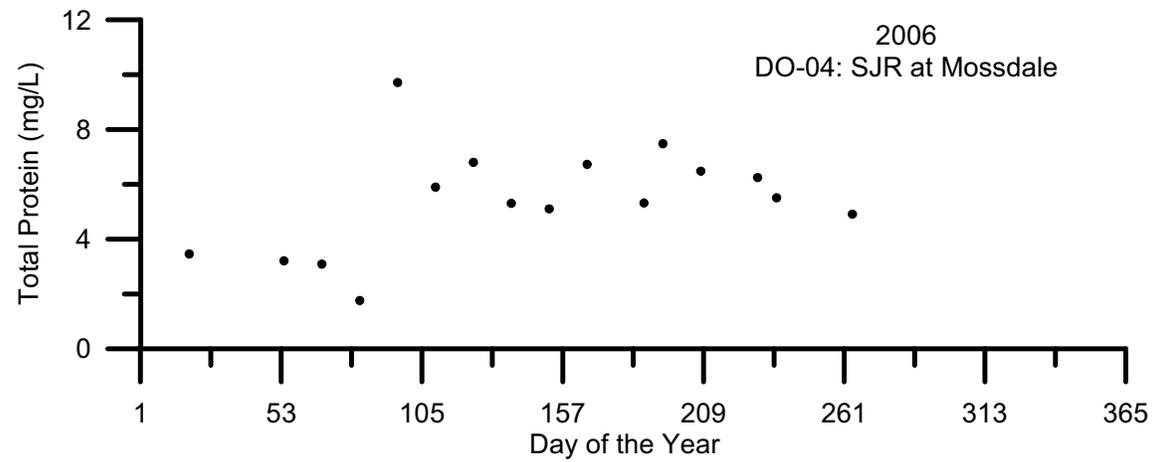
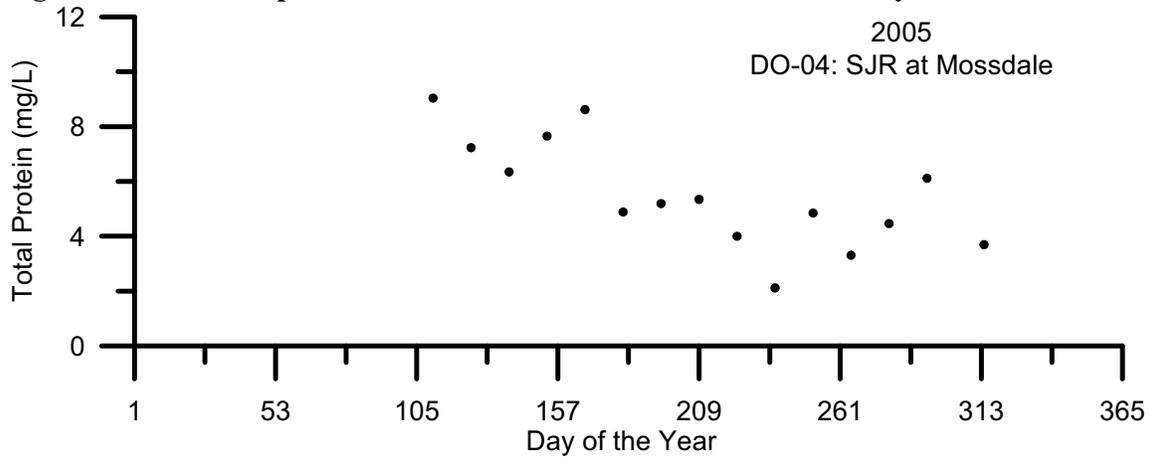
Table 1: EERP Sampling Site List

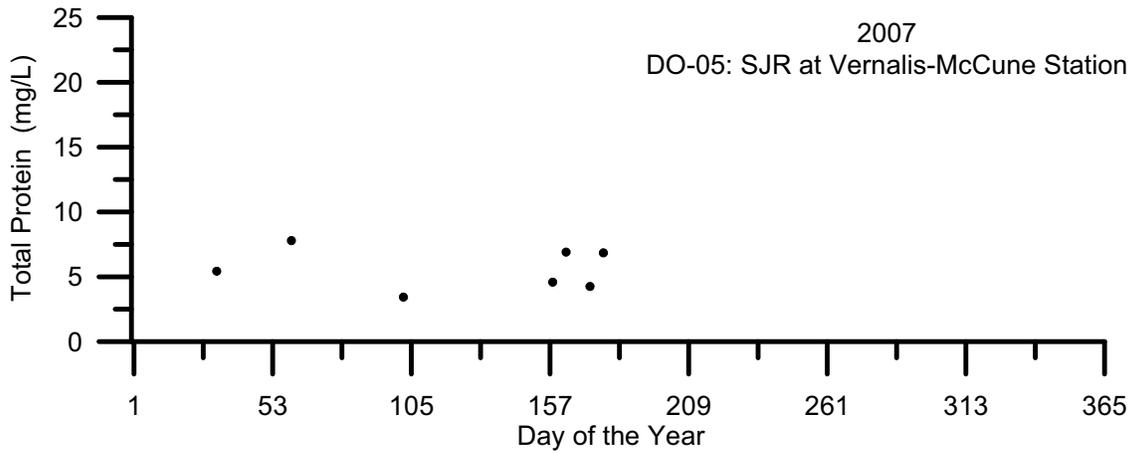
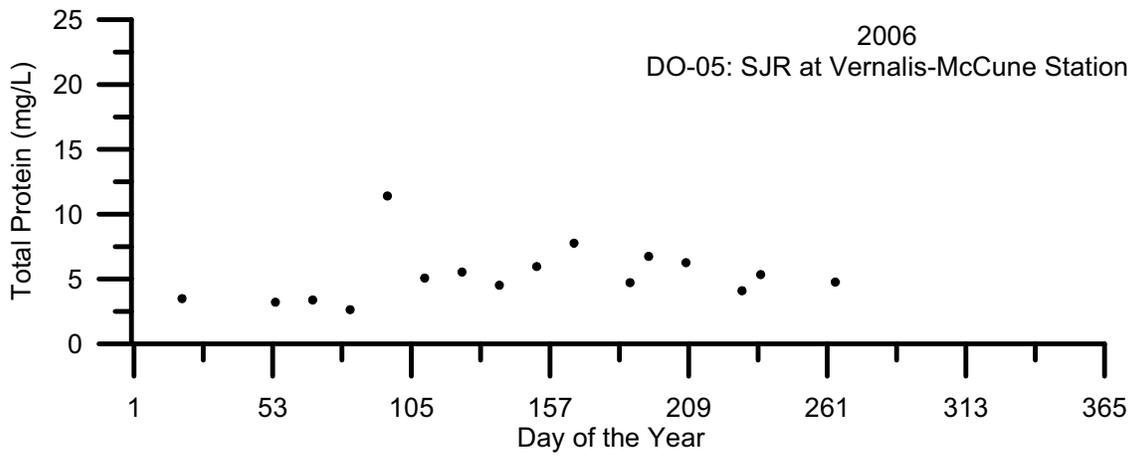
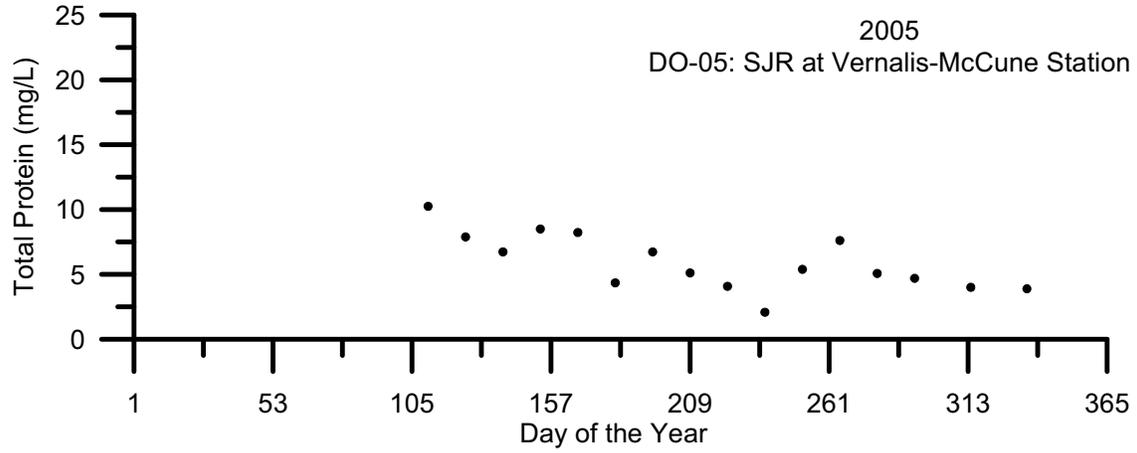
DO Number	Site Name	Type
4	SJR at Mossdale	Core sites
5	SJR at Vernalis-McCune Station (River Club)	Core sites, BMP
6	SJR at Maze	Core sites, BMP
7	SJR at Patterson	Core sites, BMP
8	SJR at Crows Landing	Core sites, BMP
10	SJR at Lander Avenue	Core sites
12	Stanislaus River at Caswell Park	Core sites
14	Tuolumne River at Shiloh Bridge	Core sites
16	Merced River at River Road	Core sites
18	Mud Slough near Gustine	Core sites, Wetland
19	Salt Slough at Lander Avenue	Core sites, Wetland
20	Los Banos Creek Flow Station	Core sites, Wetland
21	Orestimba Creek at River Road	Core sites, BMP
23	Modesto ID Lateral 5 to Tuolumne	Core sites
25	Modesto ID Main Drain to Stan. R. via Miller Lake	Core sites
28	Turlock ID Westport Drain Flow station	Core sites
29	Turlock ID Harding Drain	Core sites
30	Turlock ID Lateral 6 & 7 at Levee	Core sites
33	Hospital Creek	Intermittent, BMP
34	Ingram Creek	Core sites, BMP
36	Del Puerto Creek Flow Station	Core sites, BMP
44	San Luis Drain End	Core sites
53	Salt Slough at Wolfsen Road	Wetland
57	Ramona Lake Drain	Core sites, BMP
59	SJR Laird Park	Core sites
60	Moffit 1 South	Wetland
61	Deadman's Slough	Wetland
62	Mallard Slough	Wetland
63	Inlet C Canal	Wetland
80	South Marsh-1-Inlet	Wetland
81	South Marsh-1-Outlet	Wetland
82	South Marsh-3-Inlet	Wetland
83	South Marsh-3-Outlet	Wetland
95	Ramona drain at Ramona Lake	BMP, Intermittent

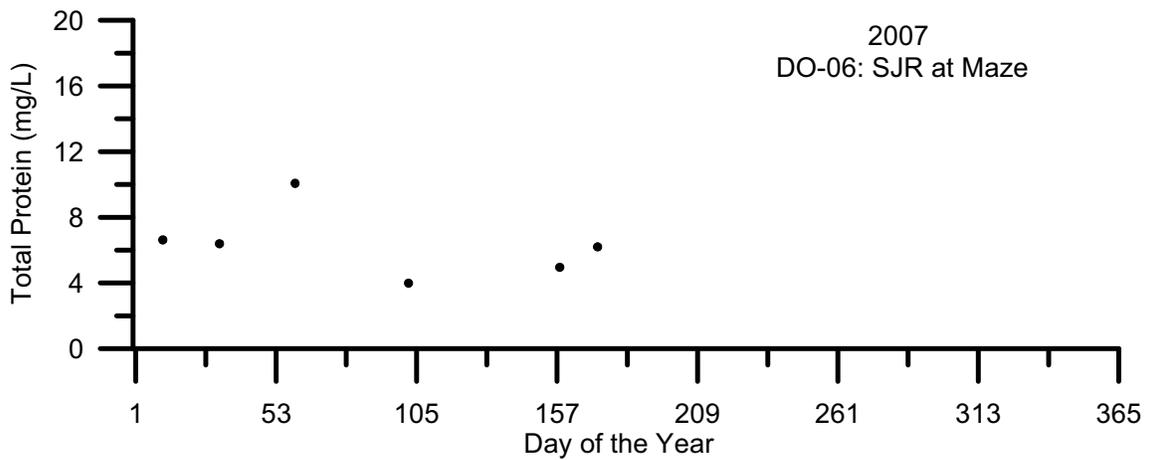
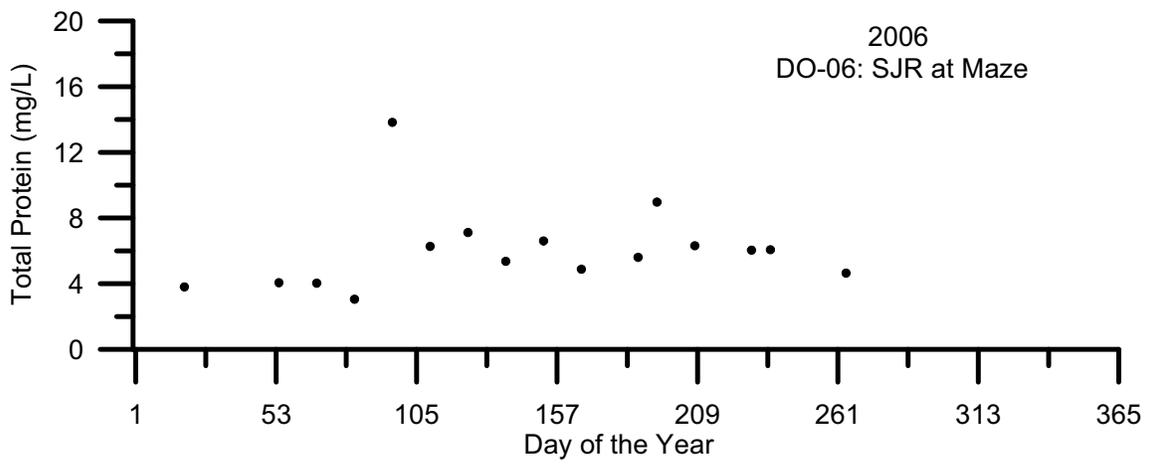
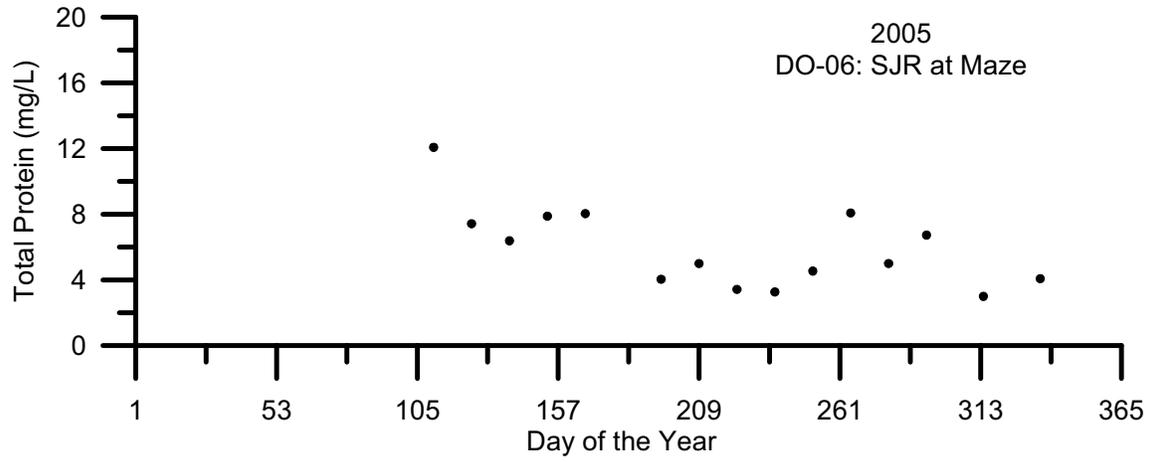
Figure 1: EERP Sampling Site Map of SJR Watershed and Tributaries

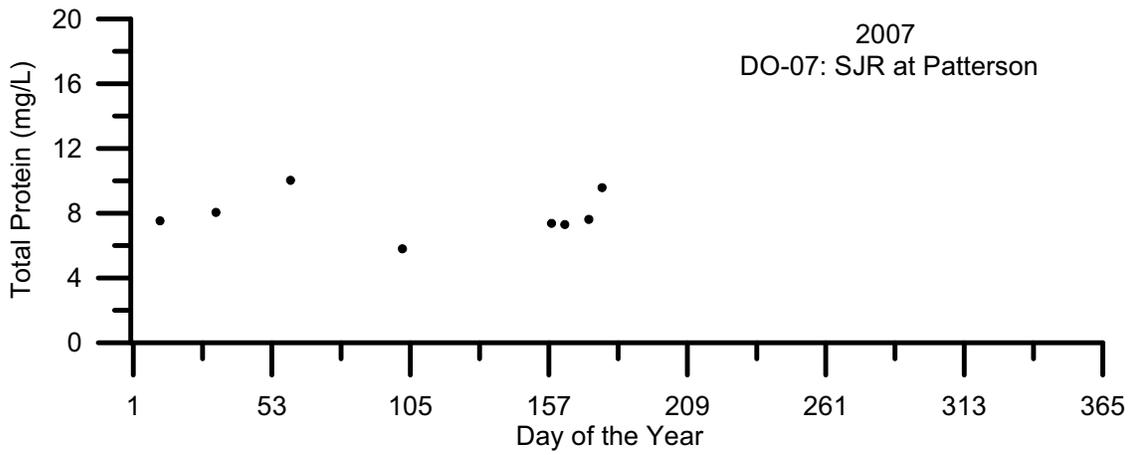
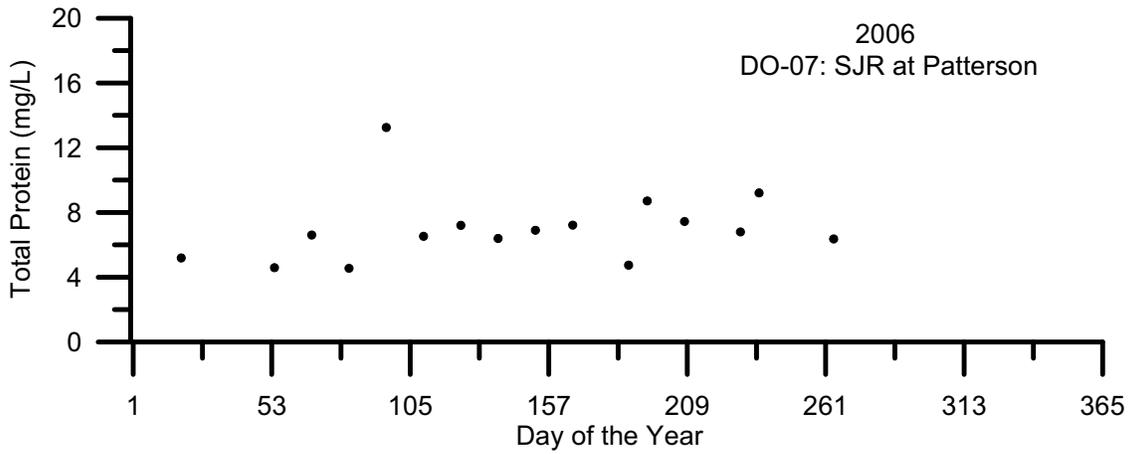
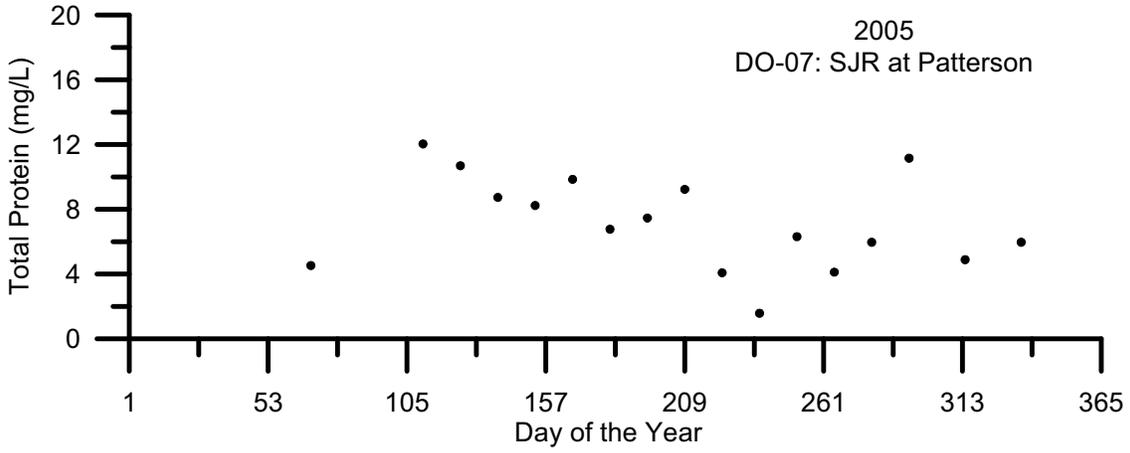


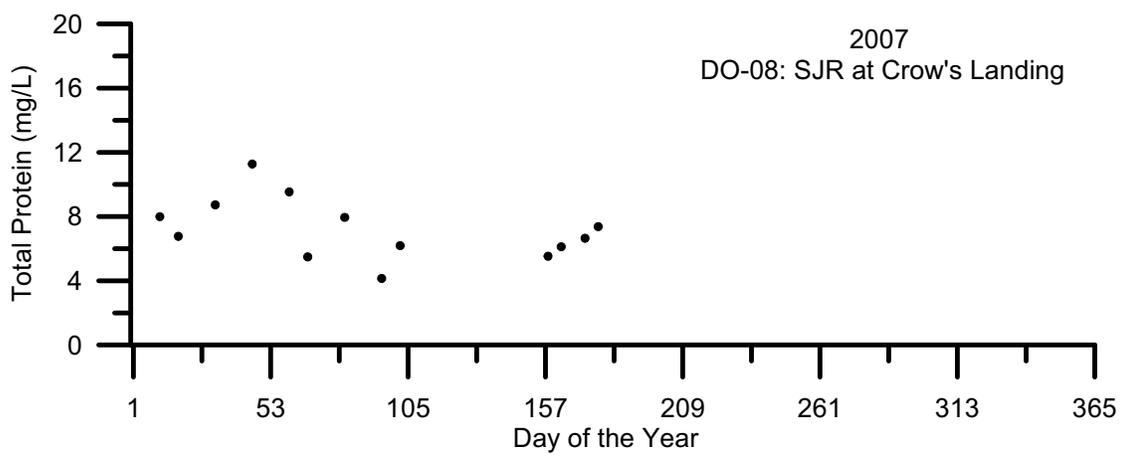
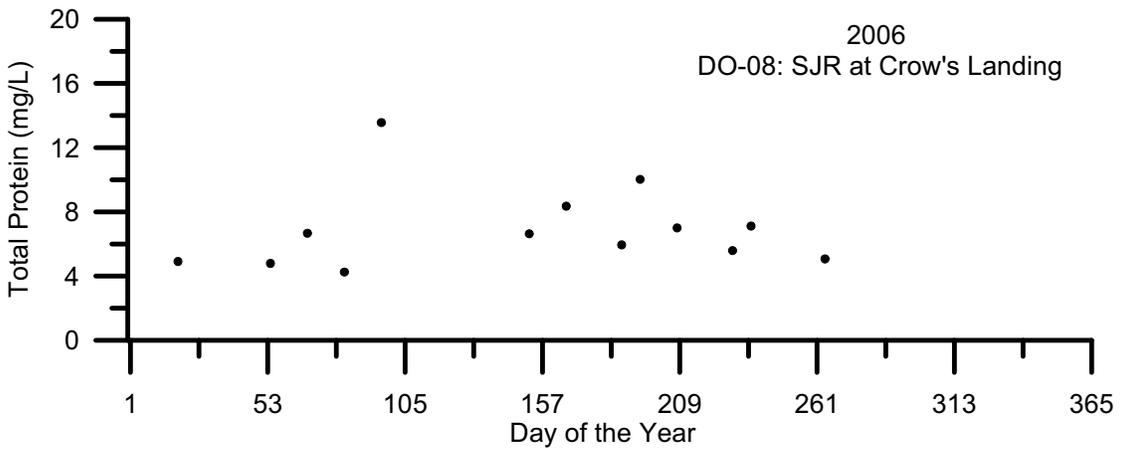
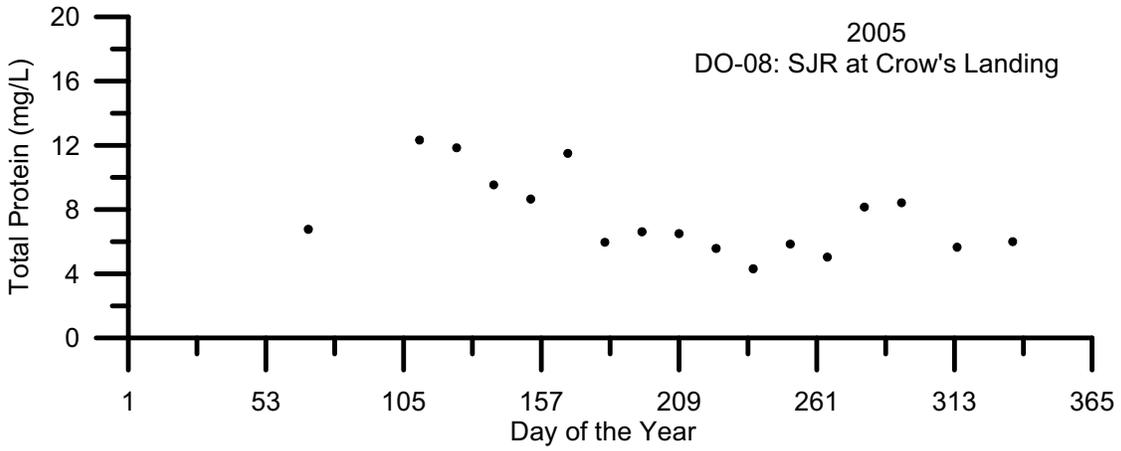
Figures 2 -101: Temporal Plots of Total Protein Concentrations By Site ID

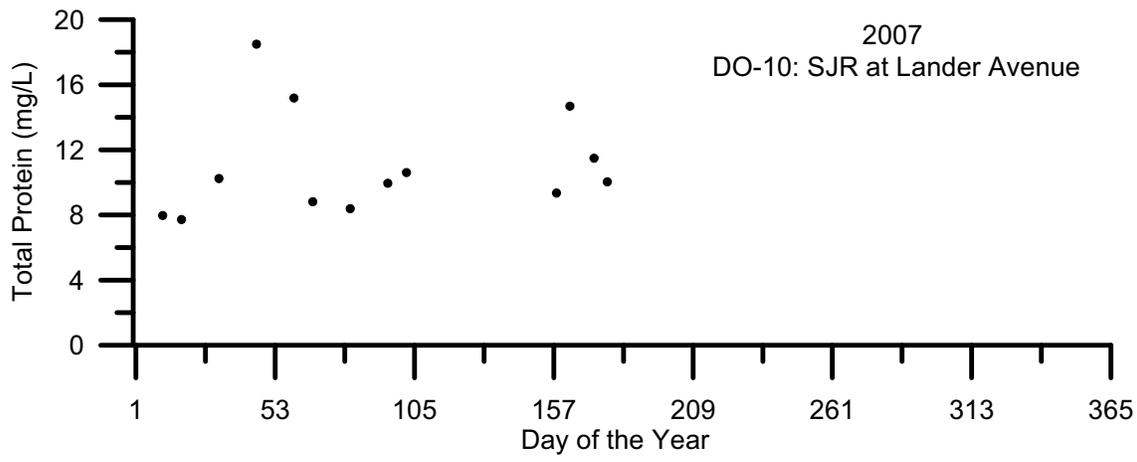
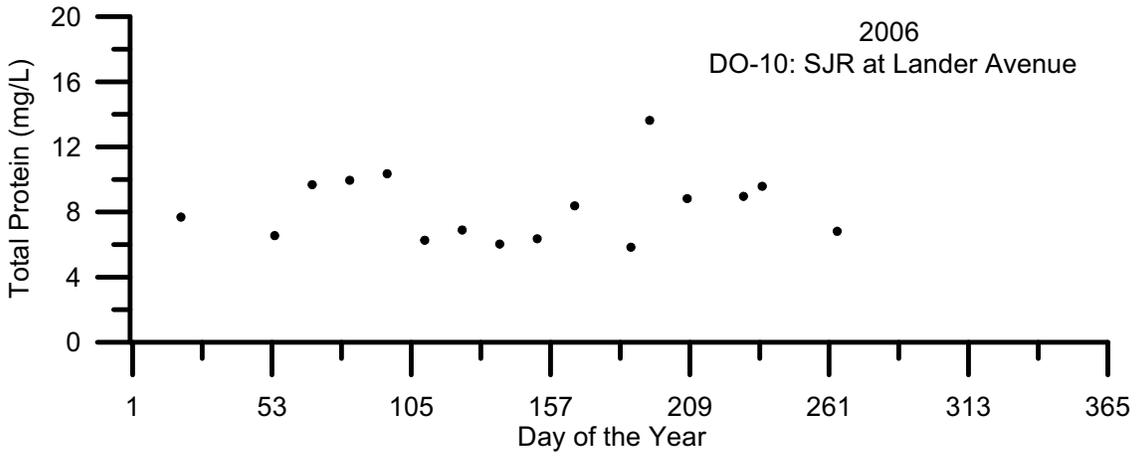
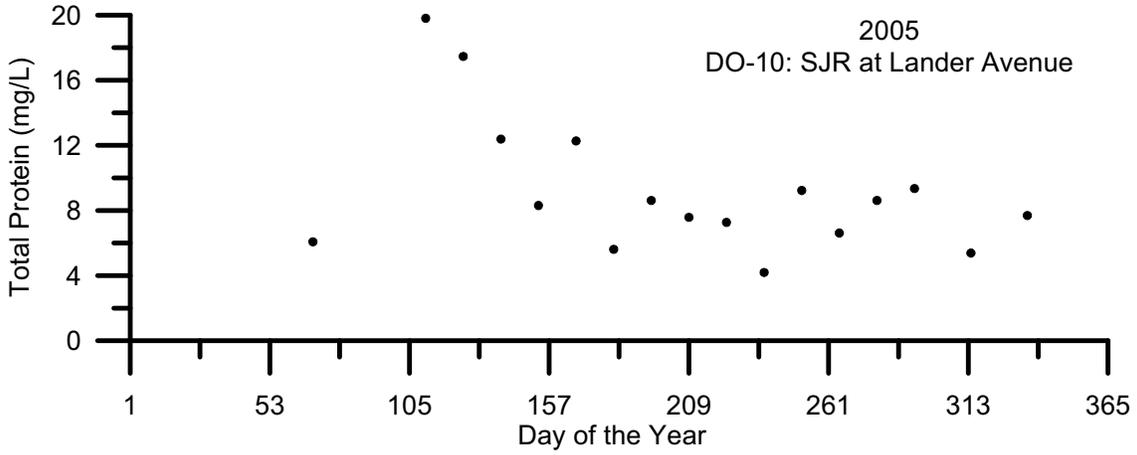


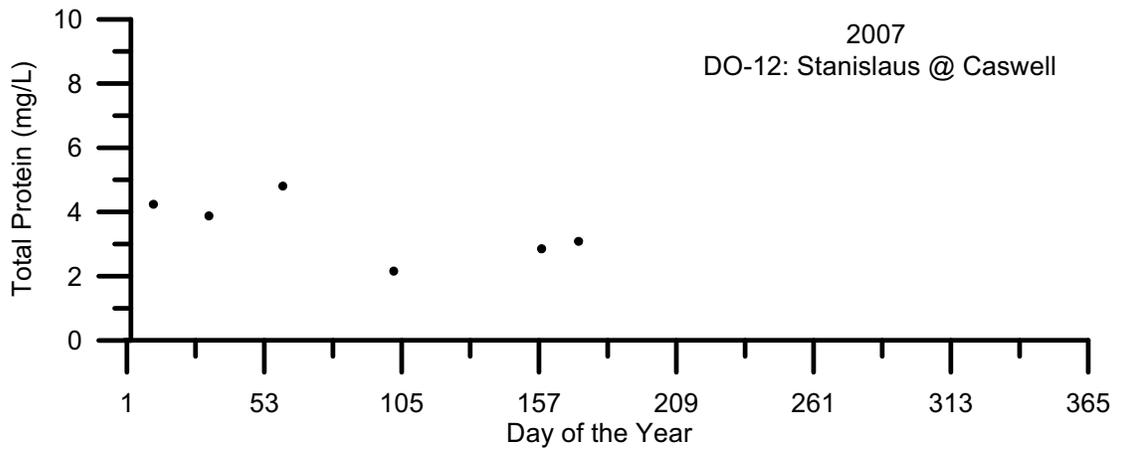
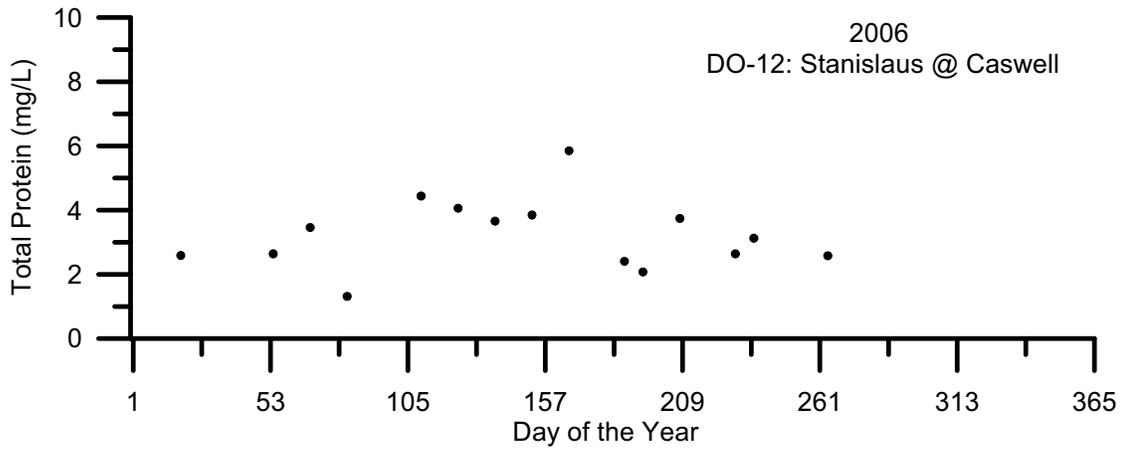
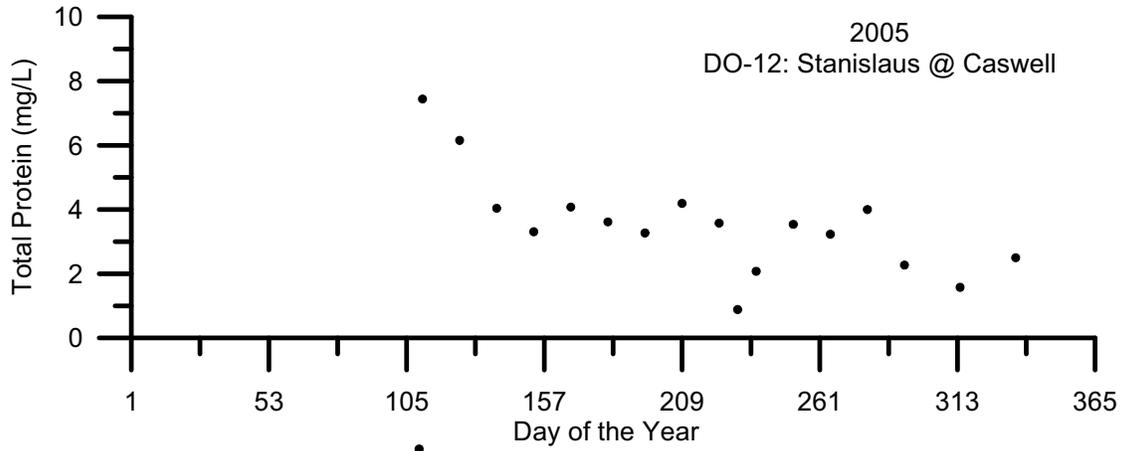


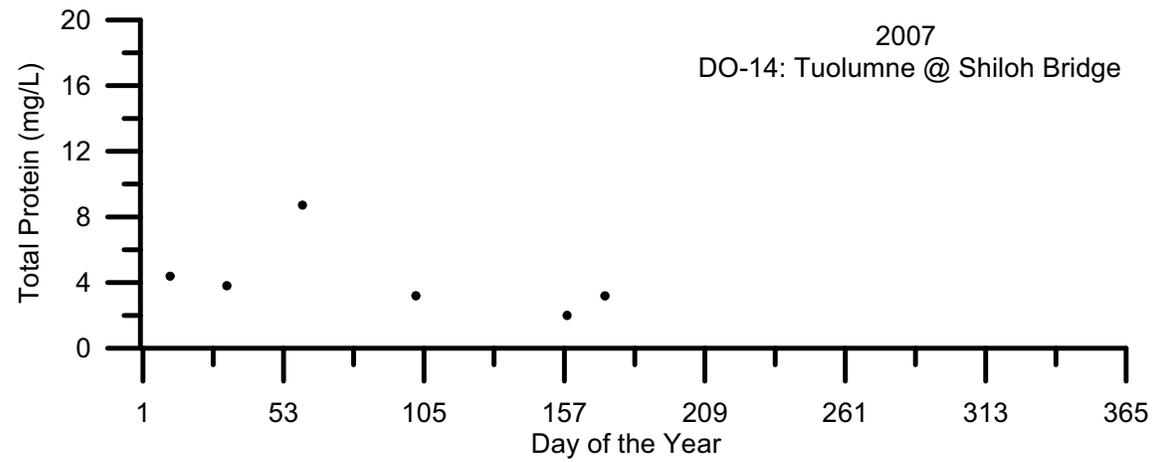
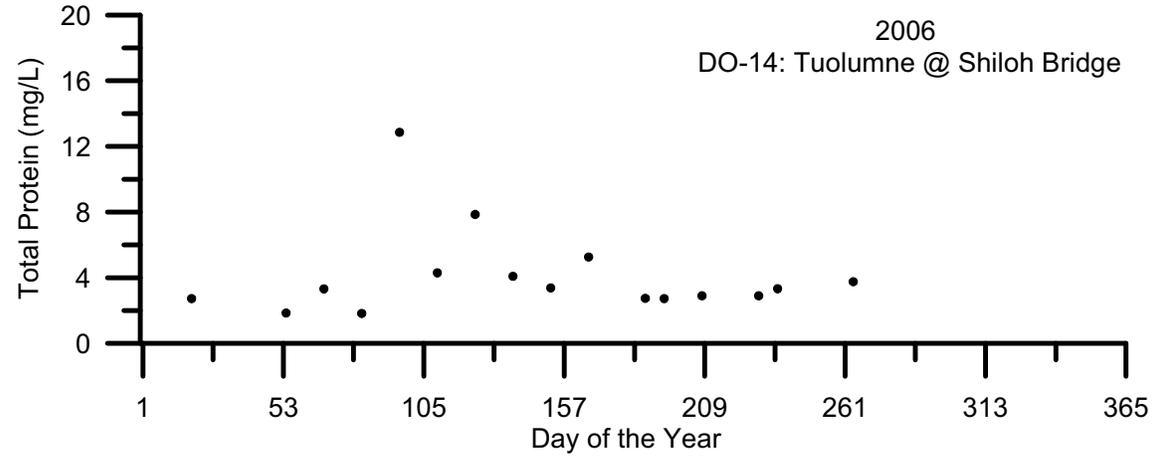
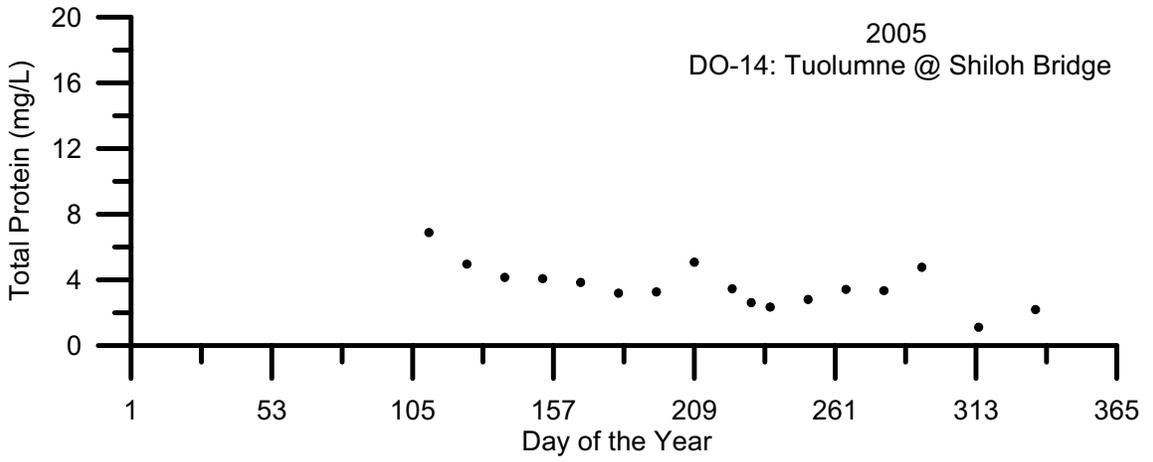


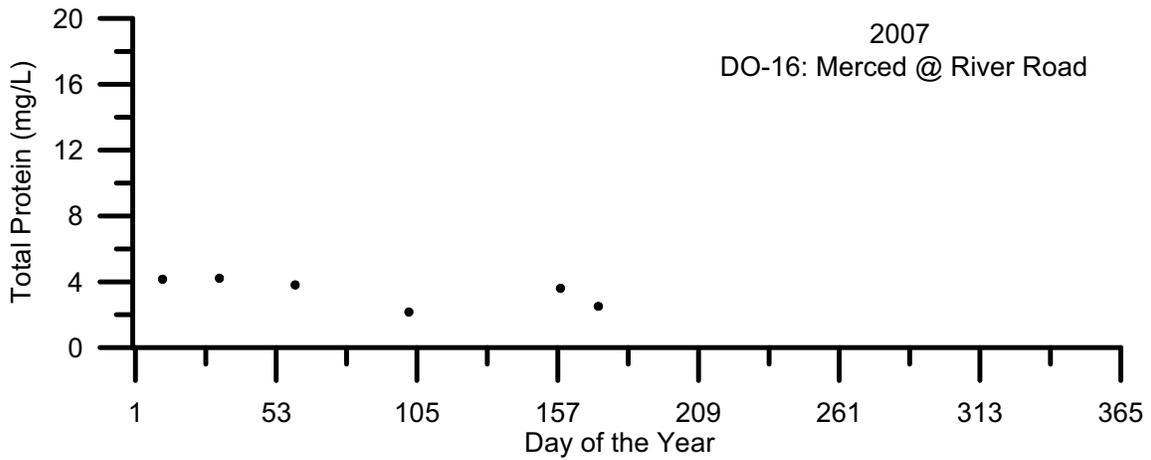
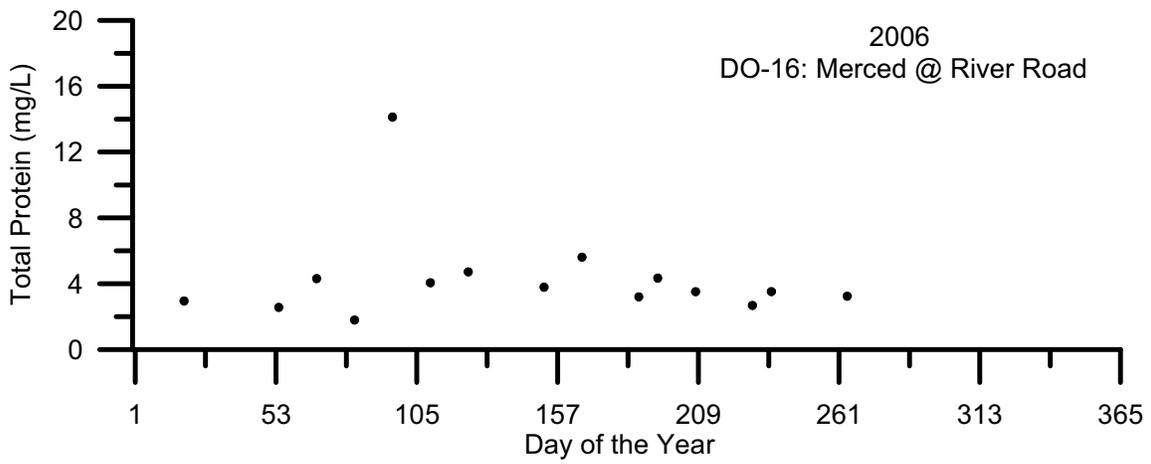
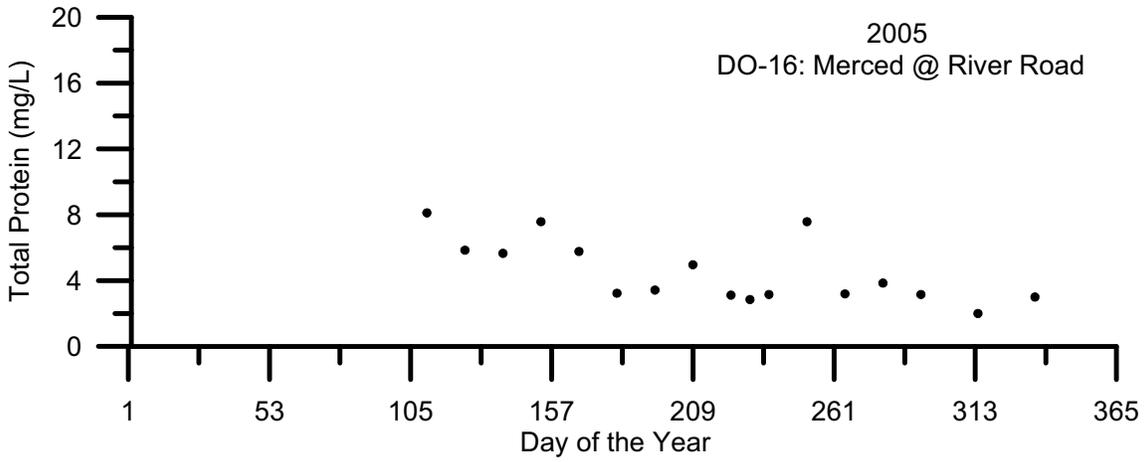


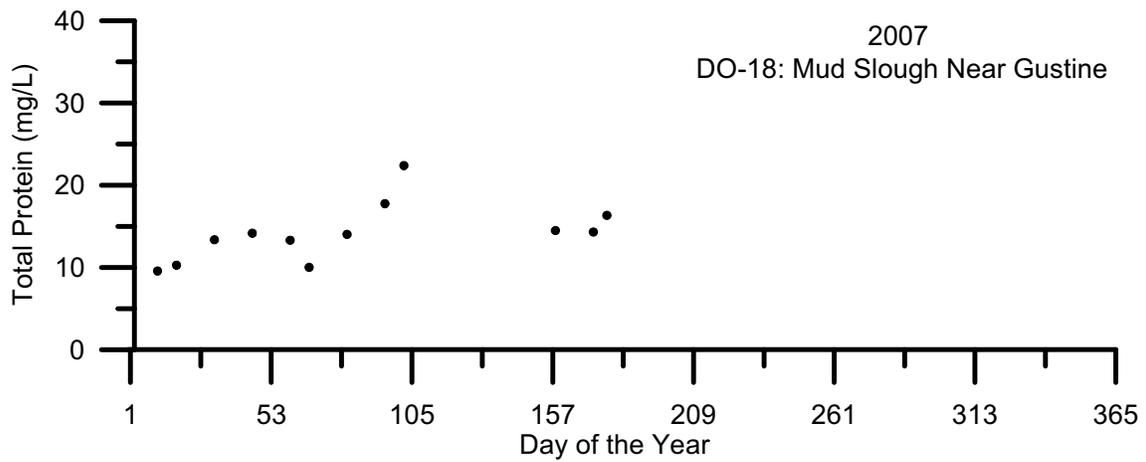
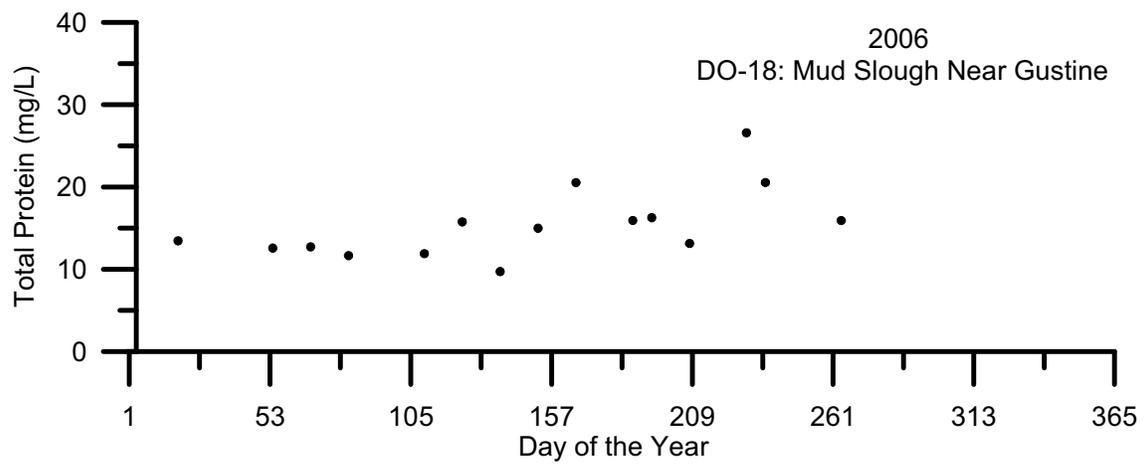
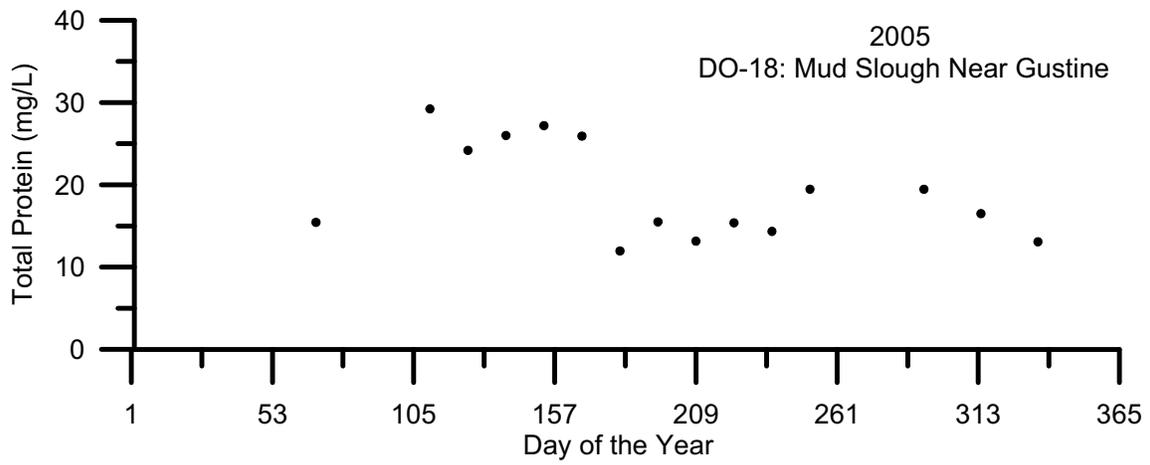


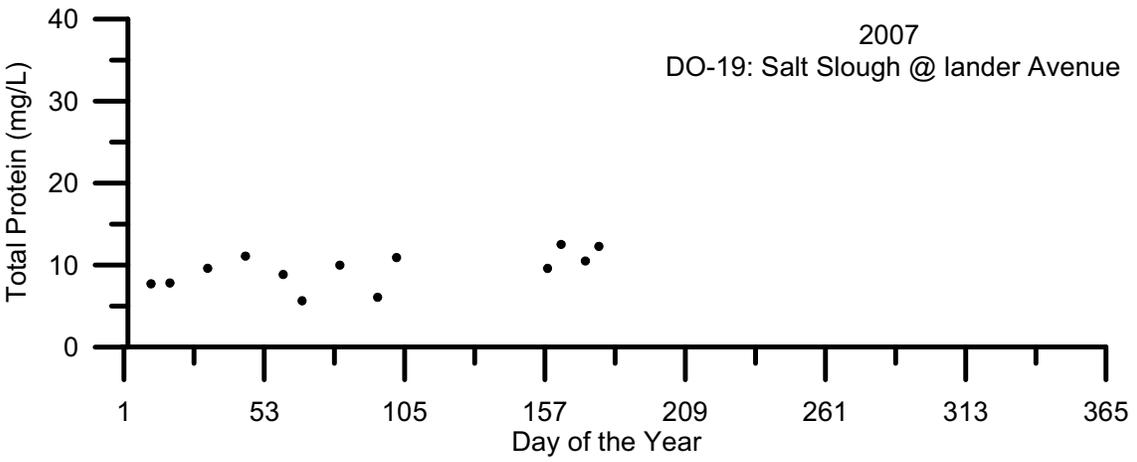
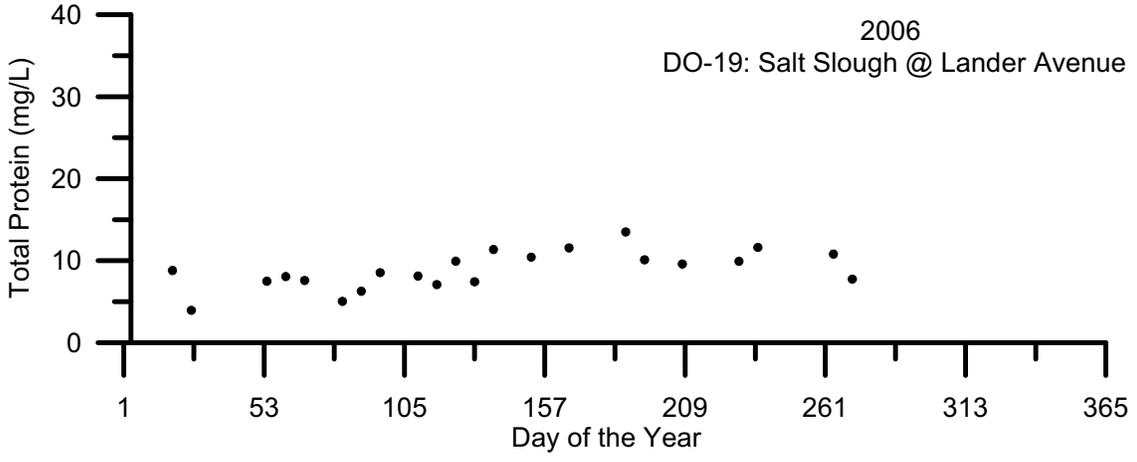
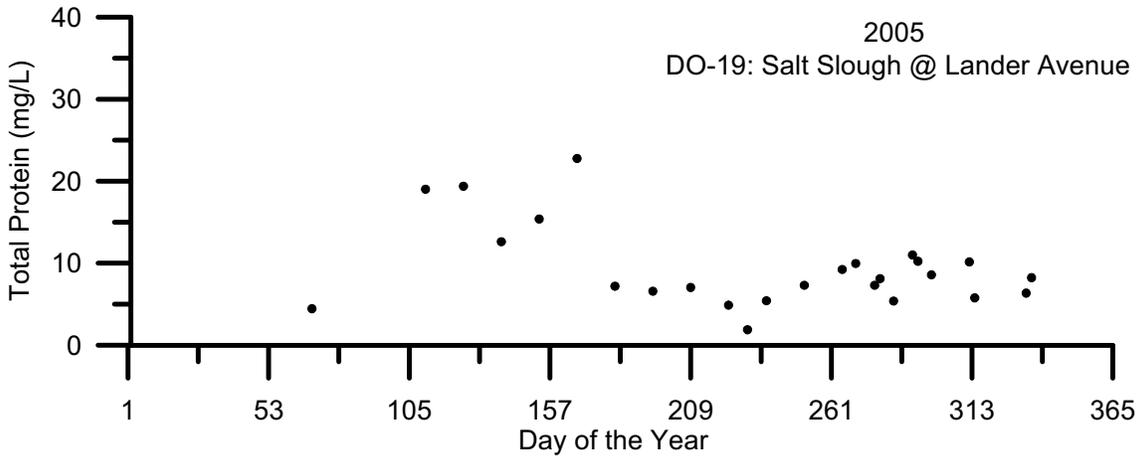


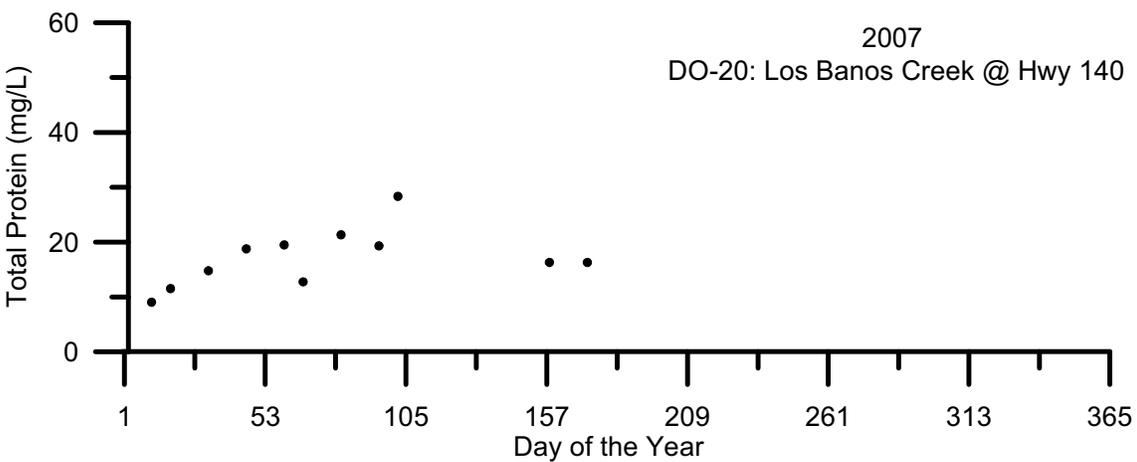
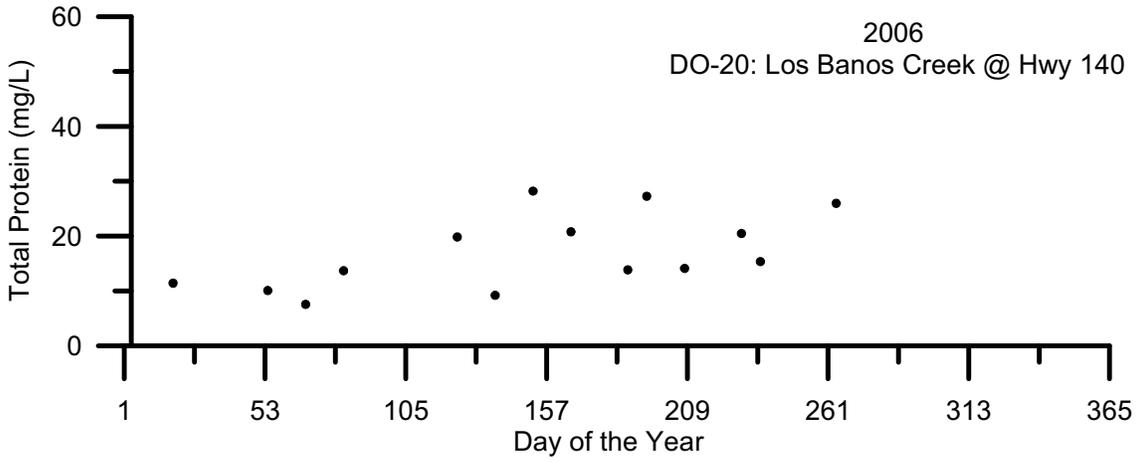
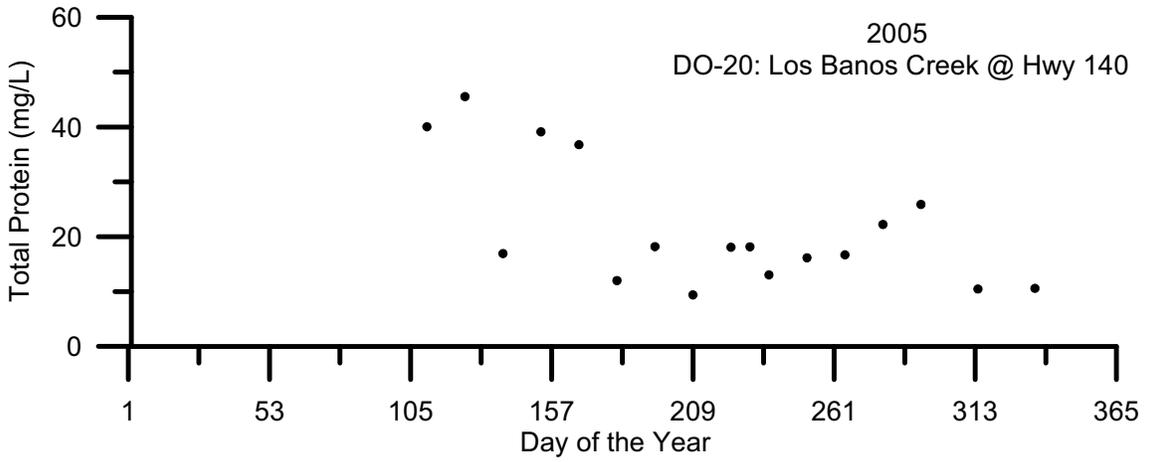


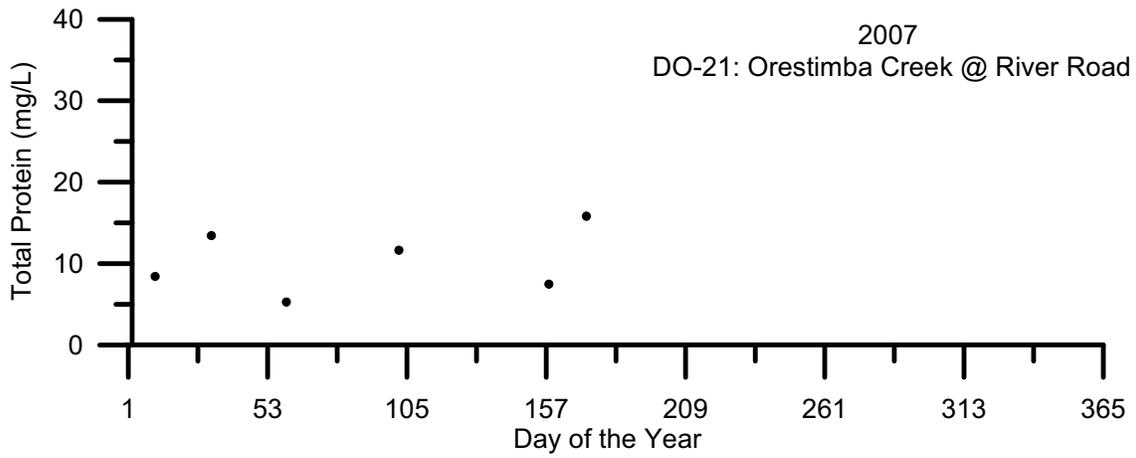
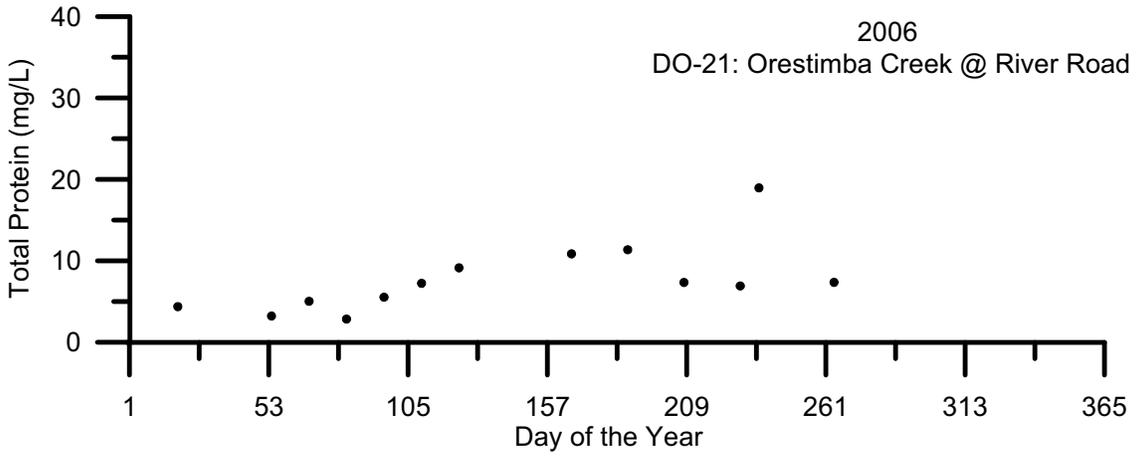
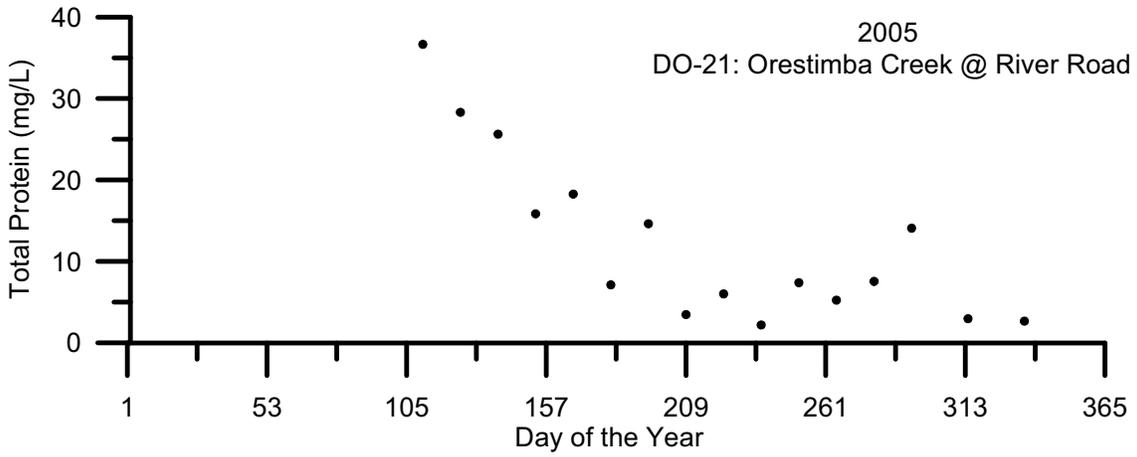


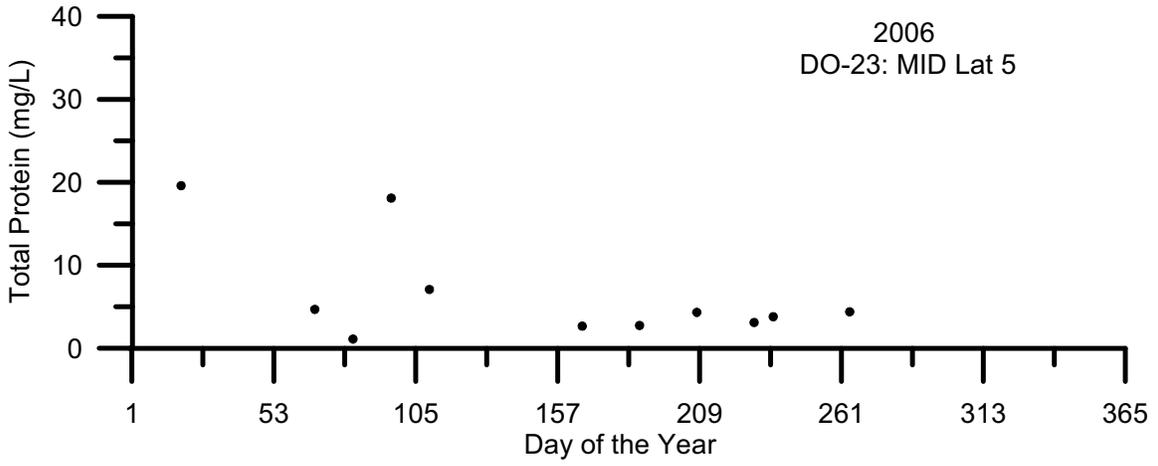
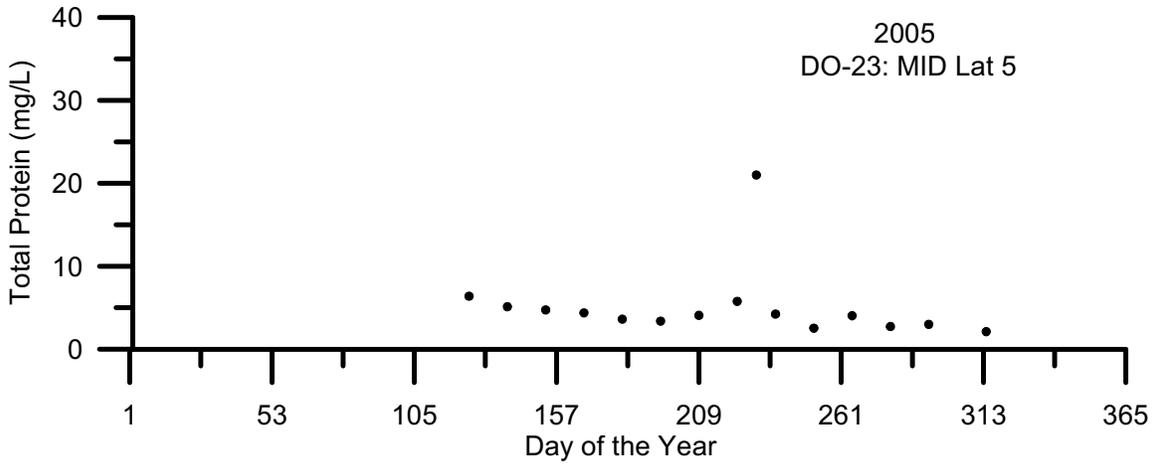


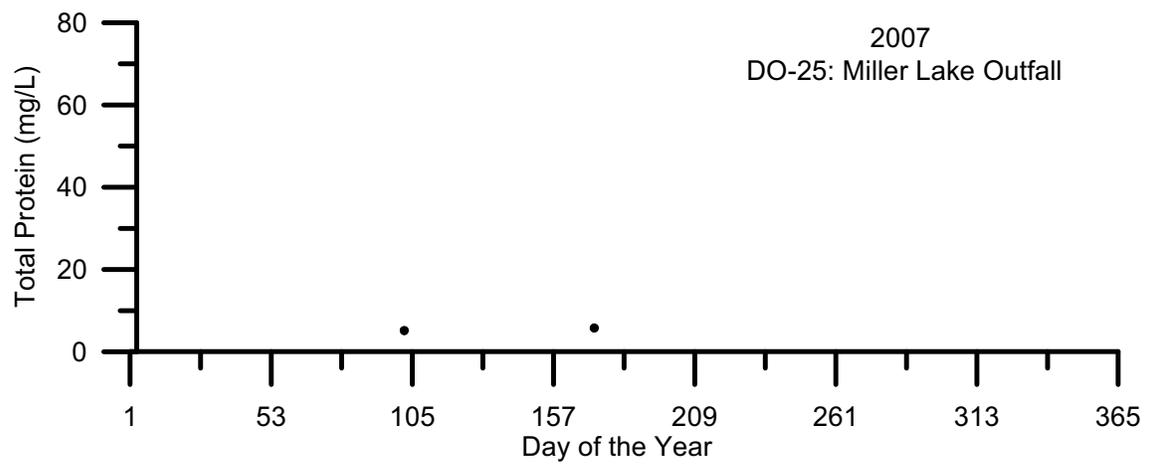
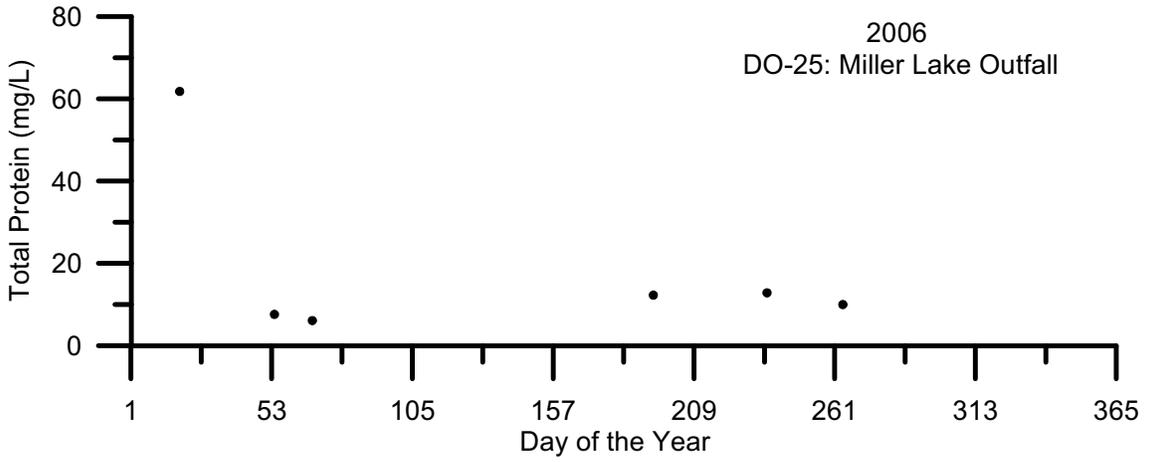
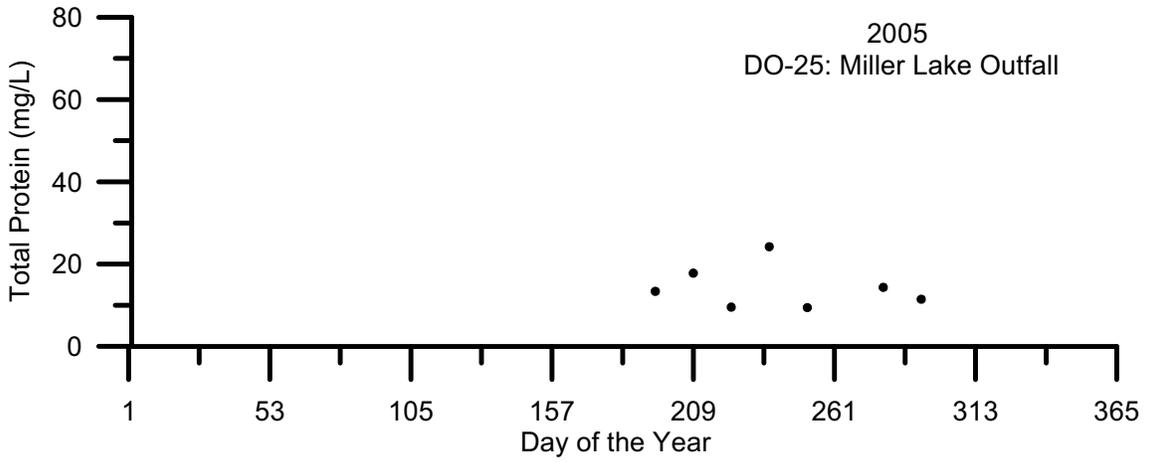


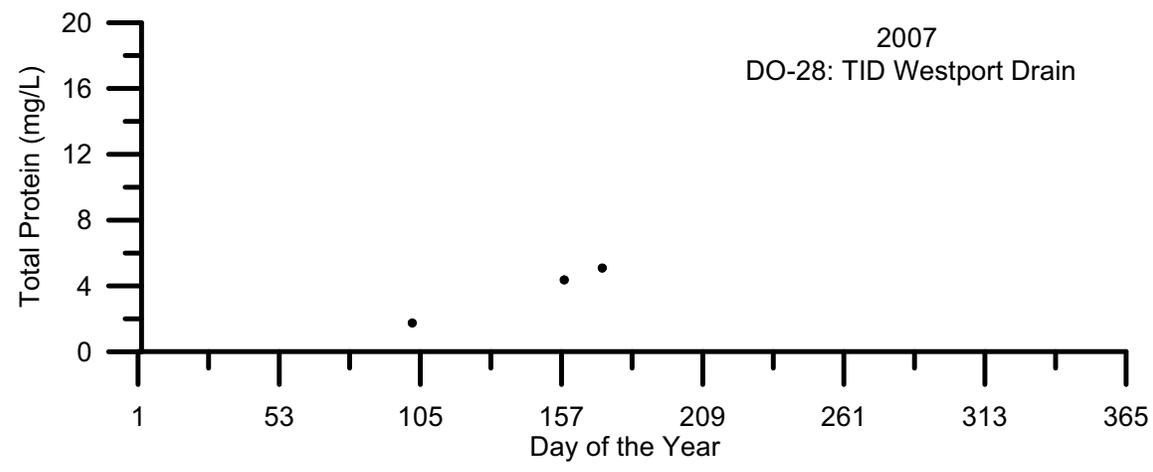
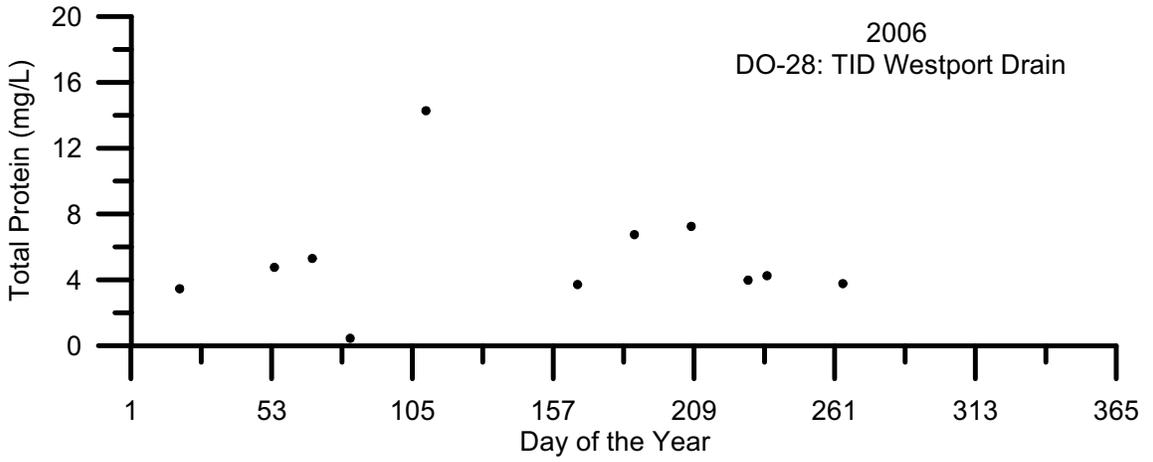
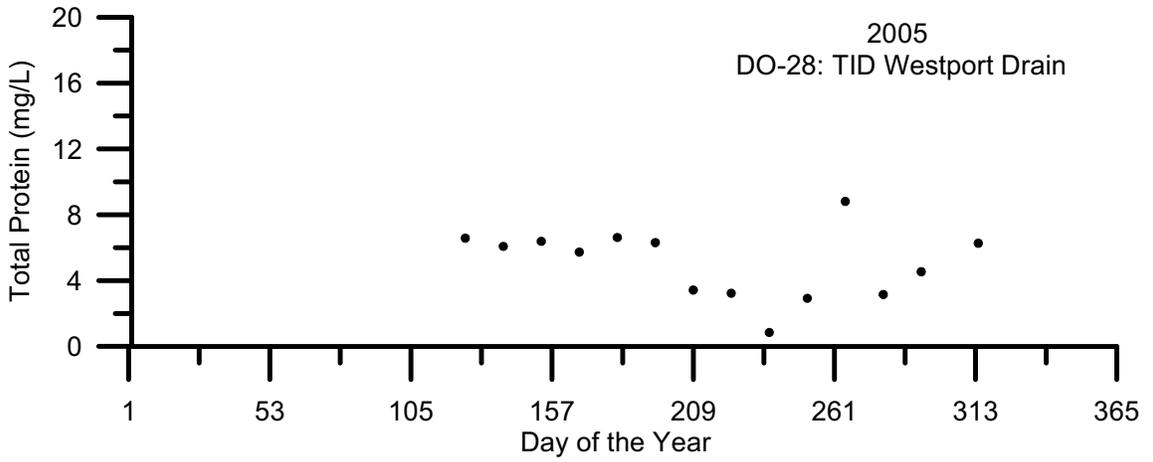


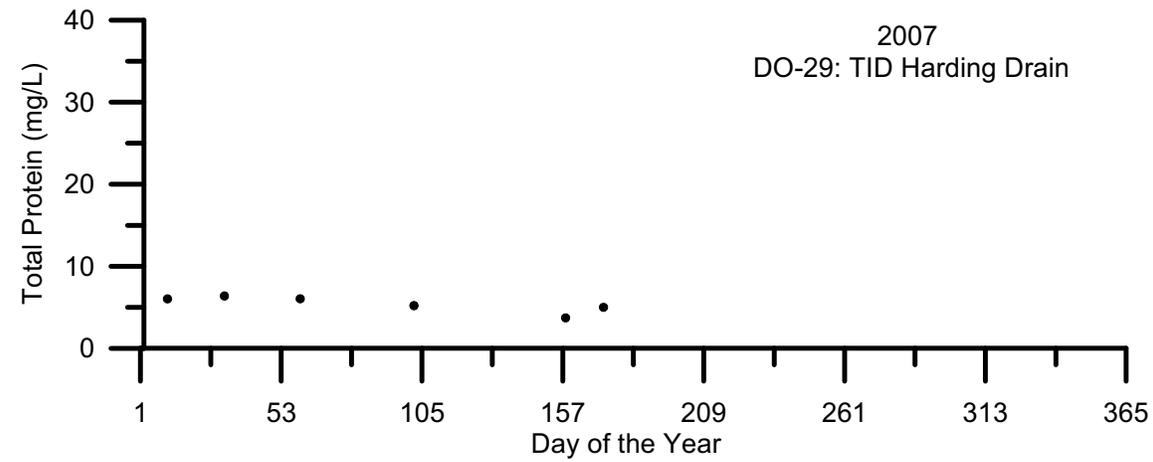
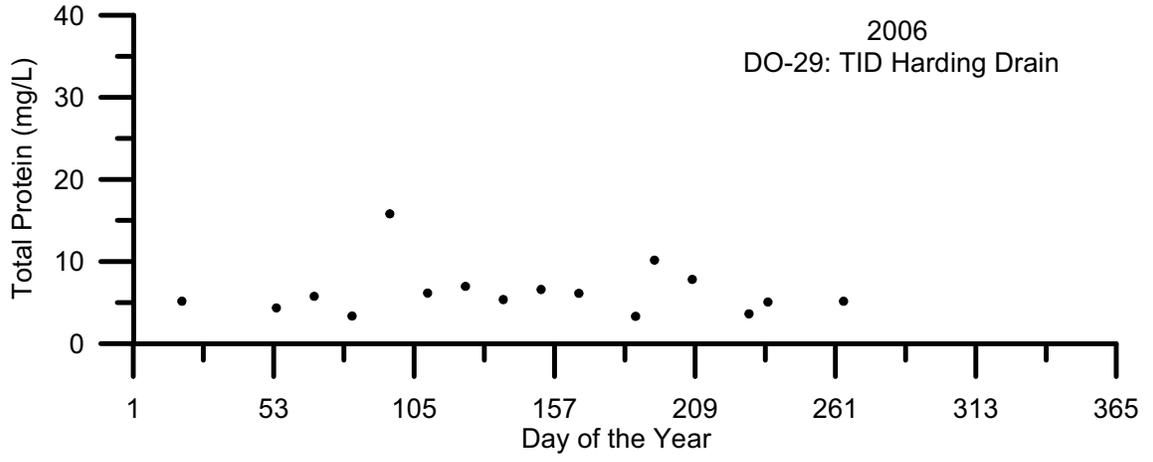
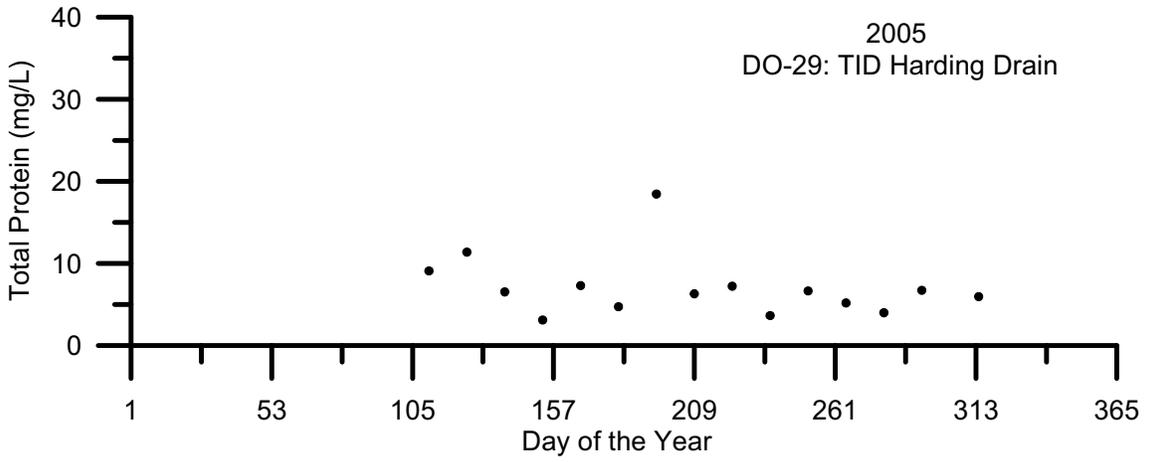


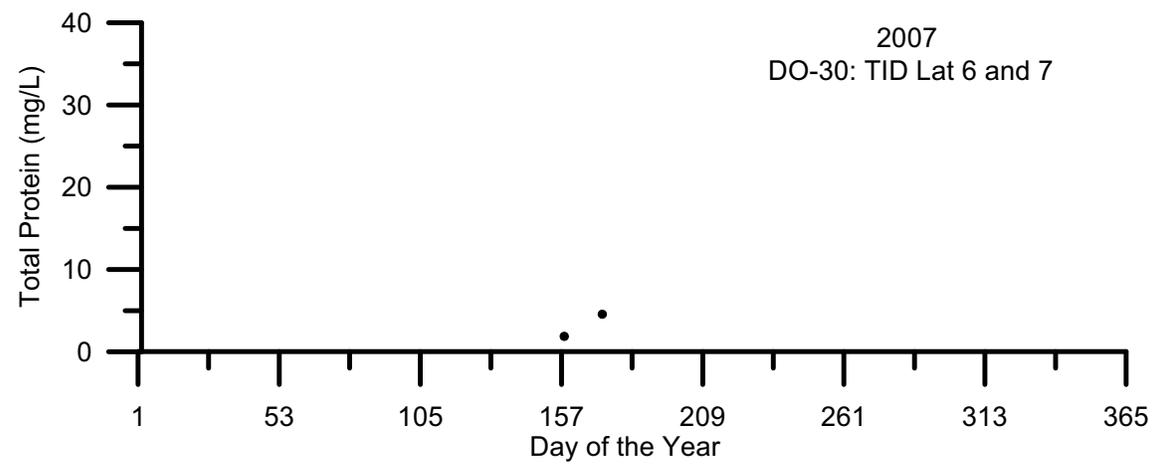
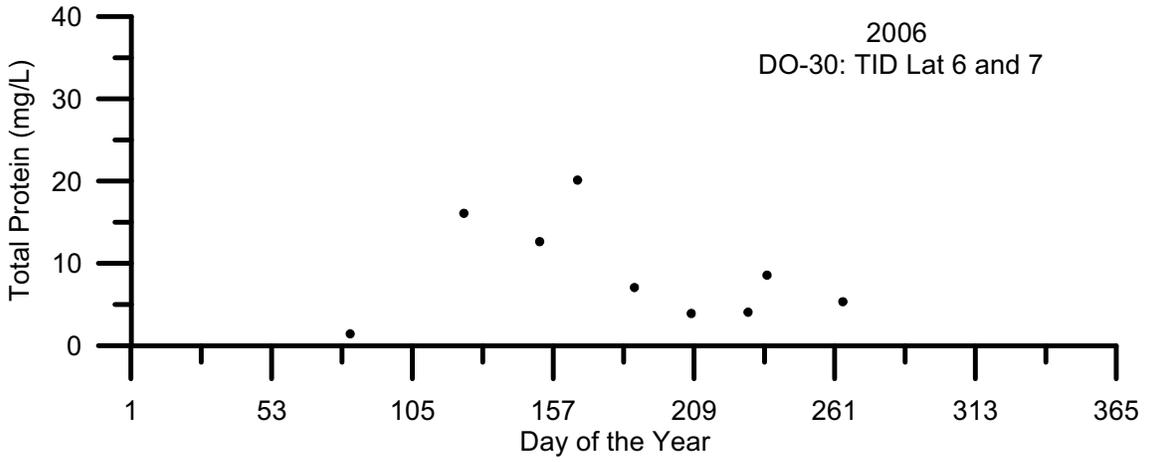
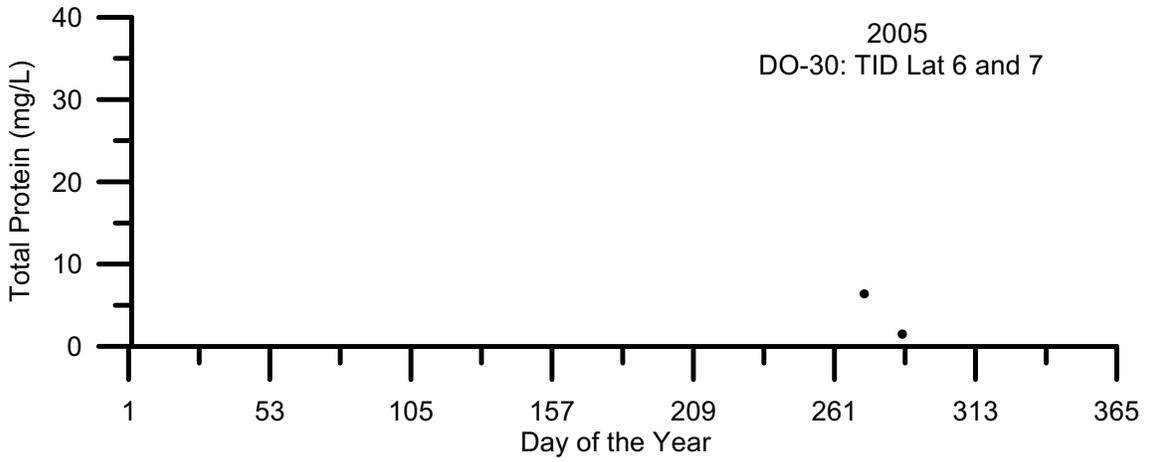


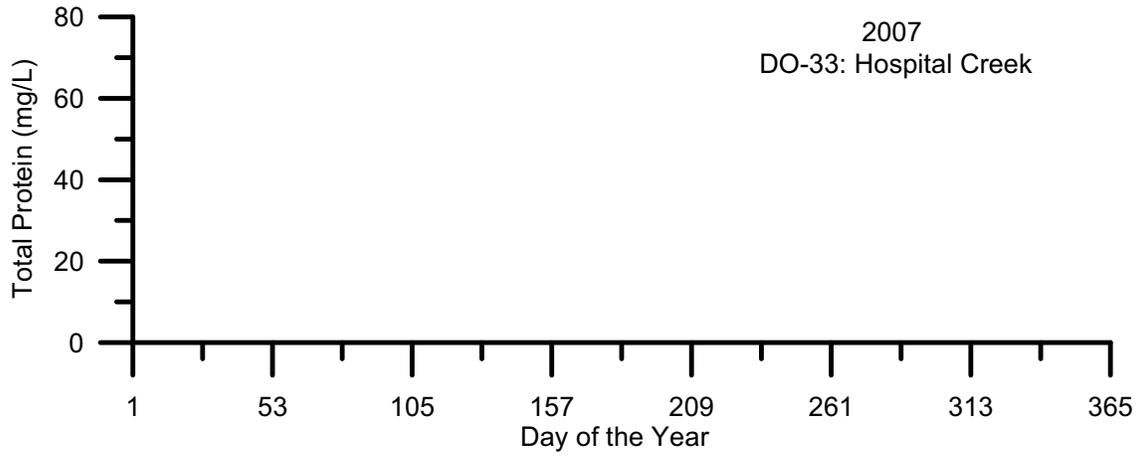
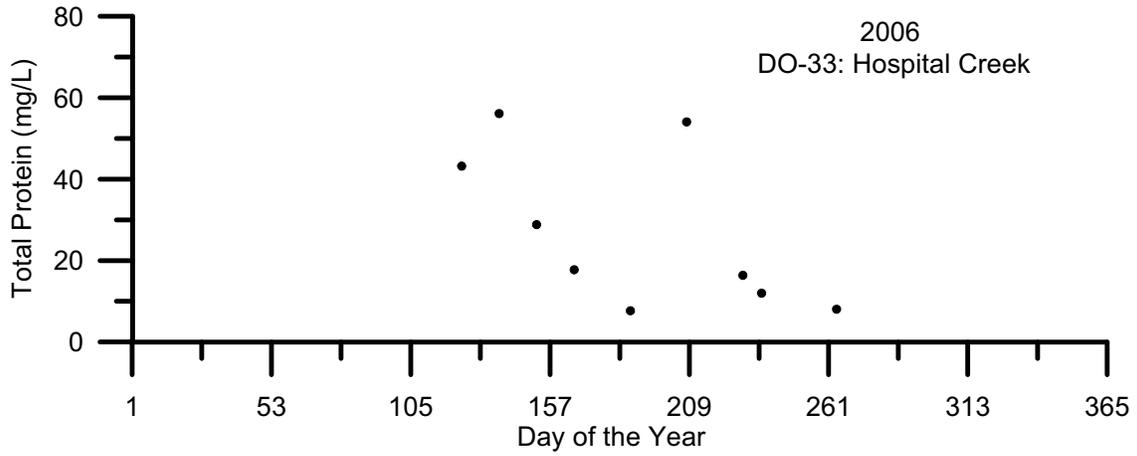
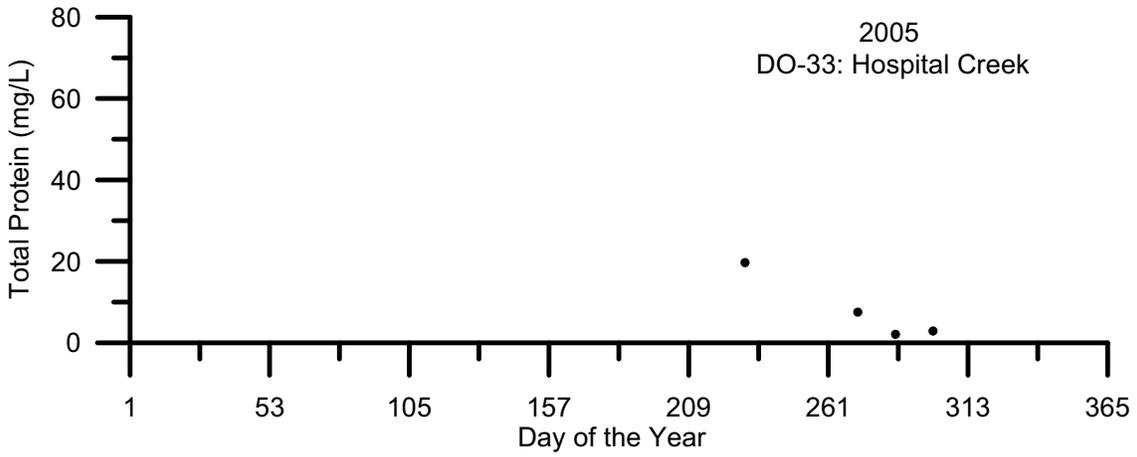


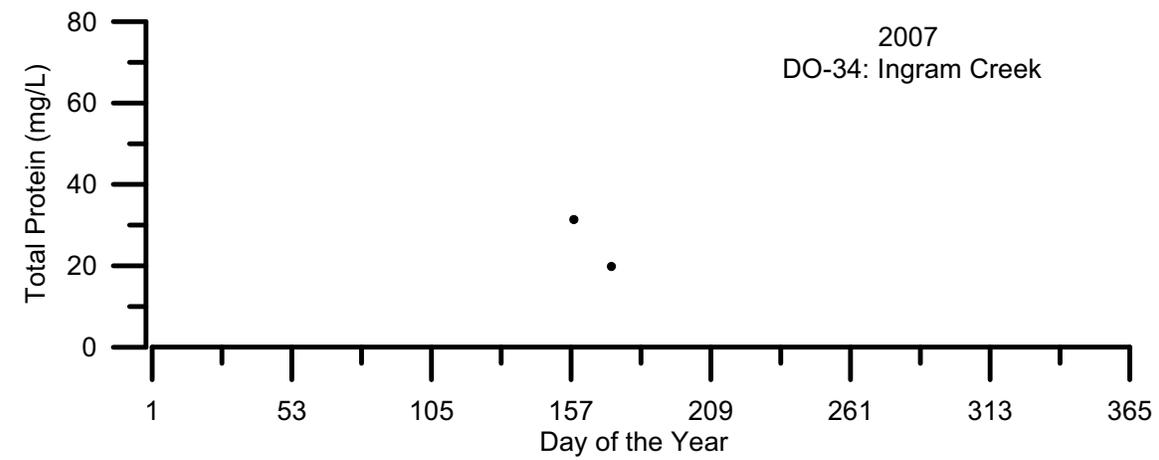
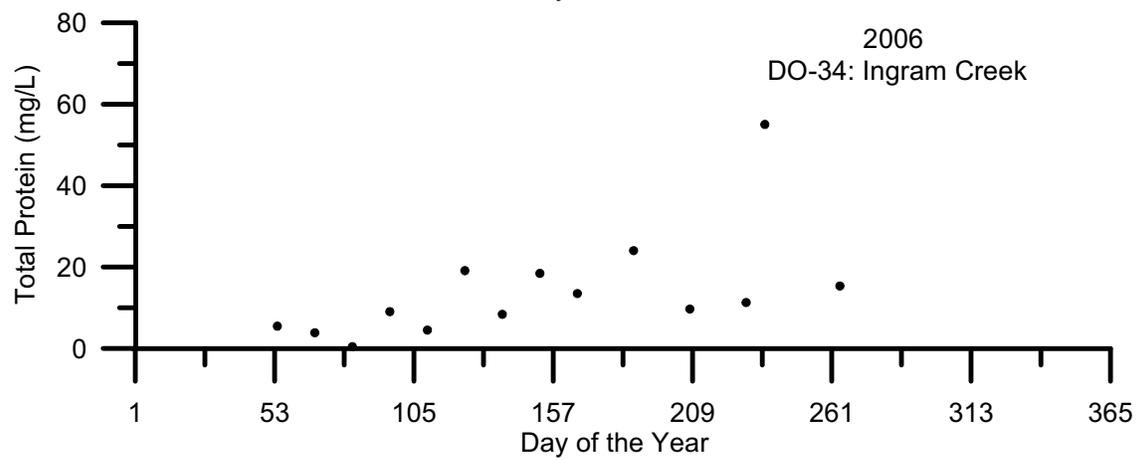
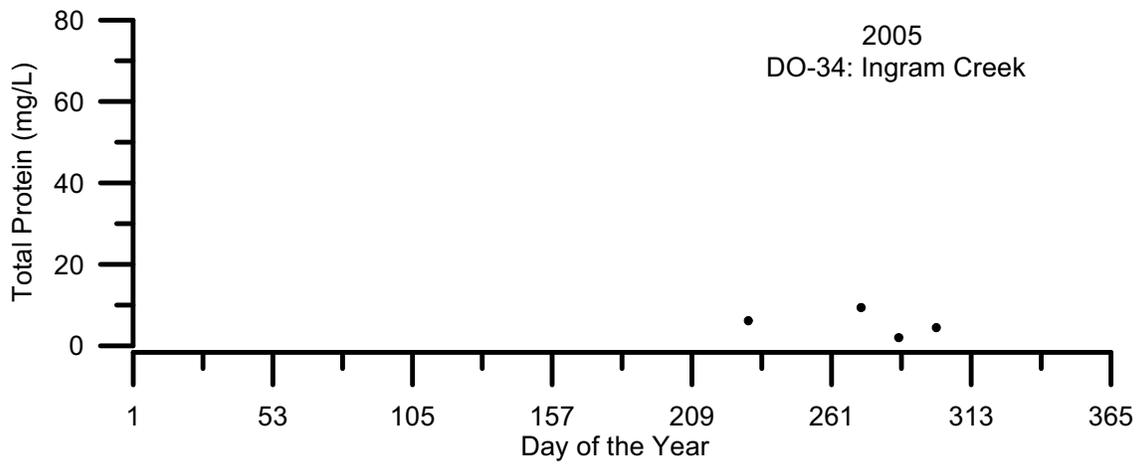


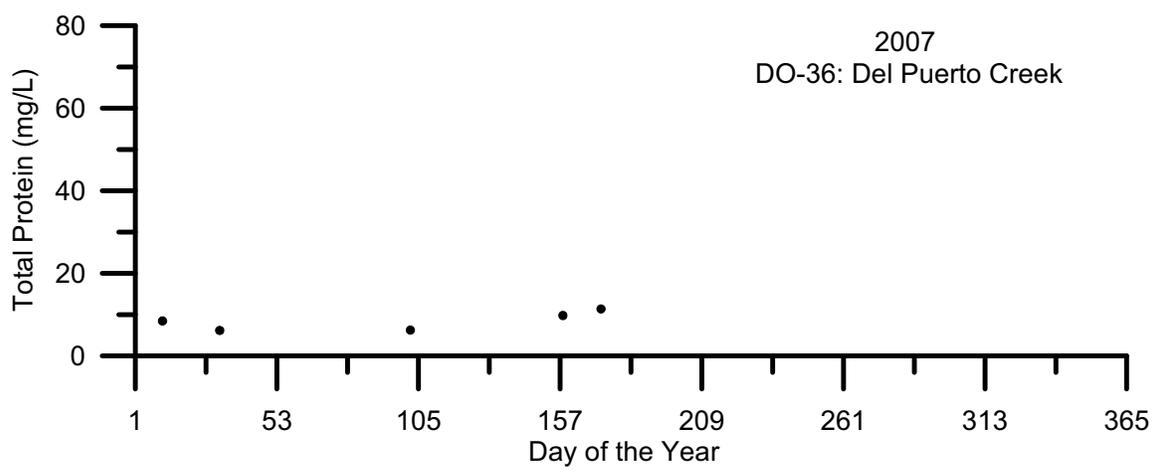
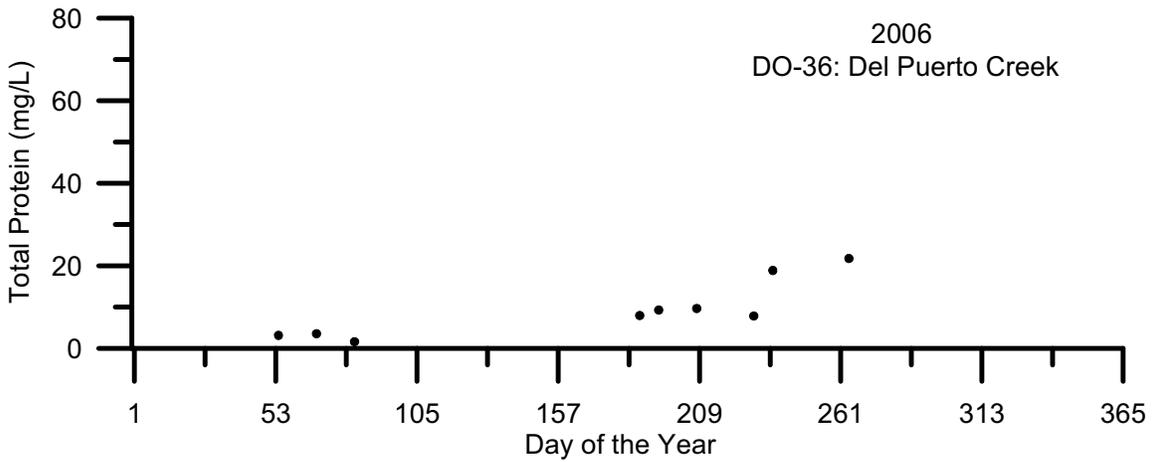
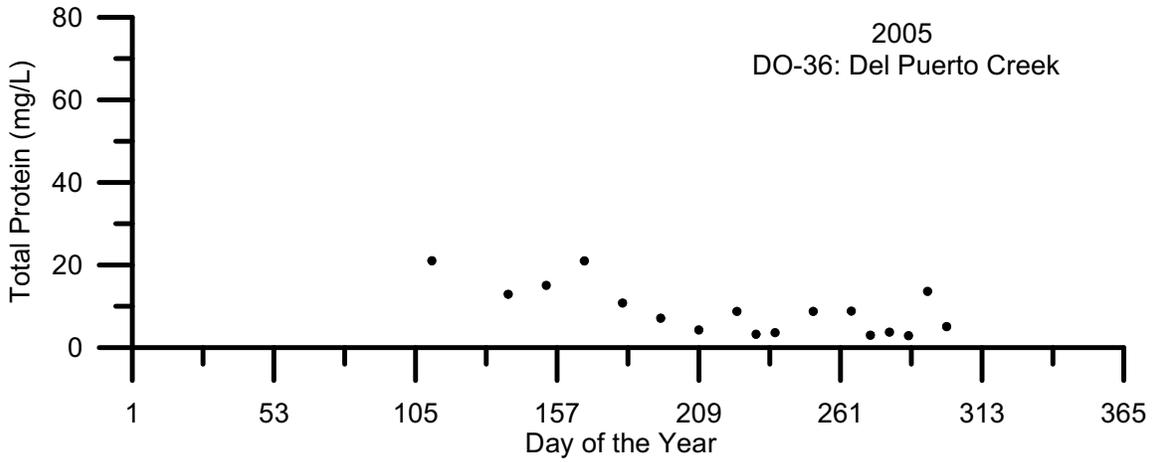


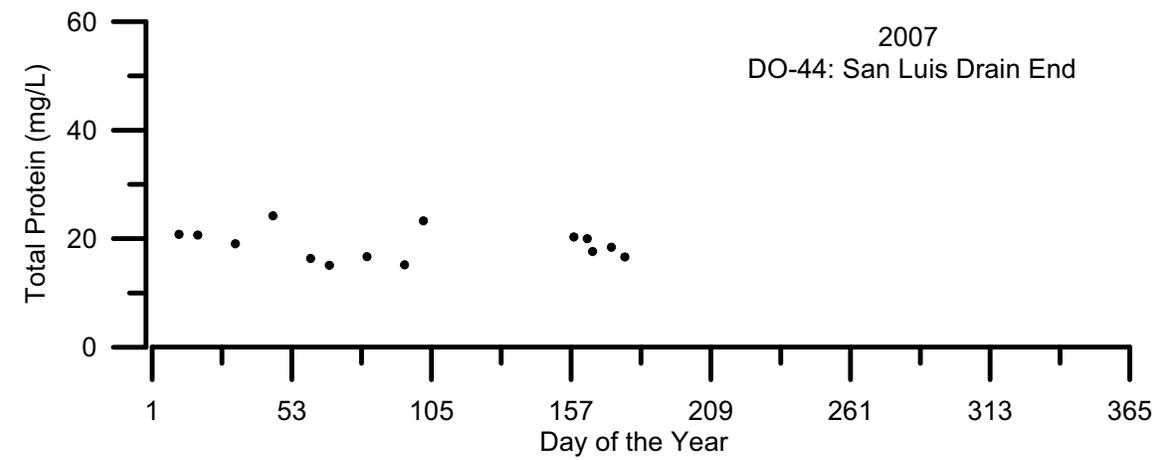
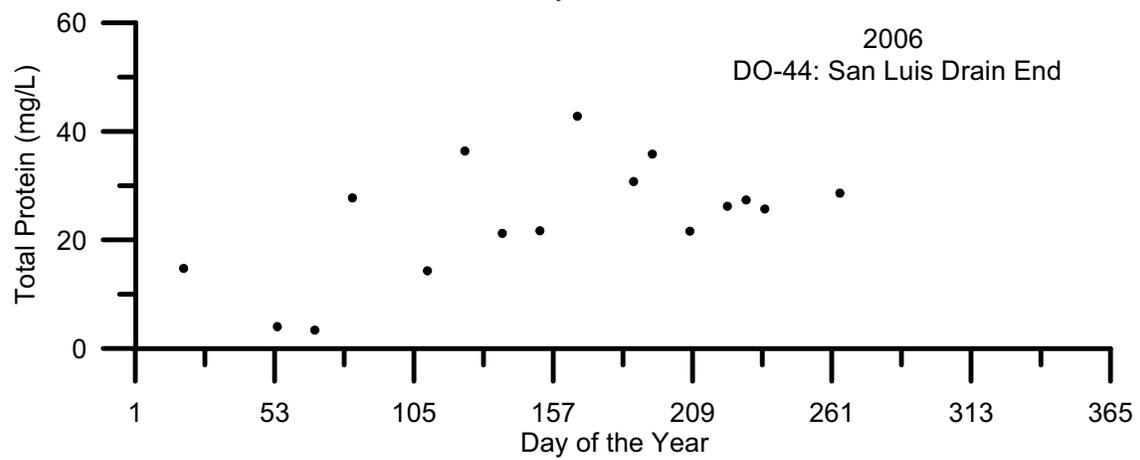
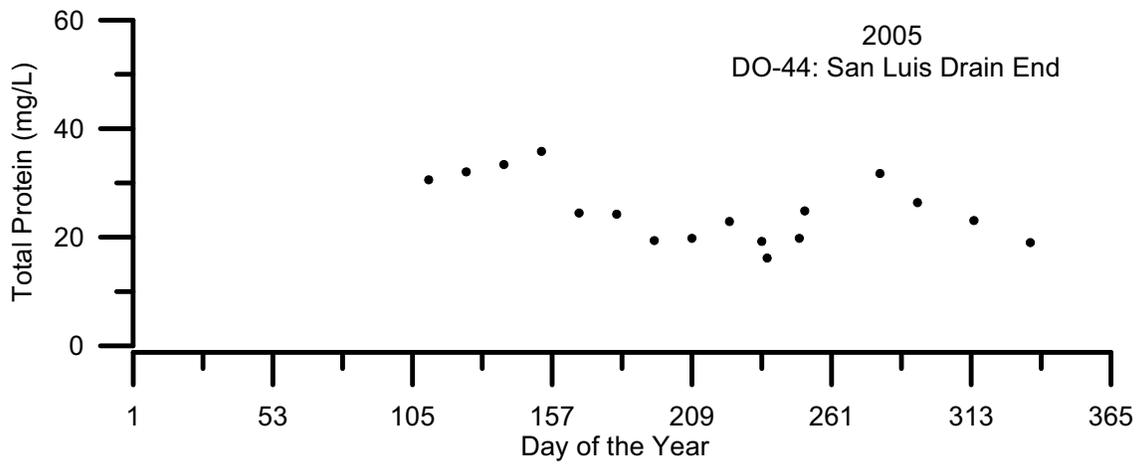


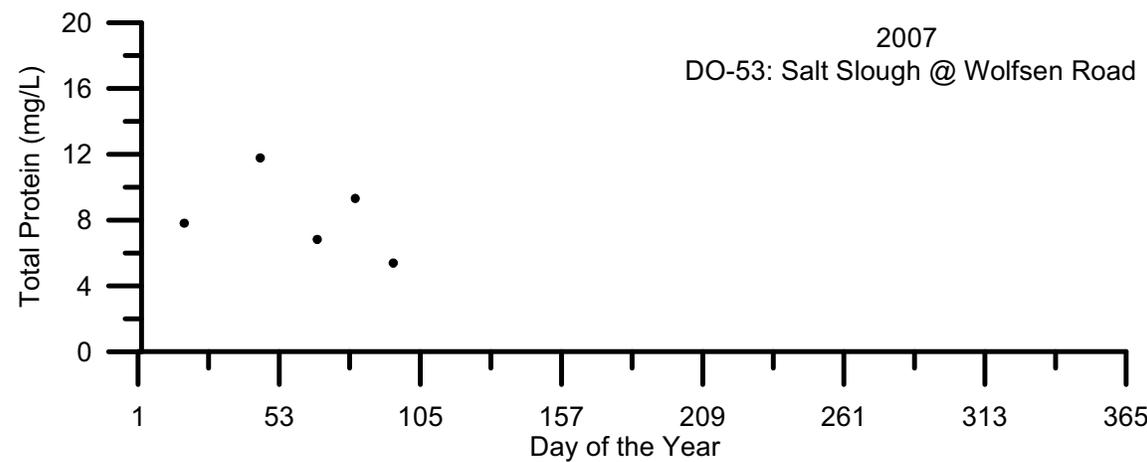
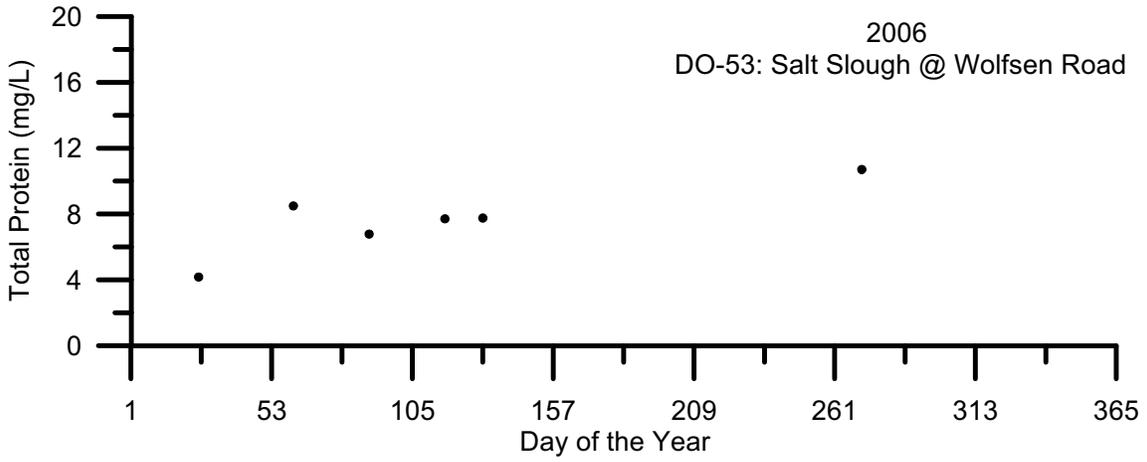
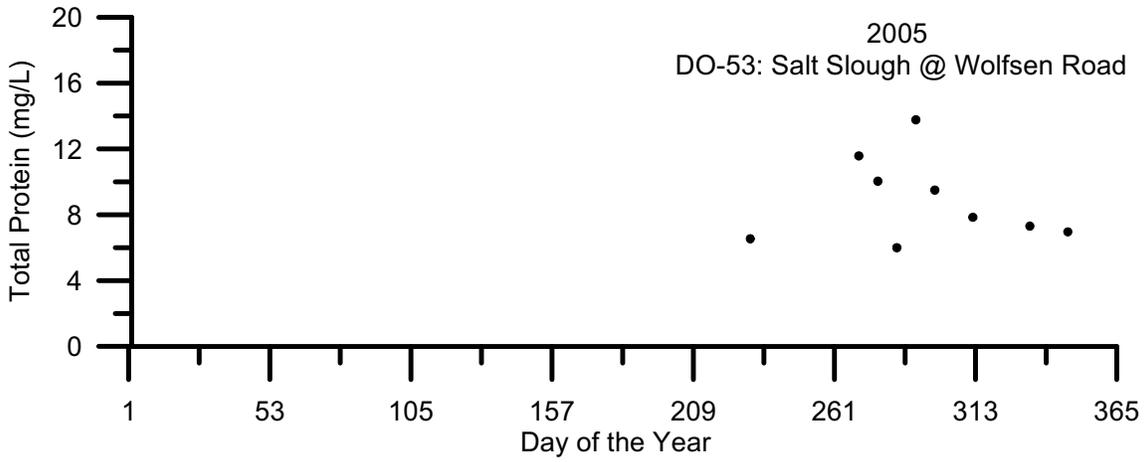


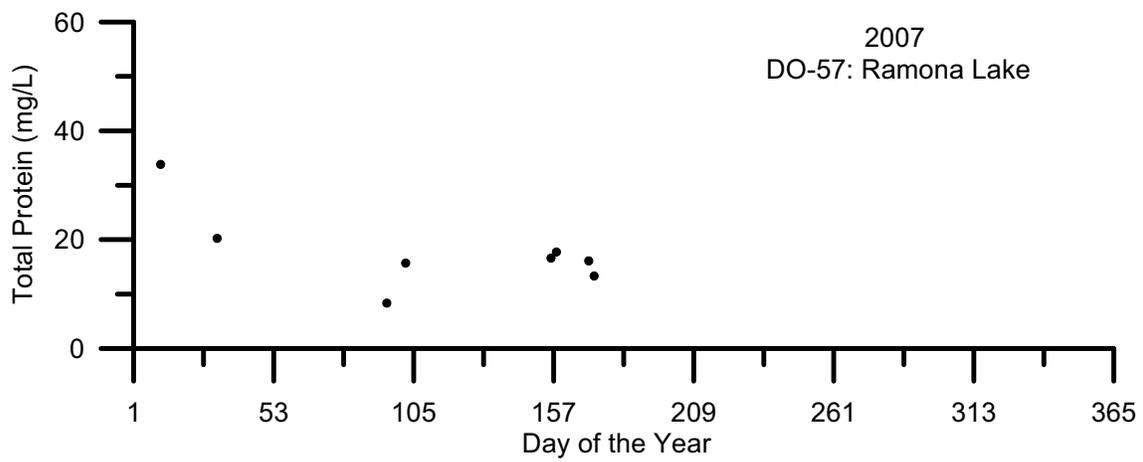
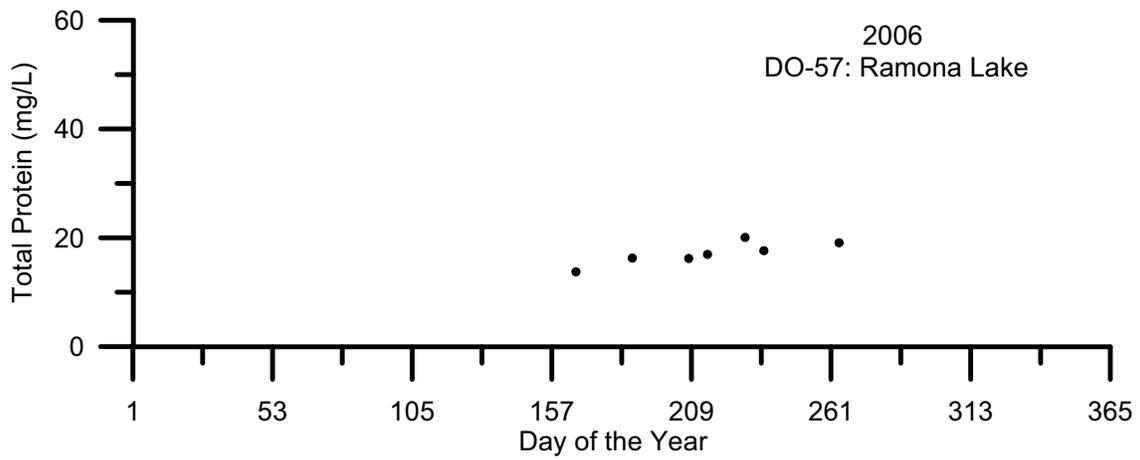


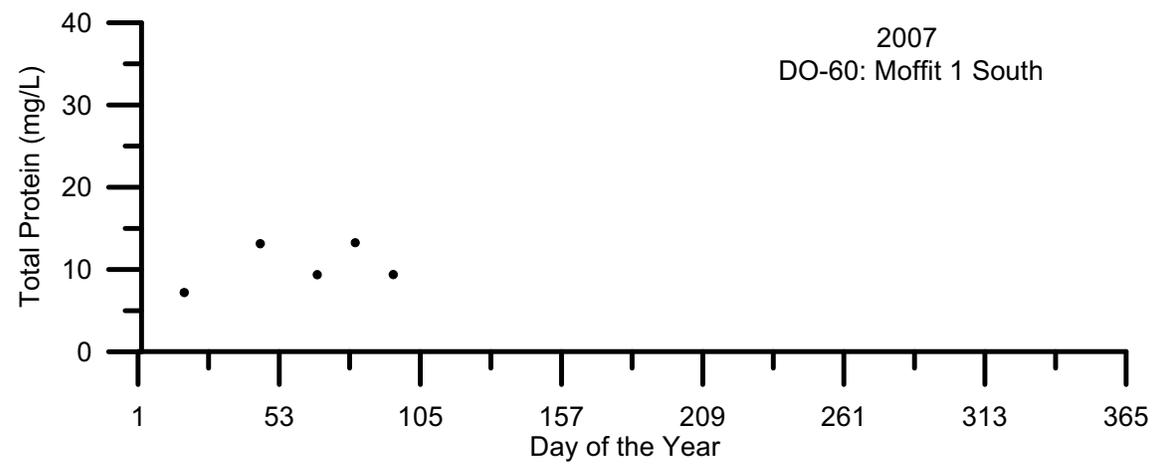
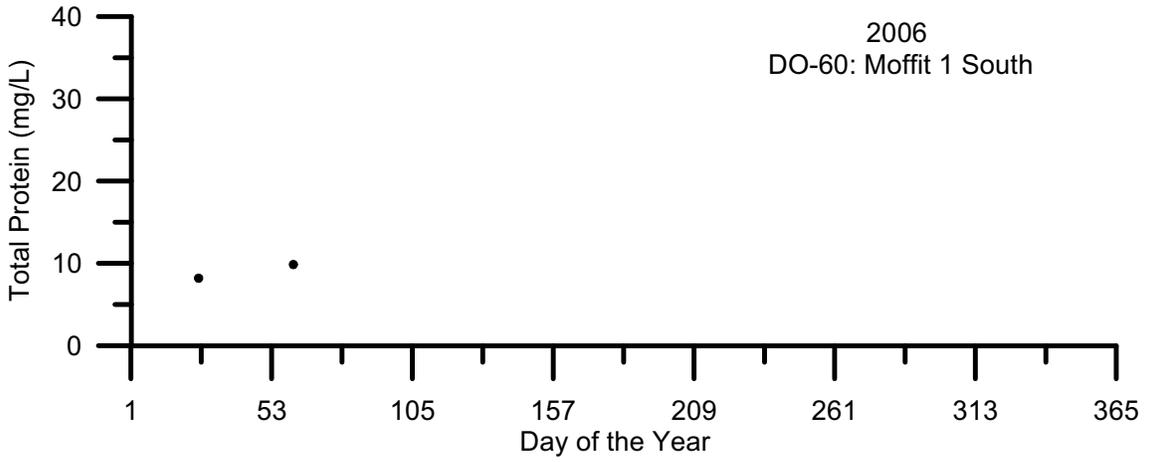
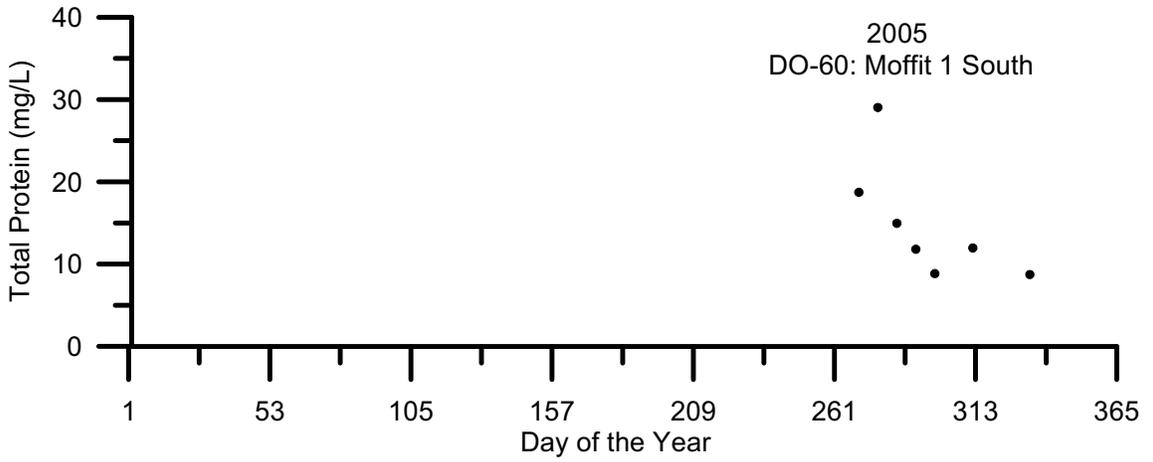


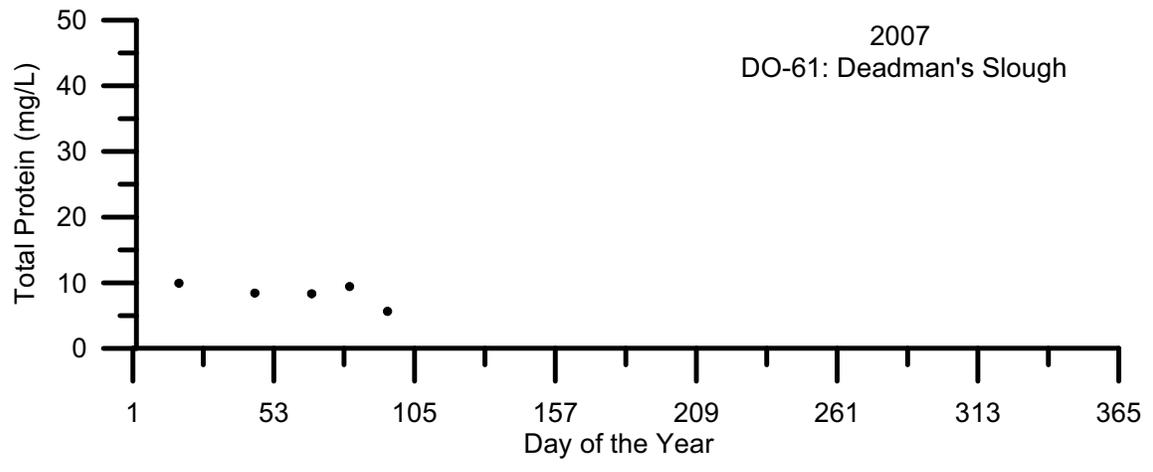
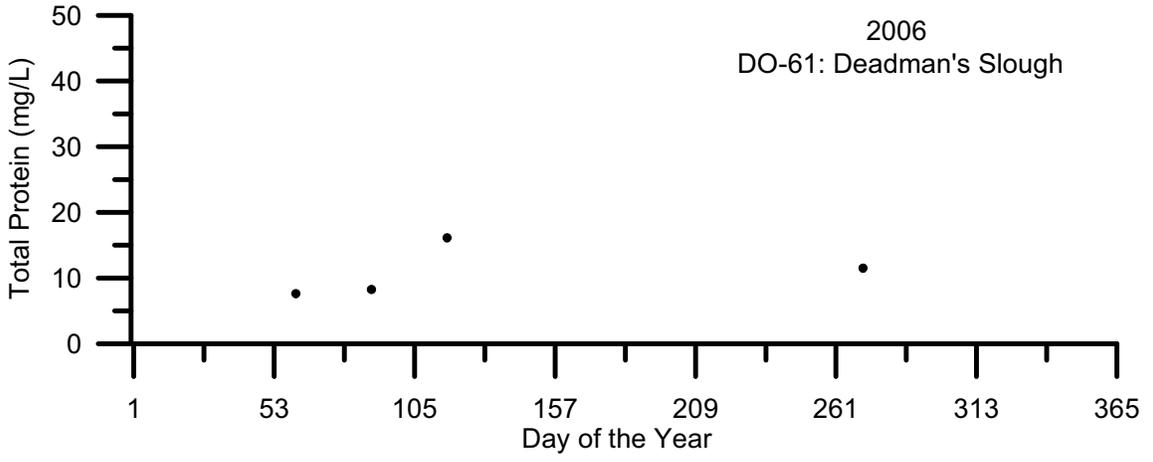
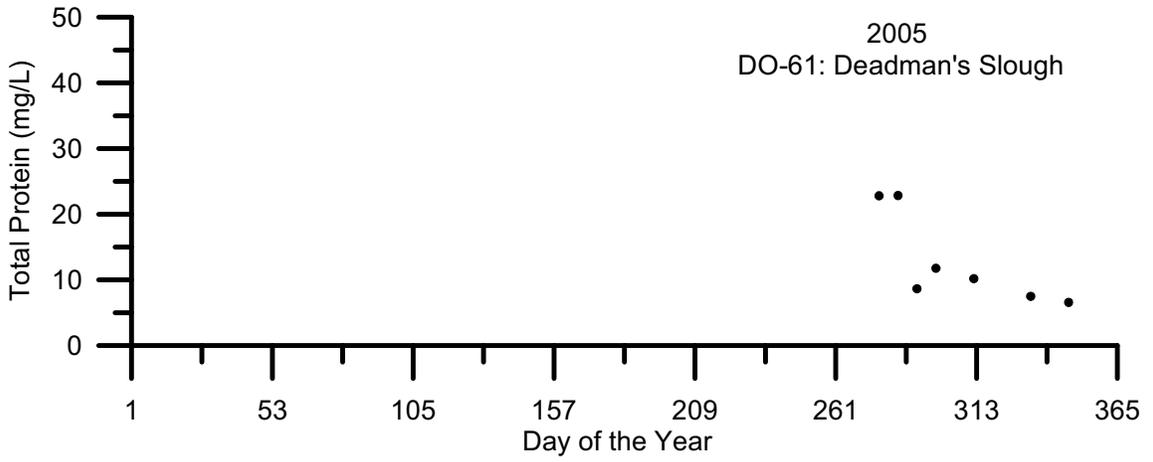


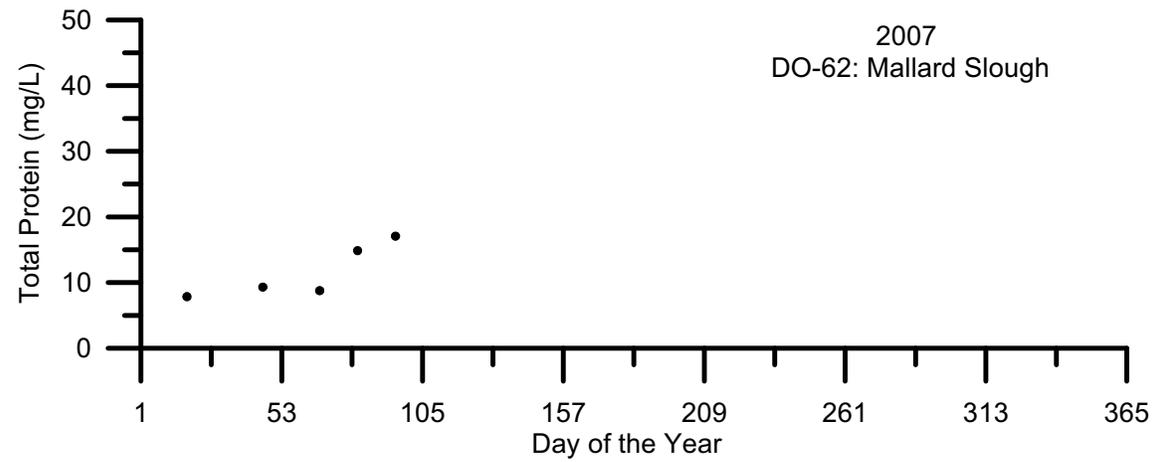
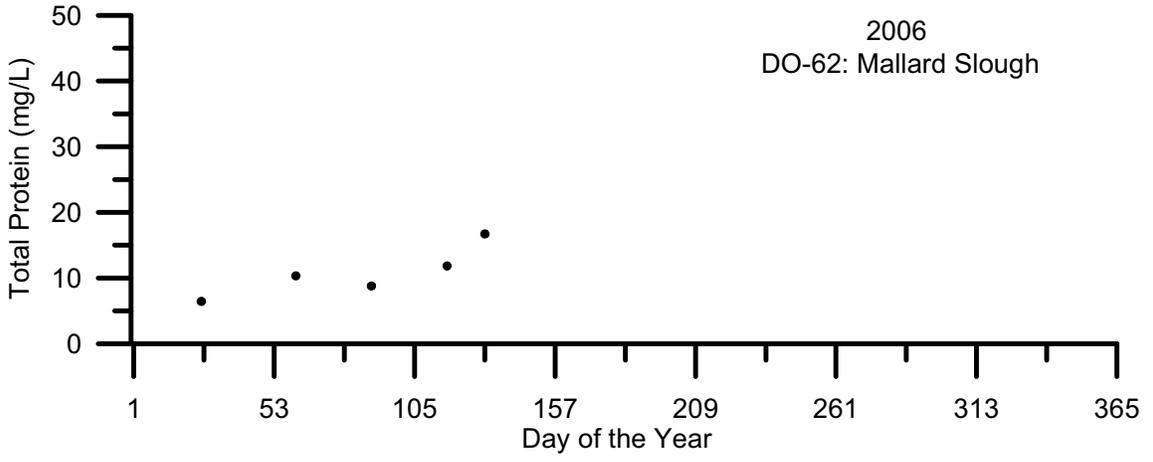
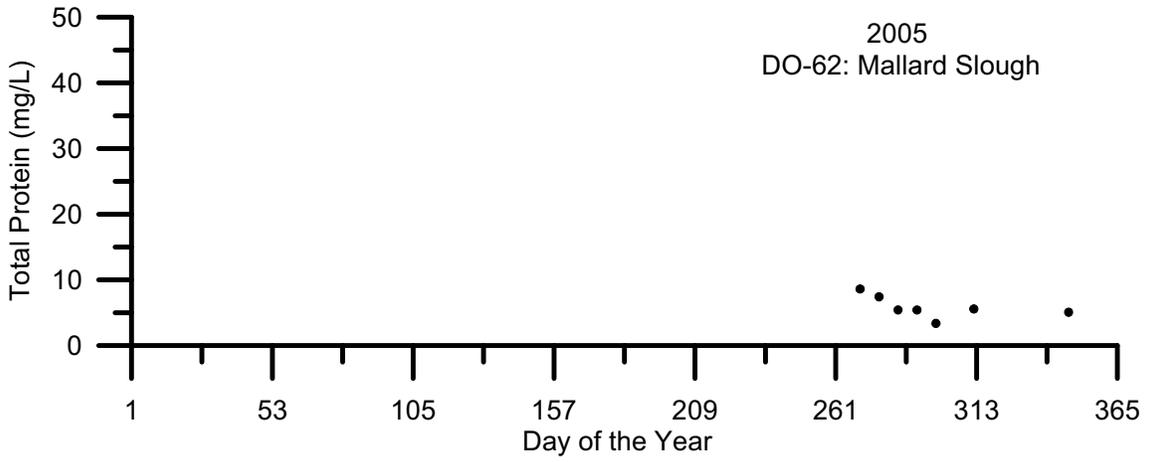


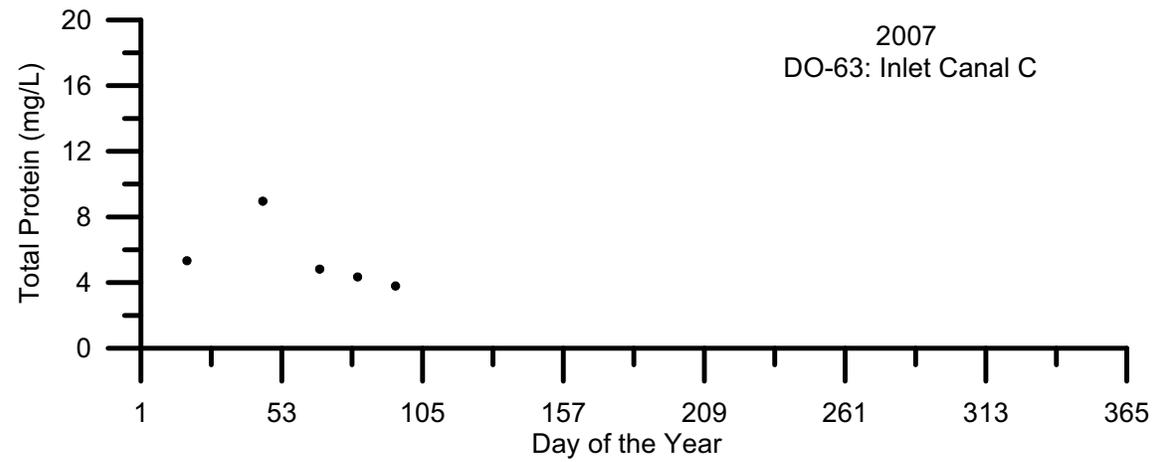
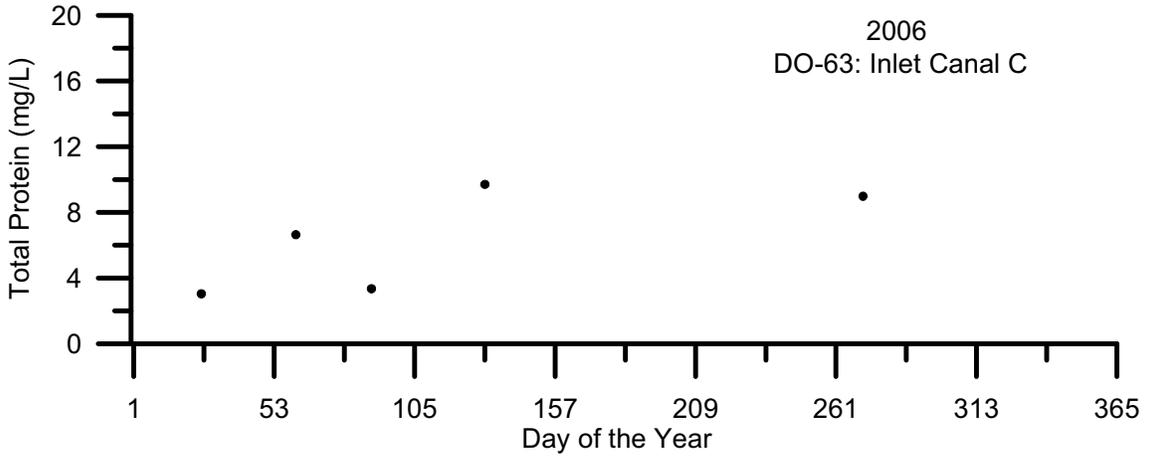
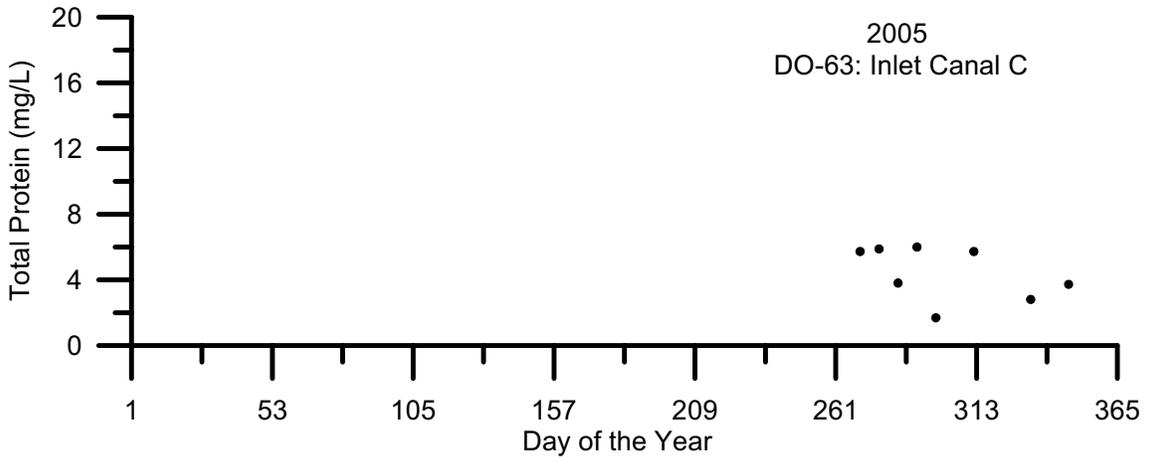


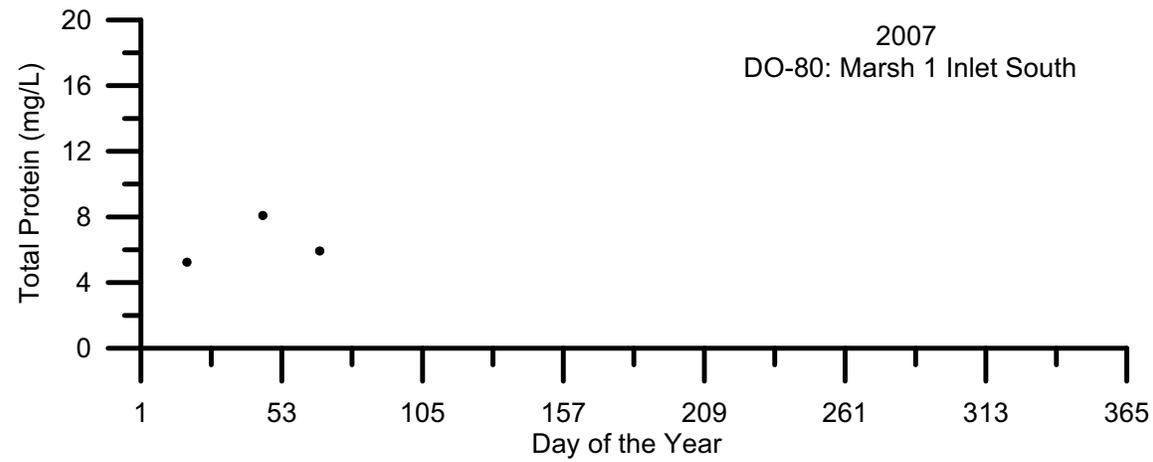
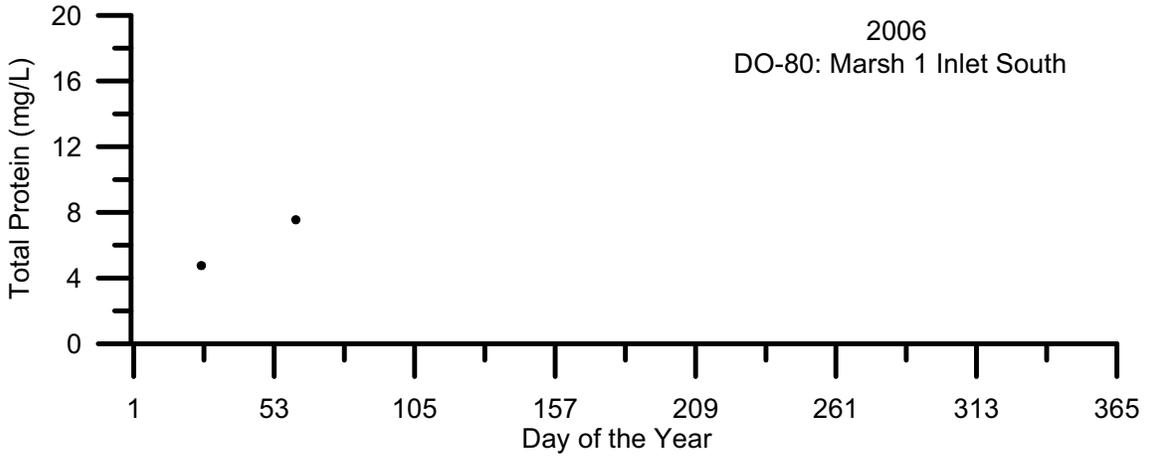
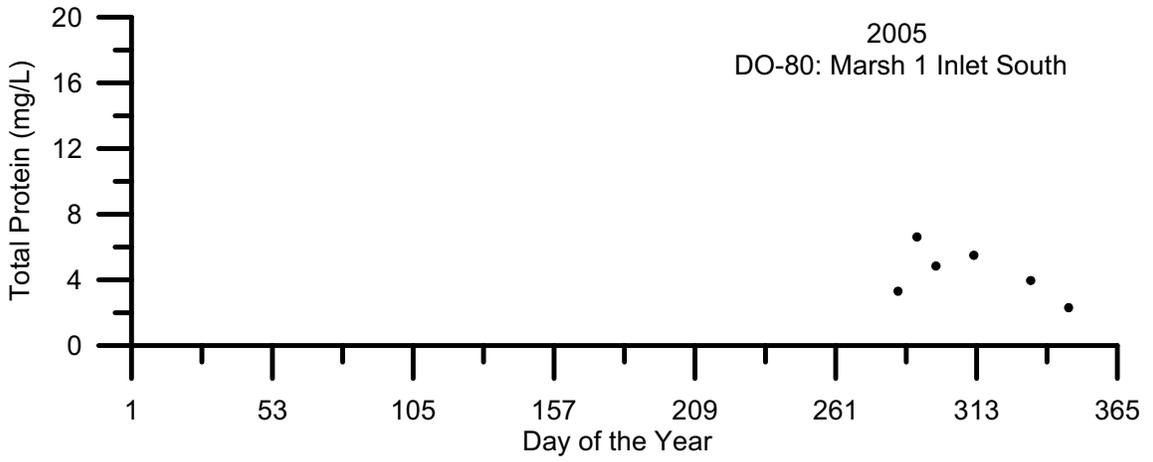


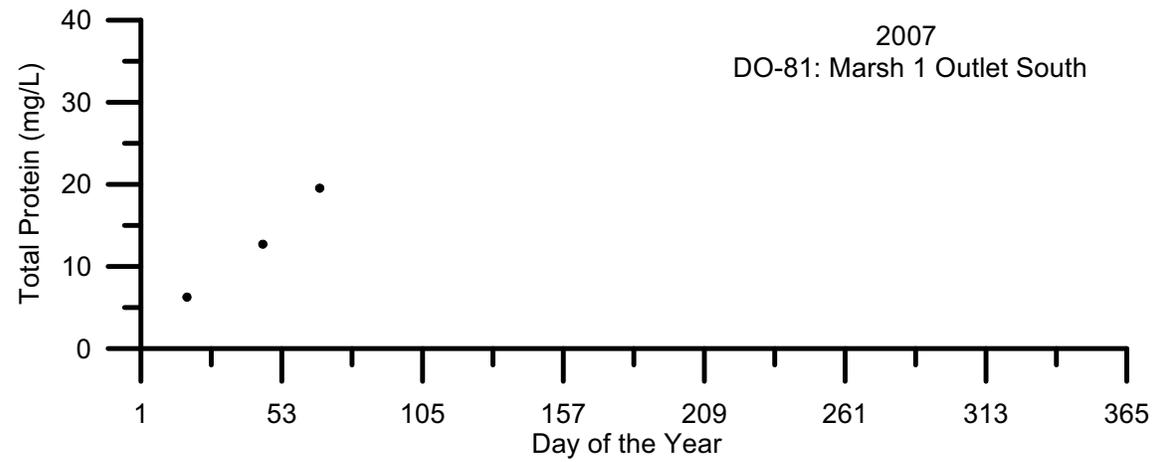
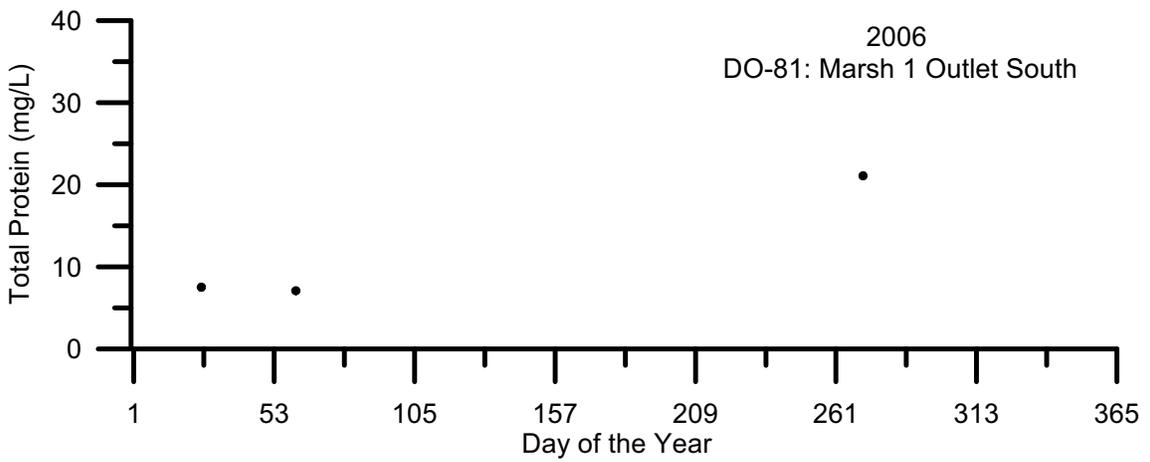
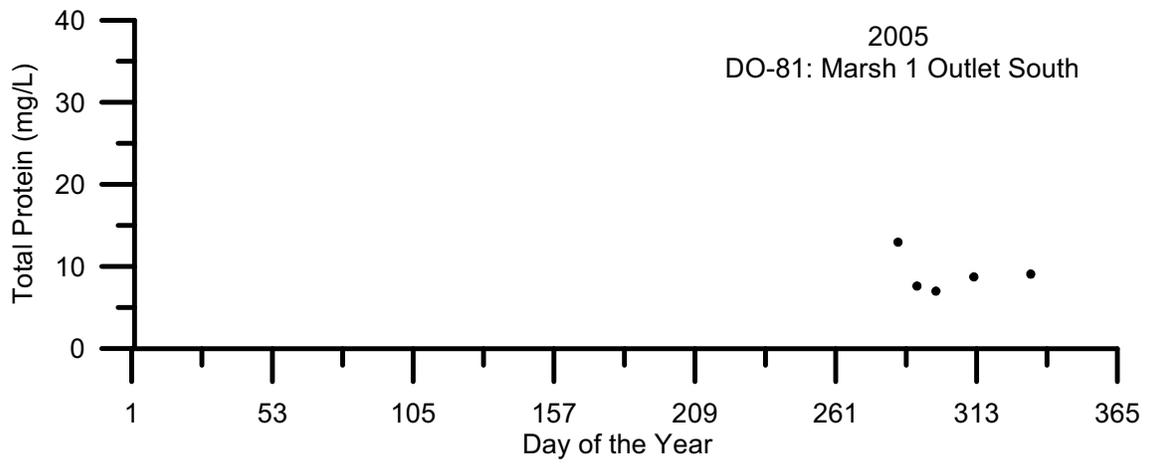


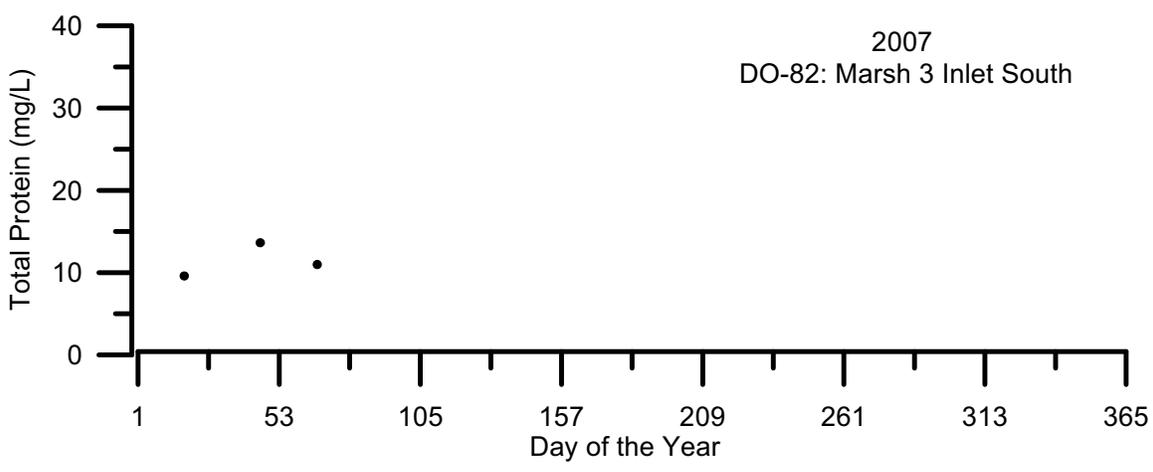
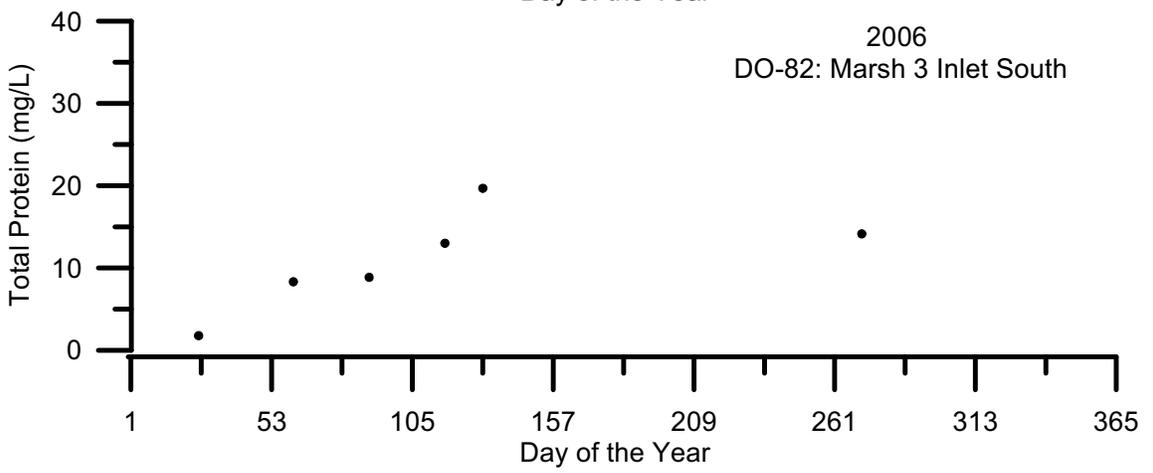
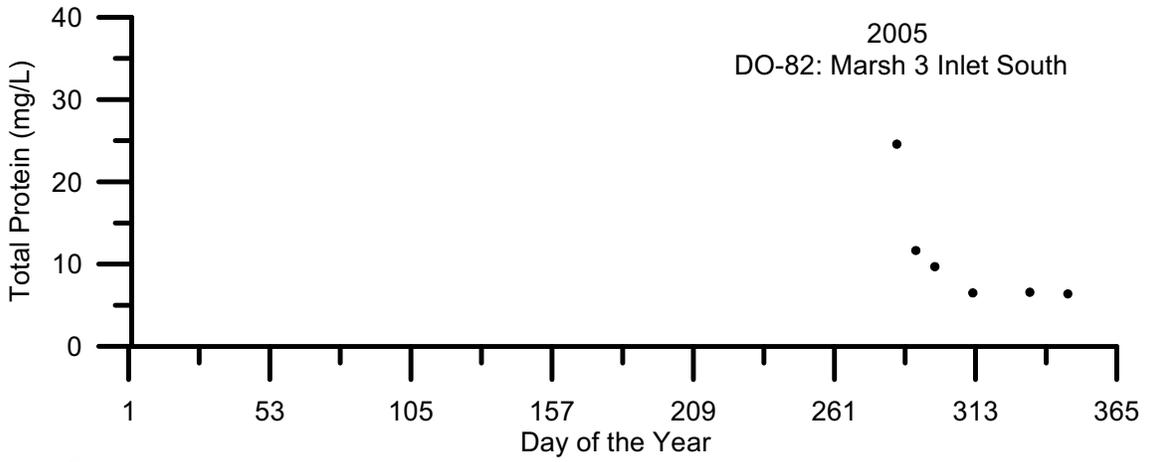


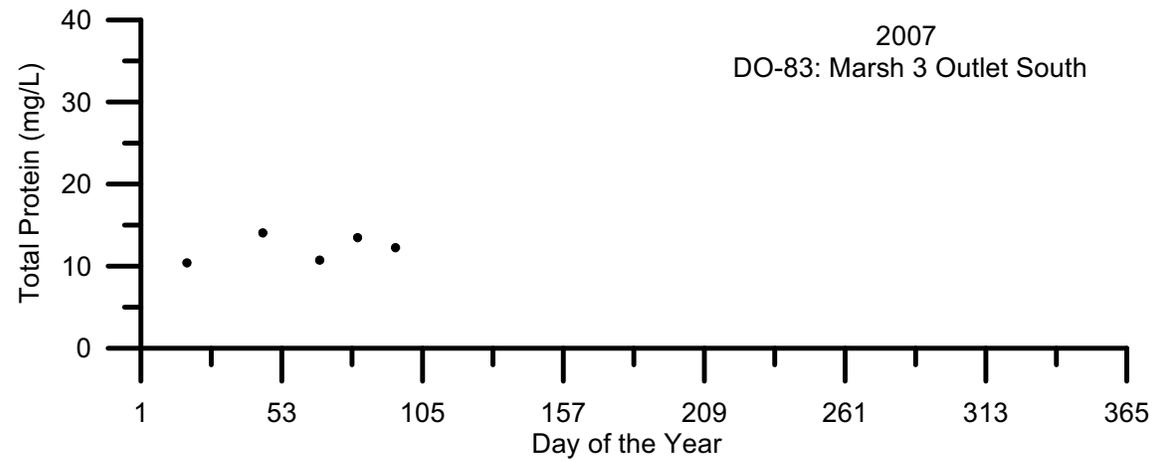
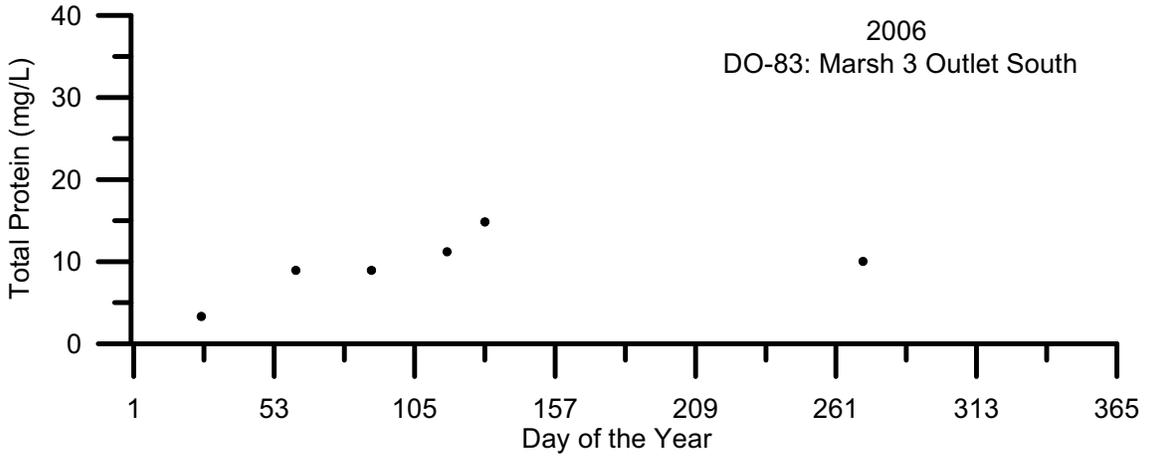
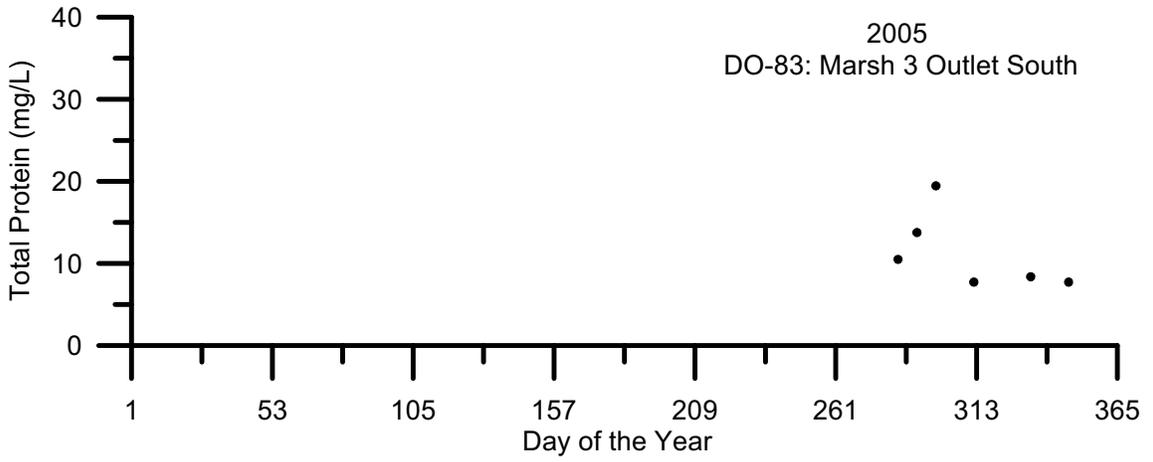


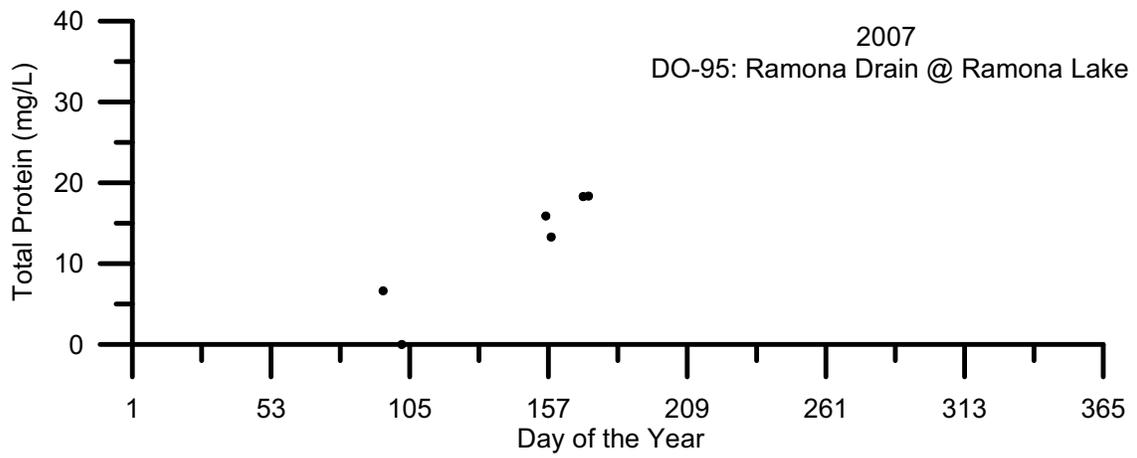
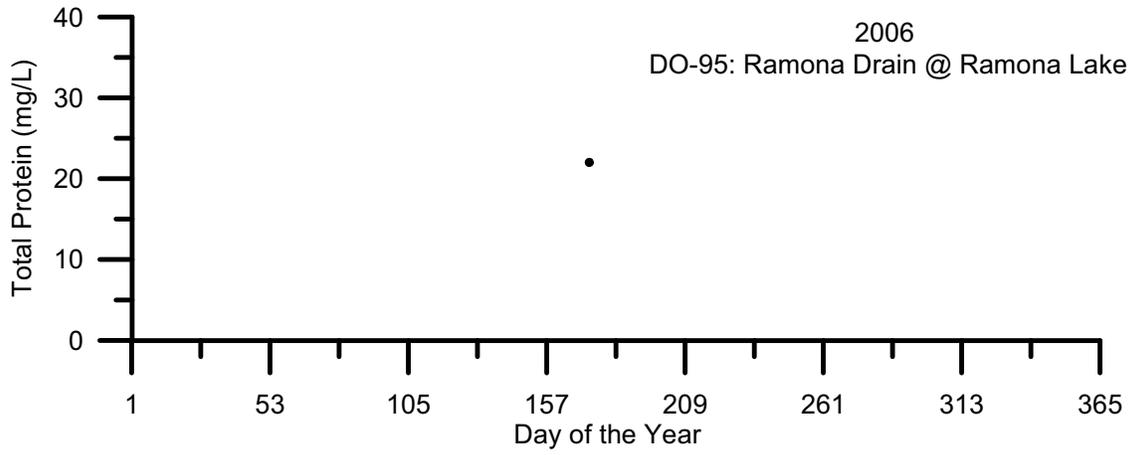














**Analysis of Total Organic Carbon Concentrations
in the San Joaquin River Watershed
2005-2007**

*Remie Burks
Chelsea Spier
Sharon Borglin
Jeremy Hanlon
Justin Graham
William Stringfellow*

February 2008

*Environmental Engineering Research Program
University of the Pacific
3601 Pacific Avenue, Sears Hall
Stockton CA 95211*

Introduction

The San Joaquin River (SJR) supports one of the most productive agricultural regions in the world and its productivity is heavily dependant on irrigated agriculture. A consequence of irrigated agriculture is the production of return flows conveyed down gradient drains that eventually discharge to surface waters. Agricultural drainage may have significant nutrient load and can impact algae growth and general water quality in the SJR. Individual farmers and agricultural organizations, such as drainage authorities, are in need of tools to manage the environmental impacts of agricultural activities (Stringfellow, 2008).

For the years 2005 through 2007, sites throughout the San Joaquin Valley watershed were sampled to assess the overall water quality in the region. One thousand nine hundred and ninety-six (1996) individual surface water samples were collected and analyzed and WQ was assessed at 113 sampling locations in the SJR basin (Borglin et al., 2008). Samples were processed and analyzed by the Environmental Engineering Research Program (EERP) laboratory at the University of the Pacific as well as at the University of California, Davis, Dahlgren Lab. This report presents temporal plots of total organic carbon (TOC) for all sites sampled in the SJR from 2005-2007.

Methods

Field sampling consisted of collecting water samples, measuring water quality with a YSI Sonde 6600 with MDS650 hand-held display, and recording of field conditions at sites within the study area per *EERP Field Protocol Book* (Graham, 2008).

Water samples were collected in glass 1000 mL bottles (Wheaton Science Products, Millville, NJ), 1000 mL HDPE Trace-Clean narrow mouth plastic bottles (VWR International), 250 mL HDPE Trace-Clean wide mouth plastic bottles (VWR International) as well as 40 mL trace clean vials with PTFE septa (ICChem, Rockwood, TN) in accordance with requirements for different lab analysis and volume requirements.

Samples were immediately stored at 4°C after sampling (cooler temperature was recorded in the lab upon delivery) and transported to the lab on the day of sampling.

Samples were received by the laboratory the same day they were sampled and stored at 4°C until filtering and analysis. Samples were collected, preserved, stored, and analyzed by methods outlined in *Standard Methods for the Analysis of Water and Wastewater*, (APHA, 2005, 1998). Total organic carbon (TOC) was analyzed on a Teledyne-Tekmar Apollo 9000 (Mason, OH) by high temperature combustion according to SM 5310 B (APHA, 2005) and quantified on using a NDIR detector. TOC samples were preserved < pH 2 with concentrated H₃PO₄ and stored at 4°C until analysis. The limits of detection for TOC and DOC is 1.00mg/L C and for IC 5.00mg/L.

Results and Discussion

This analysis was preformed routinely over the years 2005-2007 using two different Apollo 9000HS instruments. This analysis had a 99.19% passage rate for all QA parameters over the three year period and a 85.71% passage rate for proficiency check

samples during that time (Borglin et. al., 2008). These high sensitivity instruments are challenging to maintain and on a few occasions sample sets were analyzed by outside laboratories due to malfunctions of the instrument that could not be repaired within the holding time for these samples.

References

Stringfellow, W.T., et al., (2008), *Evaluation of Vegetated Ditches, Ponds, and Wetlands as BMPs for Mitigating the Water Quality Impact of Irrigated Agriculture in the San Joaquin Valley*, University of the Pacific, Stockton, CA

Graham, J., Hanlon, J.S., Stringfellow, W.T., (2008), *EERP Field Protocol Book*, University of the Pacific, Stockton, CA.

Borglin, S., W. Stringfellow, J. Hanlon. (2005), *Standard Operating Procedures for the Up-Stream Dissolved Oxygen TMDL Project*, LBNL/Pub-937.

Borglin, S., Burks, R., Hanlon, J., Graham, J., Spier, C., Stringfellow, W., and Dahlgren, R., (2008) Methods overview, quality assurance, and quality control, University of the Pacific, Stockton, CA

American Public Health Association, (2005), *Standard Methods for the Examination of Water and Wastewater*, American Public Health Association, Washington, D.C.

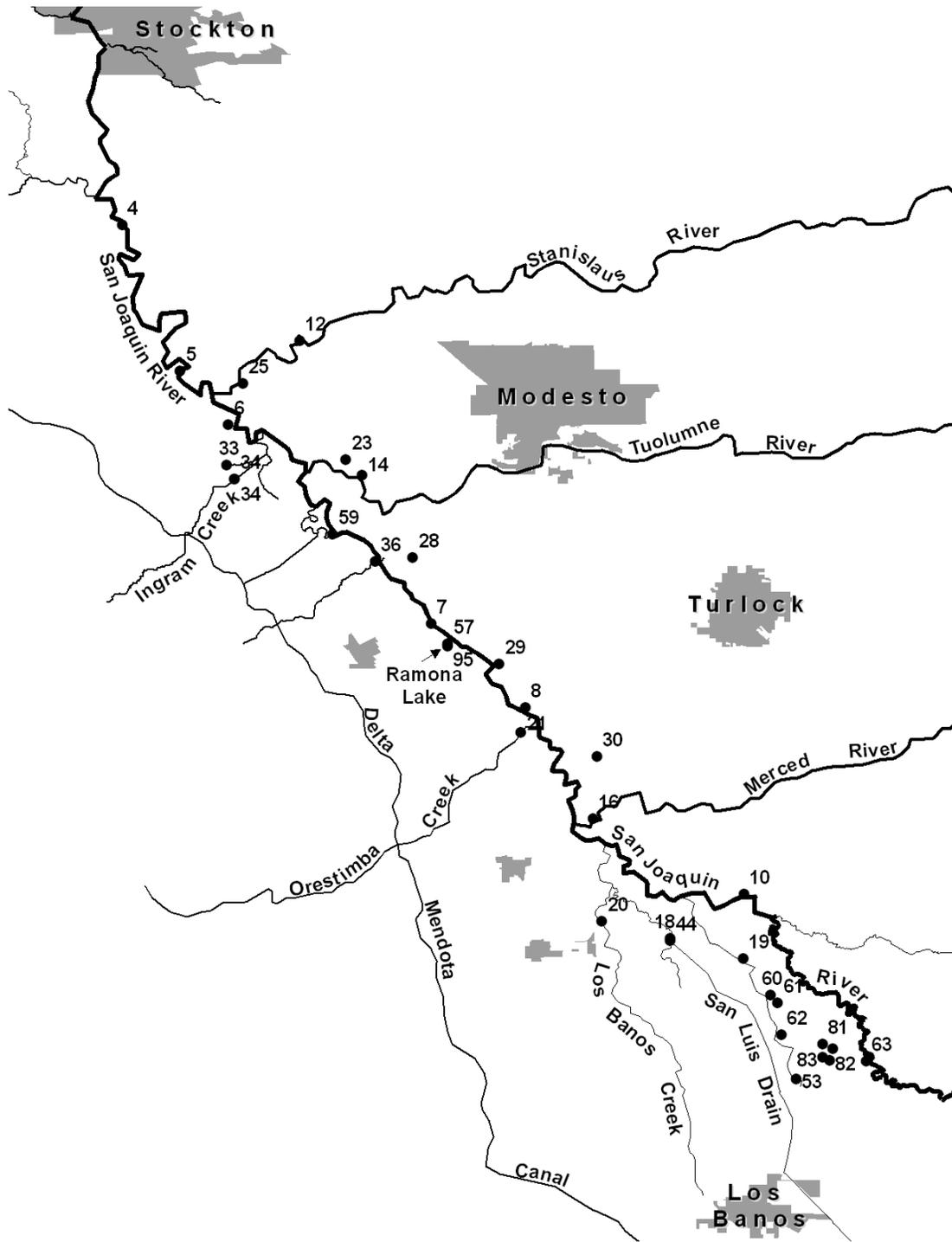
Borglin, S.E., Burks, R.D., Hanlon, J.S., Stringfellow, W.T. (2008) *EERP Lab Protocol Book*, University of the Pacific, Stockton, CA.

YSI Environmental Operations Manual, (2005), 6-Series Environmental Monitoring Systems, Yellow Springs, OH.

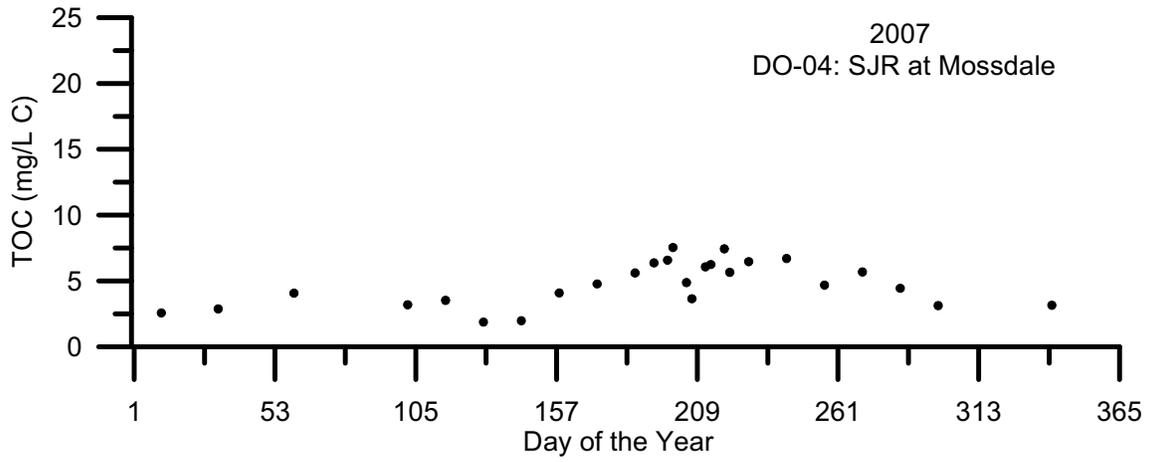
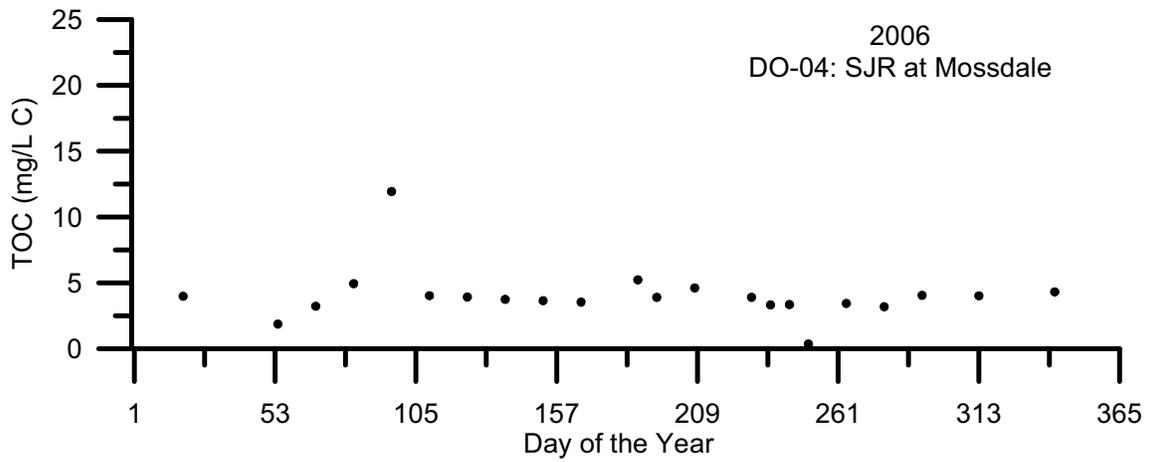
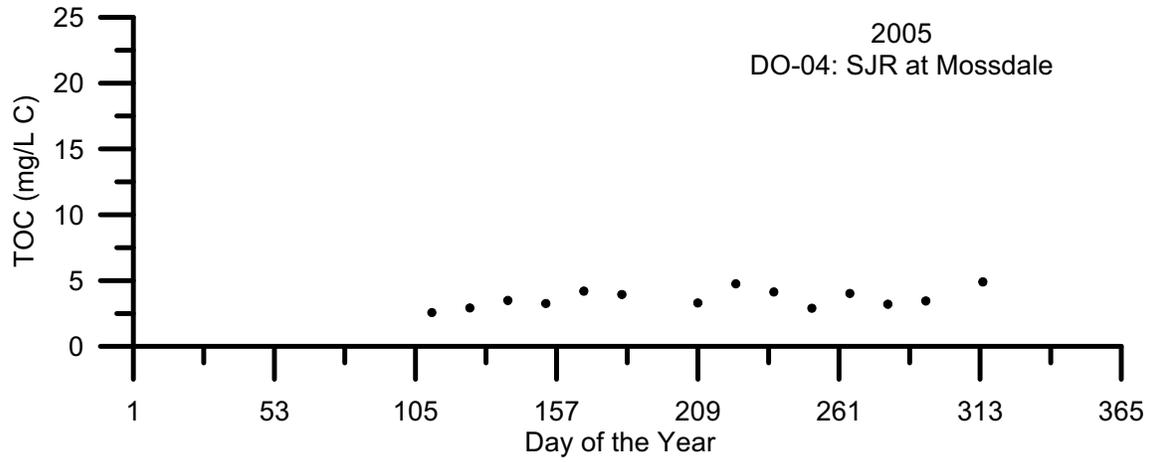
Table 1: EERP Sampling Site List

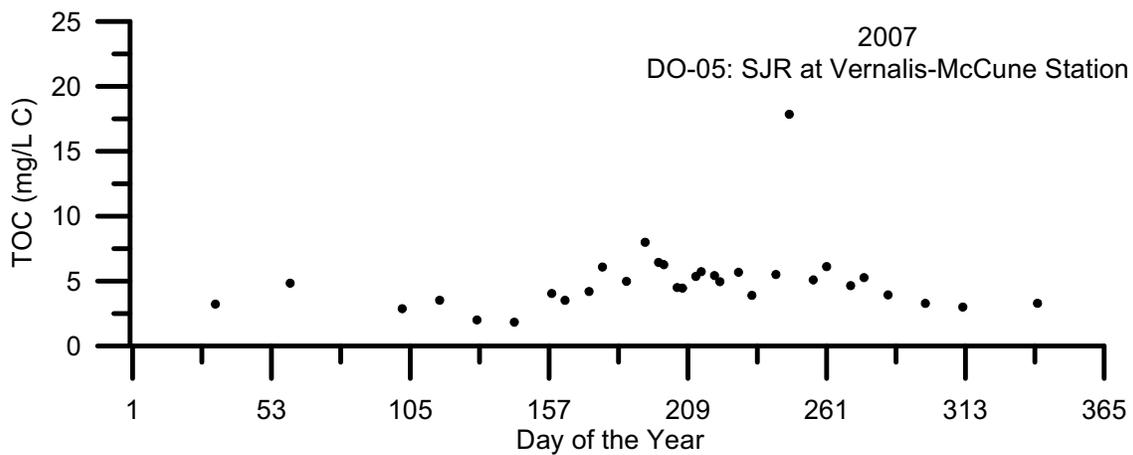
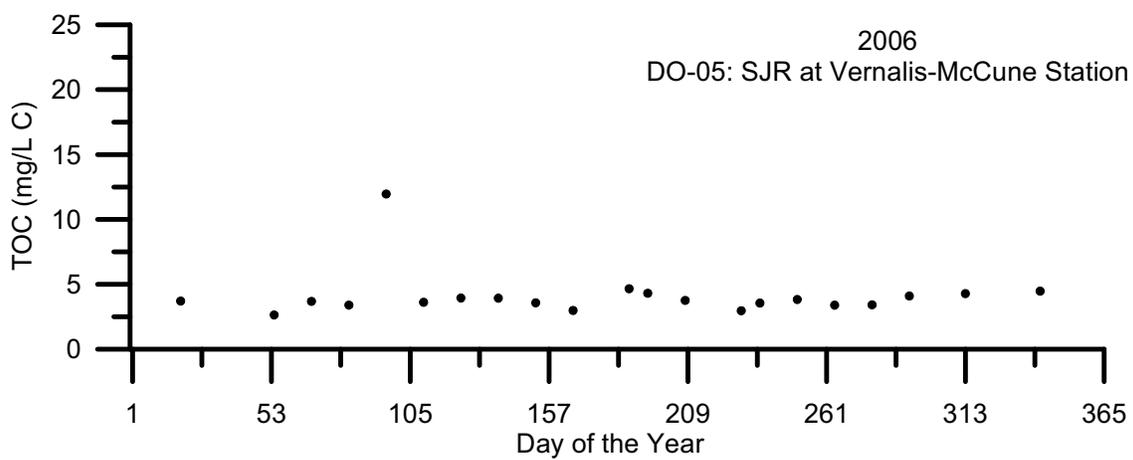
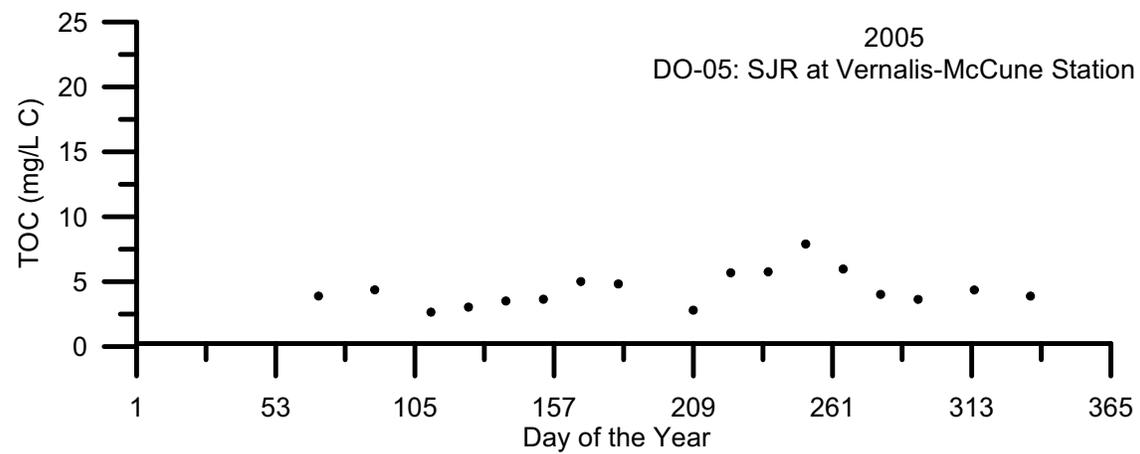
DO Number	Site Name	Type
4	SJR at Mossdale	Core sites
5	SJR at Vernalis-McCune Station (River Club)	Core sites, BMP
6	SJR at Maze	Core sites, BMP
7	SJR at Patterson	Core sites, BMP
8	SJR at Crows Landing	Core sites, BMP
10	SJR at Lander Avenue	Core sites
12	Stanislaus River at Caswell Park	Core sites
14	Tuolumne River at Shiloh Bridge	Core sites
16	Merced River at River Road	Core sites
18	Mud Slough near Gustine	Core sites, Wetland
19	Salt Slough at Lander Avenue	Core sites, Wetland
20	Los Banos Creek Flow Station	Core sites, Wetland
21	Orestimba Creek at River Road	Core sites, BMP
23	Modesto ID Lateral 5 to Tuolumne	Core sites
25	Modesto ID Main Drain to Stan. R. via Miller Lake	Core sites
28	Turlock ID Westport Drain Flow station	Core sites
29	Turlock ID Harding Drain	Core sites
30	Turlock ID Lateral 6 & 7 at Levee	Core sites
33	Hospital Creek	Intermittent, BMP
34	Ingram Creek	Core sites, BMP
36	Del Puerto Creek Flow Station	Core sites, BMP
44	San Luis Drain End	Core sites
53	Salt Slough at Wolfsen Road	Wetland
57	Ramona Lake Drain	Core sites, BMP
59	SJR Laird Park	Core sites
60	Moffit 1 South	Wetland
61	Deadman's Slough	Wetland
62	Mallard Slough	Wetland
63	Inlet C Canal	Wetland
80	South Marsh-1-Inlet	Wetland
81	South Marsh-1-Outlet	Wetland
82	South Marsh-3-Inlet	Wetland
83	South Marsh-3-Outlet	Wetland
95	Ramona drain at Ramona Lake	BMP, Intermittent

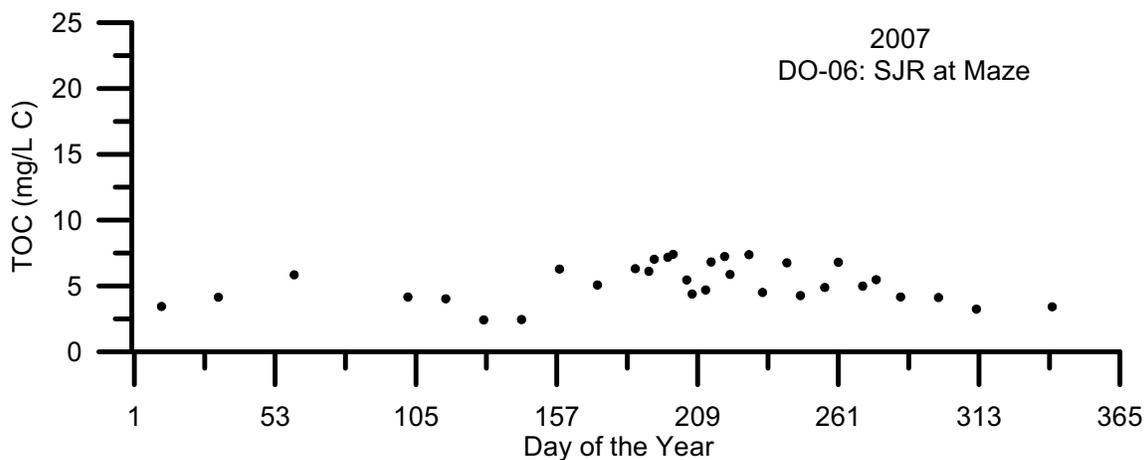
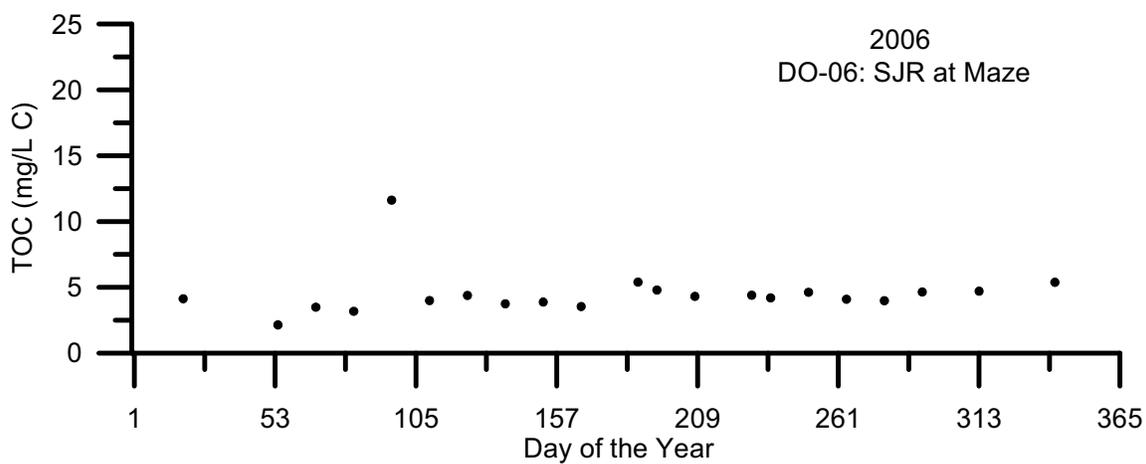
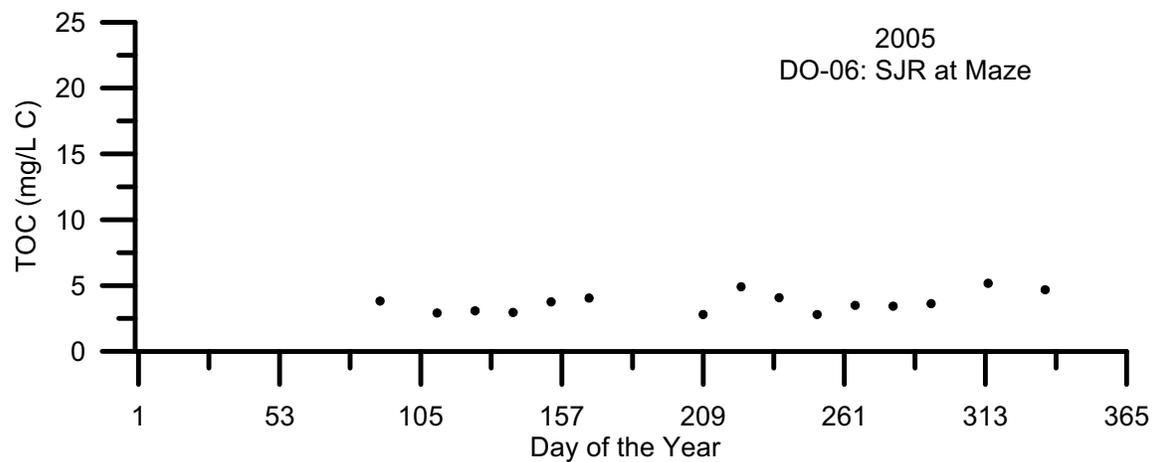
Figure 1: EERP Sampling Site Map of SJR Watershed and Tributaries

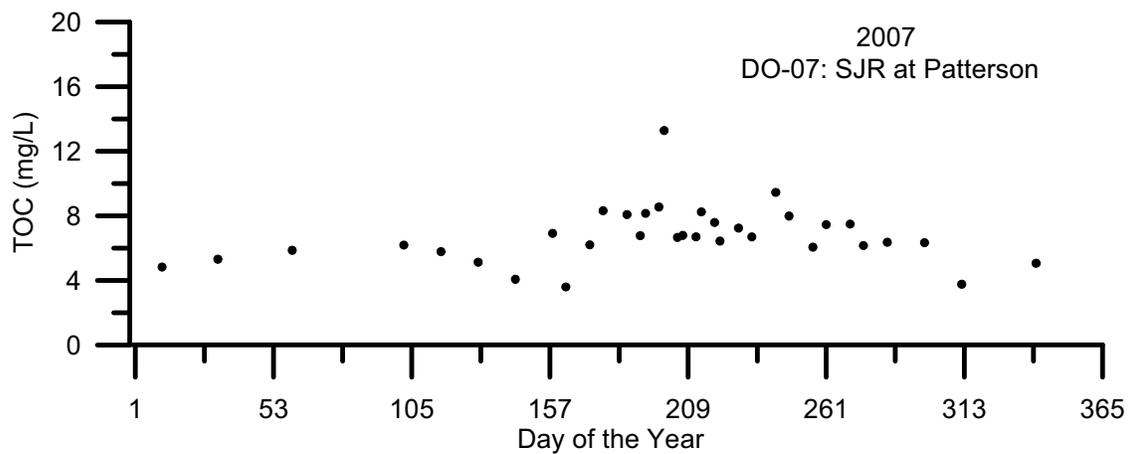
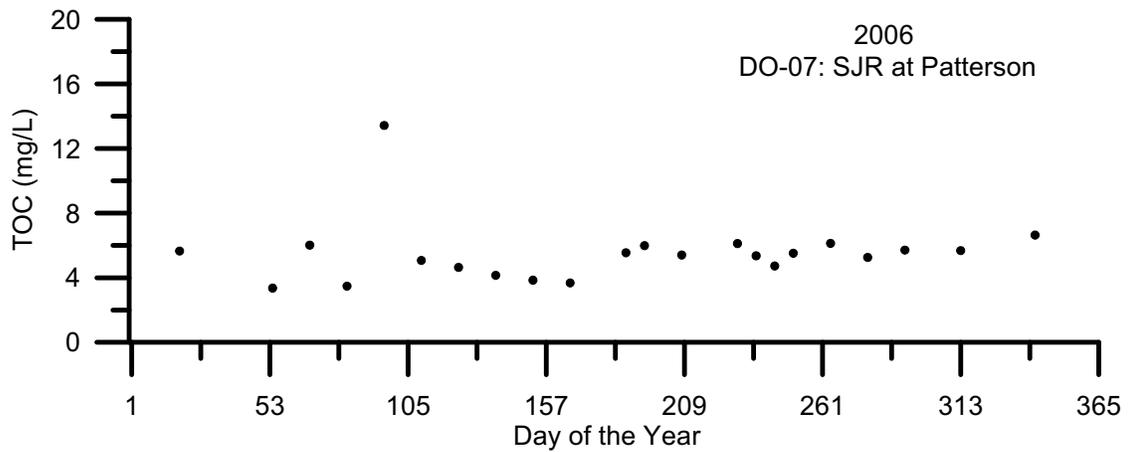
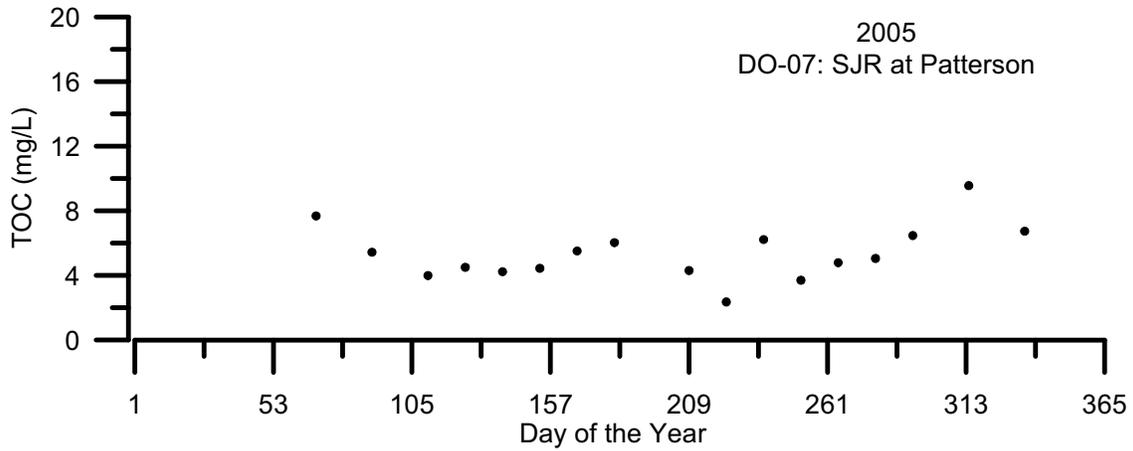


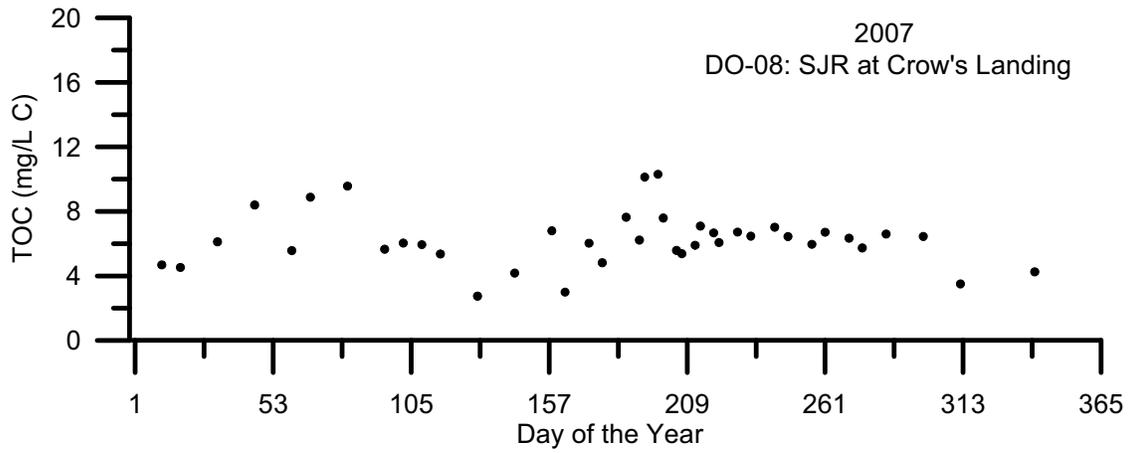
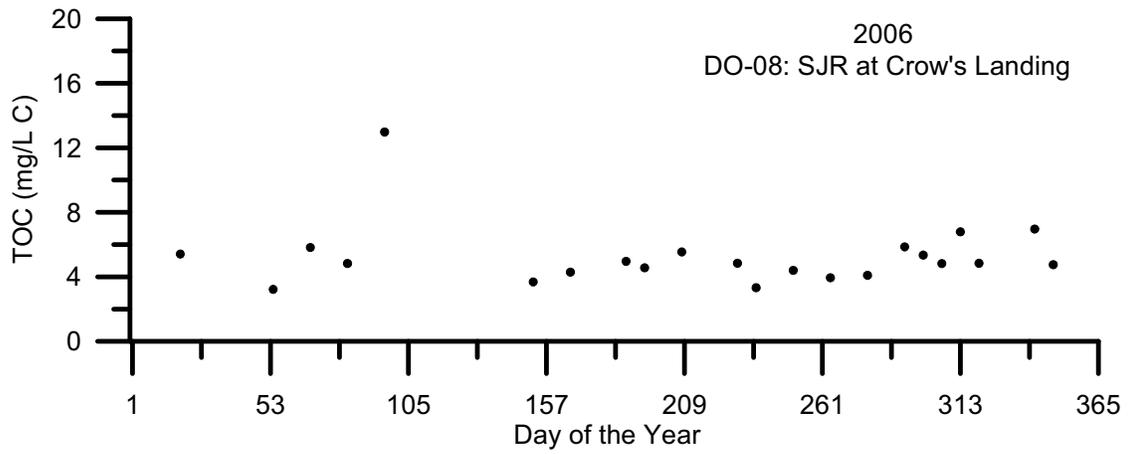
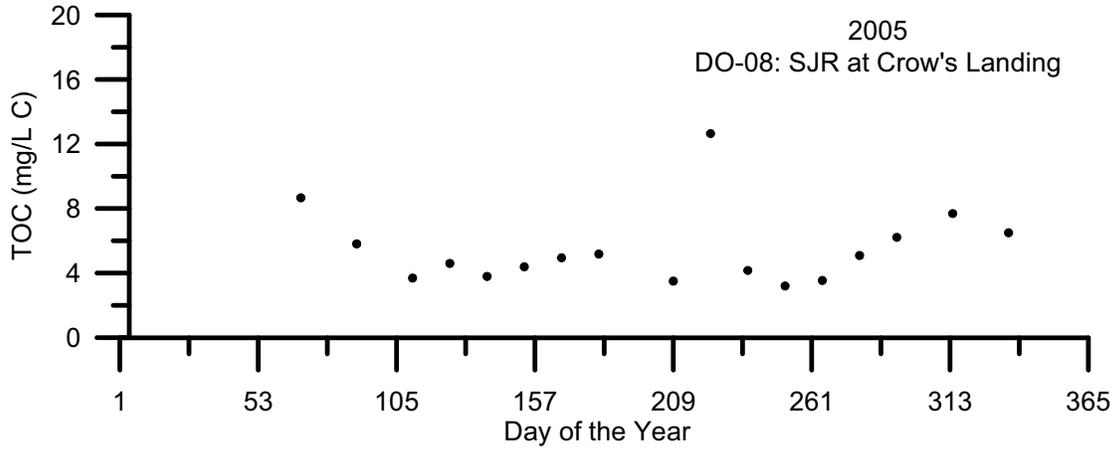
Figures 2 -101: Temporal Plots of Total Organic Carbon Concentrations By Site ID

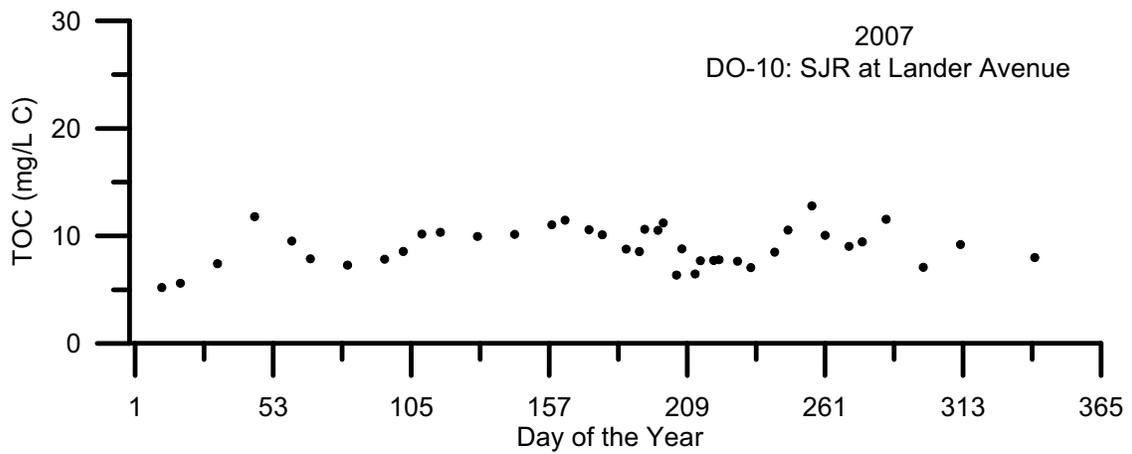
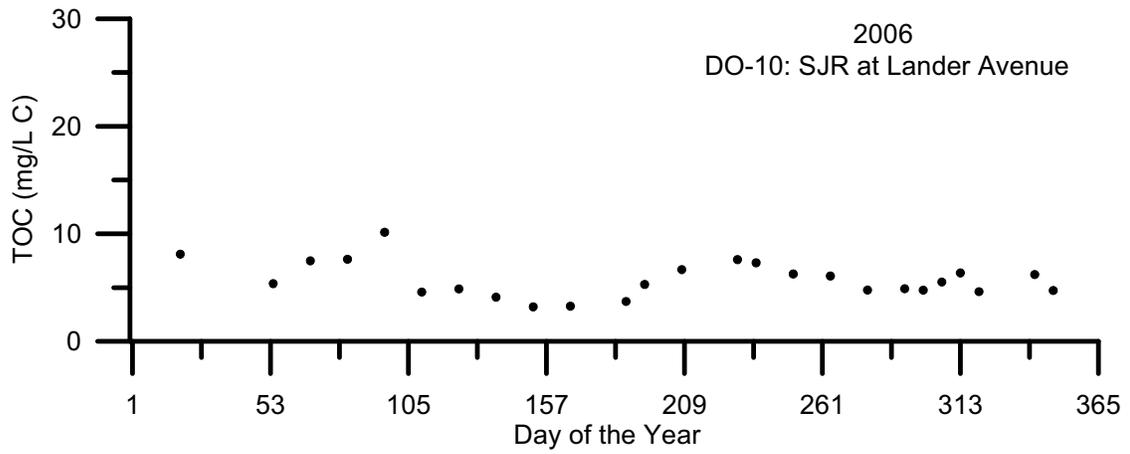
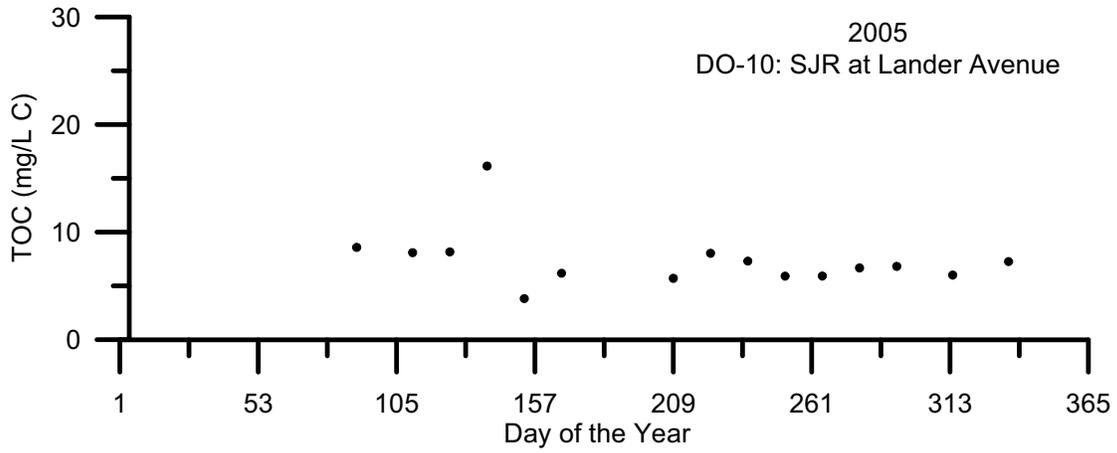


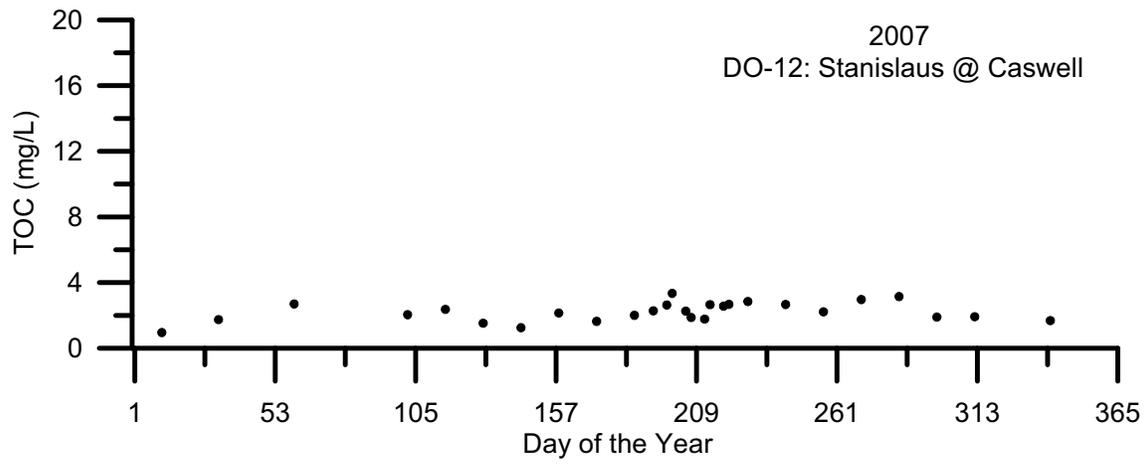
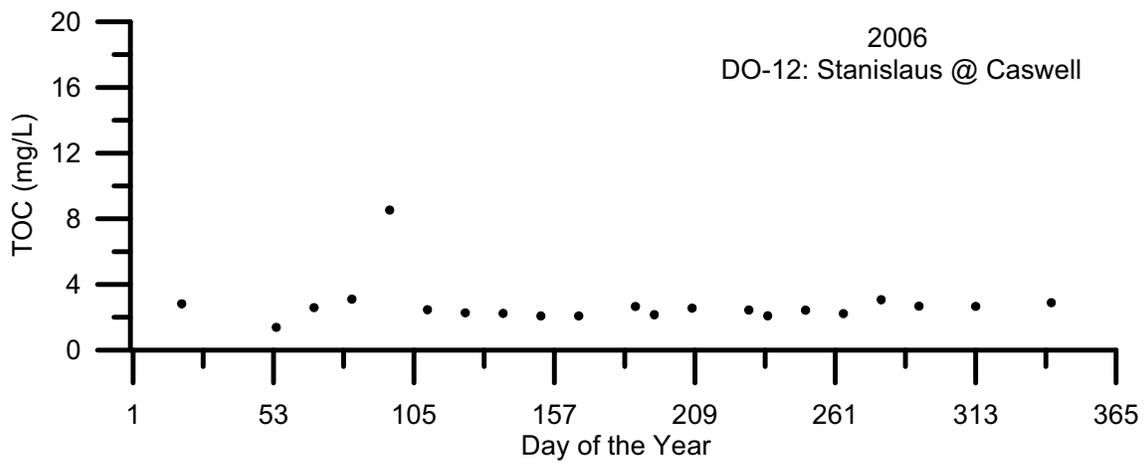
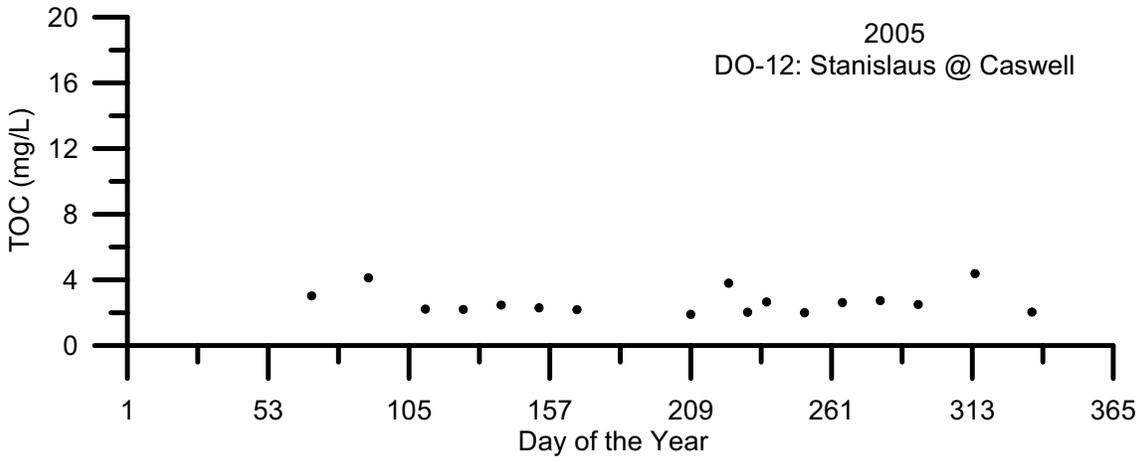


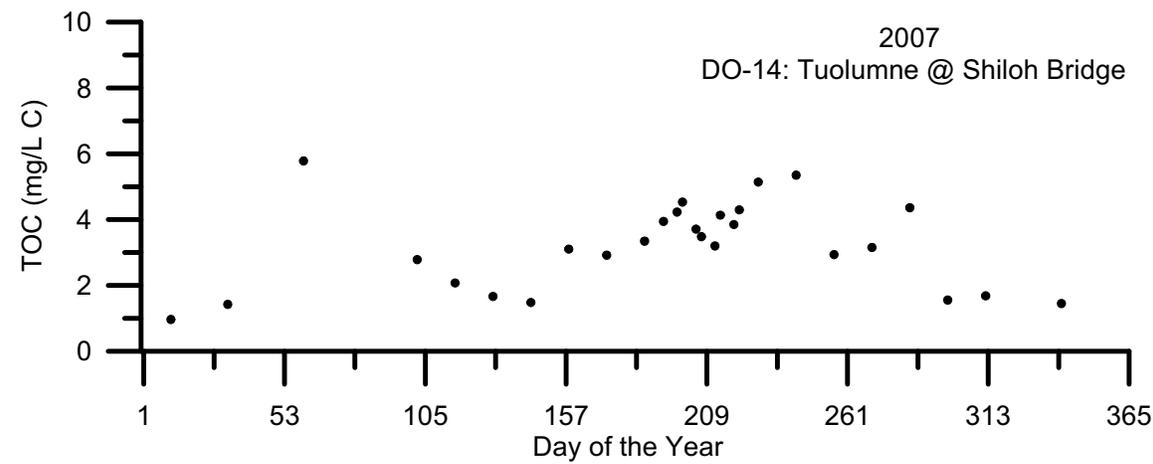
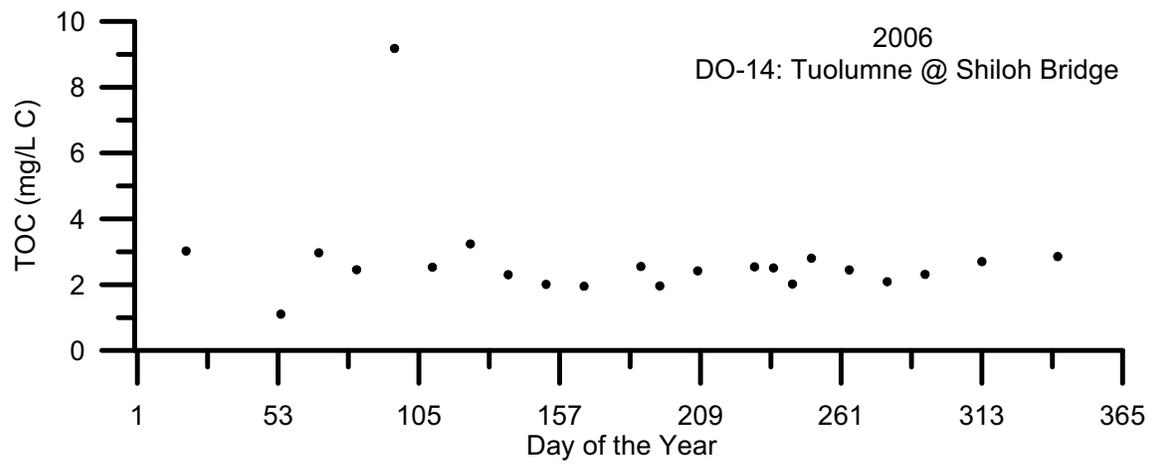
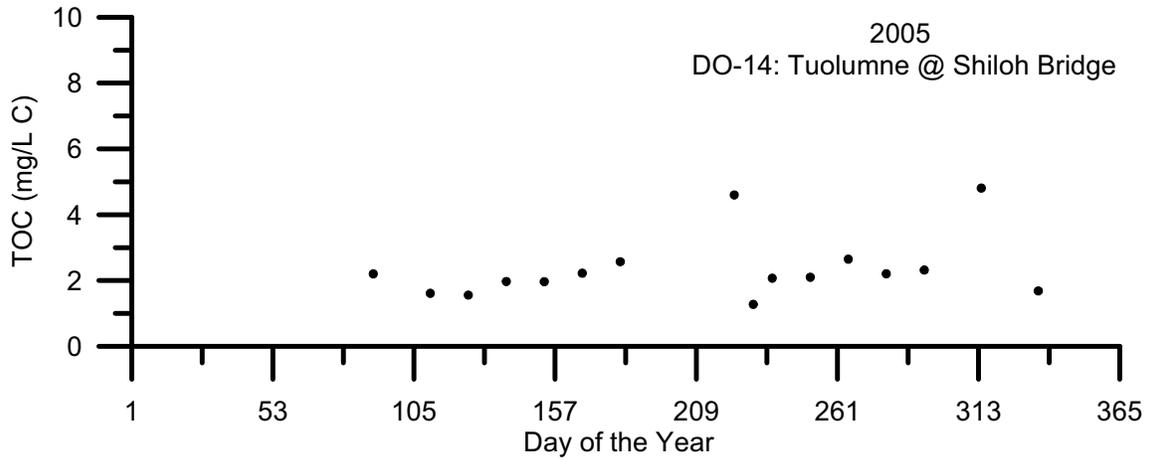


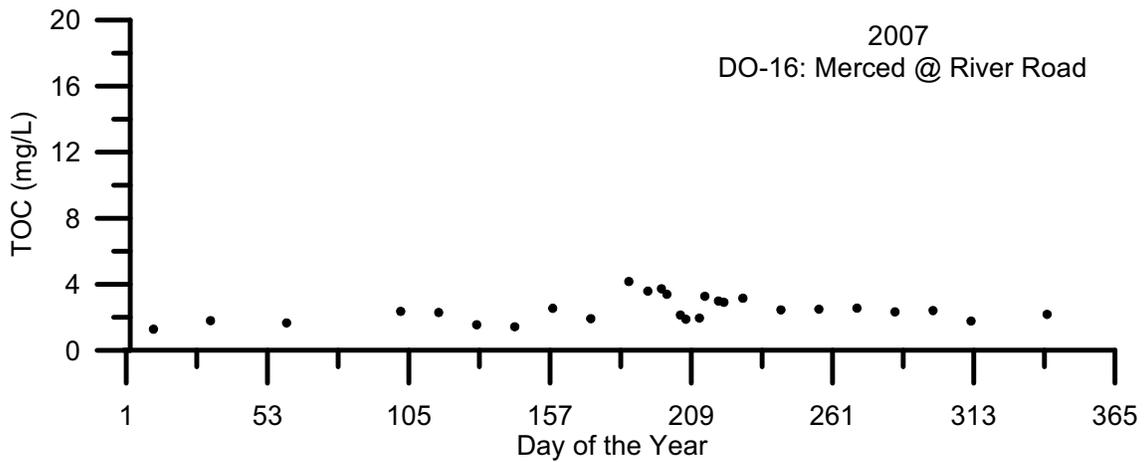
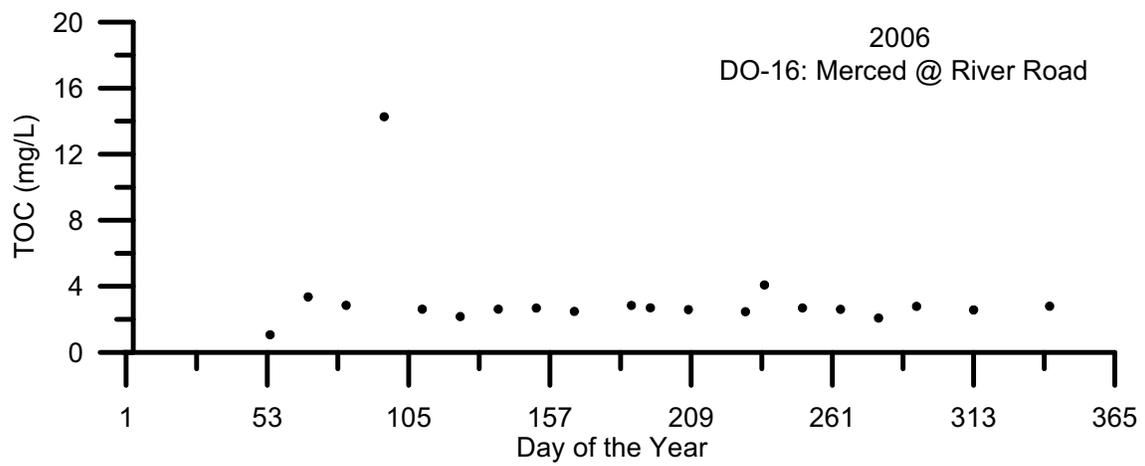
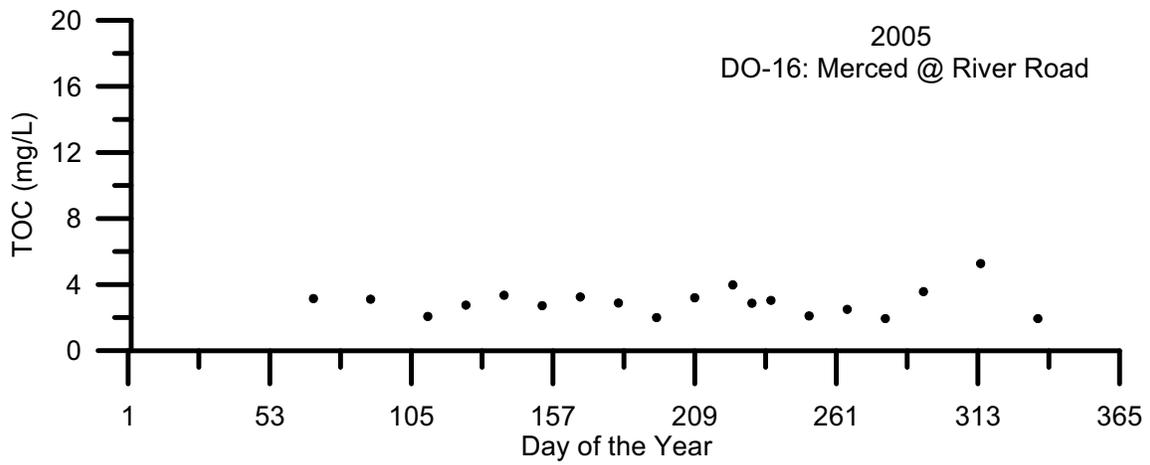


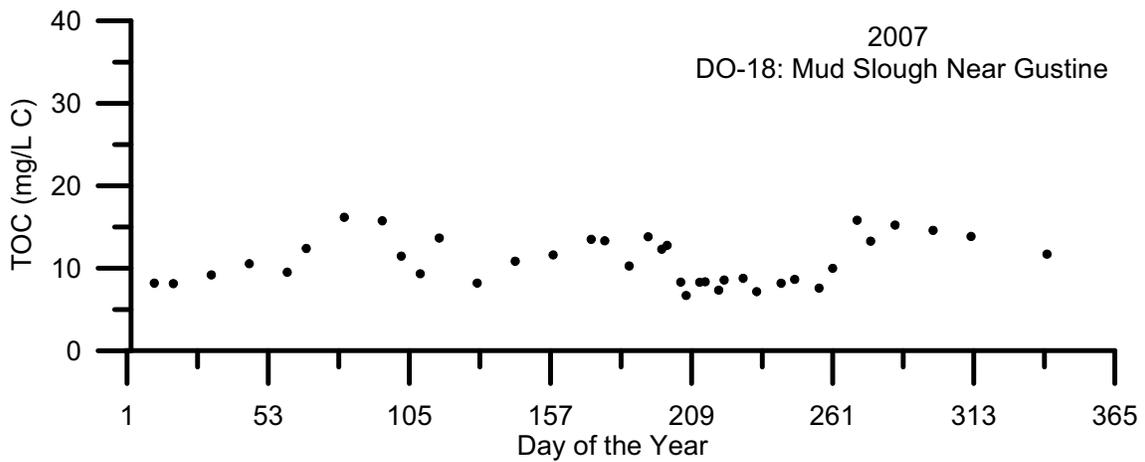
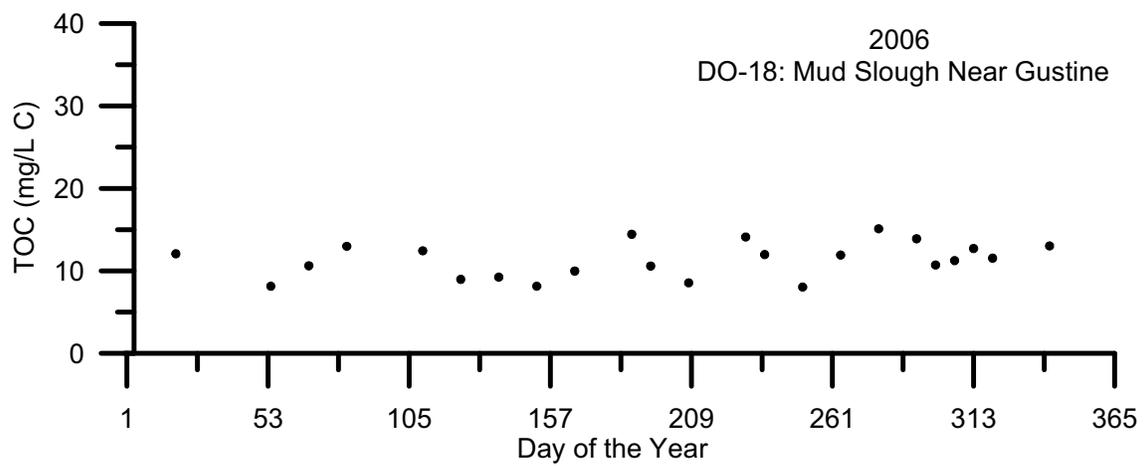
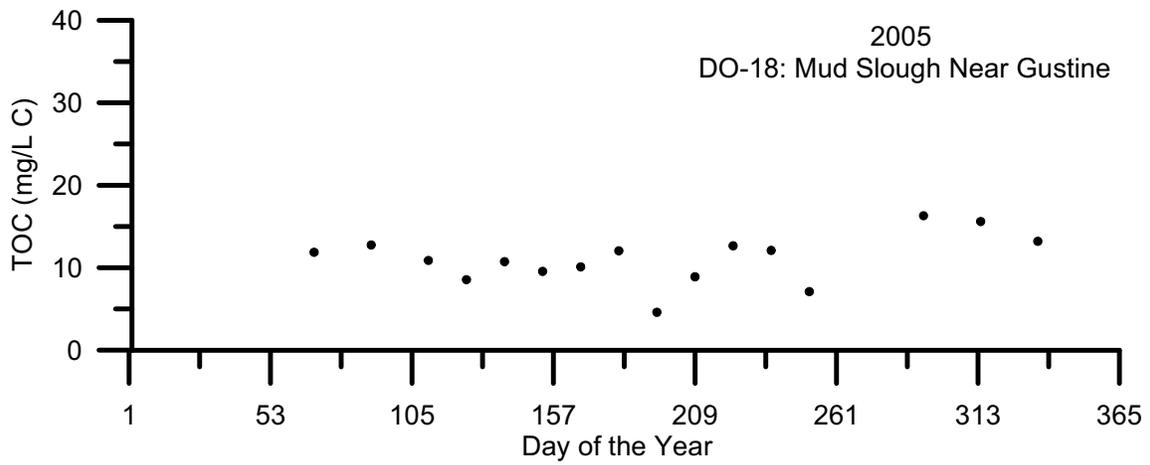


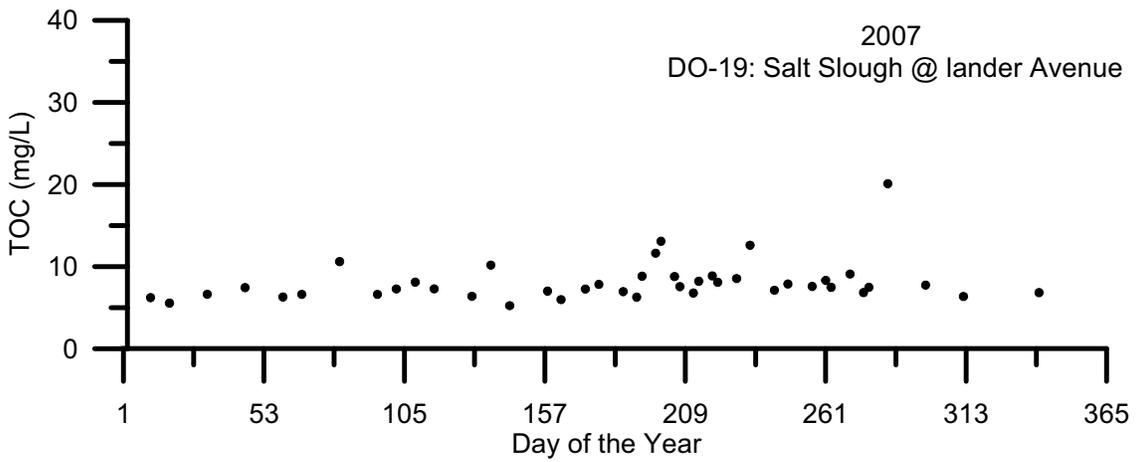
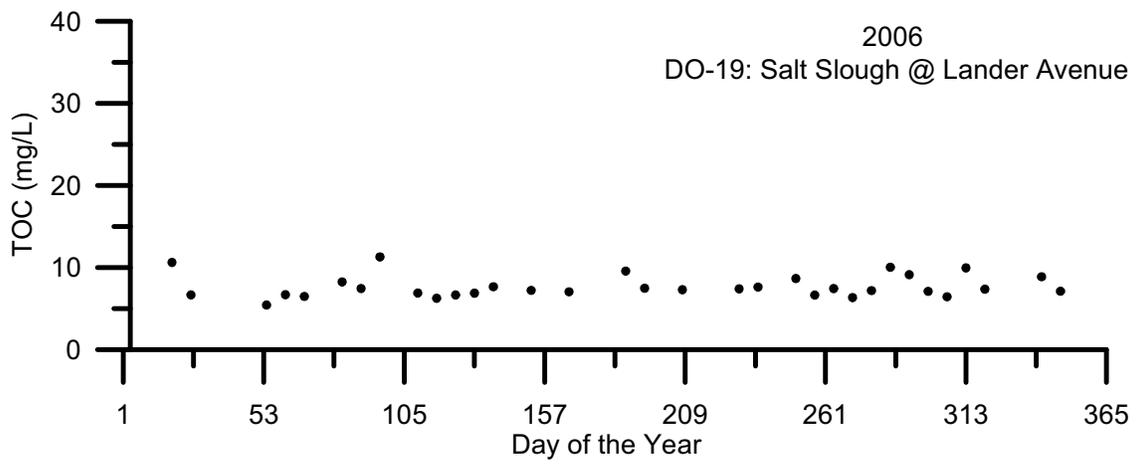
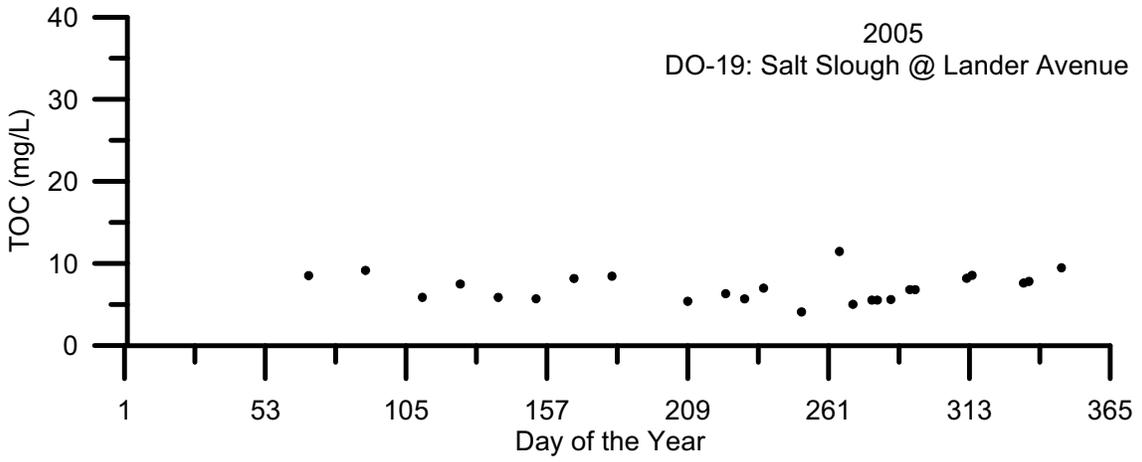


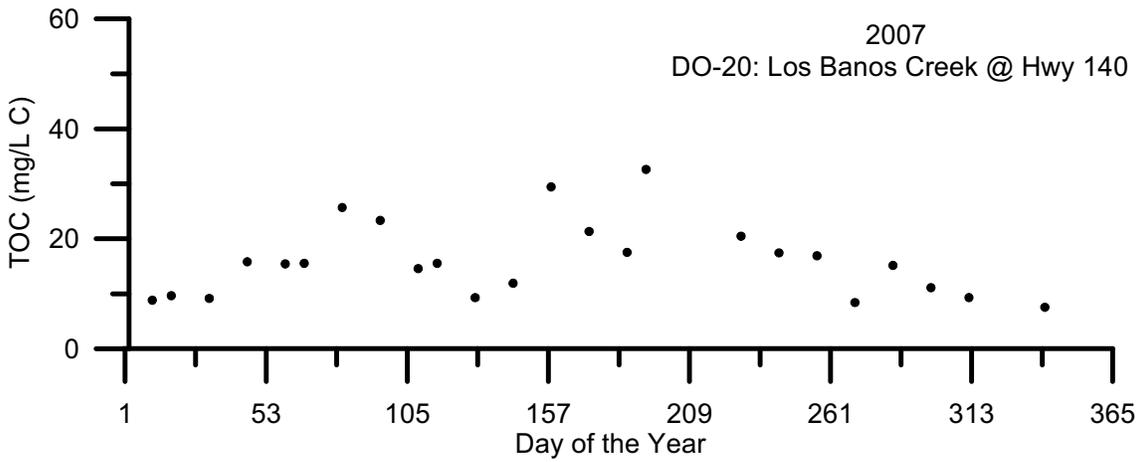
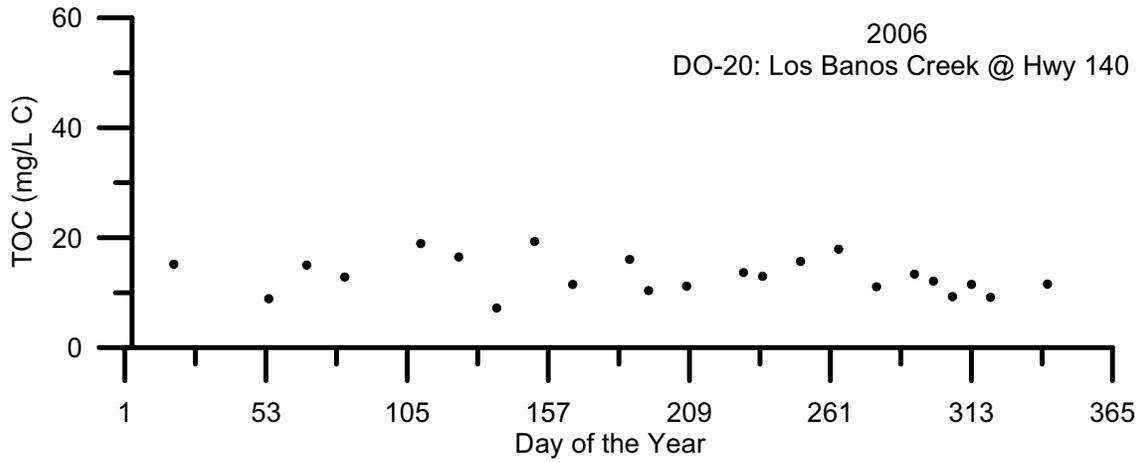
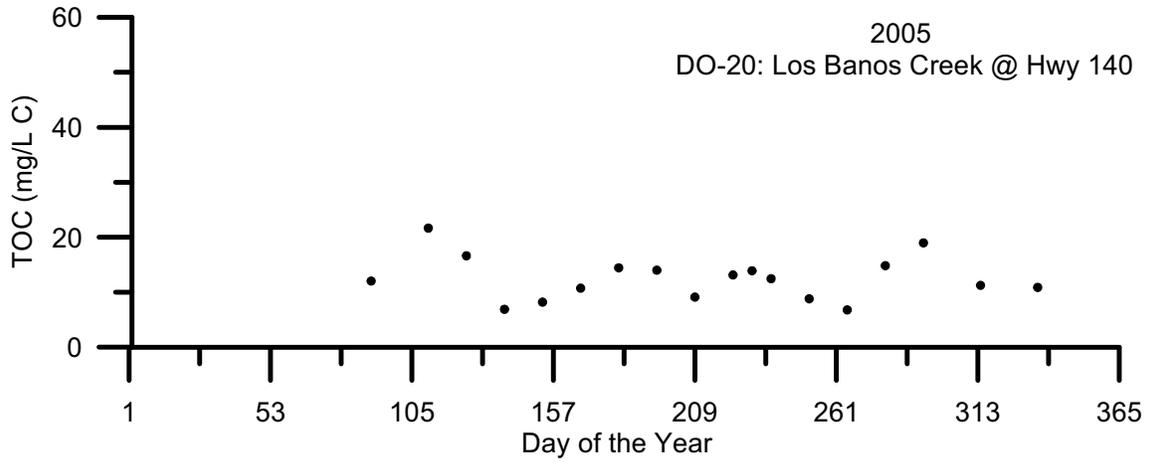


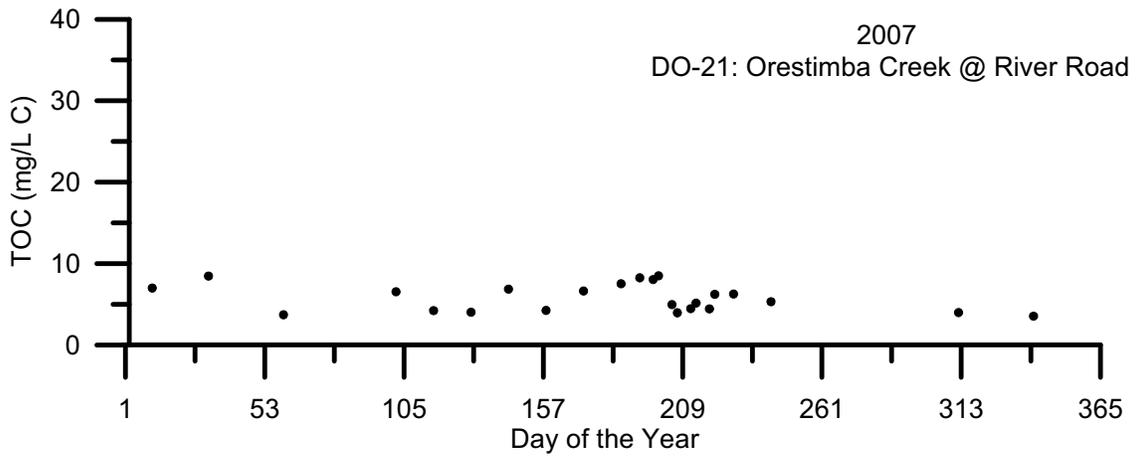
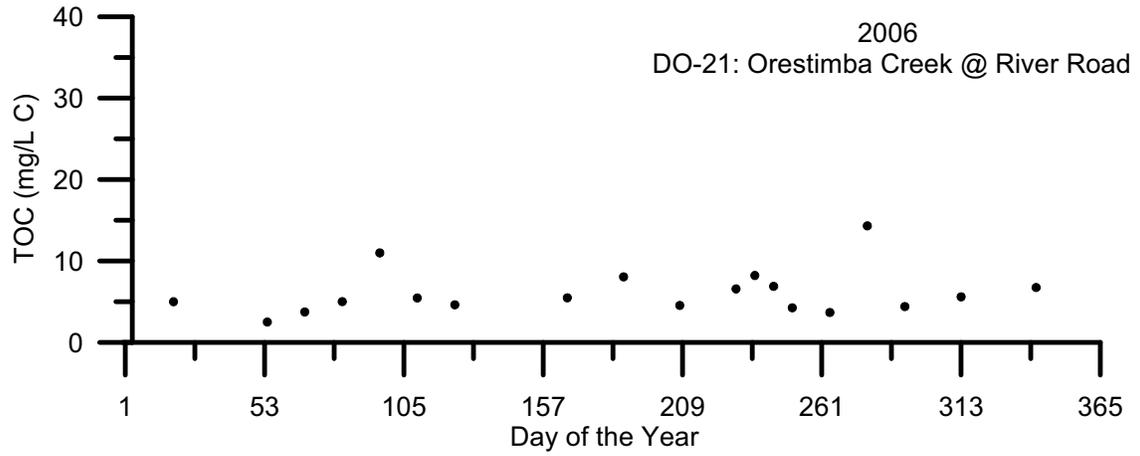
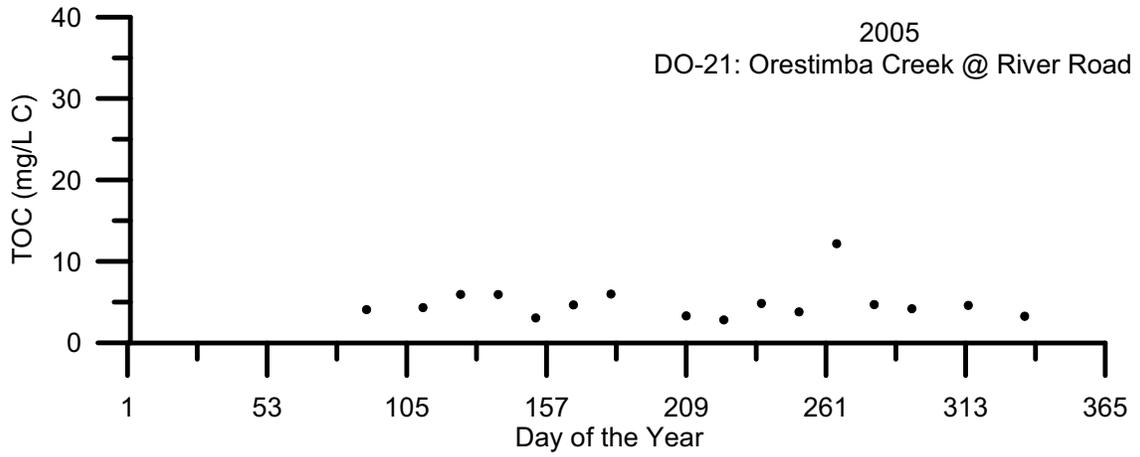


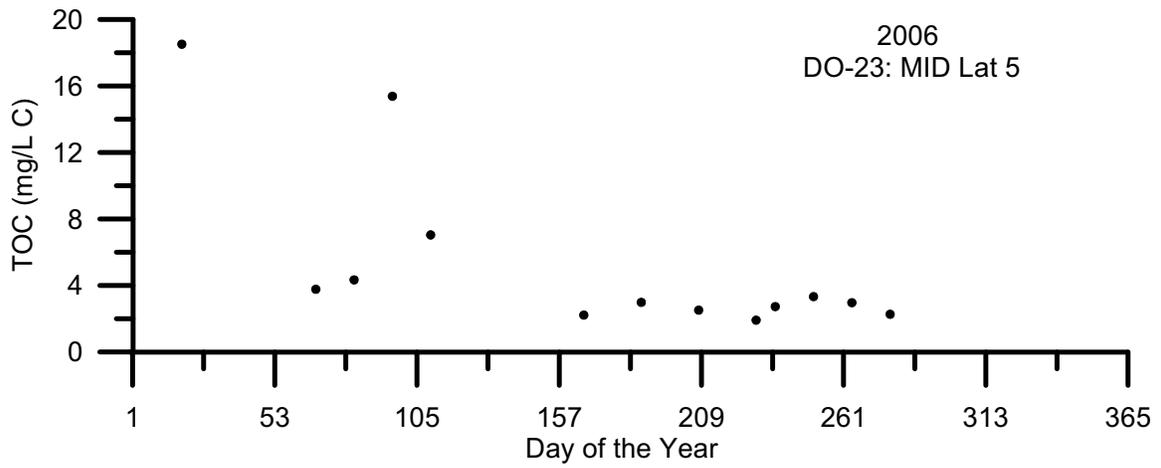
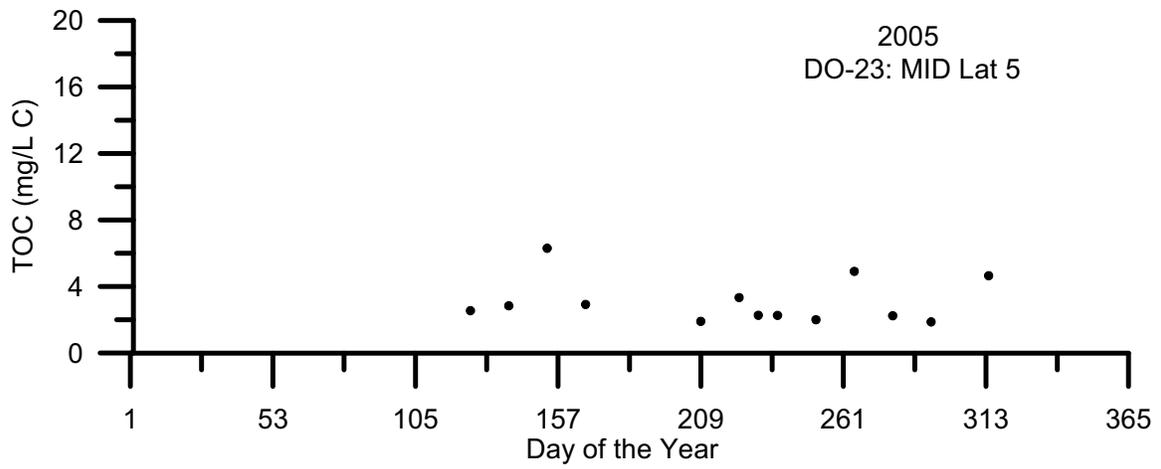


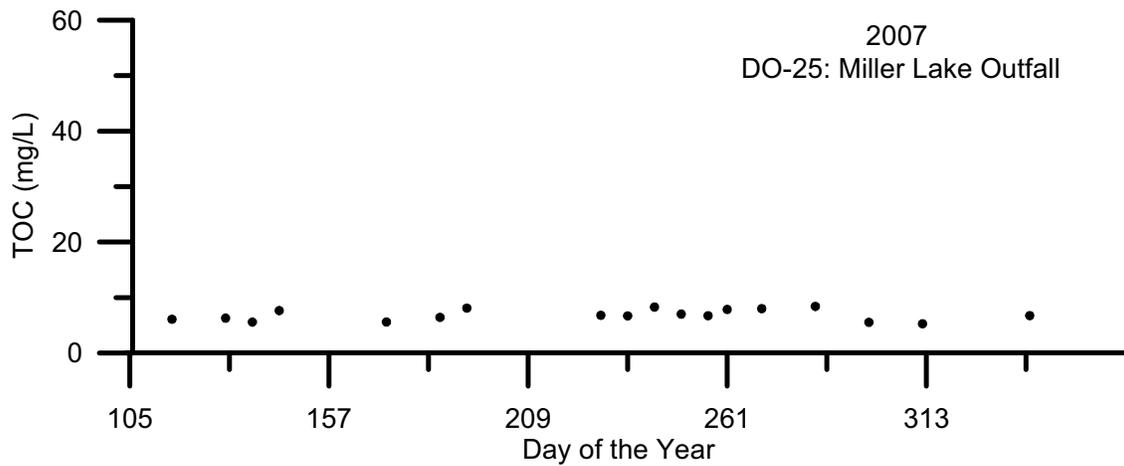
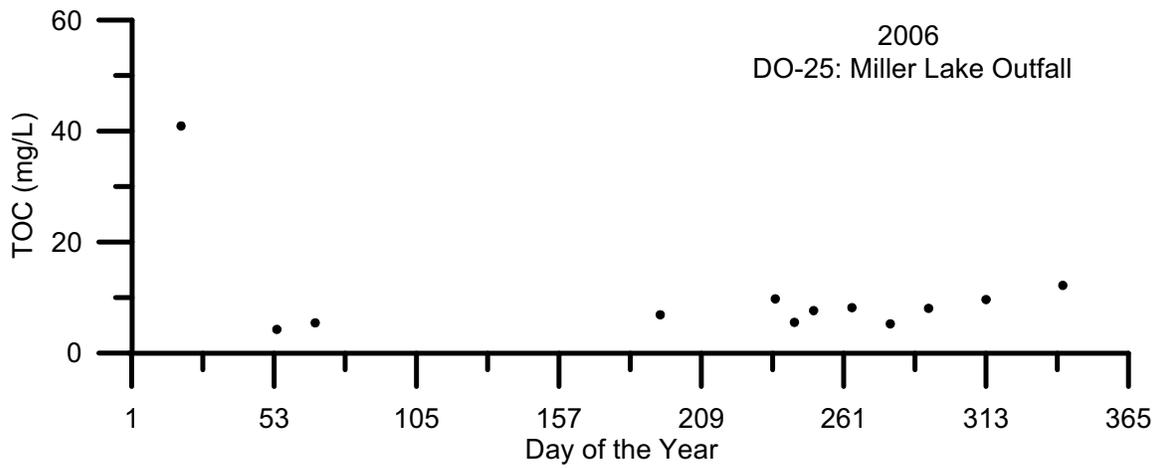
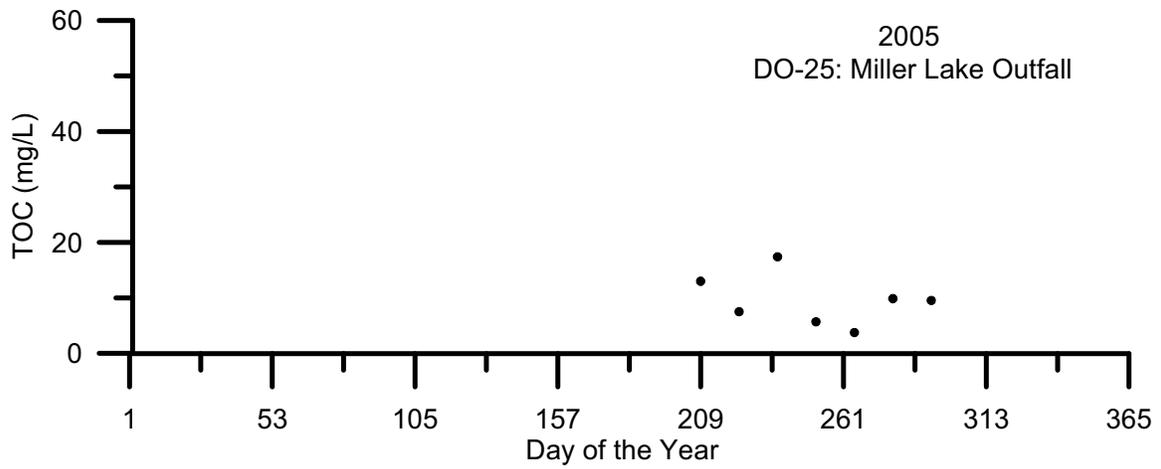


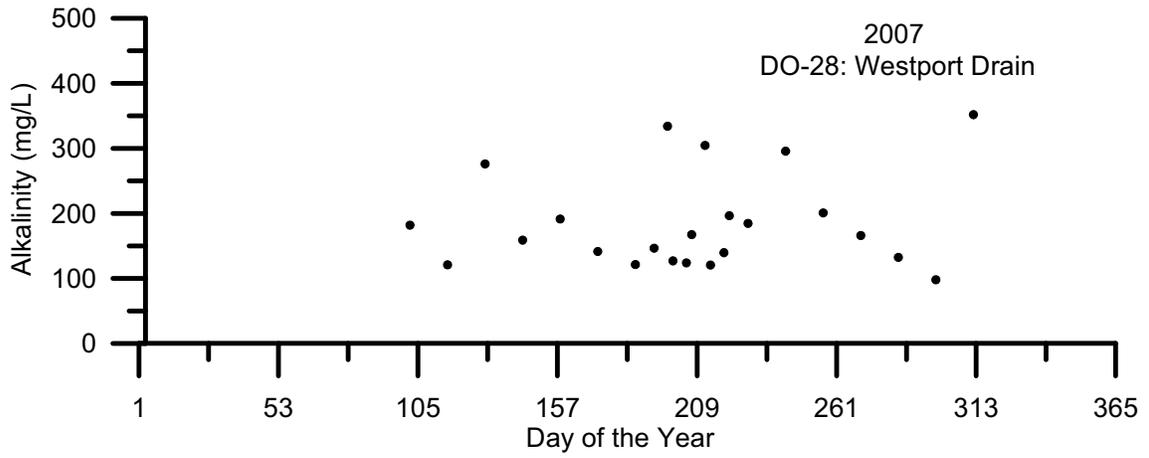
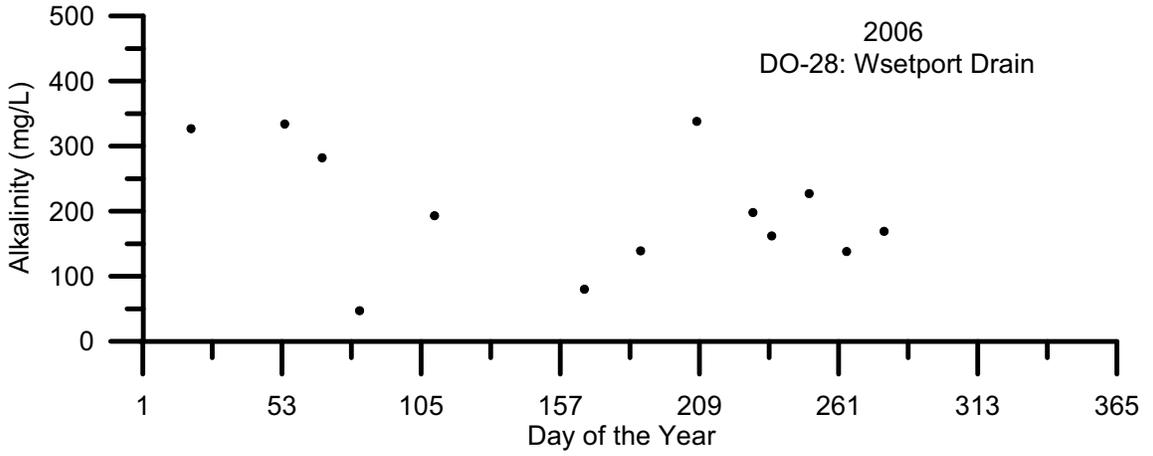
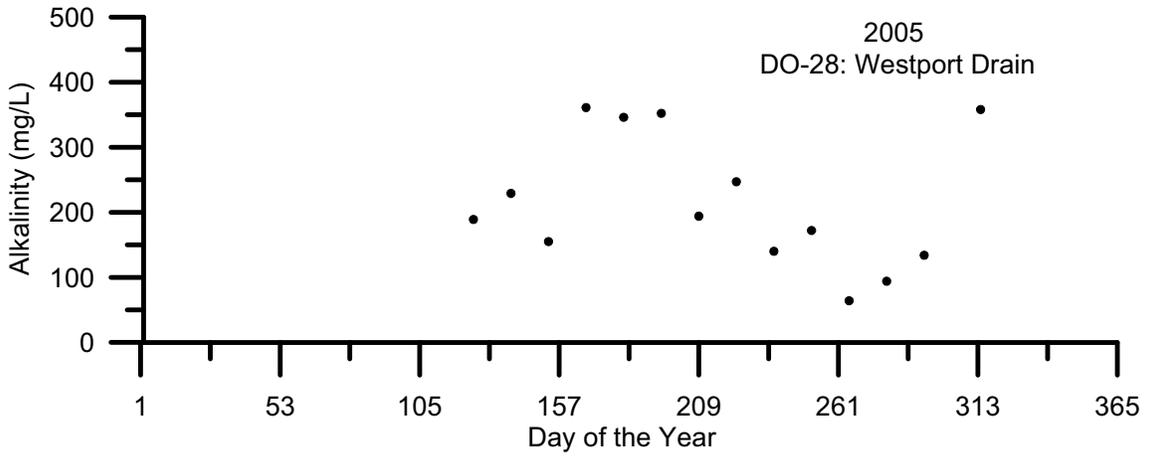


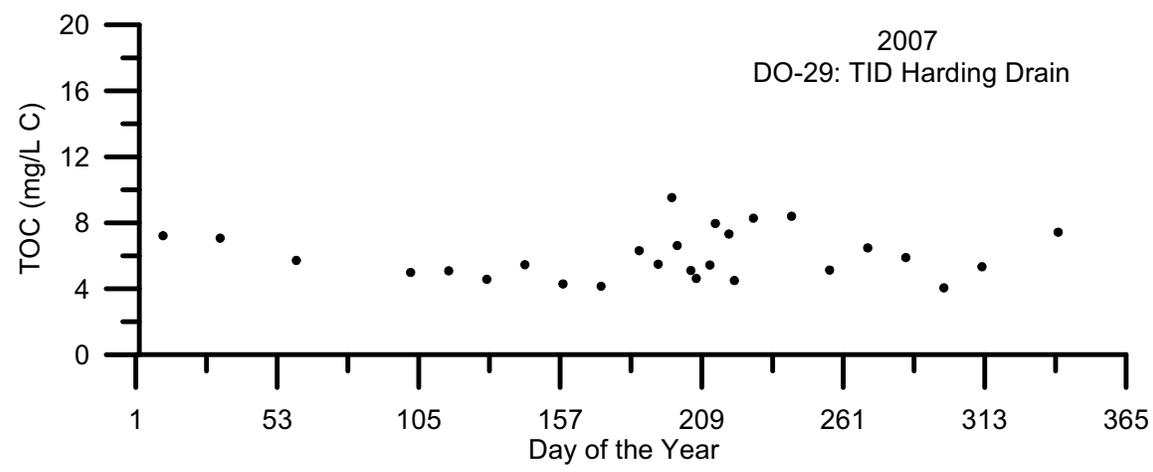
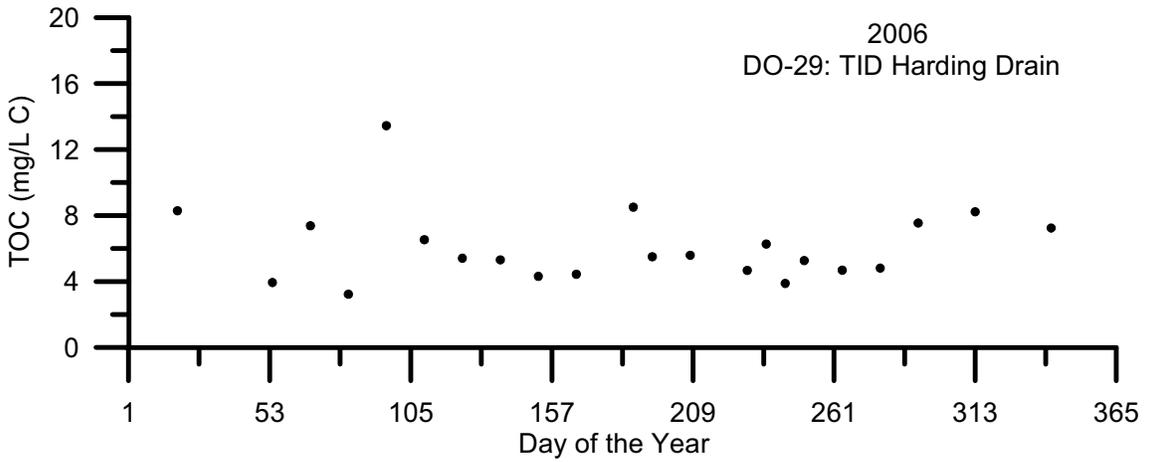
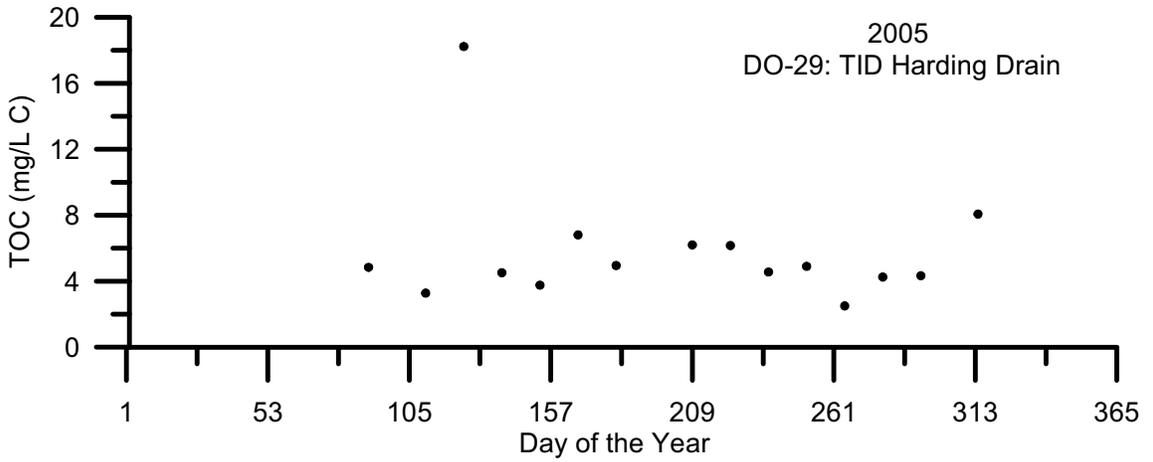


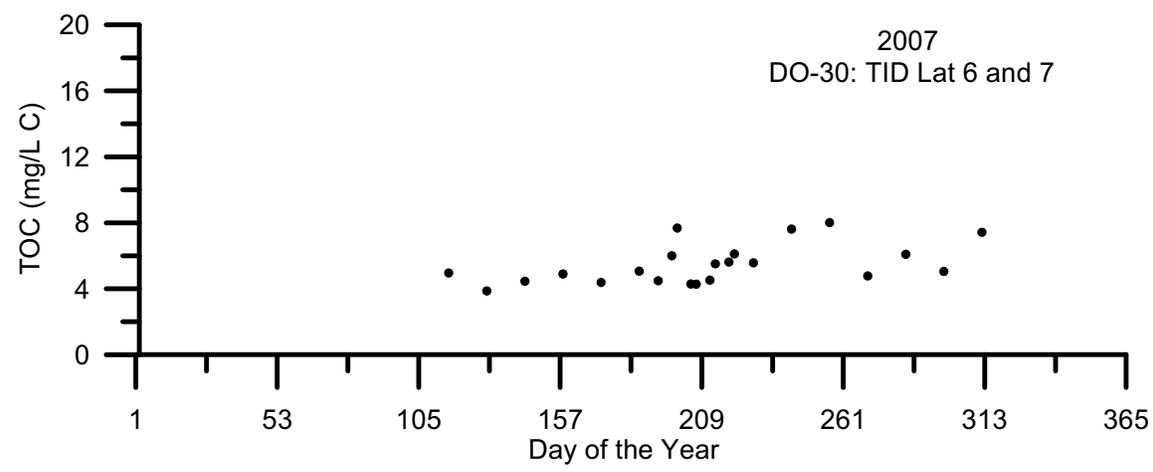
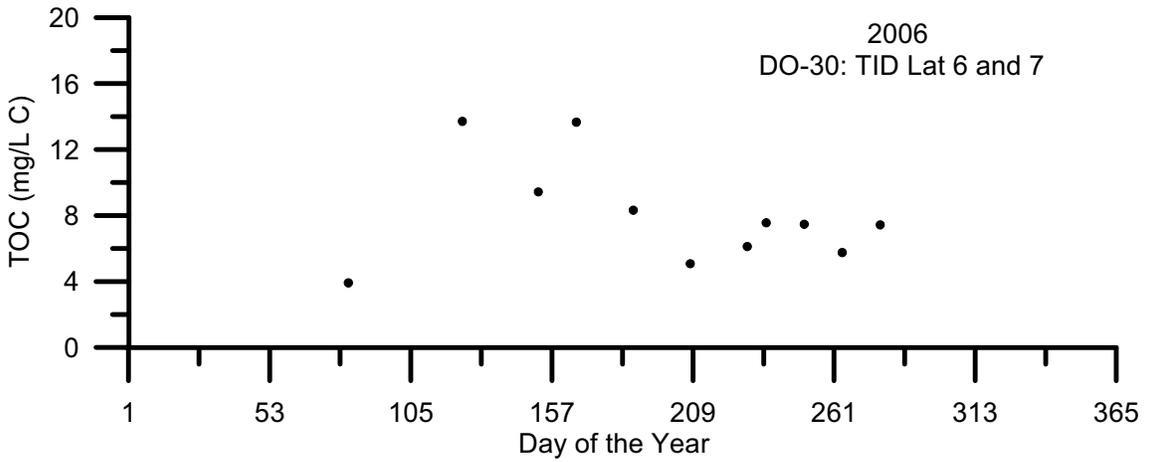
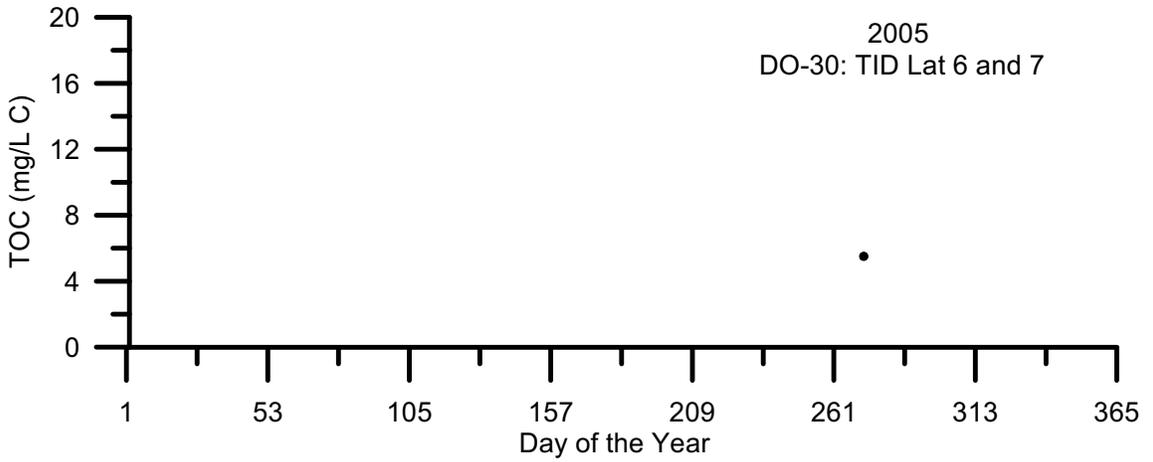


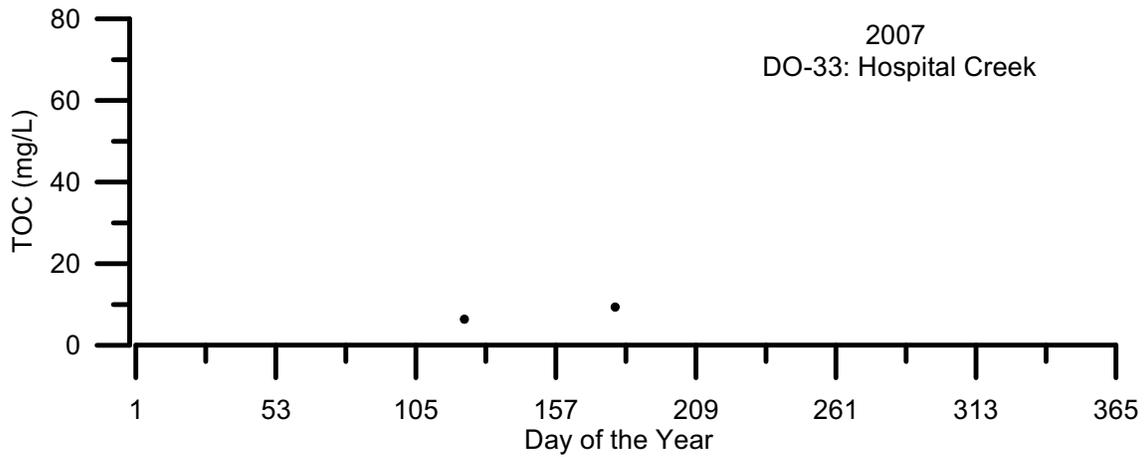
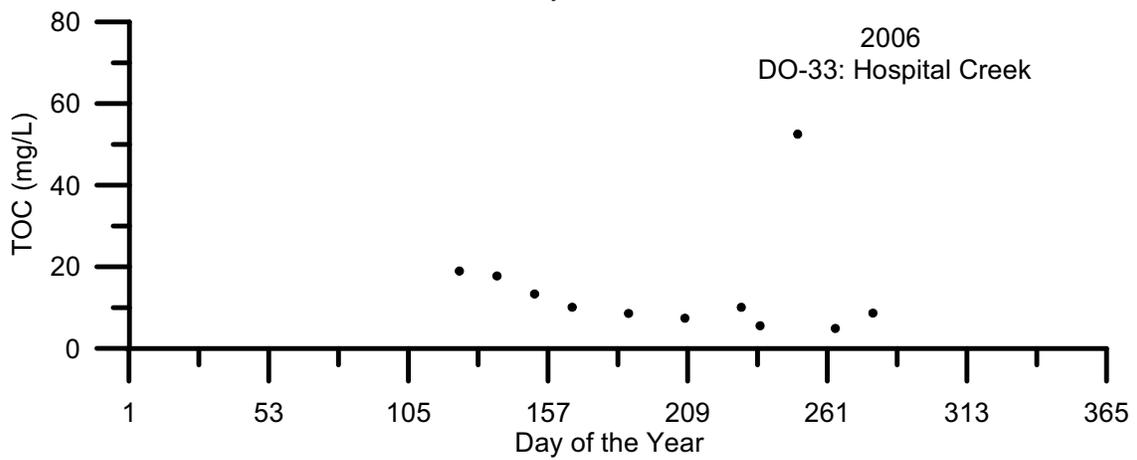
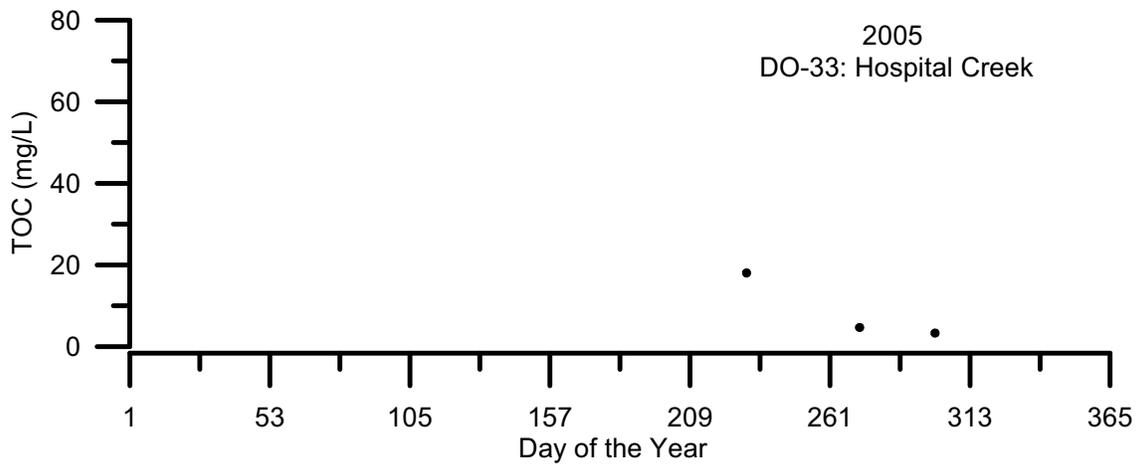


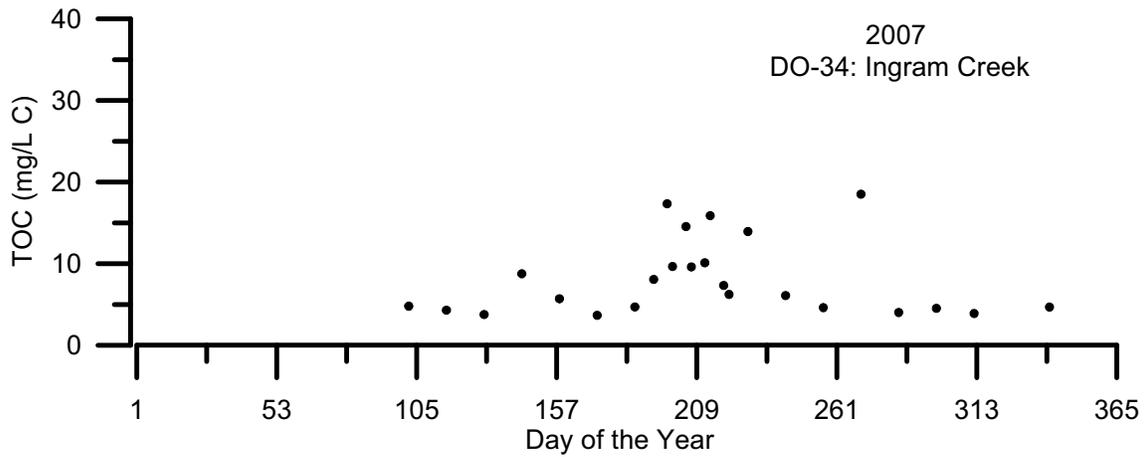
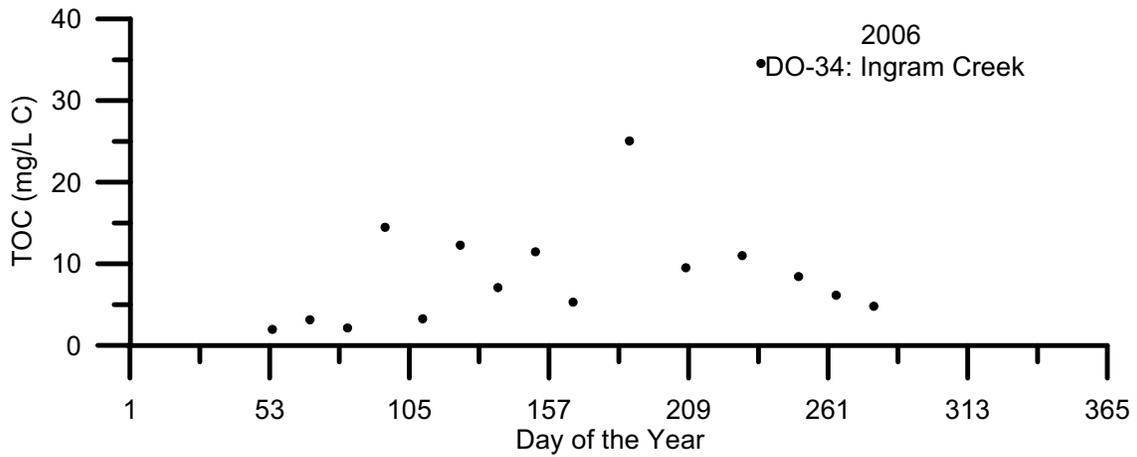
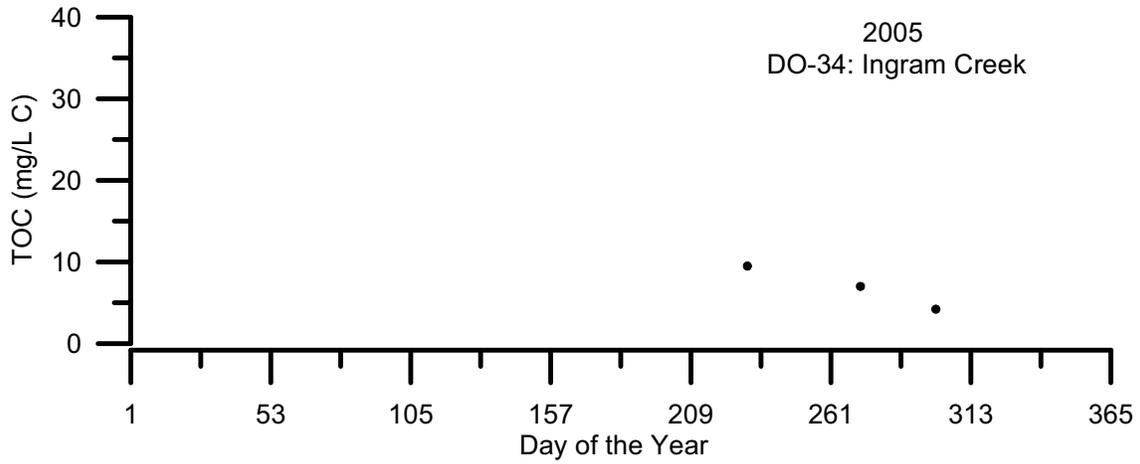


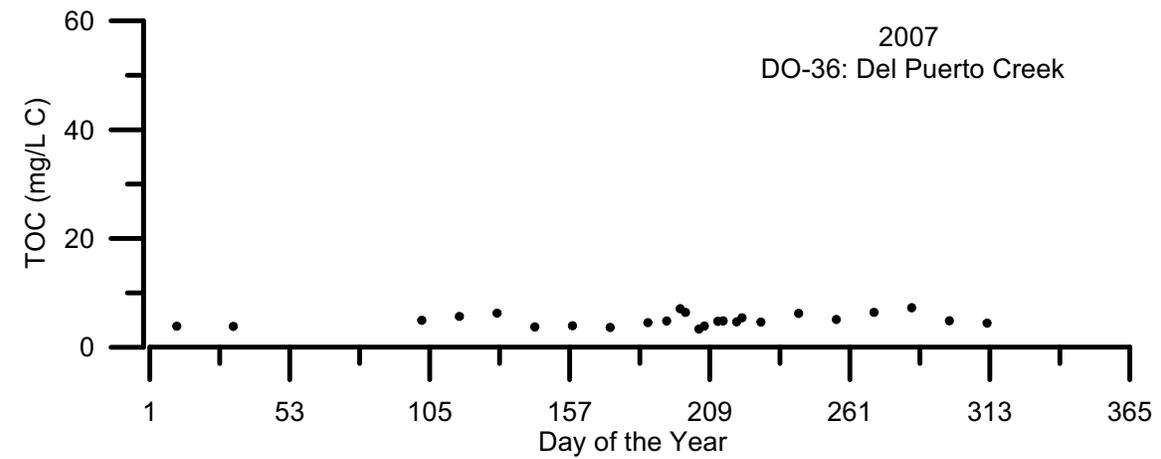
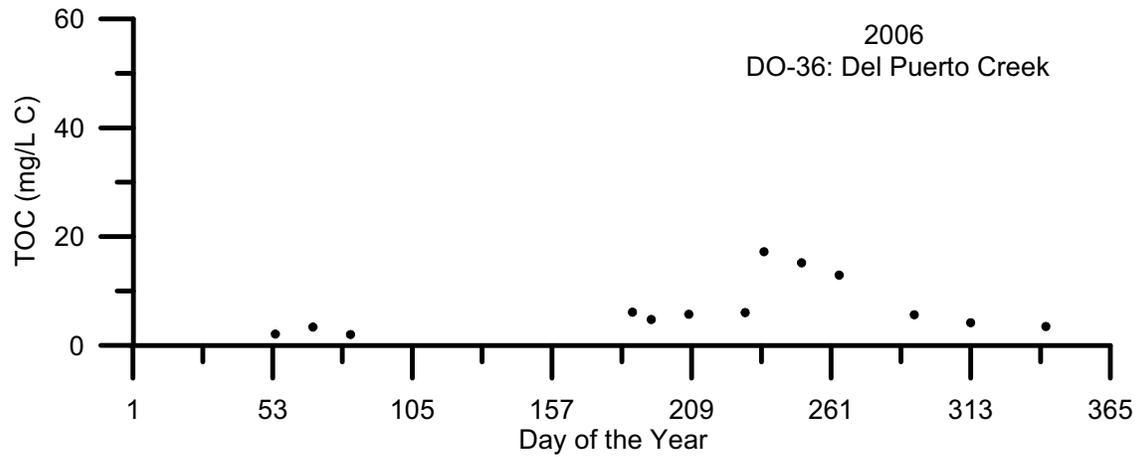
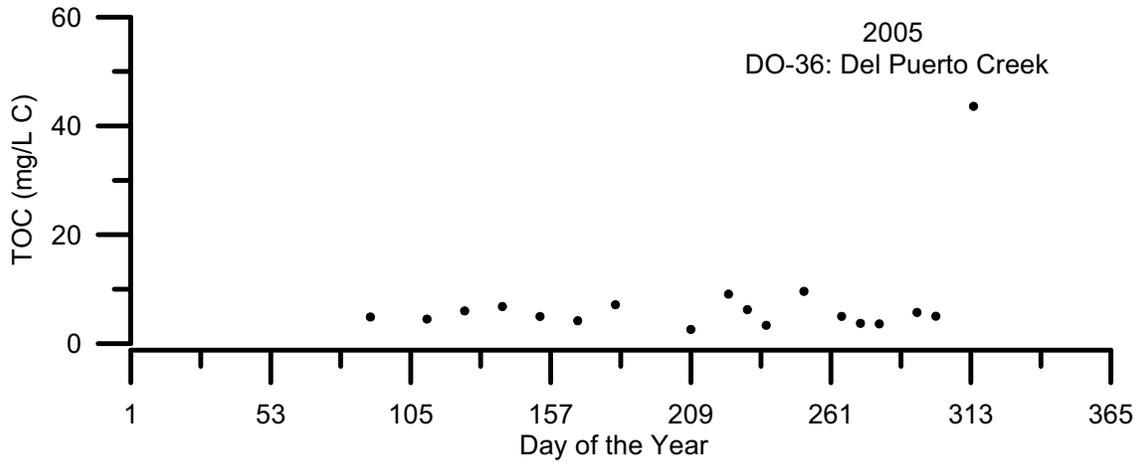


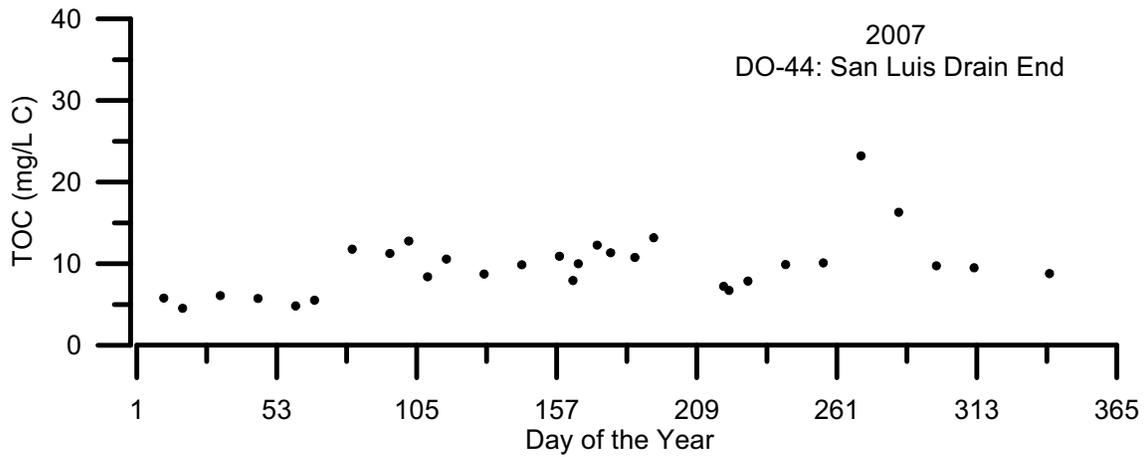
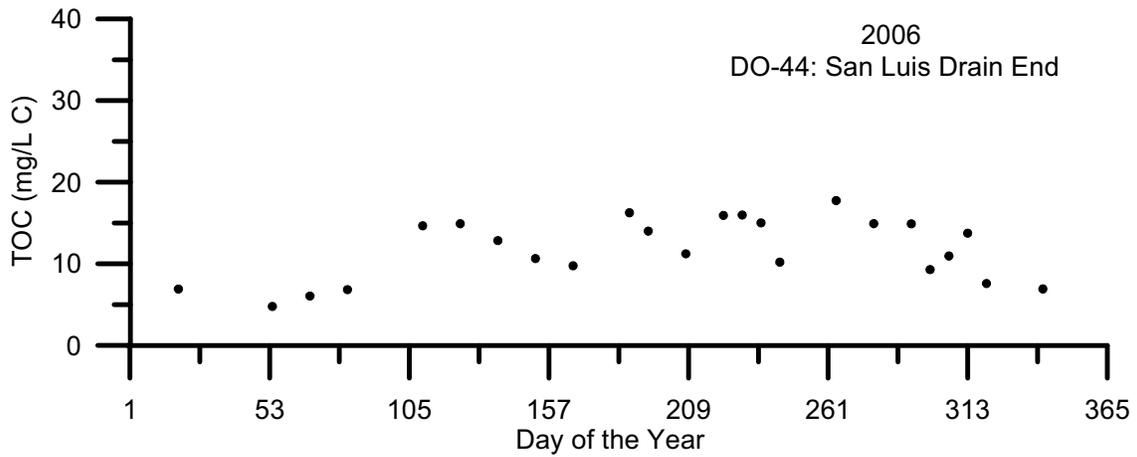
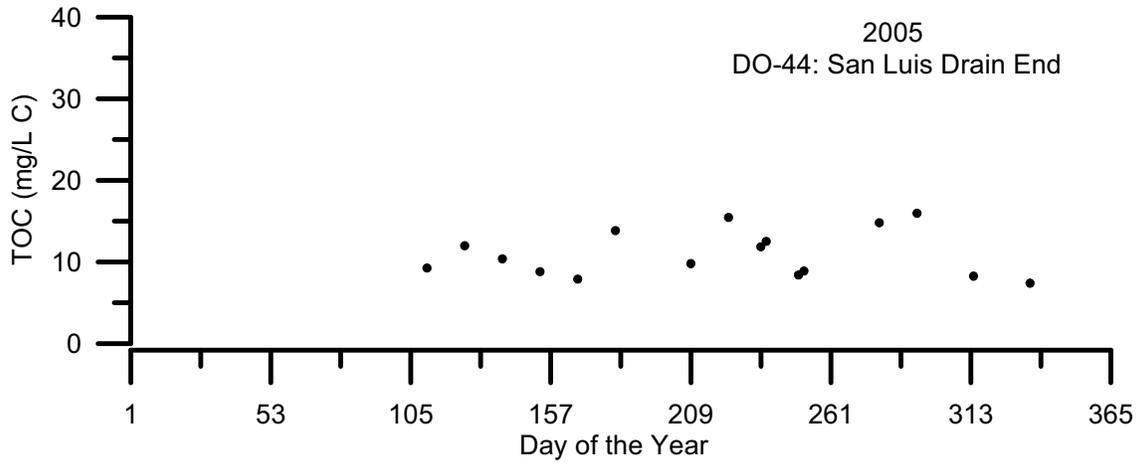


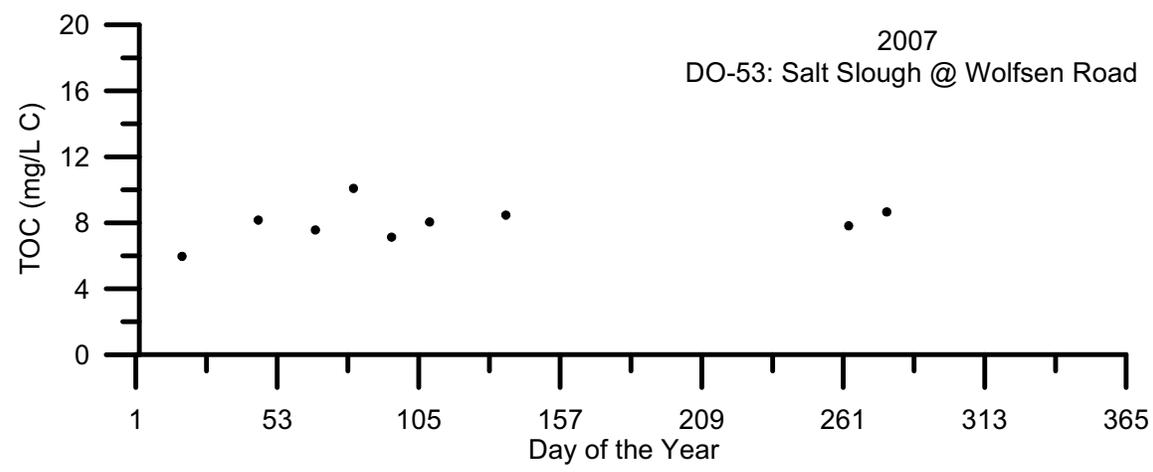
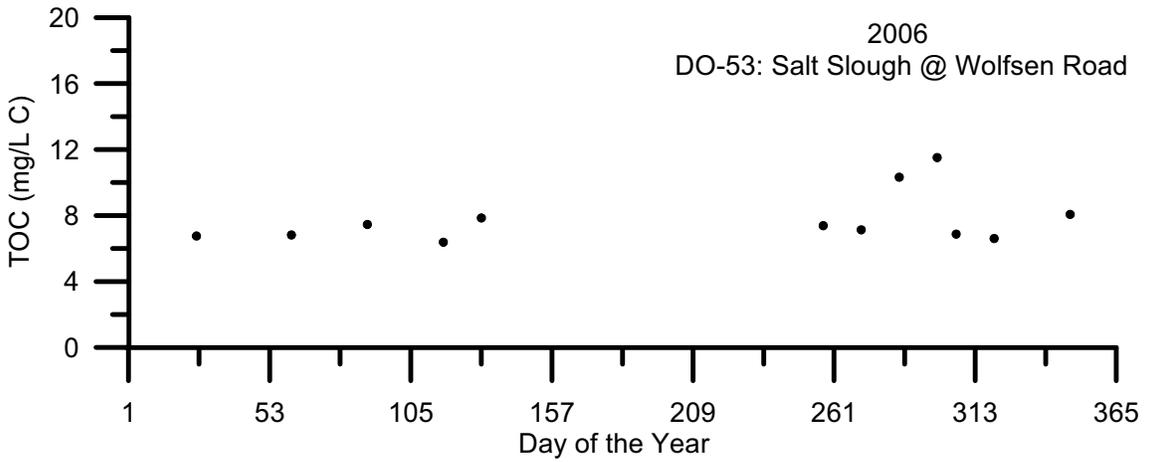
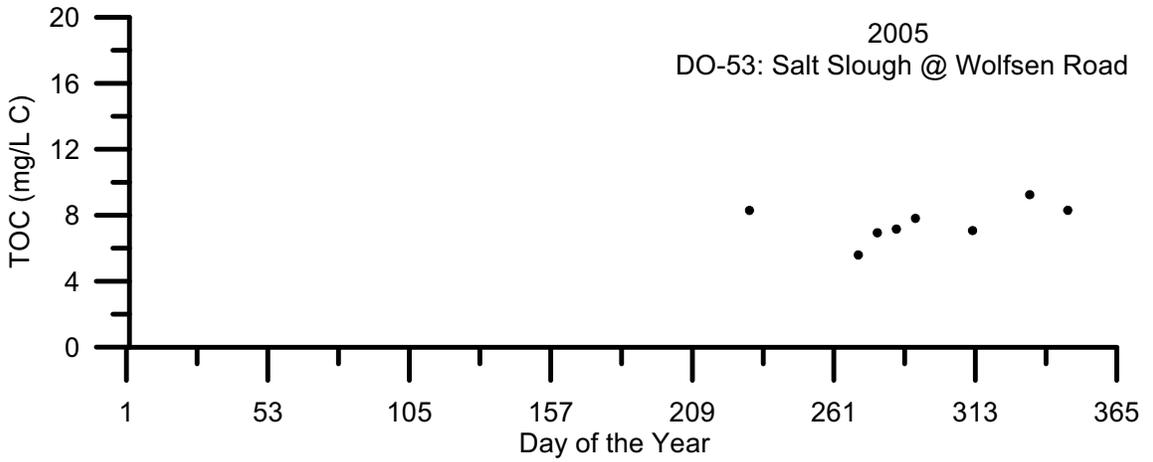


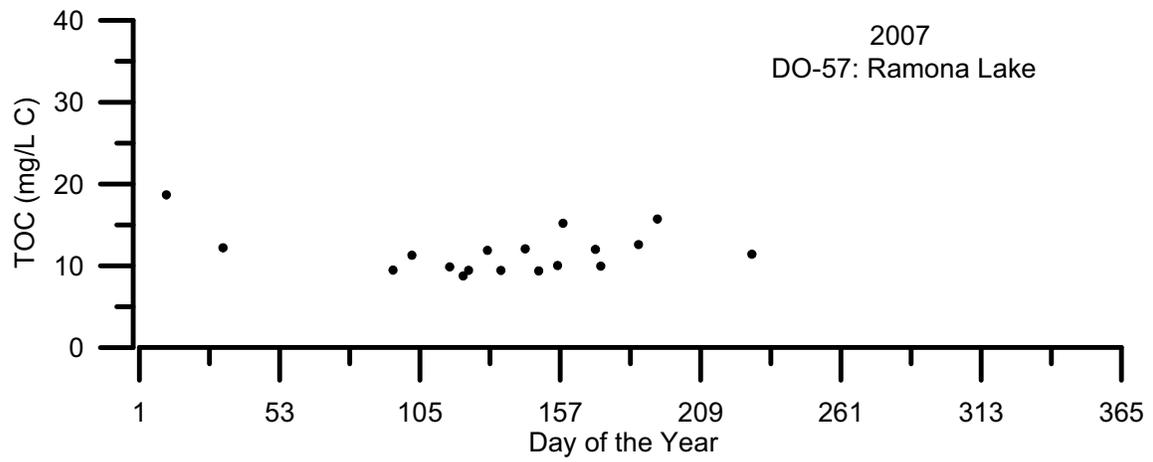
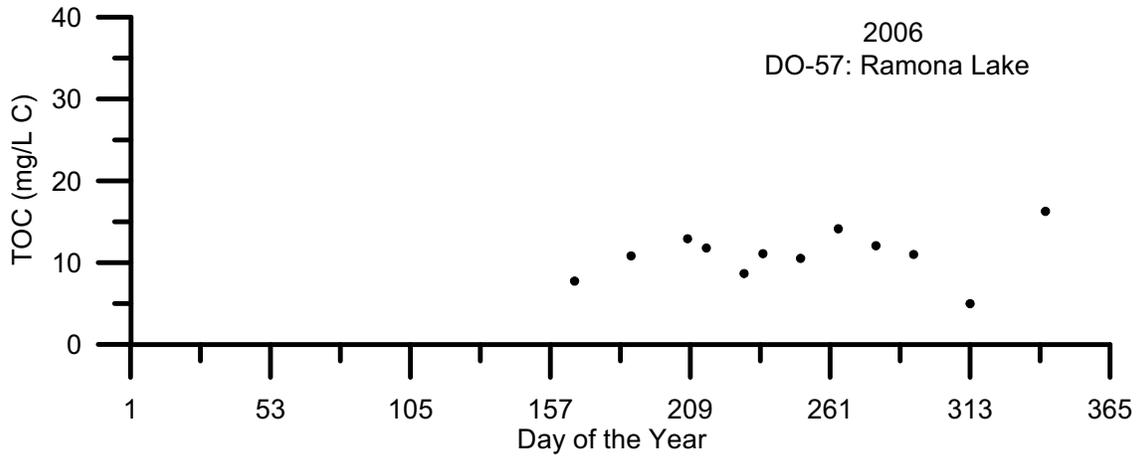


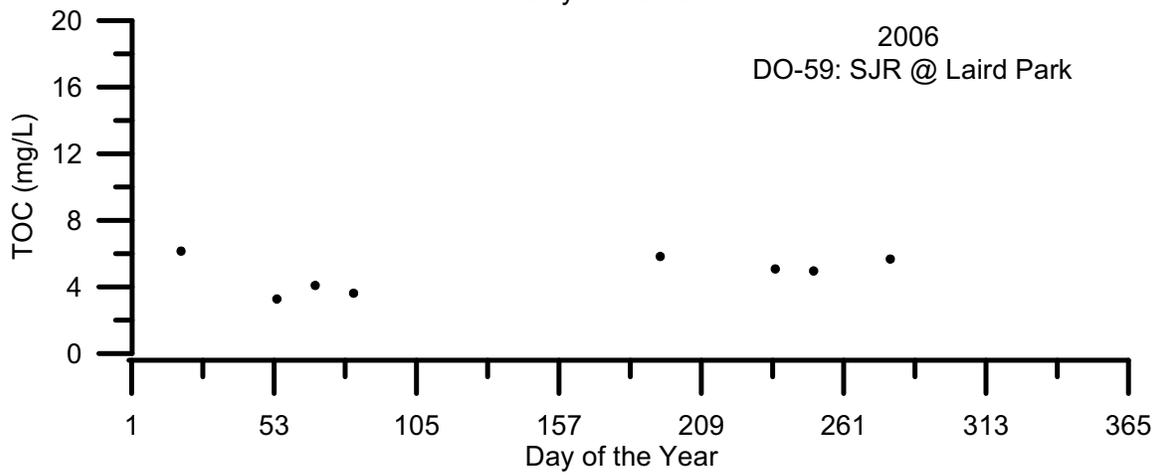
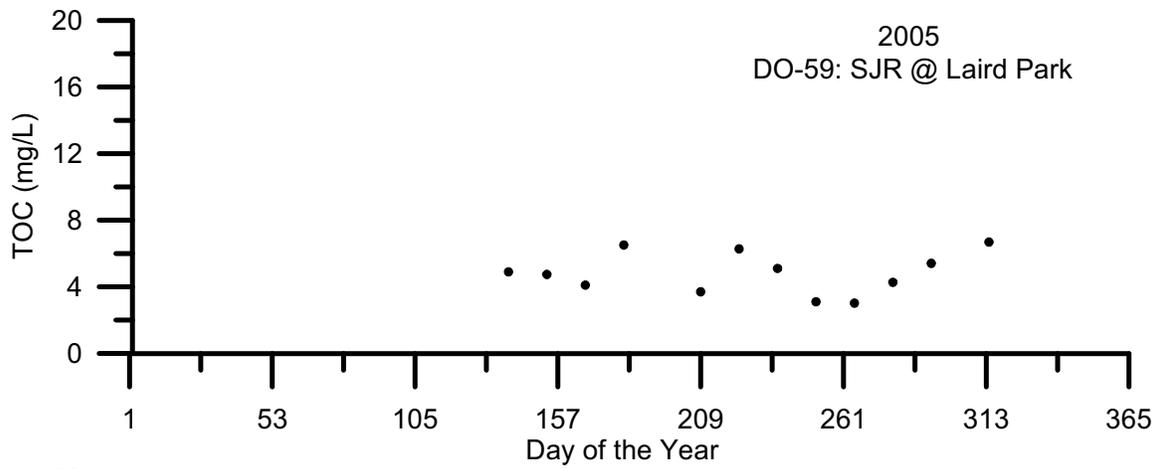


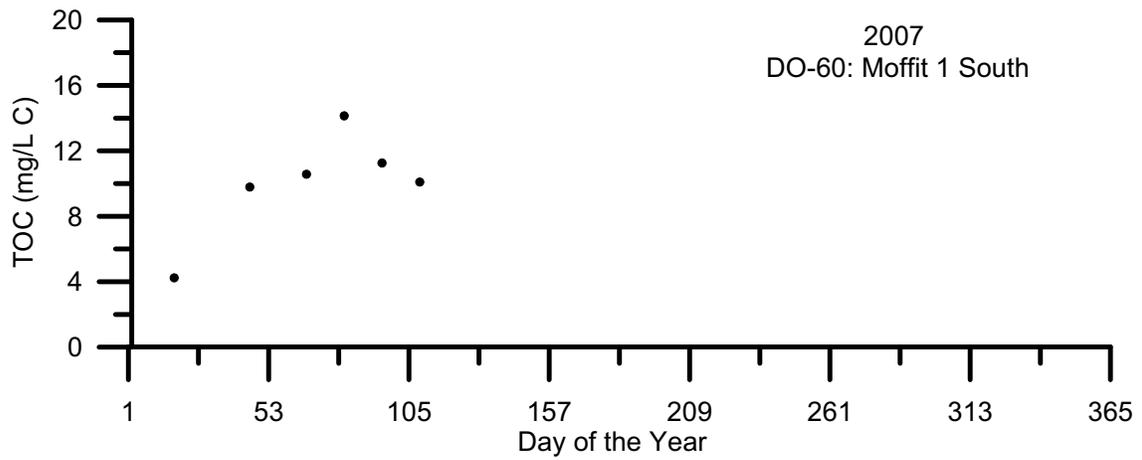
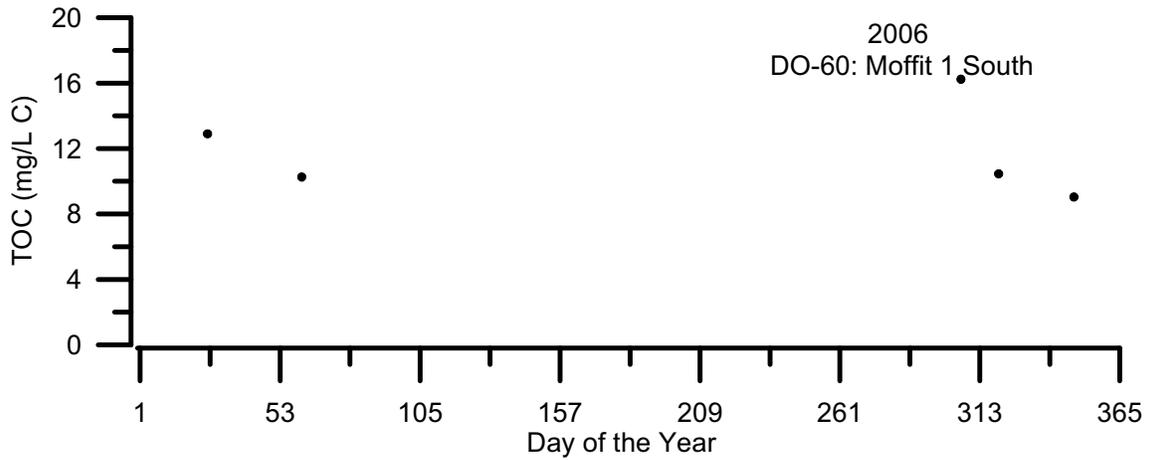
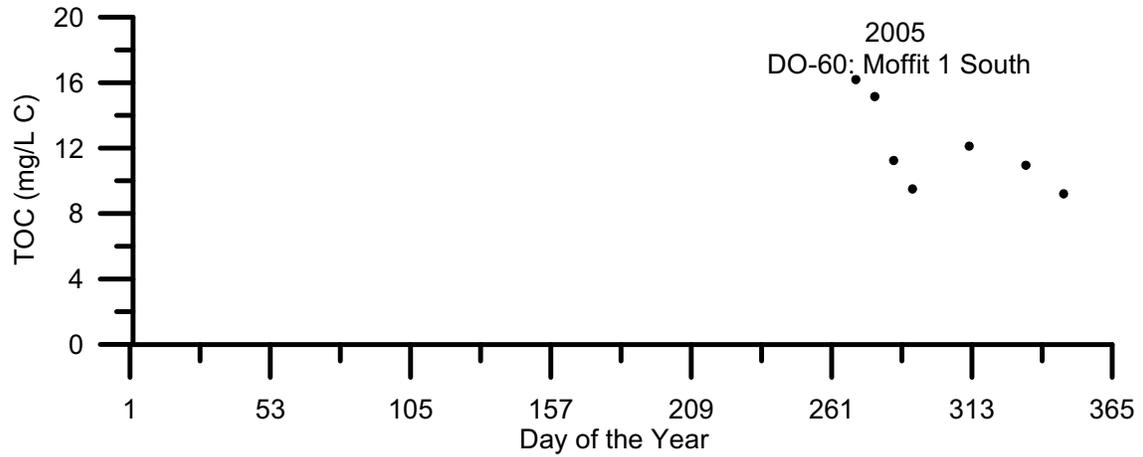


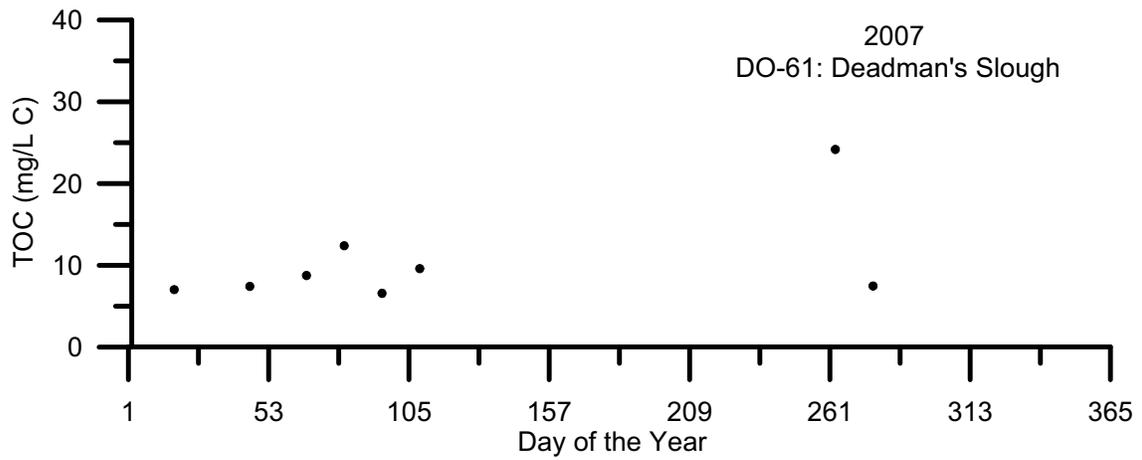
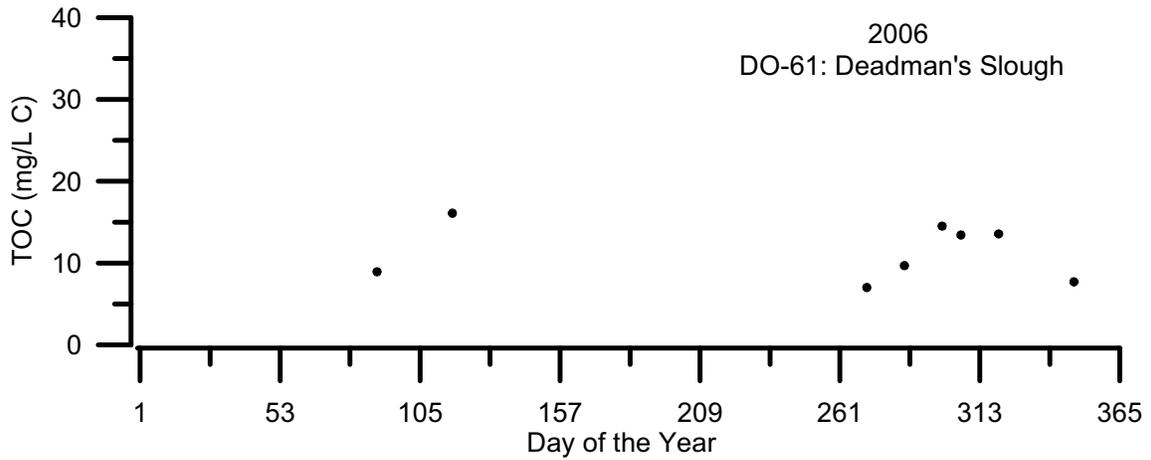
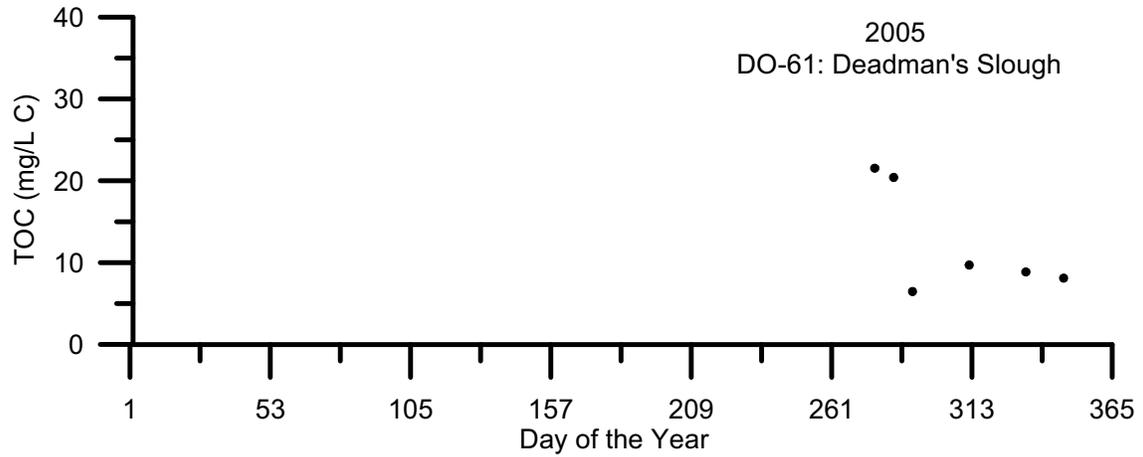


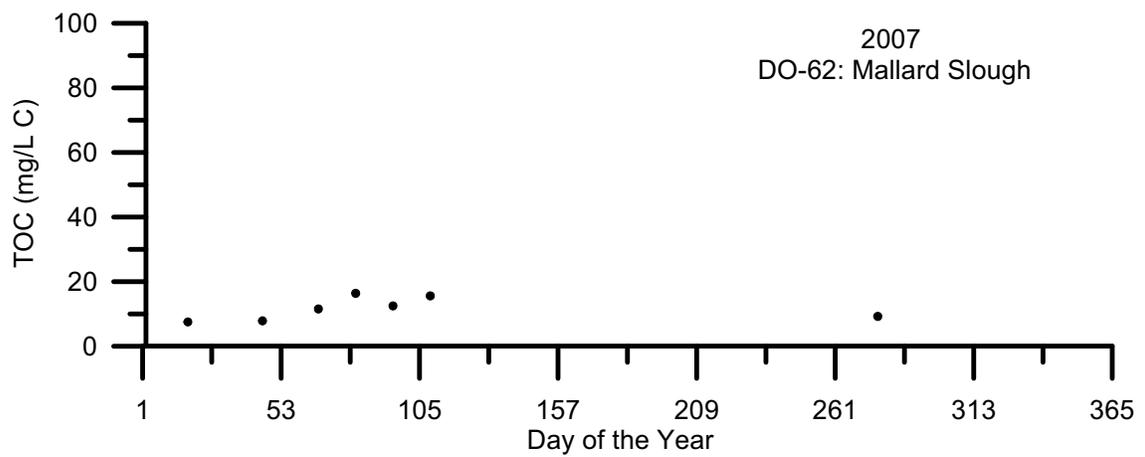
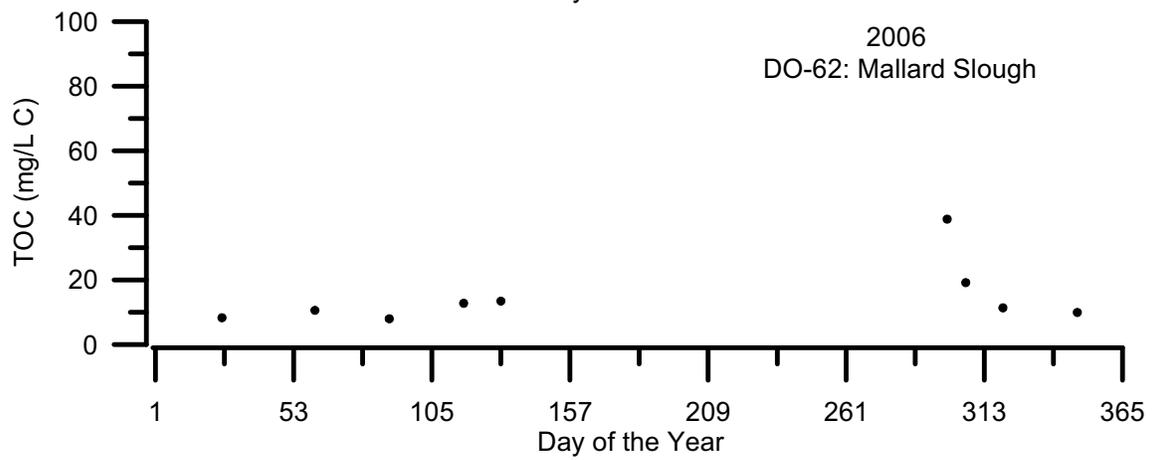
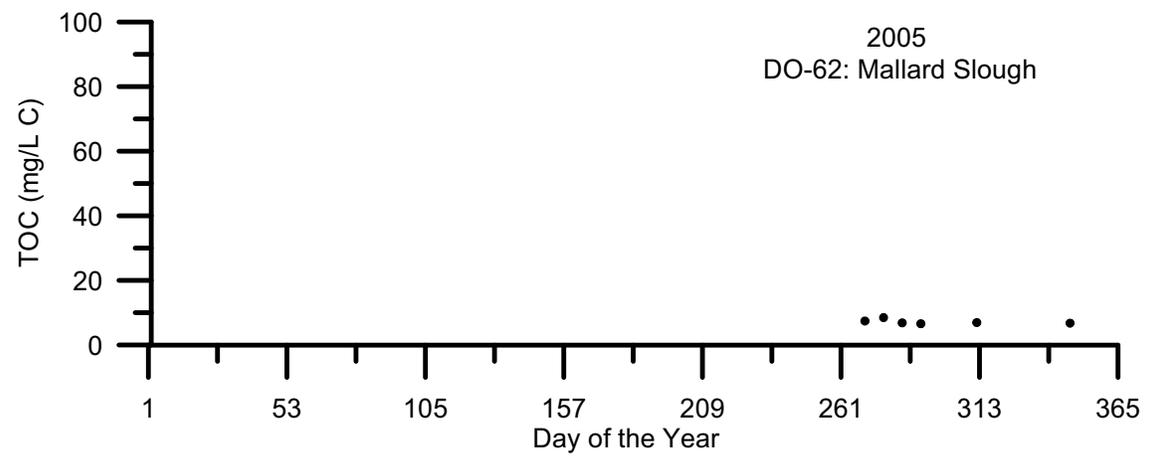


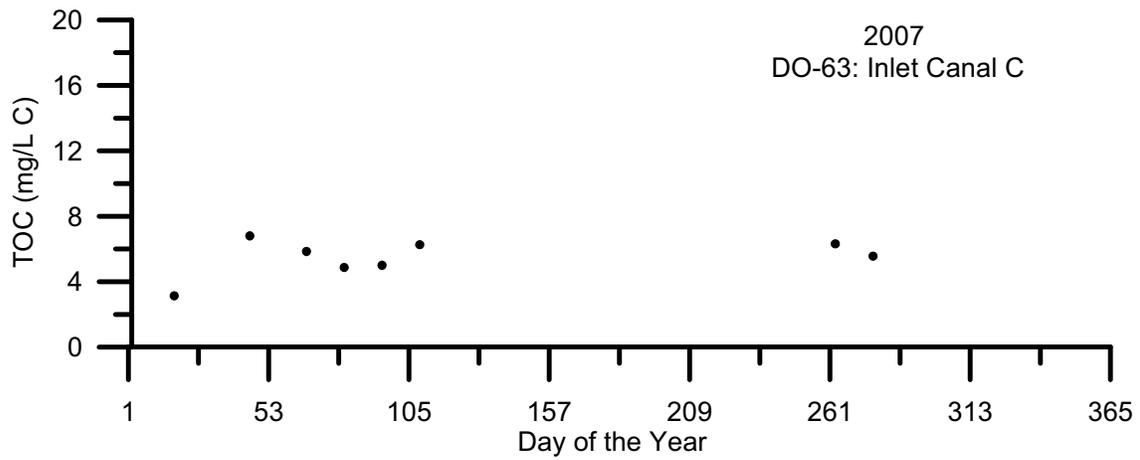
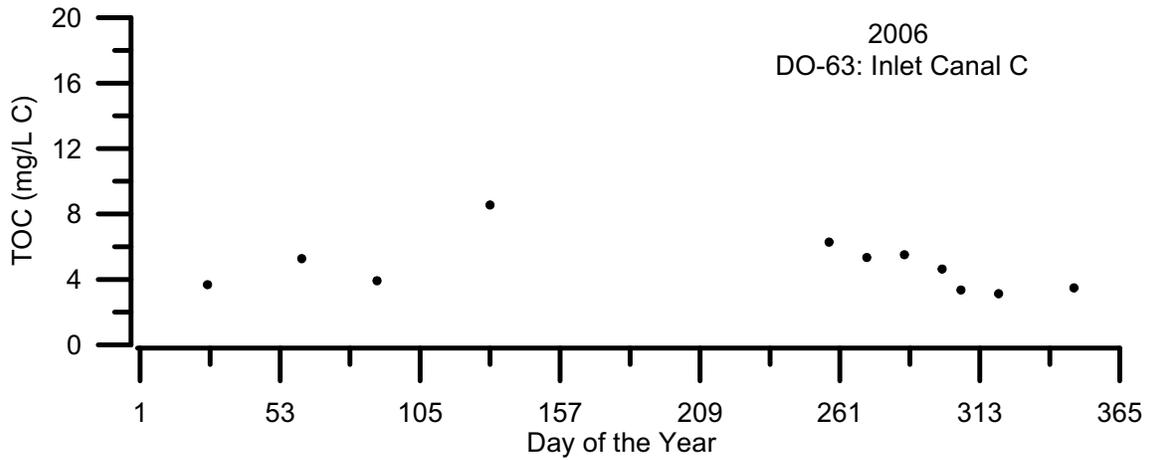
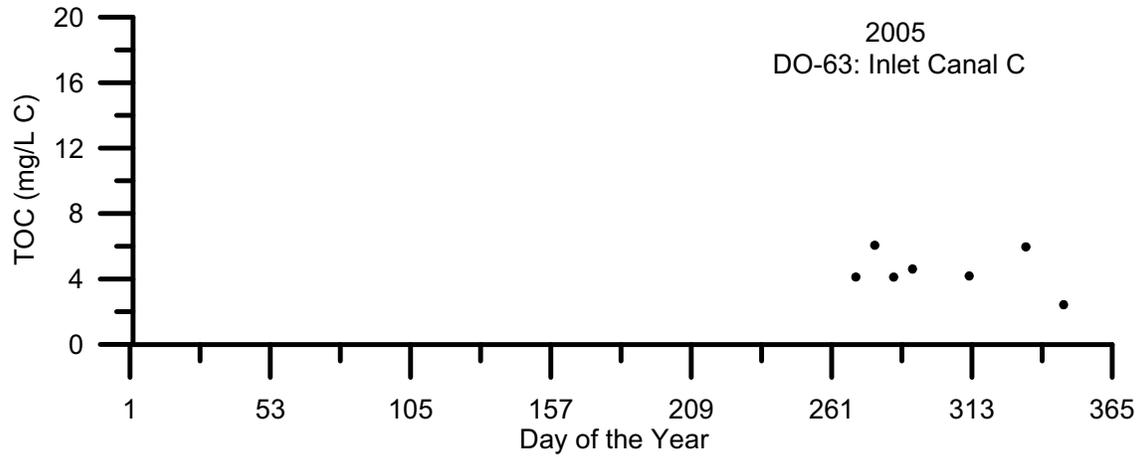


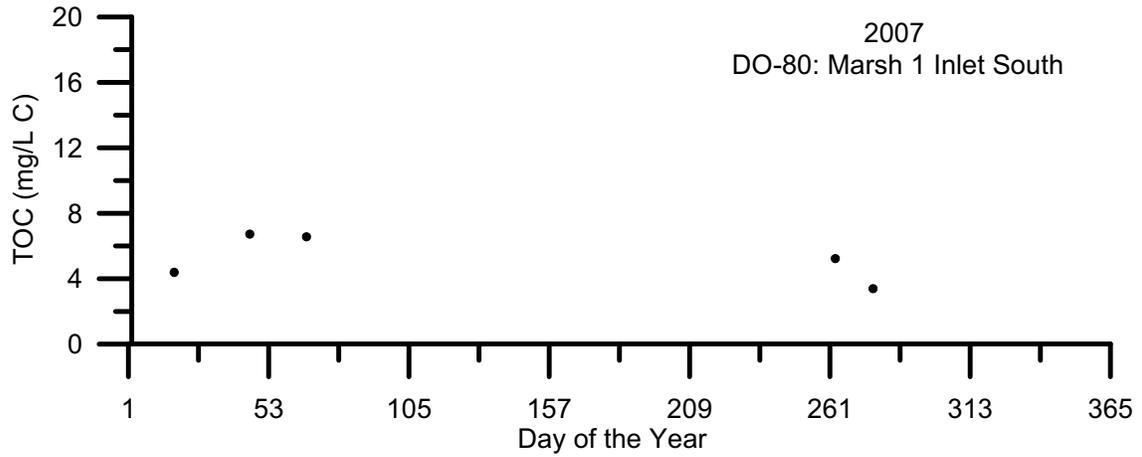
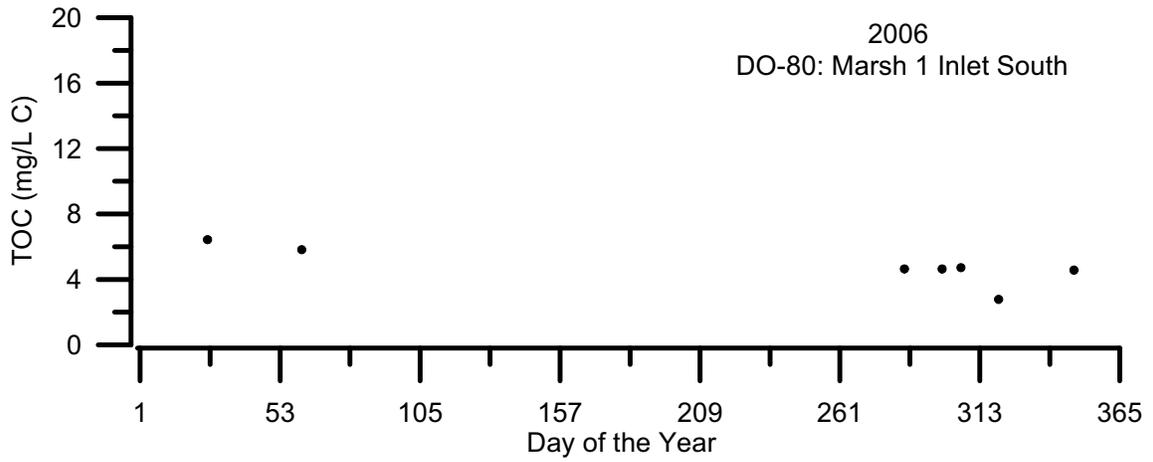
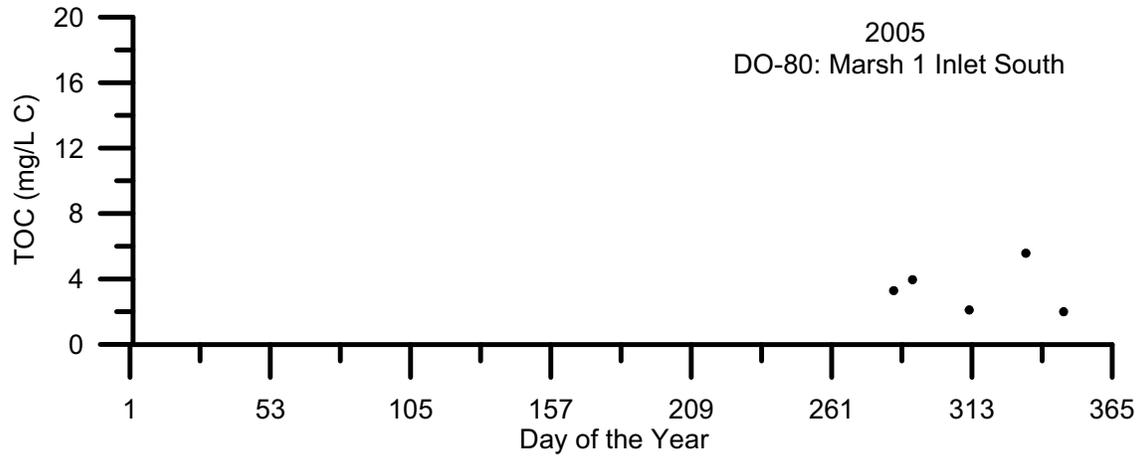


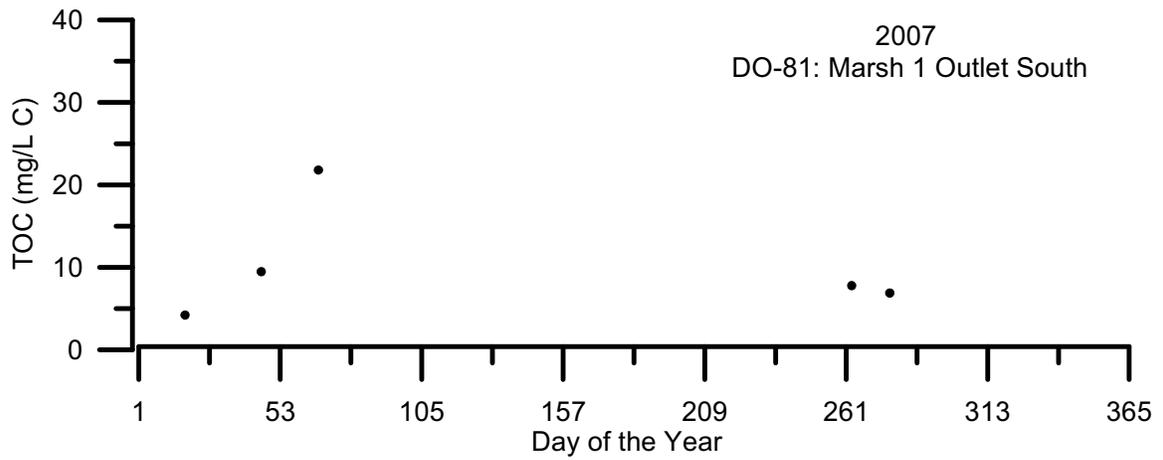
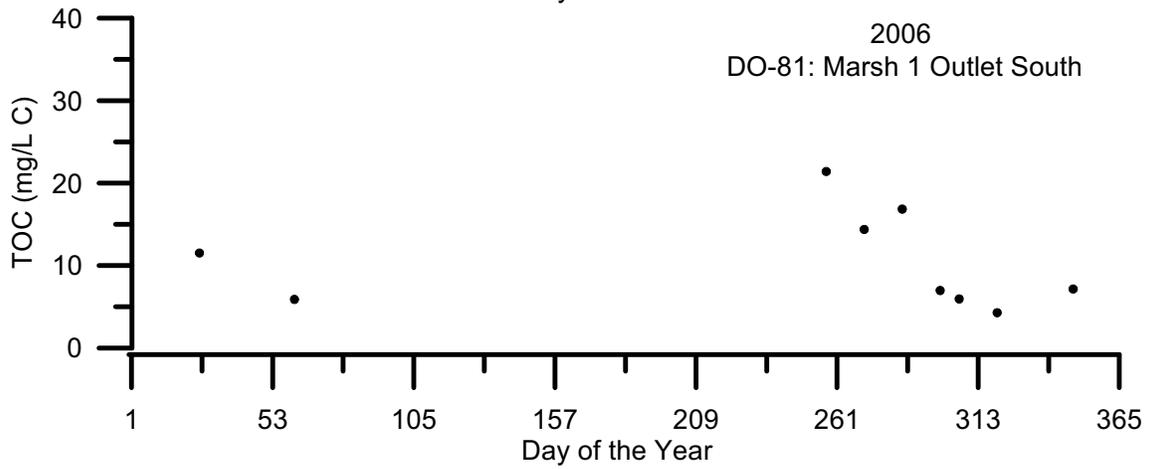
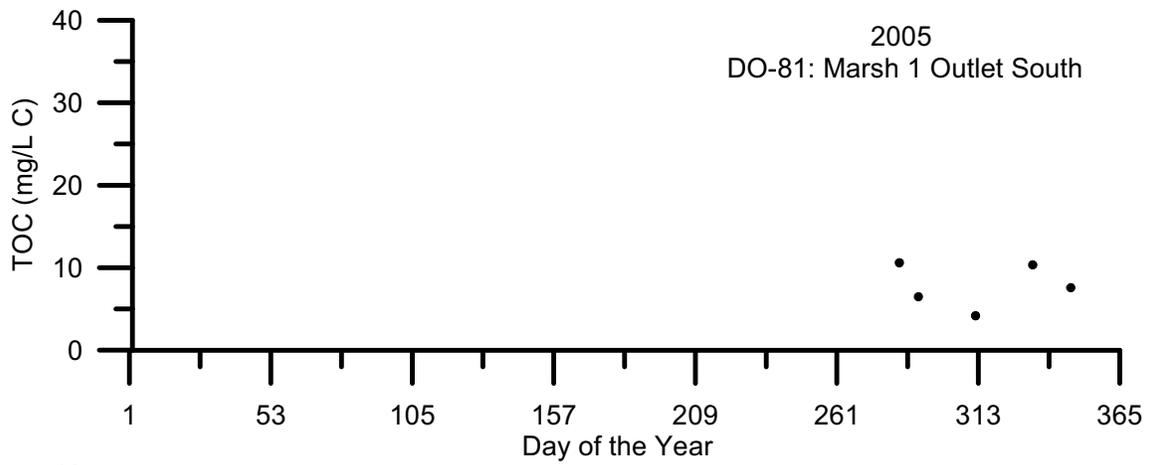


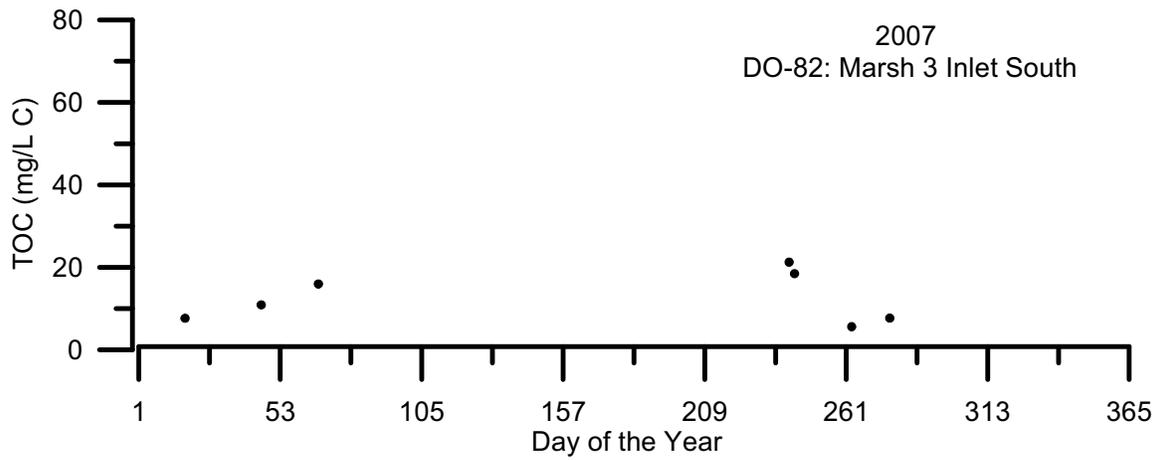
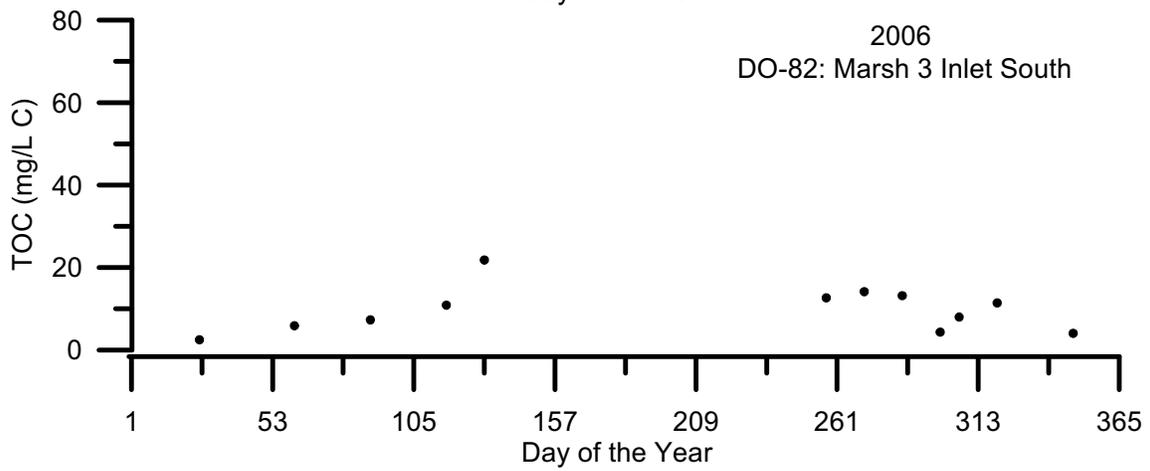
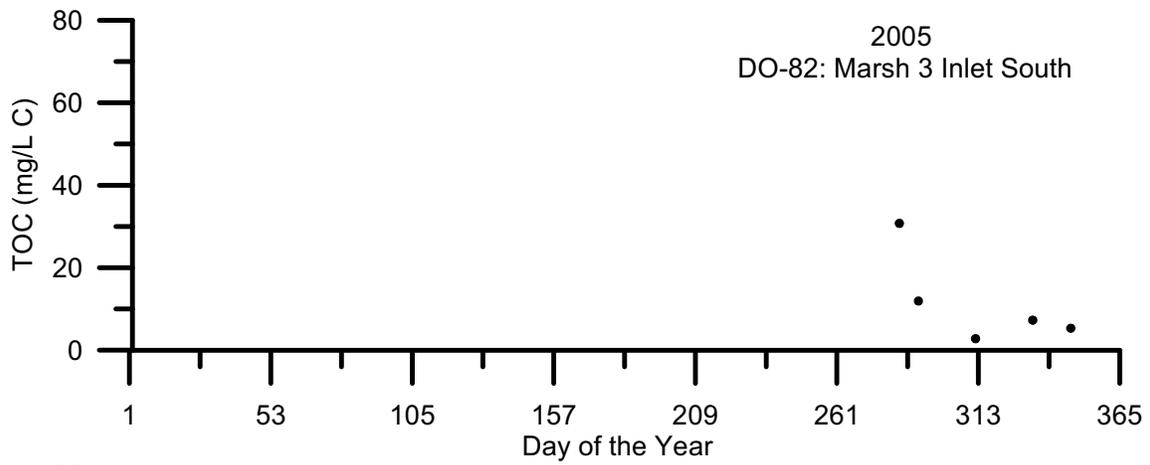


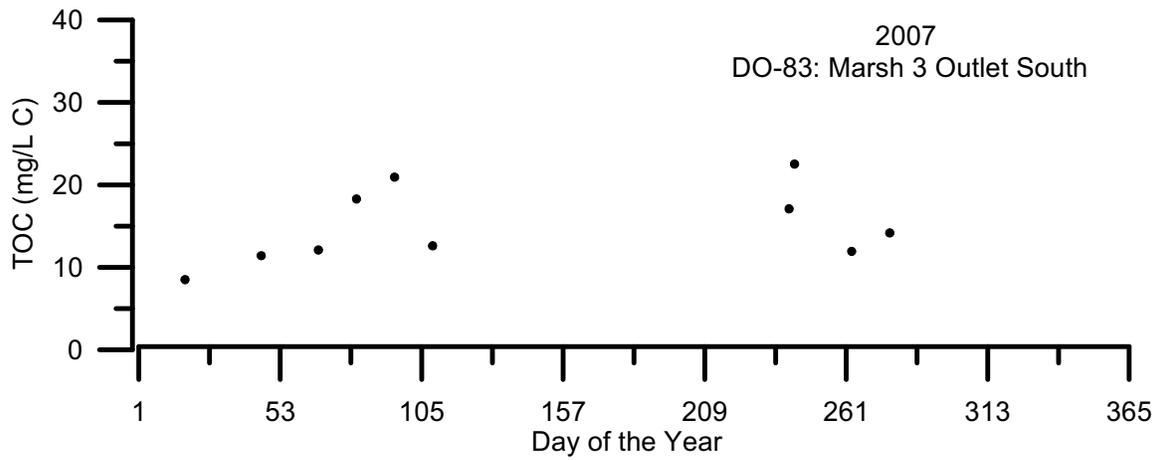
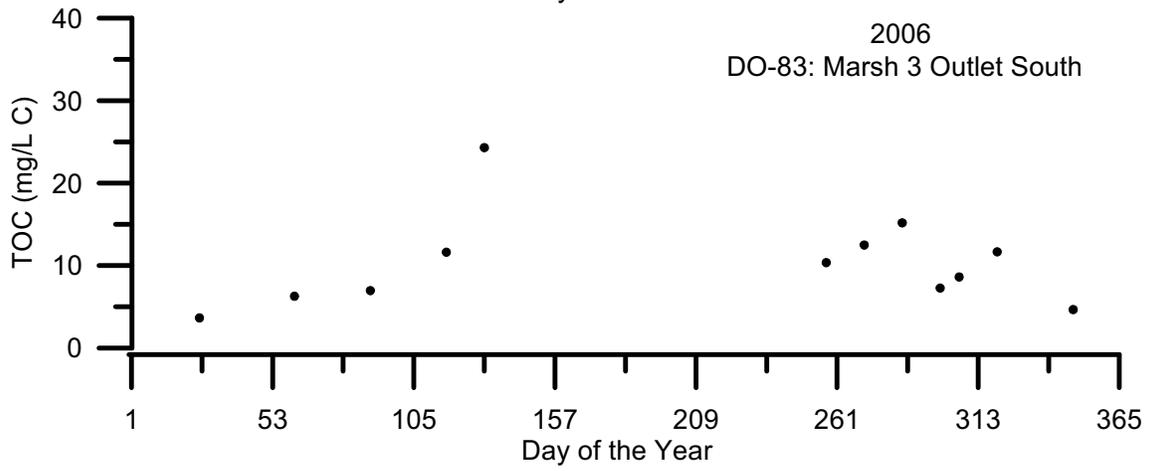
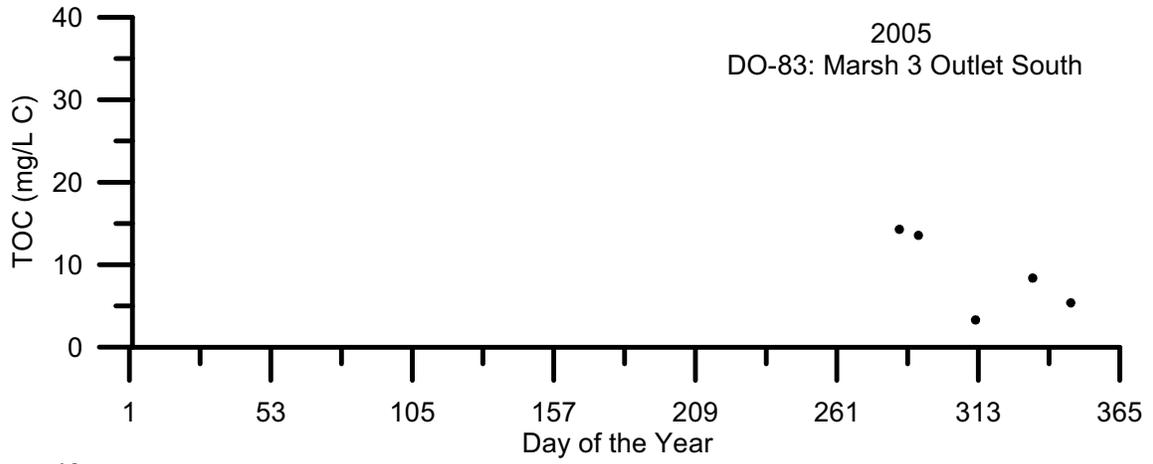


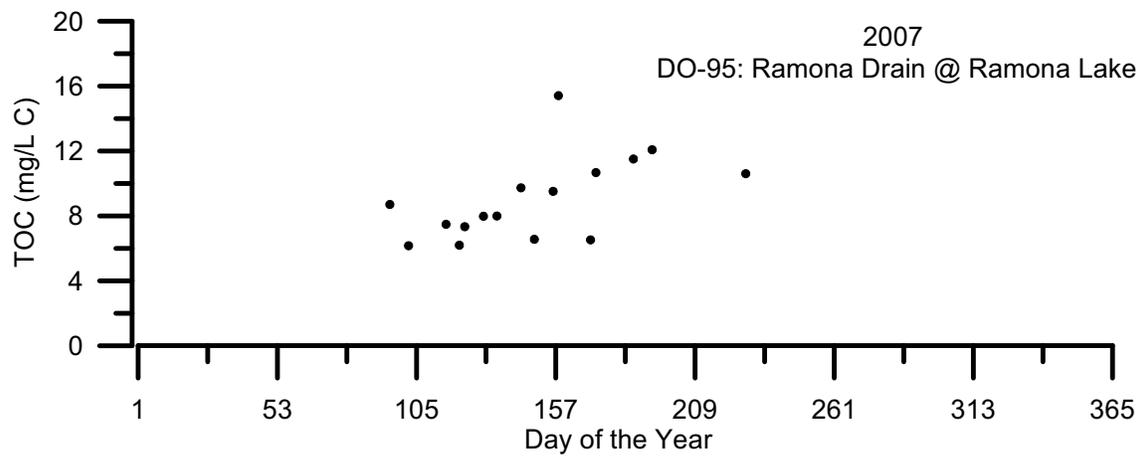
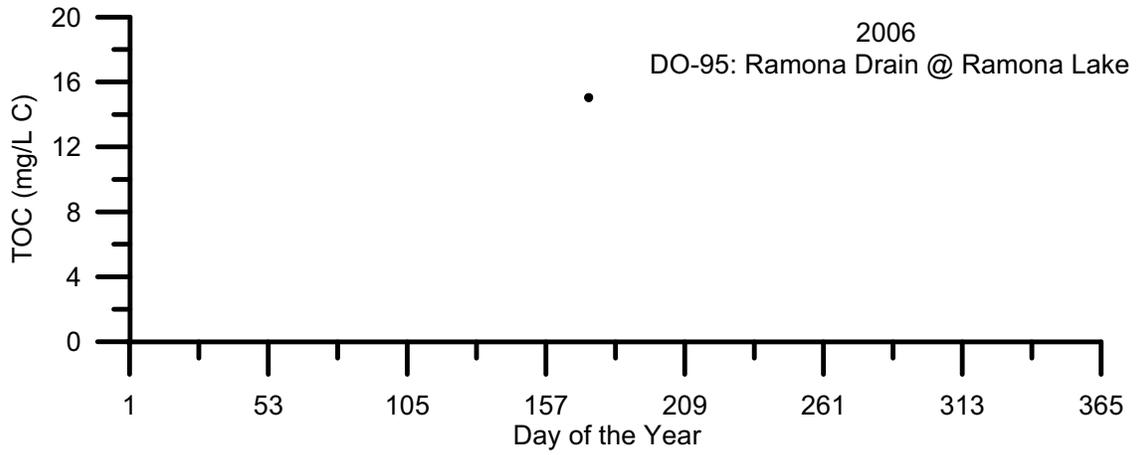














**Trichromatic Method Chlorophyll *A* Extract Data
for the San Joaquin River Watershed
2005-2007**

*Remie Burks
Chelsea Spier
Sharon Borglin
Randy Dahlgren
Jeremy Hanlon
Justin Graham
William Stringfellow*

February 2008

*Environmental Engineering Research Program
School of Engineering and Computer Science
University of the Pacific
3601 Pacific Avenue, Sears Hall
Stockton CA 95211*

Introduction

The San Joaquin River (SJR) supports one of the most productive agricultural regions in the world and its productivity is heavily dependant on irrigated agriculture. A consequence of irrigated agriculture is the production of return flows conveyed down gradient drains that eventually discharge to surface waters. Agricultural drainage may have significant nutrient load and can impact algae growth and general water quality in the SJR. Individual farmers and agricultural organizations, such as drainage authorities, are in need of tools to manage the environmental impacts of agricultural activities (Stringfellow, 2008).

For the years 2005 through 2007, sites throughout the San Joaquin Valley watershed were sampled to assess the overall water quality in the region. One thousand nine hundred and ninety-six (1996) individual surface water samples were collected and analyzed and WQ was assessed at 113 sampling locations in the SJR basin (Borglin et al., 2008). Samples were processed and analyzed by the Environmental Engineering Research Program (EERP) laboratory at the University of the Pacific as well as at the University of California, Davis, Dahlgren Lab. This report presents temporal plots of total chlorophyll *A* concentration as determined by the tri-chromatic method for all sites sampled in the SJR from 2005-2007.

Methods

Field sampling consisted of collecting water samples, measuring water quality with a YSI Sonde 6600 with MDS650 hand-held display, and recording of field conditions at sites within the study area per the *EERP Field Protocol Book*. Water samples were collected in glass 1000 mL bottles (Wheaton Science Products, Millville, NJ), 1000 mL HDPE Trace-Clean narrow mouth plastic bottles (VWR International), 250 mL HDPE Trace-Clean wide mouth plastic bottles (VWR International) as well as 40 mL trace clean vials with PTFE septa (IChem, Rockwood, TN) in accordance with requirements for different lab analysis and volume requirements. Bottles were labeled with the appropriate sample number, site name and sampling date. All bottles were rinsed with sample water prior to collection of a depth-integrated sample. Some sites required a bucket to collect sample water because of accessibility from a high bridge or platform. For these sites, the bucket was pre-rinsed with sample water and sample bottles were filled using a rinsed funnel. Care was taken to distribute water simultaneously to all sample bottles (rather than sequentially). Samples were immediately stored at 4°C after sampling (cooler temperature was recorded in the lab upon delivery) and transported to the lab on the day of sampling. All bottle numbers, meter readings, and time in and out of the sample site were recorded in the field notebook.

Samples were received by the laboratory the same day they were sampled, logged in and inspected for damage, and stored at 4°C until filtering. Chlorophyll-a (chl-a) and pheophytin-a (pha-a) were extracted and analyzed using UV absorption as described in SM 10200 H (APHA, 2005). Both the trichromatic chl-a and the pha-a methods were used for quantification. Approximately 1000 mL of samples were filtered using a

vacuum filtration onto a Whatman GF/F filter within 24 hours of sample collection. The sample was kept in the dark during storage and filtration. After the water was removed saturated $MgCO_3$ was applied to the sample on the filter and the filter was stored at $-20^{\circ}C$ for up to 21 days before analysis. Extraction was performed by grinding the filter with a Teflon tissue grinder in acetone saturated with 10% by weight $MgCO_3$. The extracted sample was centrifuged for 20 minutes at 2000 rpm and the chl-a and pha-a was quantified by measurement of the supernatant on a Perkin-Elmer Lambda 35 spectrophotometer (Wellesley, MA) using a 5 cm path length (Borglin, et al 2008).

Results and Discussion

Chlorophyll was analyzed routinely over the years 2005-2007 with no modification to the standard method (APHA, 2005). No standard solutions for this analysis were available so only duplicates and blanks could be analyzed to insure uniformity and accuracy in the application of the method and for those two QA parameters, there was an 86.07% passage rate for this analysis.

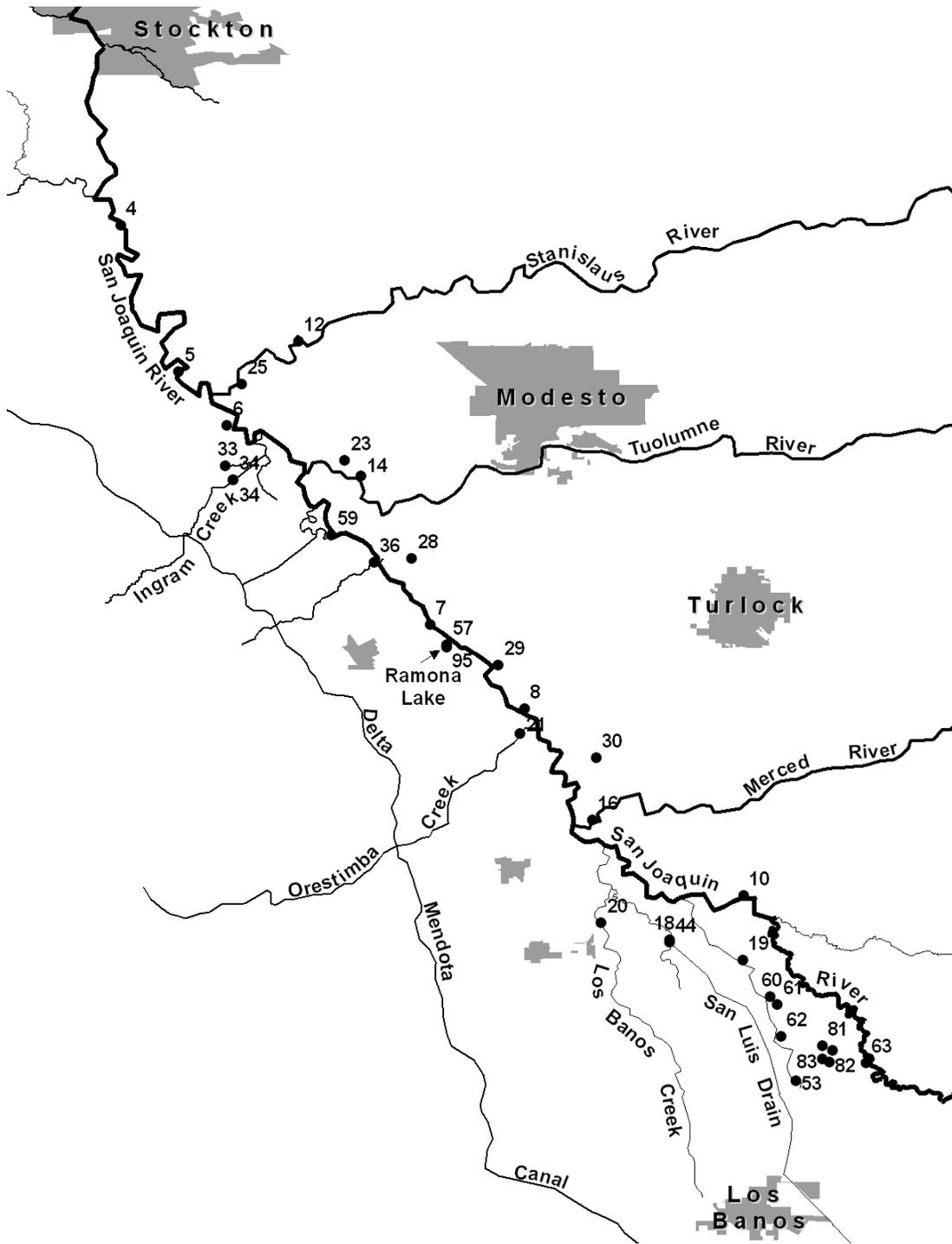
References

- Stringfellow, W.T., et al., (2008), *Evaluation of Vegetated Ditches, Ponds, and Wetlands as BMPs for Mitigating the Water Quality Impact of Irrigated Agriculture in the San Joaquin Valley*, University of the Pacific, Stockton, CA
- Graham, J., Hanlon, J.S., Stringfellow, W.T., (2008), *EERP Field Protocol Book*, University of the Pacific, Stockton, CA.
- Borglin, S., W. Stringfellow, J. Hanlon. (2005), *Standard Operating Procedures for the Up-Stream Dissolved Oxygen TMDL Project*, LBNL/Pub-937.
- Borglin, S., Burks, R., Hanlon, J., Graham, J., Spier, C., Stringfellow, W., and Dahlgren, R., (2008) *Methods overview, quality assurance, and quality control*, University of the Pacific, Stockton, CA
- American Public Health Association, (2005), *Standard Methods for the Examination of Water and Wastewater*, American Public Health Association, Washington, D.C.
- Borglin, S.E., Burks, R.D., Hanlon, J.S., Stringfellow, W.T. (2008) *EERP Lab Protocol Book*, University of the Pacific, Stockton, CA.
- YSI Environmental Operations Manual, (2005), 6-Series Environmental Monitoring Systems, Yellow Springs, OH.

Table 1: EERP Sampling Site List

DO Number	Site Name	Type
4	SJR at Mossdale	Core sites
5	SJR at Vernalis-McCune Station (River Club)	Core sites, BMP
6	SJR at Maze	Core sites, BMP
7	SJR at Patterson	Core sites, BMP
8	SJR at Crows Landing	Core sites, BMP
10	SJR at Lander Avenue	Core sites
12	Stanislaus River at Caswell Park	Core sites
14	Tuolumne River at Shiloh Bridge	Core sites
16	Merced River at River Road	Core sites
18	Mud Slough near Gustine	Core sites, Wetland
19	Salt Slough at Lander Avenue	Core sites, Wetland
20	Los Banos Creek Flow Station	Core sites, Wetland
21	Orestimba Creek at River Road	Core sites, BMP
23	Modesto ID Lateral 5 to Tuolumne	Core sites
25	Modesto ID Main Drain to Stan. R. via Miller Lake	Core sites
28	Turlock ID Westport Drain Flow station	Core sites
29	Turlock ID Harding Drain	Core sites
30	Turlock ID Lateral 6 & 7 at Levee	Core sites
33	Hospital Creek	Intermittent, BMP
34	Ingram Creek	Core sites, BMP
36	Del Puerto Creek Flow Station	Core sites, BMP
44	San Luis Drain End	Core sites
57	Ramona Lake Drain	Core sites, BMP
59	SJR Laird Park	Core sites
95	Ramona drain at Ramona Lake	BMP, Intermittent

Figure 1: EERP Sampling Site Map of SJR Watershed and Tributaries



Figures 2 -72: Temporal Plots of TC Chlorophyll A Concentration By Site ID

